

# Metabotropic glutamate receptor 5 antagonist protects dopaminergic and noradrenergic neurons from degeneration in MPTP-treated monkeys

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Degeneration of the dopaminergic nigrostriatal system and of noradrenergic neurons in the locus coeruleus are important pathological features of Parkinson's disease. There is an urgent need to develop therapies that slow down the progression of neurodegeneration in Parkinson's disease. In the present study, we tested whether the highly specific metabotropic glutamate receptor 5 antagonist, 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine, reduces dopaminergic and noradrenergic neuronal loss in monkeys rendered parkinsonian by chronic treatment with low doses of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine. Weekly intramuscular 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine injections (0.2–0.5 mg/kg body weight), in combination with daily administration of 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine or vehicle, were performed until the development of parkinsonian motor symptoms in either of the two experimental groups (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine versus 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/vehicle). After 21 weeks of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine treatment, all 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/vehicle-treated animals displayed parkinsonian symptoms, whereas none of the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine-treated monkeys were significantly affected. These behavioural observations were consistent with *in vivo* positron emission tomography dopamine transporter imaging data, and with post-mortem stereological counts of midbrain dopaminergic neurons, as well as striatal intensity measurements of dopamine transporter and tyrosine hydroxylase immunoreactivity, which were all significantly higher in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine-treated animals than in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/vehicle-treated monkeys. The 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine treatment also had a significant effect on the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced loss of norepinephrine neurons in the locus coeruleus and adjoining A5 and A7 noradrenaline cell groups. In 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/vehicle-treated animals, almost 40% loss of tyrosine hydroxylase-positive norepinephrine neurons was found in locus coeruleus/A5/A7 noradrenaline cell groups, whereas the extent of neuronal loss was lower than 15% of control values in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine-treated monkeys. Our data demonstrate that chronic treatment with the metabotropic glutamate receptor 5 antagonist, 3-[(2-methyl-1,3-thiazol-4-yl)

ethynyl] pyridine, significantly reduces 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine toxicity towards dopaminergic and noradrenergic cell groups in non-human primates. This suggests that the use of metabotropic glutamate receptor 5 antagonists may be a useful strategy to reduce degeneration of catecholaminergic neurons in Parkinson's disease.

**Keywords:** substantia nigra; locus coeruleus; striatum; neuroprotection; noradrenaline

**Abbreviations:** FECNT = 2 $\beta$ -carbomethoxy-3 $\beta$ -(4-chlorophenyl)-8-(2-[18F]-fluoroethyl)-nortropane; mGluR5 = metabotropic glutamate receptor 5; MPTP = 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; MTEP = 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl]; PET = positron emission tomography

## Introduction

Parkinson's disease is a progressive neurodegenerative disorder clinically characterized by bradykinesia, tremor, rigidity and postural instability. In addition, non-motor manifestations are increasingly recognized as being part of the wider clinical syndrome of Parkinson's disease (Aarsland *et al.*, 1999, 2004; Braak *et al.*, 2003; Zesiewicz *et al.*, 2003; Grimbergen *et al.*, 2004; Langston, 2006). Although there is compelling evidence that the parkinsonian motor symptoms are largely due to the progressive degeneration of the nigrostriatal dopaminergic system, the pathological substrate of non-motor deficits is unknown. It is possible that some of these deficits result from degeneration of noradrenergic neurons in the locus coeruleus and adjoining areas (Braak *et al.*, 2003; Remy *et al.*, 2005; Fornai *et al.*, 2007; Benarroch, 2009; Frisina *et al.*, 2009; Barone, 2010), a pathological feature described in post-mortem brain studies of patients with Parkinson's disease (Halliday *et al.*, 1990; Chan-Palay 1991; German *et al.*, 1992; Patt and Gerhard 1993; Del Tredici *et al.*, 2002; Braak *et al.*, 2003; Zarow *et al.*, 2003; Fornai *et al.*, 2007). In addition, rodent studies have suggested that degeneration of norepinephrine neurons in the locus coeruleus may precede and could contribute to the degeneration of dopaminergic cells in the ventral midbrain (Marien *et al.*, 1993; Fornai *et al.*, 1995, 1997, 2007; Rommelfanger *et al.*, 2007a, b).

The development of therapeutic approaches that could slow down the death of midbrain dopaminergic neurons has been of great interest during the past decade (Clarke, 2004; Schapira and Olanow, 2004; Biglan and Ravina, 2007; Kiebertz and Ravina, 2007; Schapira, 2008; Olanow, 2009; Pavese *et al.*, 2009; Yacoubian and Standaert, 2009). Although the mechanisms that underlie nigral dopaminergic cell loss in Parkinson's disease remain unknown (and may differ between patients), there is preclinical evidence that blockade of ionotropic glutamate receptors [i.e. 2-amino-3-(5-methyl-3-oxo-1,2-oxazol-4l)-propanoic acid and N-methyl-D-aspartic acid] protects midbrain dopaminergic neurons from toxin-induced degeneration in rodent and non-human primate models of Parkinson's disease (Turski *et al.*, 1991; Brouillet and Beal, 1993; Srivastava *et al.*, 1993; Loschmann *et al.*, 1994; Ossowska, 1994; Kanthasamy *et al.*, 1997; Lange *et al.*, 1997; Sonsalla *et al.*, 1998; Konitsiotis *et al.*, 2000; Horowitz and Greenamyre, 2010). However, presumably due to the fact that these fast acting glutamate receptors are essential for normal brain operations, chronic administration of ionotropic glutamate receptor antagonists elicits unwanted side effects in humans,

thereby limiting their usefulness as therapeutics (Marino *et al.*, 2003; Muir, 2006; Johnson *et al.*, 2009).

Because of their modulatory effects, localization specificity and potential for drug targeting at allosteric modulatory sites, the G protein-coupled metabotropic glutamate receptors have generated significant interest as new therapeutic targets for brain diseases, including Parkinson's disease (Conn *et al.*, 2005; Ossowska *et al.*, 2007; Gasparini *et al.*, 2008; Nicoletti *et al.*, 2010; Niswender and Conn, 2010). There is evidence that antagonists of metabotropic glutamate receptor 5 (mGluR5), a member of the group I metabotropic glutamate receptors, have significant antidyskinetic effects in rodent and non-human primate models of Parkinson's disease (Levandis *et al.*, 2008; Rylander *et al.*, 2009, 2010; Morin *et al.*, 2010; Johnston *et al.*, 2010). Furthermore, mGluR5 receptor antagonists have antiparkinsonian effects in 6-hydroxydopamine-treated rats (Breyse *et al.*, 2002), and reduce the toxicity of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) against midbrain dopaminergic neurons in mice (Battaglia *et al.*, 2004; Aguirre *et al.*, 2005; Armentero *et al.*, 2006; Vernon *et al.*, 2007).

In light of these promising findings, the present study addressed the potential neuroprotective effects of the highly specific mGluR5 antagonist, 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] (MTEP), against the loss of midbrain dopaminergic neurons in a chronic MPTP-treated monkey model of Parkinson's disease. Furthermore, taking into consideration that the chronic MPTP monkey model used in this study also results in damage to locus coeruleus noradrenergic cells (Masilamoni *et al.*, 2010b), combined with the fact that locus coeruleus neurons display functional mGluR5 expression (Page *et al.*, 2005; Rasmussen *et al.*, 2005; Noriega *et al.*, 2007), and that lesion of noradrenergic systems may underlie some of the non-motor symptoms of Parkinson's disease (Remy *et al.*, 2005; Fornai *et al.*, 2007; Benarroch, 2009; Frisina *et al.*, 2009), another objective of this study was to assess whether MTEP may also protect brainstem noradrenergic neurons in the locus coeruleus and adjoining cell groups against the toxic effects of chronic MPTP injections.

Some of the data reported here have been presented in abstract forms (Masilamoni *et al.*, 2009, 2010a, b).

## Materials and methods

### Animals

Ten adult female rhesus monkeys (*Macaca mulatta*, 4.5–8.5 kg) from the Yerkes National Primate Research Center colony were used in this

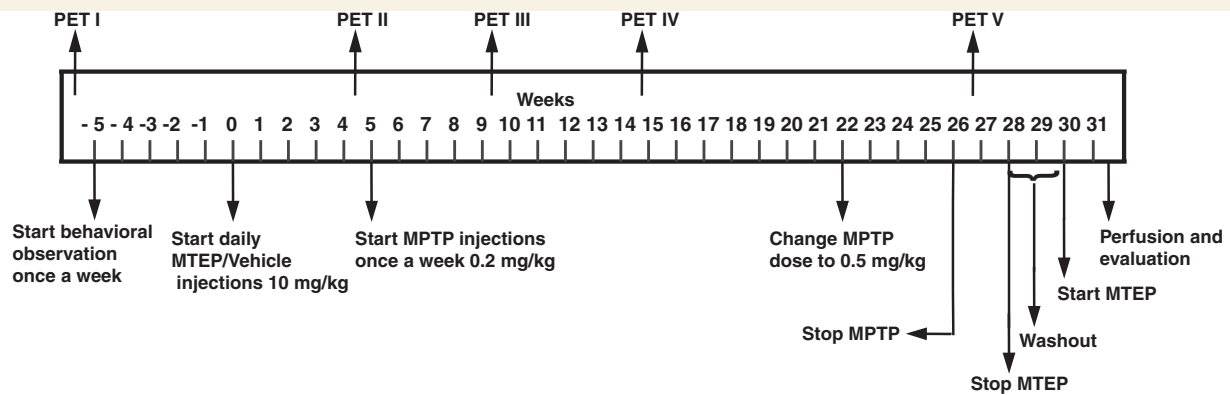


Figure 1 Study design.

study, in accordance with guidelines from the National Institutes of Health. All procedures were approved by Emory's Animal Care and Use Committee. The animals were housed in a temperature-controlled room and exposed to a 12-h light/dark cycle. They were fed twice daily with monkey chow supplemented with fruits or vegetables. The animals had free access to water.

### Experimental design

The 10 rhesus monkeys used in this study were divided into three groups. Group 1 consisted of three animals that were treated with MPTP and vehicle, Group 2 consisted of four animals treated with MPTP and MTEP, and Group 3 was comprised of three untreated monkeys that were used for tyrosine hydroxylase and dopamine transporter immunocytochemistry, and for stereological cell counting.

The temporal sequence of the experimental procedures is shown in Fig. 1. The seven monkeys in Groups 1 and 2 were first trained to sit in a primate chair, and transported to a behavioural testing room once weekly to habituate them to the testing environment. After collection of baseline behavioural data using methods described below, these monkeys received injections of either vehicle (Group 1) or MTEP (10 mg/kg, intramuscular, Group 2) for 5 weeks prior to the beginning of MPTP treatments, to assess the potential effects of mGluR5 blockade on motor behaviour and on 2β-carbomethoxy-3β-(4-chlorophenyl)-8-(2-[18F]-fluoroethyl)-nortropane (<sup>18</sup>F-FECNT) positron emission tomography (PET) imaging in naïve monkeys. The MTEP dosage was chosen based on pilot data from our laboratory (Bogenpohl *et al.*, 2006) and others showing that MTEP monotherapy is well tolerated and achieves maximum antiparkinsonian and antidyskinetic effects at 10 mg/kg (Phillips *et al.*, 2006; Morin *et al.*, 2010).

A series of weekly intramuscular MPTP injections (0.2–0.5 mg/kg body weight; Sigma-Aldrich) was then started (i.e. Week 5 in Fig. 1). This regimen was continued until significant parkinsonian motor symptoms appeared in all animals of either treatment groups. To avoid any possible unknown effects MTEP may have on MPTP pharmacokinetics and metabolism, the MTEP daily administration was performed at least 12 h prior to any weekly injections of MPTP. After 18–21 weeks of MPTP treatment, (i.e. Weeks 23–26 in Fig. 1) all Group 1 animals displayed moderate to severe parkinsonian symptoms according to the rating scale described below. In contrast, none of the MTEP-treated monkeys in Group 2 showed any significant change in their motor behaviour relative to baseline measurements (Fig. 2). At this point, the MPTP administration (cumulative dose of 7.9 mg MPTP/kg body weight) was terminated in all monkeys, followed by 2 weeks of MTEP washout before a final assessment of Parkinson's

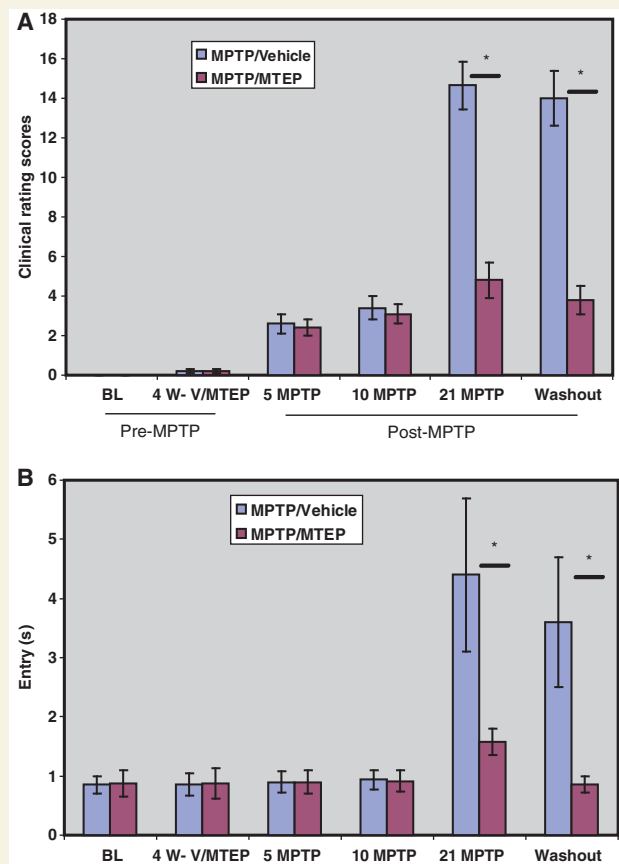


Figure 2 Histograms illustrating the progressive increase of (A) Parkinsonism rating scores and (B) food picking 'entry' time (i.e. the time between entering the apparatus and retrieval of a food object) in MPTP-treated monkeys (mean ± SD). Four weeks of vehicle (V) or MTEP (10 mg/kg) treatment (4 W-V/MTEP) did not alter behaviour. Significant motor impairments were observed in MPTP/vehicle-treated monkeys after 21 weekly MPTP injections, in the vehicle-treated group ('21 MPTP') and were maintained throughout the rest of the study period. \**P* < 0.001 for differences between the vehicle and MTEP-treated animals. No significant difference was found between control and MPTP/MTEP-treated monkeys. BL = baseline.

disease symptoms, to ensure that the differences in parkinsonian motor scores resulted from a variable degree of nigrostriatal degeneration instead of the symptomatic antiparkinsonian effects of MTEP between the two groups. The MTEP administration was then initiated again for one more week to ensure that all animals had been recently exposed to the drug treatment at the time of sacrifice (Fig. 1).

## Positron emission tomography neuroimaging

*In vivo* PET imaging with dopamine transporter ligands is the most sensitive approach to follow longitudinally the state of degeneration of the nigrostriatal dopaminergic system during the course of neuroprotective trials in Parkinson's disease (see review in Pavese and Brooks, 2009; Pavese *et al.*, 2009). We have recently shown that the PET tracer  $^{18}\text{F}$ -FECNT, is a highly reliable ligand to estimate levels of striatal and nigral dopamine transporter in the MPTP-treated monkey model of Parkinson's disease (Masilamoni *et al.*, 2010a). In order to further characterize the use of this ligand as a reliable indicator of the degree of striatal and nigral dopaminergic denervation in MPTP-treated monkeys (Masilamoni *et al.*, 2010b), and to follow dynamic changes of the nigrostriatal dopaminergic system during the course of MPTP intoxication ( $\pm$ MTEP), each monkey underwent five  $^{18}\text{F}$ -FECNT PET scans in these experiments (Fig. 1). First, we assessed possible changes in  $^{18}\text{F}$ -FECNT binding induced by MTEP prior to the beginning of MPTP administration with two PET scans (PET I and PET II) that were performed 10 weeks apart before MPTP treatment. PET I was performed prior to the beginning of any behavioural training or drug treatment. Then, baseline behavioural data were collected for five weeks prior to the beginning of MTEP or vehicle administration (i.e. Week 0 in Fig. 1). During this period, Group 1 animals received daily injections of vehicle, while Group 2 monkeys were injected daily with MTEP for 4 weeks prior to the second PET scan (PET II) and the beginning of the MPTP treatment. The third and fourth PET scans (PET III and PET IV) were performed 5 and 10 weeks after the start of the MPTP treatment regimen (i.e. Weeks 9 and 14 in Fig. 1) to monitor changes in striatal dopamine transporter binding. The fifth PET scan (PET V) was performed 21 weeks after the start of the MPTP treatment protocol (i.e. Week 26 in Fig. 1), after the development of parkinsonian symptoms in all MPTP/vehicle-treated monkeys.

PET images were acquired with a Siemens Focus 220 micro-PET scanner. The monkeys were fasted for 12 h prior to the PET scans. The animals were intubated under anaesthesia with Telazol (3 mg/kg intramuscular), and then maintained throughout the imaging session on a mixture of 1% isoflurane and 5% oxygen gas. The monkeys were positioned in the scanner and connected to monitoring equipment to measure respiratory parameters, blood pressure, pulse rate and rectal temperature. A 15-min transmission scan was obtained for attenuation correction, then a slow bolus of  $\sim 5.0$  mCi of  $^{18}\text{F}$ -FECNT (specific activity 1.5 Ci/ $\mu\text{mol}$ ) was injected over 5–6 min at a rate of 1.0 ml/min. The collection of scans began at the same time as the start of the radiotracer injection. The initial acquisition was a 28-frame dynamic sequence, starting with 30-s scans and ending with 20-min scans (total duration, 110 min). All images were reconstructed using the manufacturer-supplied software with measured attenuation correction, zoom factor 8, and Shepp–Logan reconstruction with a filter cut-off at 1 cycle/cm. The axial slice thickness was 3.375 mm. All images were decay-corrected to the time of injection. For the generation of time-activity curves for each monkey, the five different PET images taken in different experimental conditions (PET I–V) were superimposed using IDL software (ITT Visual Information

Solutions), and averaged to draw regions of interest so that the volume would be equal and comparable. Regions of interest were manually drawn on the average image, delimiting the following striatal regions: putamen/associative, putamen/motor, putamen/limbic, caudate nucleus/associative, nucleus accumbens/limbic and the substantia nigra. The regions of interest were then superimposed onto the individual images to obtain time-activity curves. Because of the minimal expression of dopamine transporter in the cerebellum, we used it as the reference region, and expressed the FECNT uptake value as region of interest/cerebellum ratio, as described in our previous studies (Votaw *et al.*, 2002; Masilamoni *et al.*, 2010a). One-way ANOVA was used to determine significance of differences between control, MPTP/vehicle and MPTP/MTEP groups.

## Behavioural observations

Changes in the animal's behavioural state were documented through observations in an observation cage of which one of the side walls was made of Plexiglas for easy visibility of the monkey. After having been brought to this cage, the animals were given a 30-min habituation period before being videotaped for 15 min/session. Their behaviour was later monitored and evaluated from videos viewed by two observers, one of them blinded to the treatment regimen, using a modified Parkinson's disease rating scale. The differences in the rating scores between the two observers were  $<6\%$ . The mean values obtained by the two experimenters were used for statistical analysis. The scale used in this study evaluated nine criteria (gross motor activity, balance, posture, arm bradykinesia, arm hypokinesia, leg bradykinesia, leg hypokinesia, arm tremor and leg tremor). Each criterion received a score between 0 and 3 (0 = normal, 1 = mild, 2 = moderate, 3 = severe), for a maximum of 27 points. The total number of points was used as the clinical score to compare the severity of parkinsonian motor symptoms across animals in the different experimental groups.

In addition, the monkeys were trained to perform a food retrieval task to assess their fine motor skills. During this test, animals were required to retrieve a raisin from a device that uses infrared beams to automatically measure the time required for the monkey to move its hand from the door to the raisin (entry) and from the raisin back to the door (exit). A minimum of 10 trials per hand was performed in each session. The average entry and exit times to perform this test were calculated from data collected in both hands because there was no significant difference in this test performance between the right and left hand.

Because of the rapid clearance of MTEP (Anderson *et al.*, 2003; Busse *et al.*, 2004; Johnston *et al.*, 2010), the behavioural tests were performed between 20 and 24 h after the daily MTEP injections to avoid any confound in the test performance induced by possible acute antiparkinsonian effects of MTEP.

## Termination of the experiments

At the end of the experiments, the monkeys were deeply anaesthetized with an overdose of pentobarbital (100 mg/kg, intravenous), and perfused transcardially with cold oxygenated Ringer's solution, followed by 2 l of fixative containing 4% paraformaldehyde and 0.1% glutaraldehyde in phosphate buffer (0.1 M, pH 7.4). After perfusion, the brains were removed from the skull and cut into 10 mm-thick blocks in the frontal plane. The blocks were further cut into 50  $\mu\text{m}$ -thick sections with a vibratome and used for post-mortem immunostaining and cell counting procedures.

## Immunostaining

To validate the extent of MTEP effects upon MPTP-induced degeneration of the dopaminergic nigrostriatal system, striatal and midbrain sections of untreated, MPTP/vehicle- and MPTP/MTEP-treated monkeys were immunostained with antibodies against dopamine transporter (monoclonal rat anti-dopamine transporter IgGs) or tyrosine hydroxylase (monoclonal mouse anti-tyrosine hydroxylase IgGs) at a concentration of 1:1000 (Millipore; catalogue numbers MAB 369 and 318, respectively). In addition, we used highly specific mouse monoclonal calbindin D28K antibodies at a concentration of 1:4000 (Sigma, catalogue number C9848) to label calbindin-positive cells in the dorsal tier of the substantia nigra pars compacta and the ventral tegmental area, which allowed us to differentiate these neurons from the calbindin-negative ventral tier substantia nigra pars compacta neurons (Masilamoni *et al.*, 2010a). The immunoperoxidase avidin biotin complex method with diaminobenzidine as chromogen was used to localize the different markers according to procedures described in previous studies (Hsu *et al.*, 1981; Smith and Bolam, 1991). Sections were then rinsed in phosphate-buffered saline (0.01 M, pH 7.4), mounted onto gelatin-coated slides, dehydrated and cover-slipped with Permount. Dopamine transporter and tyrosine hydroxylase immunoreactivity in the striatum was quantified using the MCID program (Imaging Research Inc.). Each slide was illuminated using an even-field illumination table, and macroscopic images were acquired with a Cool SNAP cf photometrics camera. The regions of interest were outlined on a computer screen, and the average pixel density measured and expressed as relative optical density. Mean values were calculated from five to seven sections per hemisphere. The SigmaStat software (Systat) was used to assess the significance of differences in labelling intensity measured from the various striatal regions of interest in the MPTP/vehicle- and MPTP/MTEP-treated animals.

## Stereological estimation of tyrosine hydroxylase-positive midbrain, locus coeruleus, A5 and A7 noradrenaline neurons

The unbiased stereological estimation of the number of dopamine neurons in the ventral substantia nigra pars compacta, dorsal substantia nigra pars compacta, ventral tegmental area, and norepinephrine neurons in locus coeruleus, A5, A7 noradrenaline cells was achieved using the optical fractionator principle (Stereoinvestigator, MicroBrightField, Inc.), a stereological approach that combines the optical disector with a fractionator sampling scheme. This sampling technique is not affected by tissue volume changes and does not require reference volume determinations. The random systematic sampling of counting areas was done using the Leica DMR microscope. To count midbrain tyrosine hydroxylase-positive neurons, we first took low power micrographs ( $\times 1.25$ ) of tyrosine hydroxylase- and calbindin-immunostained ventral midbrain sections, and manually delineated the borders of the ventral substantia nigra pars compacta, dorsal substantia nigra pars compacta, and ventral tegmental area, based on the presence or absence of calbindin-positive neurons (Masilamoni *et al.*, 2010a). Then, the borders of the different ventral midbrain regions were manually delineated on tyrosine hydroxylase-immunolabelled slides adjacent to those immunostained for calbindin. The noradrenergic cell groups were delineated based on the expression of tyrosine hydroxylase-immunoreactive neurons using the following anatomical landmarks: (i) locus coeruleus: tyrosine hydroxylase-positive neurons

located above the dorsal limit of the medial longitudinal fasciculus medial to the superior cerebellar peduncle; (ii) A7 noradrenaline cells: tyrosine hydroxylase-positive neurons located between the dorsal border of medial longitudinal fasciculus and the medial olivary nucleus; and (iii) A5 noradrenaline cells: tyrosine hydroxylase-positive neurons surrounding the medial olivary nucleus (Fig. 8A). Counts of tyrosine hydroxylase-positive cells were generated using a  $\times 100$  oil-immersion objective. To perform unbiased stereology, counting frames ( $65 \times 65 \mu\text{m}$ ) were randomly placed by the stereology software within the chosen region of interest. The software also controlled the position of the x–y stage of the microscope, so that the entire brain region could be scanned by successively meandering between counting frames. On average, 12 sections were analysed and  $\sim 300$  cells were counted. The software calculated the estimated total number of cells in each region of interest per hemisphere. The cell counts were performed by two investigators, one of them blinded to the drug treatment. The differences in the total number of neurons counted by these investigators were  $< 5\%$ . The mean of values obtained by each experimenter was used for statistical analysis.

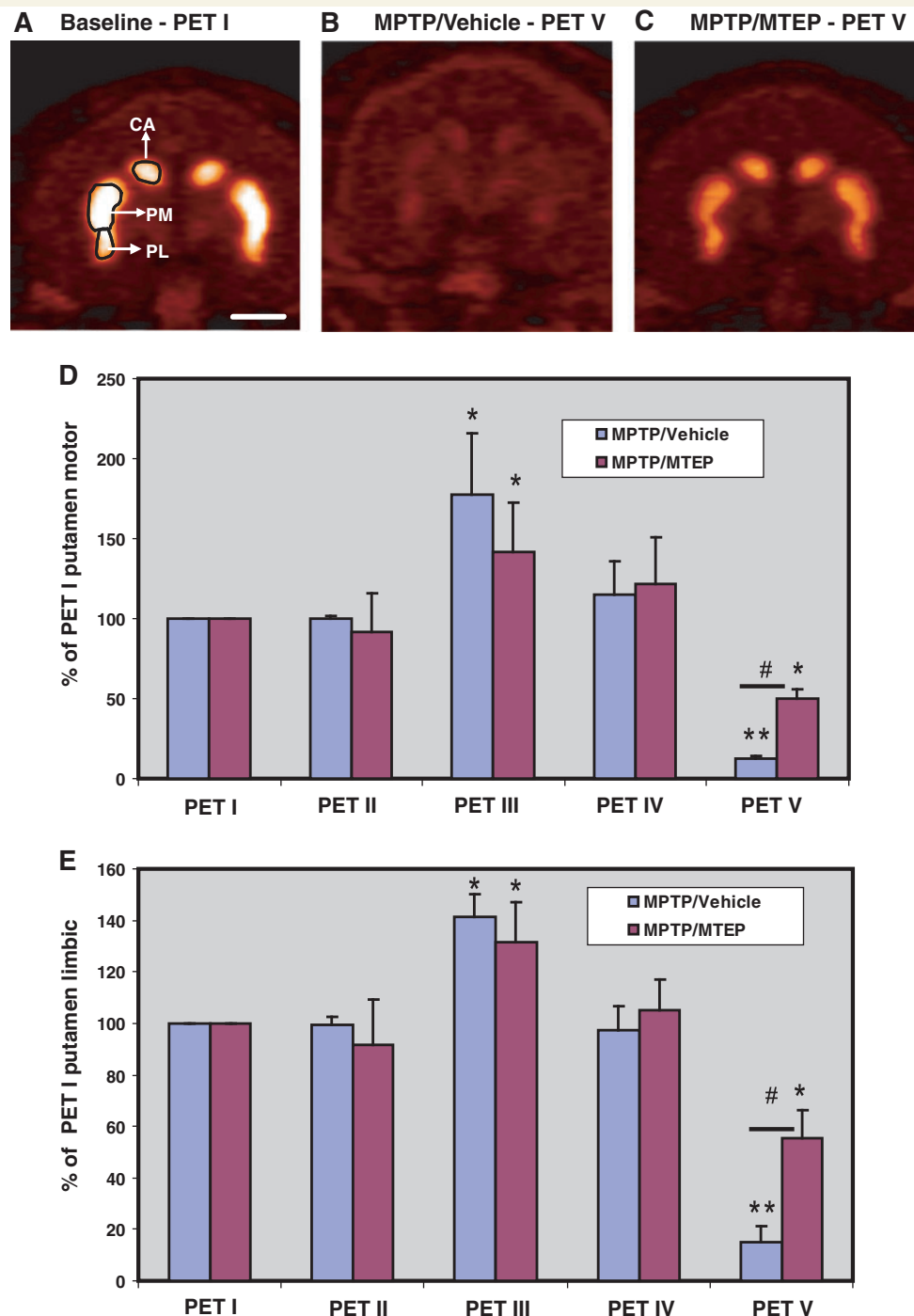
## Results

### Comparison of changes in striatal $^{18}\text{F}$ -2 $\beta$ -carbomethoxy-3 $\beta$ -(4-chlorophenyl)-8-(2-[ $^{18}\text{F}$ ]-fluoroethyl)-nortropine positron emission tomography imaging data with changes in behaviour

PET I served to determine the baseline  $^{18}\text{F}$ -FECNT uptake values in the striatum and the substantia nigra of each animal that received subsequent MPTP injections, while PET II served to assess potential effects of MTEP treatment upon  $^{18}\text{F}$ -FECNT binding. This scan was performed 20–24 h after the last MTEP (or vehicle) administration, to avoid possible acute MTEP effects upon FECNT binding. The uptake values measured in PET II were not significantly different from those obtained under baseline conditions (PET I) (Figs 3D and E, 4D–F and 5D), indicating that MTEP alone does not significantly affect  $^{18}\text{F}$ -FECNT uptake measurements at the striatal and midbrain levels. Four weeks of MTEP treatment also did not alter scores on the parkinsonian rating scale, or performance in the food picking task in these monkeys (Fig. 2A and B).

However, the subsequent PET scan (PET III), performed 5 weeks after the beginning of weekly MPTP treatment, revealed a significant increase ( $P < 0.001$ ) in  $^{18}\text{F}$ -FECNT uptake in all striatal regions of interest, except in the nucleus accumbens (nucleus accumbens/limbic) in the MPTP/vehicle- and MPTP/MTEP-treated animals (Figs 3D and E and 4D–F). This increase in PET measurement was not associated with changes in the parkinsonism rating score and performance in the food picking task (Fig. 2A and B). The transient increase in FECNT uptake was no longer apparent in the next PET scan, performed 5 weeks later (PET IV; Figs 3D and E and 4D and E).

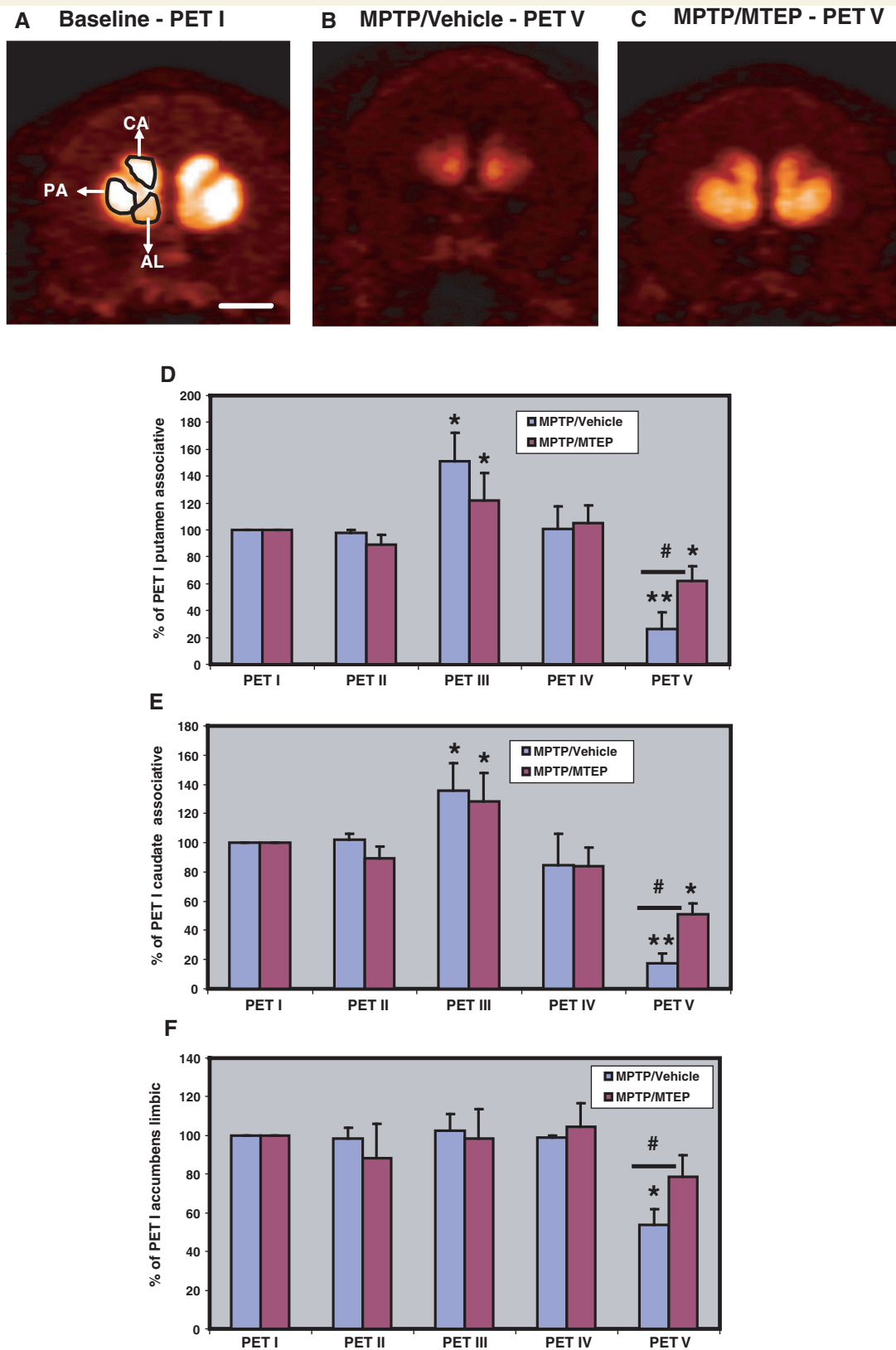
After 18–21 weeks of MPTP treatment, all MPTP/vehicle-treated monkeys displayed moderate to severe parkinsonian



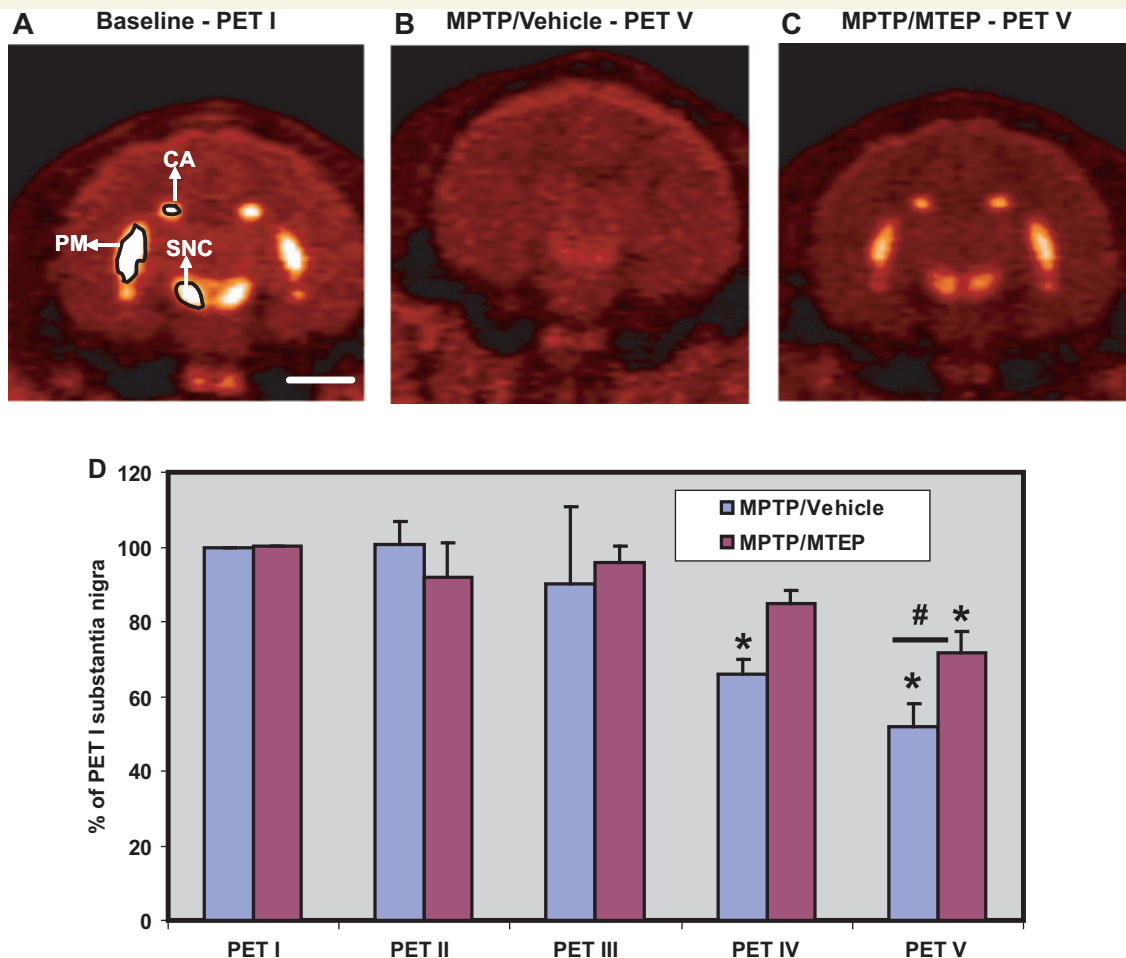
**Figure 3** Representative post-commissural coronal *in vivo*  $^{18}\text{F}$ -FECNT PET images. (A) Baseline, (B) MPTP/vehicle, (C) MPTP/MTEP-treated monkeys. (D, E) Bar graphs showing the percentages of striatocerebellar  $^{18}\text{F}$ -FECNT uptake ratios (means  $\pm$  SD) for the motor putamen (D) and the limbic putamen (E), obtained from the five different PET scans collected throughout the study.  $***P < 0.001$ ;  $*P < 0.05$  for differences between control, MPTP/vehicle- and MPTP/MTEP-treated animals.  $\#P < 0.05$  for differences between vehicle- and MTEP-treated animals. CA = associative caudate nucleus; PM = motor putamen; PL = limbic putamen. Scale bar = 10 mm (A–C).

symptoms, while monkeys treated with MPTP/MTEP did not display significant changes in their parkinsonism rating scores and performance in the food picking task (Fig. 2A and B). The different behavioural response to MPTP treatment was correlated with a significantly lower ( $P < 0.001$ ) level of  $^{18}\text{F}$ -FECNT uptake in all striatal regions of interest in MPTP/vehicle- than in MPTP/

MTEP-treated monkeys (PET V; Figs 3 and 4). It is worth noting that the striatal  $^{18}\text{F}$ -FECNT uptake values were below the baseline values in both MPTP/vehicle- and MPTP/MTEP-treated animals at this time point, suggesting that the daily MTEP treatment did not completely prevent a decrease in striatal dopamine transporter binding (Figs 3 and 4).



**Figure 4** Representative pre-commissural coronal planes of *in-vivo* <sup>18</sup>F-FECNT PET images. (A) Baseline, (B) MPTP/vehicle, (C) MPTP/MTEP-treated monkeys. (D–F) Bar graphs showing the percentages of striatocerebellar <sup>18</sup>F-FECNT uptake ratios (means ± SD) for the pre-commissural putamen (D), caudate nucleus (E) and the nucleus accumbens (F) obtained from the five PET scans throughout the study. \*\*\**P* < 0.001; \**P* < 0.05 for differences from control, MPTP/vehicle-, MPTP/MTEP-treated animals. #*P* < 0.05 for differences between the vehicle and MTEP-treated animals. No significant difference was found in the nucleus accumbens between control and MPTP/MTEP-treated monkeys. AL = limbic nucleus accumbens; CA = associative caudate nucleus; PA = associative putamen. Scale bar = 10 mm (A–C).



**Figure 5** Representative midbrain coronal *in-vivo*  $^{18}\text{F}$ -FECNT PET images. (A) Baseline, (B) MPTP/vehicle, (C) MPTP/MTEP-treated monkeys. (D) Bar graph showing the percentage of nigrocerbellar  $^{18}\text{F}$ -FECNT uptake ratios (means  $\pm$  SD) for the substantia nigra pars compacta obtained from the five PET scans throughout the study. \* $P < 0.05$  for differences from control, MPTP/vehicle, MPTP/MTEP-treated animals. # $P < 0.05$  for differences between the vehicle and MTEP-treated animals. CA = associative caudate nucleus; PM = motor putamen; SNC = substantia nigra pars compacta. Scale bar = 10 mm (A–C).

### Effects of 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine treatment on 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced changes in midbrain $^{18}\text{F}$ -2 $\beta$ -carbomethoxy-3 $\beta$ -(4-chlorophenyl)-8-(2-[ $^{18}\text{F}$ ]-fluoroethyl)-nortropine binding

The pattern of changes of  $^{18}\text{F}$ -FECNT uptake in the substantia nigra pars compacta/ventral tegmental area complex over time was slightly different from what was found in the striatum. Specifically, there was no transient increase of uptake measured in PET III in the substantia nigra pars compacta/ventral tegmental area region (Fig. 5). In addition, MPTP/vehicle-treated monkeys displayed a significant decrease in FECNT uptake ( $P < 0.05$ ) in the substantia nigra pars compacta/ventral tegmental area, compared to the baseline in PET IV. Such a change was not observed in

MPTP/MTEP-treated monkeys (Fig. 5D). In the final PET scan (PET V), both groups of animals displayed lower substantia nigra pars compacta/ventral tegmental area  $^{18}\text{F}$ -FECNT uptake than at baseline, with a greater decrease in the MPTP/vehicle-treated animals (Fig. 5). However, the differences in uptake values between the two groups were not as pronounced in the substantia nigra pars compacta as they were in the striatum (compare PET V values in Fig. 5D with Figs 3D and E and 4D–F).

### 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine-induced protection of dopaminergic neurons against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-mediated neurotoxicity

To determine whether the lack of parkinsonism in MPTP/MTEP-treated animals after 21 weeks of MPTP treatment was



due to MTEP treatment-related preservation of the nigrostriatal dopaminergic denervation, or antiparkinsonian effects of MTEP (Bogenpohl *et al.*, 2006), we compared the various behavioural measures taken after 21 weeks of MPTP treatment (i.e. Week 26 in Fig. 1) with those obtained following a 2 weeks washout of MTEP (i.e. Week 30 in Fig. 1). These data revealed no significant difference in behavioural scores between the two time points for either experimental group (Fig. 2), suggesting that the MTEP effects were not due to its symptomatic antiparkinsonian properties.

### Post-mortem assessment of nigrostriatal dopaminergic denervation in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/vehicle and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine/3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine-treated animals

Brain sections from perfusion-fixed monkeys in each group were processed for light microscopic localization of tyrosine hydroxylase and dopamine transporter immunoreactivity. The data presented in Fig. 6A–F show different pre- and post-commissural striatal territories that were immunostained with a dopamine transporter antibody from control (Fig. 6A and D), MPTP/vehicle- (Fig. 6B and E) and MPTP/MTEP-treated (Fig. 6C and F) animals. MPTP/vehicle-treated animals displayed a significantly more severe decrease in striatal dopamine transporter immunoreactivity in all regions of interest compared with the MPTP/MTEP-treated monkeys (Fig. 6A–G). The degree of dopamine transporter depletion in the MPTP/vehicle-treated monkeys was most severe in the post-commissural putamen regions of interest (putamen/motor, putamen/limbic) (Fig. 6G). Similar findings were found in striatal tissue immunostained for tyrosine hydroxylase (Fig. 6H).

The degree of tyrosine hydroxylase-positive cell loss in the ventral midbrain, assessed with unbiased stereological cell counts, was different across the three groups of dopaminergic neurons examined (ventral substantia nigra pars compacta, dorsal substantia nigra pars compacta, ventral tegmental area). In the ventral substantia nigra pars compacta, both MPTP/vehicle- and MPTP/MTEP-treated monkeys displayed a significant loss of tyrosine hydroxylase-immunoreactive neurons compared with controls ( $P < 0.001$ ). In the dorsal substantia nigra pars compacta and ventral tegmental area, a significant loss of tyrosine hydroxylase-immunoreactive neurons was only seen in the MPTP/vehicle-treated animals ( $P < 0.05$ ). The decrease in the number of tyrosine hydroxylase-immunoreactive neurons was significantly more pronounced ( $P < 0.001$ ) in the MPTP/vehicle-treated monkeys than in MPTP/MTEP-treated animals in all midbrain dopaminergic cell groups (ventral substantia nigra pars compacta:  $68 \pm 17\%$  loss in vehicle-treated animals versus  $31 \pm 8\%$  loss in MTEP-treated animals; dorsal substantia nigra

pars compacta:  $57 \pm 21\%$  versus  $25 \pm 10\%$ ; ventral tegmental area:  $48 \pm 14\%$  versus  $11 \pm 13\%$ ) (Fig. 7A–D).

### 3-[(2-methyl-1,3-thiazol-4-yl) ethynyl] pyridine-mediated protection of noradrenergic neurons against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine toxicity

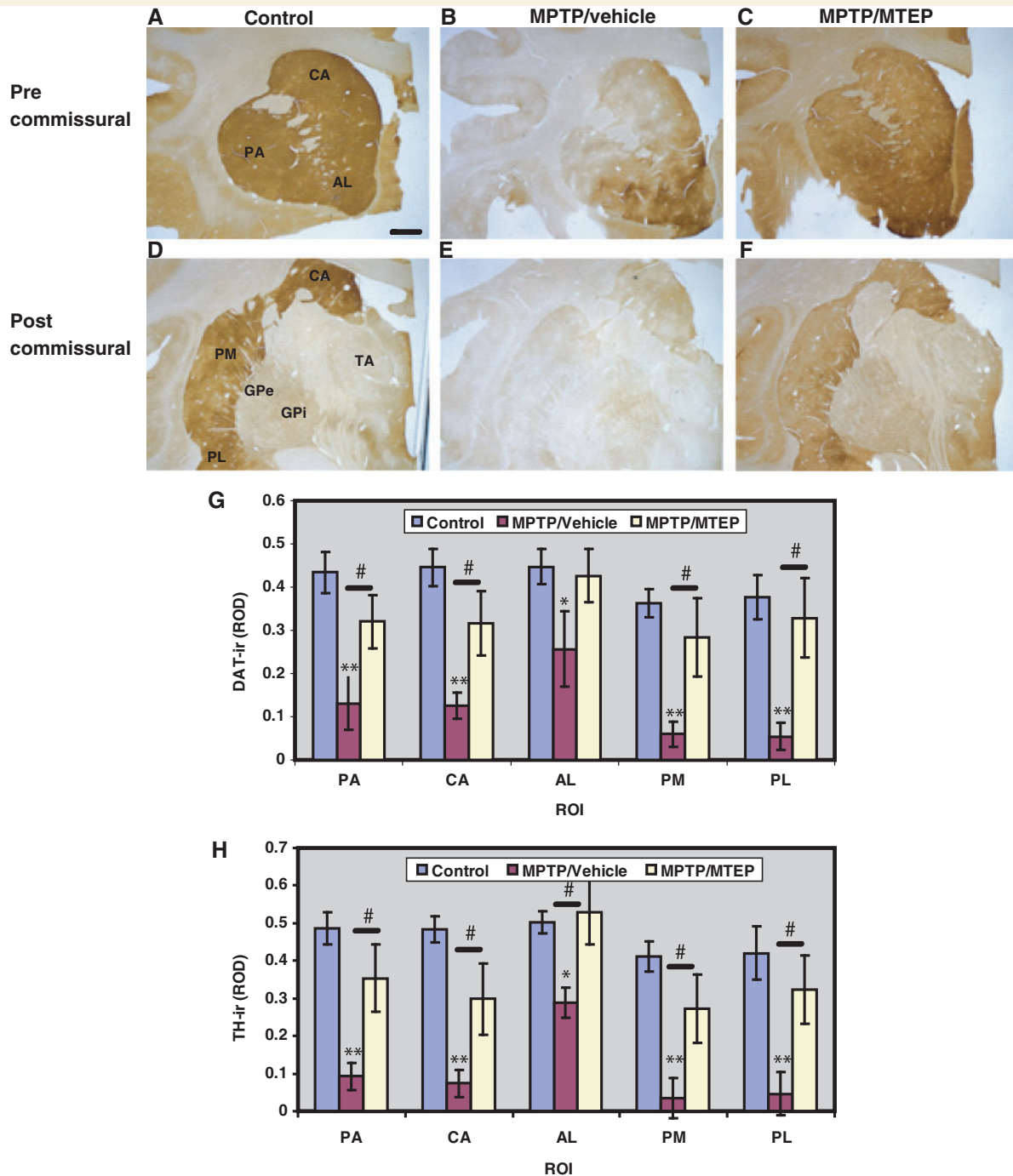
Taking advantage of the chronic MPTP regimen used in the present study that induces significant loss of noradrenergic neurons in the locus coeruleus and related A5 and A7 noradrenaline cell groups (Masilamoni *et al.*, 2010b), we determined the effect of MTEP treatment towards noradrenergic cell loss in MPTP-treated monkeys. As shown in Fig. 8, MPTP treatment resulted in a more significant reduction of the number of tyrosine hydroxylase-positive neurons in the locus coeruleus, A5 and A7 noradrenaline cells in vehicle-treated than in MTEP-treated animals (locus coeruleus:  $38.7 \pm 8.8\%$  loss in vehicle-treated animals versus  $15.2 \pm 9.4\%$  loss in MTEP-treated animals; A5 noradrenaline cells:  $48.3 \pm 7.2\%$  versus  $13.8 \pm 3.7\%$ ; A7 noradrenaline cells:  $45.1 \pm 7.8\%$  versus  $16.4 \pm 4.7\%$ ). Although the percentage of cell loss in MPTP/vehicle-treated animals was most severe in the locus coeruleus, it reached significance in all neuronal groups ( $P < 0.001$ ). The total number of labelled neurons in the MPTP/MTEP-treated animals was not significantly different from the controls (Fig. 8;  $P > 0.05$ ).

## Discussion

The present study shows that the mGluR5 antagonist, MTEP, has significant protective effects against MPTP-induced degeneration of midbrain dopaminergic neurons and noradrenergic locus coeruleus/A5/A7 neurons in non-human primates. Together, these findings suggest that mGluR5 receptor antagonists may be useful agents to delay the demise of dopaminergic and noradrenergic systems in Parkinson's disease. Furthermore, our data demonstrate that the dopamine transporter ligand,  $^{18}\text{F}$ -FECNT is a sensitive and highly reliable biomarker to assess the state of the striatal dopamine innervation and the extent of midbrain dopaminergic cell loss during the course of nigrostriatal dopaminergic denervation in parkinsonism (Masilamoni *et al.*, 2010a).

### Potential mechanisms by which metabotropic glutamate receptor 5 antagonists may protect midbrain dopaminergic neurons from 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine toxicity in monkeys

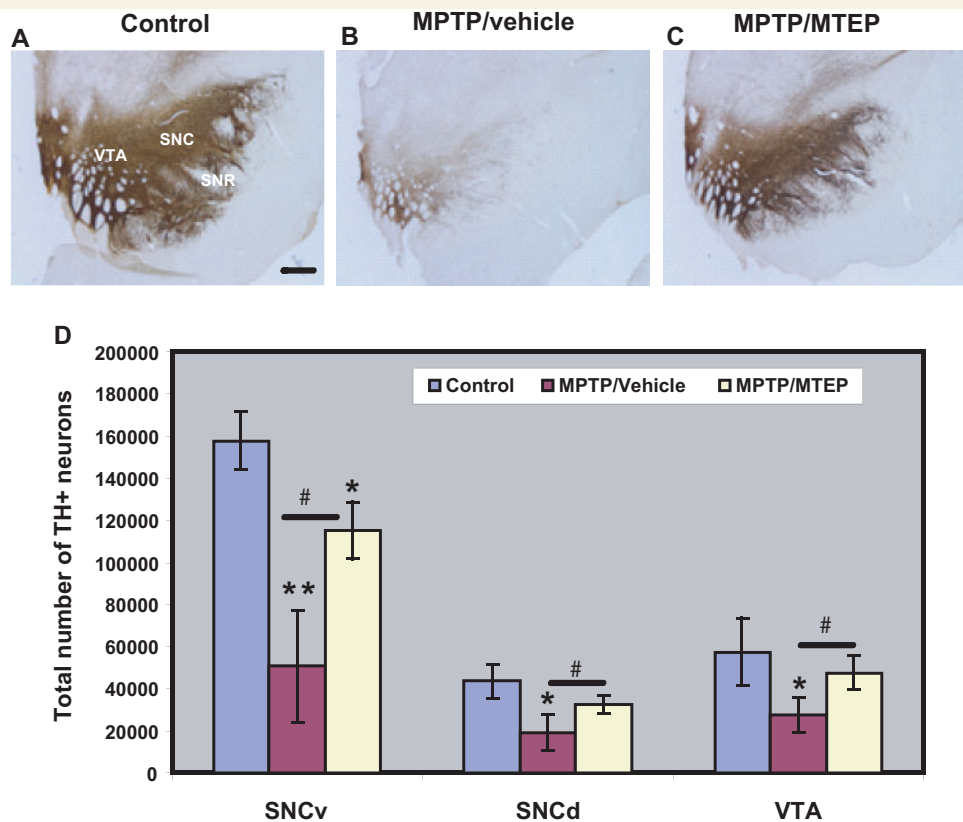
The present data provide neuropathological and behavioural evidence that prolonged systemic administration of the mGluR5 specific antagonist, MTEP, protects dopaminergic neurons in the



**Figure 6** Representative coronal sections of pre-commissural (A–C) and post-commissural (D–F) striatum showing dopamine transporter (DAT) immunoreactivity in Controls, MPTP/vehicle and MPTP/MTEP-treated monkeys. (G–H) Densitometry analysis of control, MPTP/vehicle and MPTP/MTEP-treated monkeys. \* $P < 0.001$  and \*\* $P < 0.05$  for differences between control and MPTP/vehicle-treated monkeys. # $P < 0.05$  for differences between the vehicle and MTEP-treated animals. No significant difference was found between control and MPTP/MTEP-treated monkeys. AL = limbic nucleus accumbens; CA = associative caudate nucleus; GPe = external pallidal segment; GPi = internal pallidal segment; PA = associative putamen; PL = limbic putamen; PM = motor putamen; ROD = relative optical density; ROI = region of interest; TA = thalamus; TH-ir = tyrosine hydroxylase immunoreactivity. Scale bar = 5 mm (A–F).

substantia nigra pars compacta/ventral tegmental area against MPTP-induced degeneration in rhesus monkeys. These findings are consistent with previous data showing that mice treated with the mGluR5 receptor antagonist MPEP, or mice that lack

the mGluR5 receptor gene, are significantly protected against MPTP-induced nigrostriatal dopamine denervation (Battaglia *et al.*, 2004). The exact mechanisms underlying these beneficial effects remain unknown, although various possibilities can be

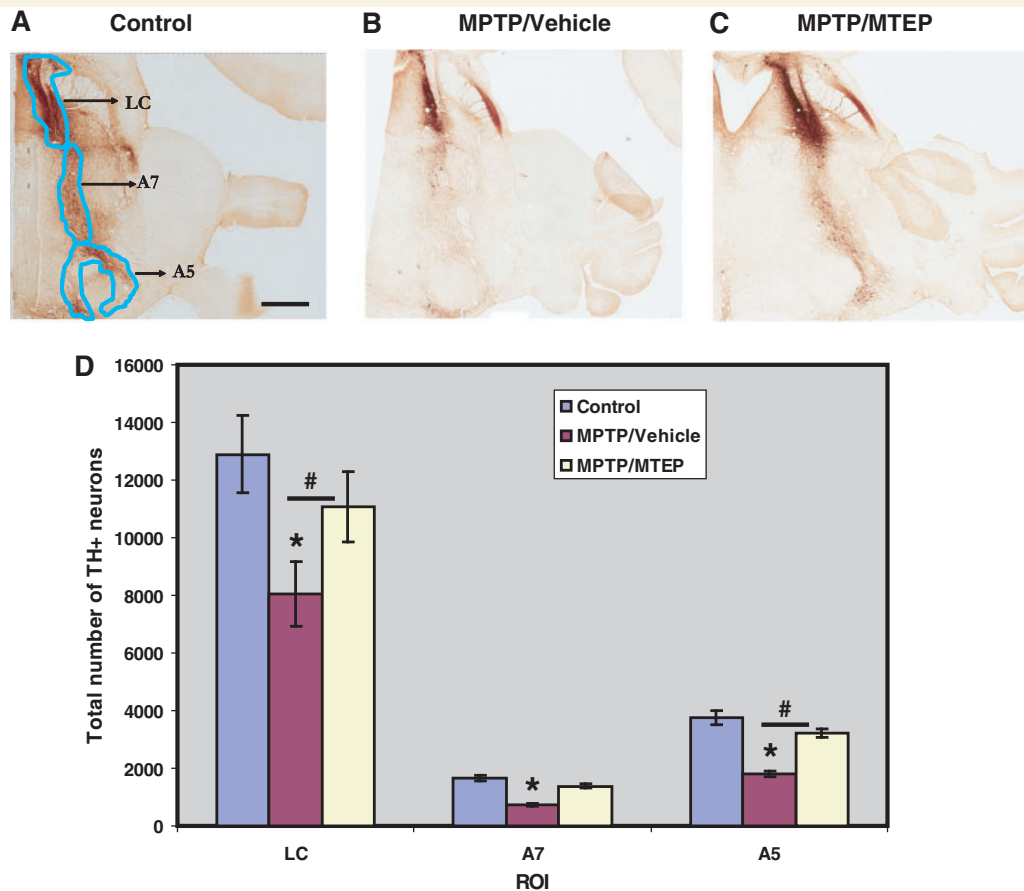


**Figure 7** Photomicrographs showing tyrosine hydroxylase labelling in the mesencephalon. (A) Control, (B) MPTP/vehicle, (C) MPTP/MTEP-treated monkeys. (D) Stereological estimate of the total number of tyrosine hydroxylase-positive (TH+) neurons (means  $\pm$  SD) in the ventral substantia nigra pars compacta, dorsal substantia nigra pars compacta and ventral tegmental area regions of control, MPTP/vehicle and MPTP/MTEP-treated monkeys. \* $P < 0.001$ ; \*\* $P < 0.05$  for differences between control, MPTP/vehicle and MPTP/MTEP-treated animals by using one-way ANOVA with Tukey's *post hoc* test. There was no significant difference found in the dorsal substantia nigra pars compacta and ventral tegmental area between control and MPTP/MTEP-treated monkeys. SNcd = dorsal substantia nigra compacta; SNcv = ventral substantia nigra compacta; SNR = substantia nigra pars reticulata; VTA = ventral tegmental area. Scale bar = 5 mm (A–C).

suggested. Thus, in light of the literature suggesting that calcium mishandling, mitochondrial respiration impairment and excitotoxicity contribute to the loss of midbrain dopaminergic cells in Parkinson's disease (Sherer *et al.*, 2002; Maesawa *et al.*, 2004; Wallace *et al.*, 2007; Caudle and Zhang 2009; Chan *et al.*, 2009; Winklhofer and Haass 2009; Cannon and Greenamyre, 2010; Van Laar and Berman 2010), the blockade of mGluR5 on the surface of midbrain dopaminergic neurons (Hubert *et al.*, 2001; Hubert and Smith, 2004) may provide its protective effects upon substantia nigra pars compacta neurons through a reduction of mGluR5-mediated intracellular calcium release from internal stores, one of the key mechanisms by which mGluR5 activation mediates its excitatory effects in various neuronal populations (Kim *et al.*, 1994; Pin and Duvoisin, 1995; Conn and Pin, 1997; Nakanishi *et al.*, 1998; Sala *et al.*, 2005; Zhang *et al.*, 2005). However, this hypothesis awaits confirmation of intracellular calcium increase in response to mGluR5 activation on nigral dopaminergic neurons. In addition to these postsynaptic effects, mGluR5 blockers may also reduce the activity of glutamatergic inputs to the substantia nigra pars compacta, specifically those

from subthalamic nucleus neurons, which strongly respond to mGluR5 activation and display abnormal increased activity in parkinsonism (Rodriguez *et al.*, 1998; Bezard *et al.*, 1999; Awad *et al.*, 2000; Breyse *et al.*, 2003; Fazal *et al.*, 2003; Shimo and Wichmann 2009; Piallat *et al.*, 2011).

Another mechanism by which MTEP protection could be mediated is through the regulation of the potentiating effects of mGluR5 upon N-methyl-D-aspartic acid receptor function (Calabresi *et al.*, 1998; Tu *et al.*, 1999; Awad *et al.*, 2000; Alagarsamy *et al.*, 2002). Because N-methyl-D-aspartic acid receptors represent a main source of calcium entry in midbrain dopaminergic neurons, a reduced mGluR5-mediated potentiation of these receptors may have a significant impact on the level of intracellular calcium handled by dopaminergic neurons upon glutamatergic activation. In fact, N-methyl-D-aspartic acid receptor blockers have been considered as neuroprotective agents against toxin-induced degeneration of midbrain dopaminergic neurons in animal models of parkinsonism (Turski *et al.*, 1991; Srivastava *et al.*, 1993; Loschmann *et al.*, 1994; Ossowska, 1994; Kanthasamy *et al.*, 1997; Sonsalla *et al.*, 1998), though their clinical use is limited due to



**Figure 8** Tyrosine hydroxylase labelling in noradrenergic cells groups. (A) Control, (B) MPTP/vehicle, (C) MPTP/MTEP-treated monkeys. (D) Stereological estimate of the total number of tyrosine hydroxylase-positive (TH+) neurons (means  $\pm$  SD) in locus coeruleus, A5 and A7 regions of control, MPTP/vehicle and MPTP/MTEP-treated monkeys, representing noradrenergic neurons. \* $P < 0.05$  for differences between control, MPTP/vehicle- and MPTP/MTEP-treated animals. # $P < 0.05$  for differences between the vehicle and MTEP-treated animals. LC = locus coeruleus; A5 and A7 = catecholaminergic areas A5 and A7; ROI = region of interest. Scale bar = 5 mm (A–C).

unwanted side effects, as discussed above. Finally, knowing the significant glial expression of mGluR5 in the ventral midbrain (Hubert *et al.*, 2001; Hubert and Smith, 2004), and its upregulation in some inflammatory processes (Byrnes *et al.*, 2009; Loane *et al.*, 2009; Drouin-Ouellet *et al.*, 2011), the blockade of these receptors in astrocytes and microglia may modulate inflammatory processes that contribute to MPTP-induced toxicity towards midbrain dopaminergic neurons.

On the other hand, one must also consider the possibility that the MTEP-mediated neuroprotective effects could have been induced by interference of MTEP with the conversion mechanisms of MPTP into the toxin 1-methyl-4-phenylpyridinium+, thereby reducing exposure of dopaminergic and noradrenergic terminals in MTEP-treated animals to toxic levels of 1-methyl-4-phenylpyridinium+. However, such interfering mechanism is very unlikely based on recent mouse data showing that MPEP administration 30 min prior to MPTP (10 mg/kg) injections does not have any significant effect on the intrastriatal levels of 1-methyl-4-phenylpyridinium+ measured at various time points *in vivo* from striatal homogenates (Battaglia *et al.*, 2004).

## Biphasic regulation in striatal dopamine transporter binding during the course of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-mediated nigrostriatal dopaminergic degeneration

Our longitudinal monitoring of striatal dopamine transporter binding along the course of MPTP treatment revealed a transient increase in  $^{18}\text{F}$ -FECNT uptake after 5 weeks of MPTP treatment in both MPTP/saline- and MPTP/MTEP-treated animals. To our knowledge, this represents the first evidence of increased dopamine transporter binding in the striatum of animals intoxicated with MPTP. However, dopamine transporter upregulation has previously been reported in mice and baboons treated with acute low doses of organochlorine insecticides, or manganese, respectively (Kirby *et al.*, 2001; Chen *et al.*, 2006). The underlying substrate of this increased PET signal remains unclear, but it is possible that the affinity of the remaining dopamine transporter for FECNT was transiently increased, that the synthesis of dopamine transporter in the remaining dopaminergic terminals was increased, or that

dopamine transporter-containing dopaminergic axons and terminals showed an excessive degree of sprouting early in the course of degeneration of the nigrostriatal system (Song and Haber, 2000; Kirby *et al.*, 2001; Gillette and Bloomquist, 2003; Chen *et al.*, 2006). It is noteworthy that binding of D2-like dopamine receptors also shows a biphasic regulation during the course of MPTP intoxication in monkeys (Bezard *et al.*, 2001), although in contrast to dopamine transporter, D2-like receptor binding shows an initial decrease followed by an upregulation (Bezard *et al.*, 2001). Together, these findings demonstrate the complex and dynamic regulatory processes of the nigrostriatal dopaminergic system during MPTP-induced toxicity. It remains to be established whether such changes represent pathological or compensatory mechanisms in response to MPTP administration.

## Metabotropic glutamate receptor 5 antagonist effects on 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced degeneration of noradrenergic neurons

In addition to its protective effects against midbrain dopaminergic cell loss, MTEP significantly reduced the extent of norepinephrine cell loss in the locus coeruleus and the adjoining A5 and A7 noradrenergic cell groups in all chronically MPTP-treated monkeys examined in our study. Although loss of locus coeruleus neurons is a well recognized pathological feature of human Parkinson's disease (Chan-Palay, 1991; German *et al.*, 1992; Patt and Gerhard, 1993; Del Tredici *et al.*, 2002; Braak *et al.*, 2003; Zarow *et al.*, 2003), the development of such pathology in animal models of parkinsonism has been controversial (Burns *et al.*, 1983; Langston *et al.*, 1984; Mitchell *et al.*, 1985; Forno *et al.*, 1986, 1993; Miyoshi *et al.*, 1988; Gibb *et al.*, 1989; Herrero *et al.*, 1993; Rose *et al.*, 1993). In light of our findings and those reported in mice implanted with MPTP-releasing mini pumps (Fornai *et al.*, 2005), it appears that the regimen of MPTP administration used in these studies may account for the differences in norepinephrine cell loss, with chronic administration regimes favouring the loss of locus coeruleus neurons. In MPTP/vehicle-treated monkeys, our findings revealed a 40% decrease in the number of tyrosine hydroxylase-positive neurons in the locus coeruleus/A5/A7 noradrenergic cell groups. However, only about 15% loss was found in animals that received MTEP treatment, suggesting that MTEP protected the norepinephrine cells from MPTP-induced damage. At present, the cellular mechanisms of these protective effects remain unclear, but it is tempting to speculate that they may be mediated in part by a reduction of intracytoplasmic calcium levels, which are normally increased by mGluR5-mediated activation of IP3 receptors (Pin and Duvoisin, 1995; Sala *et al.*, 2005). There is, indeed, evidence that locus coeruleus neurons strongly express functional mGluR5, which are known to elicit excitatory effects and to potentiate N-methyl-D-aspartic acid-mediated excitation in these cells (Singewald *et al.*, 1994, 1995, 1996; Page *et al.*, 2005; Rasmussen *et al.*, 2005; Noriega *et al.*, 2007). As in the ventral

midbrain, the potential downregulation of glutamatergic afferents, and the possible attenuation of inflammatory processes mediated by glial mGluR5 activation are other mechanisms that could explain the protective effects of MTEP in locus coeruleus/A5/A7 noradrenergic cell groups (see above).

Norepinephrine neurons are known to be involved in many autonomic and cognitive functions, and it is thought that dysfunction in cortical noradrenergic transmission contributes to depression (Remy *et al.*, 2005; Arakawa *et al.*, 2008; Samuels and Szabadi 2008; Lockrow *et al.*, 2011), a common non-motor symptom in Parkinson's disease (Picillo *et al.*, 2009; Brooks and Pavese, 2010; Chaudhuri and Odin, 2010). The discovery of neuroprotective approaches that could attenuate norepinephrine cell loss in Parkinson's disease would be highly interesting. The need for such preventive therapy is further strengthened by recent studies suggesting that norepinephrine brainstem neurons may die earlier than dopaminergic neurons in Parkinson's disease, and that the loss of norepinephrine cells may even contribute to the later demise of substantia nigra pars compacta neurons (Fornai *et al.*, 1997; Archer *et al.*, 2006; Rommelfanger *et al.*, 2007a, b).

## Conclusion

The metabotropic glutamate receptors have become of great interest for the development of pharmacotherapeutic agents in various brain diseases. Their allosteric modulatory sites combined with their selective pharmacology and specific enrichment in particular brain areas provide them the characteristics that overcome the limitations faced by ionotropic glutamate receptor antagonists when used as brain therapeutics (Conn *et al.*, 2005; Marino and Conn, 2006; Johnson *et al.*, 2009; Niswender and Conn, 2010). In light of recent rodent and monkey behavioural data, mGluR5 has been recognized as a highly relevant antiparkinsonian and antidyskinetic target (Breyse *et al.*, 2002; Johnston *et al.*, 2010; Morin *et al.*, 2010; Rylander *et al.*, 2010). These positive results are further amplified by the finding of neuroprotective effects in monkeys, as documented in our study. However, in light of recent failures in translating positive neuroprotective results gathered from animal model studies to successful clinical trials in patients with Parkinson's disease (Olanow, 2009), our data must be interpreted cautiously. Although various issues were raised to explain these failures (Olanow, 2009), we believe that the translation of our findings to the human trial setting must await the development of biomarkers that will allow the pretreatment of potential candidates for the future development of Parkinson's disease before the appearance of motor symptoms and significant degeneration of dopaminergic and noradrenergic neurons. While the currently available mGluR5 antagonists, MTEP and MPEP, may not be ideal clinical treatment candidates because of their rapid brain clearance (Anderson *et al.*, 2003; Lea and Faden, 2006; Johnston *et al.*, 2010; Niswender and Conn, 2010), the future development of new negative allosteric modulators of mGluR5 with a better pharmacokinetic profile may pave the way for future human trials (Homayoun *et al.*, 2004; Berry-Kravis *et al.*, 2009; Bird and Lawrence, 2009; Niswender and Conn, 2010; Rodriguez *et al.*, 2010; Stefani and Moghaddam 2010).

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