DOI: 10.5665/SLEEP.1106

## Measuring In Vivo Metabolite Levels in Brain

Commentary on Dworak et al. Sleep and brain energy levels: ATP changes during sleep. Journal of Neuroscience 2010;30:9007-16.

David F. Wilson, PhD

Department of Biochemistry and Biophysics, University of Pennsylvania, Philadelphia, PA

The paper presented several interesting but surprising experimental results. Most important are measurements showing large increases in ATP, up to more than 3 times that of awake animals, during sleep. Reports of large changes in ATP in the brain, except in severe hypoxia/ischemia, are rare.

The author's description of the procedure used to prepare the samples includes:

"The rats were killed by decapitation, and brains were removed. Coronal slices (2 mm thick) were carefully placed on a dry ice ( $-78.5^{\circ}$ C)-containing covered Petri dish for rapid freezing and subsequent dissection." and "Extreme care was exercised to complete this process rapidly, with an average time of  $80 \pm 9$  s for tissue collection..."

The rat brain consumes approximately  $77 \times 10^{-9}$  moles  $O_2/g$  sec.<sup>1</sup> Assuming most of this is used for oxidative phosphorylation and 6 ATP are made per  $O_2$ , ATP is being synthesized/consumed at  $0.46 \times 10^{-6}$  moles/g sec. The brain has approximately  $2.6 \times 10^{-6}$  moles ATP/g,<sup>2,3</sup> so on the average all of the ATP in the brain is consumed and resynthesized (turned over) every 6-10 seconds. Interruption of oxygen delivery results in loss of consciousness in about 10 sec.

For trapping the metabolic state in accessible tissues, such as heart or liver, Wollenberger clamps<sup>4</sup> have been used. These are a pair of tongs about 2 feet long with flat aluminum plates with much greater mass than the tissue sample. The aluminum plates are cooled in liquid  $N_2$  (-196°C), and then the tissue, without prior interruption of perfusion or blood flow, is clamped between the plates, compressing the tissue to 1 and 2 mm thick. The low temperatures, high rate of transfer of heat to the metal plates, and the thin samples results in freezing (quenching) times of < 1 sec. Krebs and coworkers<sup>5,6</sup> and Williamson and coworkers,<sup>7-9</sup> as examples, used this method when studying metabolism in liver and heart, were able to measure metabolite changes occurring within a few seconds.

Brain is a challenge because the Wollenberger clamps crush the bone and soft tissue together. To overcome this limitation, other methods were developed. Veech and coworkers, <sup>2,3</sup> for example, designed a "brain blower" in which pressurized gas blew the soft brain tissue out through a burr hole in the skull and onto an aluminum plate at –196°C. Quenching occurred in

< 1 sec. Four different methods for rapid quenching of metabolism in brain were compared by Lust et al.<sup>3</sup> It was concluded that the brain blower, with quenching in < 1 sec, resulted in tissue samples giving significantly more physiological metabolite measurements than could be attained using the somewhat slower quenching methods.

The method Dworak and coworkers used to prepare the brain tissue is suitable for measuring gene expression and protein levels, but not for trapping metabolic intermediates such as ATP or enzyme phosphorylation at *in vivo* levels. The reported values represent metabolism occurring in the brain after decapitation and not of the brain *in vivo*.

## **DISCLOSURE STATEMENT**

Dr. Wilson has indicated no financial conflicts of interest.

## **REFERENCES**

- Hägerdal M, Harp J, Nilsson L, Siesjö BK. The effect of induced hypothermia upon oxygen consumption in the rat brain. J Neurochem 1975;24:311-6.
- Ogushi S, Lawson JWR, Dobson GP, Veech RL, Uyeda K. A new transient activator of phosphofructokinase during initiation of rapid glycolysis in brain. J Biol Chem 1990;265:1094-9.
- Lust WD, Passonneau JV, Veech RL. Cyclic adenosine monophosphate, metabolites, and phosphorylase in neural tissue: A comparison of methods of fixation. Science 1973;181:280-2.
- Wollenberger A, Ristbau O, Schoffa G. A simple technic for extremely rapid freezing of large pieces of tissue. Pflugers Archiv fur die gesamte Physiologie des Menschen und der Tiere. 1960;270:399-412.
- Brosnan JT, Krebs HA, Williamson, DH. Effects of ischaemia on metabolite concentrations in rat liver. Biochem J 1970;117:91-6.
- Williamson DH, Lund P,Krebs HA. The redox state of free nicotinamideadenine dinucleotide in the cytoplasm and mitochondria of rat liver. Biochem J 1967;103:514-27.
- Williamson JR. Glycolytic control mechanisms: II. Kinetics of intermediate changes during the aerobic-anoxic transition in perfused rat heart. J Biol Chem 1966;241:5026-36.
- Scholz R, Thurman RG, Williamson JR, Chance B, Butchery T. Flavin and pyridine nucleotide oxidation-reduction changes in perfused rat liver. I. Anoxia and subcellular localization of fluorescent flavoproteins. J Biol Chem 1969;244:2317-24.
- Safer B, Williamson JR. Mitochondrial-cytosolic interactions in perfused rat heart: Role of coupled transamination in repletion of citric acid cycle intermediates. J Biol Chem 1973;248:2570-9.

Address correspondence to: David F. Wilson, Department of Biochemistry and Biophysics, University of Pennsylvania, Philadelphia, PA 19104