

Imaging of programmed cell death in arrhythmogenic right ventricle cardiomyopathy/dysplasia

Maria E. Campian · Hanno L. Tan ·
Astrid F. van Moerkerken · Raymond Tukkie ·
Berthe L. F. van Eck-Smit · Hein J. Verberne

Received: 25 October 2010 / Accepted: 6 April 2011 / Published online: 7 May 2011
© The Author(s) 2011. This article is published with open access at Springerlink.com

Abstract

Background Arrhythmogenic right ventricular cardiomyopathy/dysplasia (ARVC/D) is a myocardial disease that predominantly affects the right ventricle (RV). Its hallmark feature is fibrofatty replacement of the RV myocardium. Apoptosis in ARVC/D has been proposed as an important process that mediates the slow, ongoing loss of heart muscle cells which is followed by ventricular dysfunction. We aimed to establish whether cardiac apoptosis can be assessed noninvasively in patients with ARVC/D.

Methods Six patients fulfilling the ARVC/D criteria were studied. Regional myocardial apoptosis was assessed with ^{99m}Tc -annexin V scintigraphy.

Results Overall, the RV wall showed a higher ^{99m}Tc -annexin V signal than the left ventricular wall ($p=0.049$) and the interventricular septum ($p=0.026$). However, significantly increased uptake of ^{99m}Tc -annexin V in the RV was present in only three of the six ARVC/D patients ($p=0.001$,

compared to ^{99m}Tc -annexin V uptake in the RV wall of the other three patients).

Conclusion Our results are suggestive of a chamber-specific apoptotic process. Although the role of apoptosis in ARVC/D is unsolved, the ability to assess apoptosis noninvasively may aid in the diagnostic course. In addition, the ability to detect apoptosis in vivo with ^{99m}Tc -annexin V scintigraphy might allow individual monitoring of disease progression and response to diverse treatments aimed at counteracting ARVC/D progression.

Keywords Arrhythmogenic right ventricle cardiomyopathy/dysplasia · Scintigraphy · Apoptosis · ^{99m}Tc -annexin V scintigraphy

Abbreviations

^{99m}Tc	technetium 99m
ARVC/D	arrhythmogenic right ventricular cardiomyopathy/dysplasia
IVS	inter-ventricular septum
LV	left ventricle
PS	phosphatidylserine
ROI	region of interest
RV	right ventricle
VT	ventricular tachycardia

M. E. Campian · H. L. Tan
Heart Failure Research Center, Academic Medical Center,
University of Amsterdam,
1100 DE Amsterdam, The Netherlands

H. L. Tan
Department of Cardiology, Academic Medical Center,
University of Amsterdam,
1100 DE Amsterdam, The Netherlands

A. F. van Moerkerken · B. L. F. van Eck-Smit ·
H. J. Verberne (✉)
Department of Nuclear Medicine, Academic Medical Center,
University of Amsterdam,
PO box 22700, 1100 DE Amsterdam, The Netherlands
e-mail: h.j.verberne@amc.uva.nl

R. Tukkie
Department of Cardiology, Kennemer Gasthuis,
Haarlem, The Netherlands

Introduction

Arrhythmogenic right ventricular cardiomyopathy/dysplasia (ARVC/D) is a disease that predominantly affects the right ventricle (RV), although biventricular involvement may occur in advanced disease [1]. ARVC/D is characterized by structural derangements that may cause a broad range of

signs and symptoms. Yet, disease expression is highly variable and incomplete in most patients, confounding both the diagnostic process and clinical management, particularly during early disease stages [2].

The histopathological hallmark of ARVC/D is fibrofatty replacement of the RV myocardium. Apoptosis has been proposed as an important mechanism that mediates the slow, ongoing loss of heart muscle cells which is followed by ventricular dysfunction [3]. How fibrofatty replacement and apoptosis are related in ARVC/D is a matter of speculation. The possibility to detect apoptosis *in vivo* in ARVC/D may lead to a better understanding of the pathophysiological mechanism underlying disease progression [4]. *In vivo* imaging of cardiac apoptosis with the use of ^{99m}Tc -annexin V has been proven feasible, as ^{99m}Tc -annexin V binds to exposed phosphatidylserine (PS) on the outer surface of apoptotic cells [5]. Accordingly, ^{99m}Tc -annexin V has been effectively used to noninvasively visualize regions of apoptosis in patients with various pathologies [6–9], as well as in experimental models [10, 11]. We aimed to establish whether cardiac apoptosis can be assessed noninvasively in patients with ARVC/D.

Methods

Patients

The institutional review board approved the study protocol and informed consent was obtained from all study subjects. Six ARVC/D patients were examined. The patients, who fulfilled the ARVC/D Task Force criteria [12], were randomly taken from the cohort of ARVC/D patients at our institution. Patients were evaluated when they were in a clinically stable condition (no ventricular tachyarrhythmias or heart failure symptoms during the 2 months prior to inclusion). In all patients, molecular genetic analysis was performed and focused on known mutations related to ARVC/D; these included plakophilin-2 (PKP2), desmoplakin (DSP), desmoglein-2 (DSG2), desmocollin-2 (DSC2), plakoglobin (JUP), and transmembrane protein 43 (TMEM43) [13, 14]. No patient had a history of coronary artery disease, diabetes or hypertension.

Scintigraphy

Patients were intravenously injected with 600 MBq of ^{99m}Tc HYNIC-rh-annexin V (^{99m}Tc -annexin V), and 4 h after administration, single photon emission computed tomography (SPECT) scans were acquired using a dual-headed gamma camera equipped with a 3/8" NaI(Tl) crystal and combined with a low-dose CT scanner (Infinia; General Electric Medical Systems, Haifa, Israel). SPECT scans were

acquired with low-energy, high-resolution collimators, a 15% energy window on the 140 keV photopeak, according to a step-and-shoot protocol with a total of 90 frames and 30 s per frame in a 128×128 matrix and a zoom of 1.28. SPECT images were iteratively reconstructed (OSEM) and corrected for attenuation using the low-dose CT scans from the Infinia scanner (no intravenous contrast material).

Analysis of scintigraphic data

To define the anatomical borders of the myocardium within the thorax, anatomical tomographic images are essential and the low-dose CT images of the Infinia could not be used for this purpose. Therefore, tomographic anatomical images from contrast-enhanced CT or cardiac MR imaging performed prior to implantable cardioverter defibrillator (ICD) implantation and within 2 months of ^{99m}Tc -annexin V scintigraphy were reviewed for all subjects. To align the anatomical images with the SPECT data, first the matrix size of the anatomical images was adjusted to the matrix size of the SPECT data (128×128) and second the images were automatically aligned (MultiModality; HERMES Medical Solutions, Sweden). To semiquantify ^{99m}Tc -annexin V myocardial uptake, three regions of interest (ROI) including the RV wall, interventricular septum (IVS) and left ventricle (LV) free wall were drawn on three summed mid-myocardial horizontal long axis anatomical images. To correct for background activity (i.e. nonspecific uptake), a separate region was drawn on both lungs. As there were no differences between the two lung regions, these values were aggregated to one value (mean counts per pixel). The ROIs were determined on the anatomical images and were subsequently copied to the aligned SPECT images. ^{99m}Tc -annexin V uptake in each separate ROI was calculated as the ratio of the mean counts per pixel in the specific myocardial region to the mean counts per pixel in the total myocardium (i.e. the sum of all three ROIs). Both the regional and the total myocardial activity were corrected for background activity by subtraction of nonspecific uptake. The attenuation-corrected SPECT data were used for analysis. The reader was blinded to the clinical information.

Follow-up

Long-term follow-up data were obtained from at least one of three sources: visit to the outpatient clinic; review of the patient's hospital records; personal communication with the patient's physician. This analysis focused on the occurrence of ventricular arrhythmias, appropriate ICD discharge, and sudden cardiac death. One patient was lost to follow-up. The mean follow-up was 27±8 months (range 18–57 months).

Table 1 Clinical characteristics of patients with ARVC/D

Patient no.	Gender	Age at scintigraphy (years)	Symptoms at diagnosis	Age at diagnosis (years)	Mutation	Medication	ARVC/D Task Force criteria										
							Family history		ECG depolarization/ conduction		ECG repolarization		Arrhythmias		RV dysfunction		
							Disease confirmed at necropsy (major)	Sudden cardiac death (minor) ^a	ARVC/D (minor)	Epsilon wave (major)	Late potential (minor)	Negative T wave (minor)	LBBB-VT (minor)	>1,000 PVC/24 h (minor)		Severe (major)	Mild (minor)
1	M	24	Syncope	21	PKP2: C796R ^b	Sotalol	-	+	+	-	+	+	+	+	-	-	+
2	F	55	VT	49	No	-	-	+	+	-	NA	+	+	+	-	-	+
3	F	48	VT	38	No	Sotalol	-	-	-	-	NA	+	+	+	-	-	+
4	M	33	VT	30	No	Sotalol	-	-	+	-	-	+	+	+	-	-	+
5	M	19	VT	16	No	Sotalol	-	-	-	-	NA	+	+	-	-	-	+
6	M	41	VT	27	DSG2: T335A ^c	-	-	-	-	-	+	+	+	+	+	+	-

LBBB left bundle branch block, NA not analysed, PVC premature ventricular complex.

^a Death before 35 years of age due to suspected ARVC/D.

^b C796R missense mutation in plakophilin-2.

^c T335A missense mutation in desmoglein-2.

Statistical analysis

Data are presented as means \pm SD. Mean values were compared for differences using the (un)paired Student's *t*-test when appropriate. For multiple comparisons, means were compared for differences using analysis of variance (ANOVA) with a post-hoc Bonferroni correction (SPSS for Windows 16.0.2.1; SPSS, Chicago, IL). A *p* value <0.05 was considered to indicate statistical significance.

Results

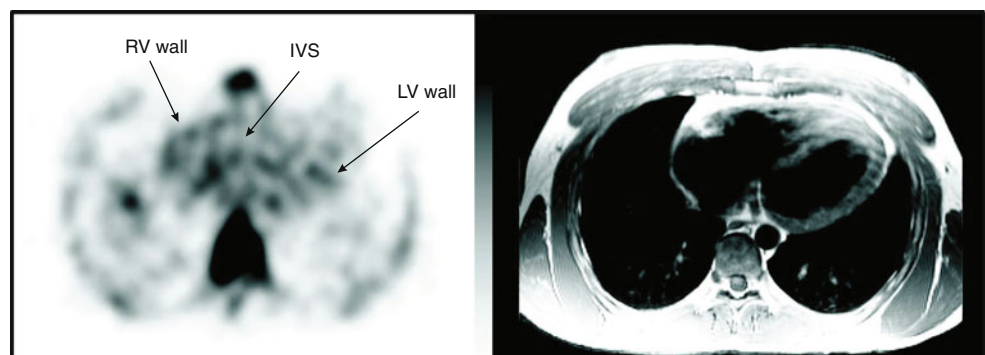
Clinical spectrum

Table 1 summarizes the demographic/clinical data. All patients fulfilled the ARVC/D Task Force criteria [12]. Their mean age at clinical presentation was 36.7 ± 13.9 years (range 19–55 years) and 33% (two) were women. In five patients, ventricular tachycardia (VT) with left bundle branch block morphology was the first expression of ARVC/D. One patient presented with syncope. Two patients had a positive family history of premature sudden cardiac death. All patients had normal LV function by echocardiography and all patients had an ICD. All patients had a history of haemodynamically unstable VT. Four patients were on antiarrhythmic agents. One patient had the C796R mutation in PKP2, while one had the T335A mutation in DSG2. In the remaining patients, no DNA mutations were found. One patient (patient 6) had severe segmental dilatation of the RV on echocardiography (major ARVC/D Task Force criterion [12]). The other five patients had regional RV hypokinesia (patients 2, 3 and 5), mild segmental dilatation of the RV (patient 4) and mild global RV dilatation with normal LV function (patient 1) on echocardiography (minor ARVC/D Task Force criteria [12]).

Myocardial ^{99m}Tc -annexin V uptake

Figure 1 shows a typical example of a patient who exhibited increased ^{99m}Tc -annexin V uptake in the RV wall

Fig. 1 Coregistered transaxial images of patient 3 (left cardiac MR image, right ^{99m}Tc -annexin SPECT scintigraphy image). There is increased ^{99m}Tc -annexin V uptake in the RV wall (IVS interventricular septum, LV left ventricular free wall)



(patient 2). Overall, the RV wall showed a higher ^{99m}Tc -annexin V uptake (1.328 ± 0.437) than the LV wall (0.936 ± 0.175 , $p=0.049$) or the IVS (0.902 ± 0.222 , $p=0.026$). There was no difference in ^{99m}Tc -annexin V uptake between the LV wall and the IVS ($p=0.986$). However, the overall higher uptake of ^{99m}Tc -annexin V in the RV wall could be explained by the fact that 50% of patients (patients 3, 5 and 6) showed increased ^{99m}Tc -annexin V uptake in the RV compared to the other three patients (patients 1, 2 and 4; 1.788 ± 0.133 vs 0.983 ± 0.034 respectively, $p=0.001$; Fig. 2).

Within 2 months of ^{99m}Tc -annexin V scintigraphy, cardiac MR images were available in three patients (patients 2, 3 and 4). Only patient 3 showed increased uptake of ^{99m}Tc -annexin V. The increased uptake of ^{99m}Tc -annexin V was located in the lateral wall of the RV, while the MR images showed an overall dilated RV with regional dyskinesia of the apex. It is therefore not possible to draw any conclusions as to a potential correlation between cardiac MRI RV abnormalities and the location of increased uptake of ^{99m}Tc -annexin V.

Follow-up

The extent of ^{99m}Tc -annexin V uptake in the RV wall did not distinguish patients with arrhythmias within 2 years after ^{99m}Tc -annexin V scintigraphy from those without, nor did it distinguish patients in whom a gene mutation was found from those in whom it was not (Table 2).

Discussion

Apoptosis is a significant pathophysiological feature of ARVC/D and is a consistent post-mortem finding in both the RV and LV [1, 15]. In this study, ^{99m}Tc -annexin V scintigraphy was performed with the purpose of establishing whether apoptosis can be visualized in vivo in patients with ARVC/D. Our results demonstrated increased ^{99m}Tc -annexin V uptake in the RV free wall of three ARVC/D patients, suggestive of RV-specific apoptotic activity in

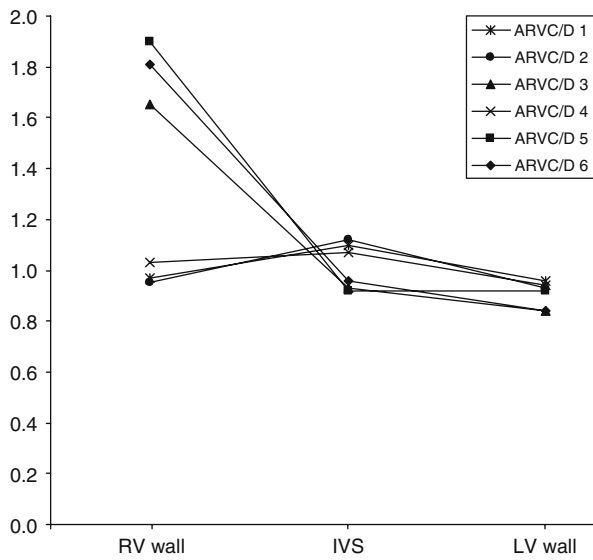


Fig. 2 ^{99m}Tc-annexin uptake in the RV wall, IVS and LV wall calculated as the ratio of the uptake (mean counts per pixel) in the vvp particular myocardial region to the uptake in the total myocardium (i.e. the sum of all three ROIs)

these patients. The variation in myocardial uptake of ^{99m}Tc-annexin V between patients is not surprising and might partly be explained by the random distribution and the episodic nature of the apoptotic process [16]. Furthermore, patients differed with respect to the time since diagnosis and severity of morphological abnormalities. These variations were probably reflected by differences in myocardial uptake of ^{99m}Tc-annexin V.

All our ARVC/D patients had a history of documented VT episodes. Mallat et al. speculated that apoptosis in ARVC/D might result from repetitive ventricular tachyarrhythmia episodes [15]. Furthermore, apoptotic myocytes are frequently found in regions of the myocardium which are not subjected to invasion by adipocytes and fibrosis, suggesting that the loss of myocytes through apoptosis occurs as a primary process before adipocytes and fibrous tissues fill the vacant cellular space. Also, Valente et al. have reported that apoptosis is present in endomyocardial biopsy samples of patients with ARVC/D, especially in the early symptomatic phase of the disease [17].

The exposure of PS on the cell surface is a general marker of apoptotic cells. Externalization of nonapoptotic PS is induced by several activation stimuli, including engagement of immunoreceptors. Externalized PS is observed in apoptotic, injured, infected, senescent and necrotic cells, and becomes a target for recognition by phagocytes [18–20]. Thus, in addition to acting as a marker of apoptosis, annexin V may be a marker of inflammation and cell stress. Accordingly, the myocardial uptake of ^{99m}Tc-annexin V is most likely not only a marker of

Table 2 Follow-up in ARVC/D patients on the occurrence of ventricular arrhythmias, appropriate ICD discharge and sudden cardiac death

Patient no.	^{99m} Tc-annexin V uptake	Year of diagnosis	VT 2005		2006		2007		2008		2009		2010	
			ICD shock	Sudden cardiac death	VT	ICD shock	Sudden cardiac death	VT	ICD shock	Sudden cardiac death	VT	ICD shock	Sudden cardiac death	
1	Normal	2001	-	-	-	-	-	-	-	-	-	-	-	-
2	Increased	1998	-	-	-	-	-	-	-	-	-	-	-	-
3	Increased	1994	-	-	-	-	-	-	-	-	-	-	-	-
4	Normal	2001	Lost to follow-up	-	-	-	-	-	-	-	-	-	-	-
5	Increased	2001	+	+	+	+	-	-	-	-	-	-	-	-
6	Normal	1990	-	-	-	-	+	-	-	-	-	-	-	-

apoptosis, but may also partly reflect local inflammation. Patchy inflammatory infiltrates in RV are consistently reported in ARVC/D, both in in vitro and in in vivo examinations [3, 21, 22].

Patchy cell death combined with inflammatory infiltration is a common histological finding in ARVC/D [23]. The inflammation might be a reaction to proinflammatory cytokines induced by cell death and/or apoptosis or caused by an infectious myocarditis (e.g. viral infection) [21, 24]. Although it is most likely that these factors are, at least to some extent, interrelated, it is not known whether there is a causal relationship between inflammation and cell death in ARVC/D. However, it remains unclear whether myocarditis in ARVC/D is disease-initiating (a primary event) or a reaction to processes initiated by ARVC/D.

Study limitations and clinical usefulness

The first limitation of our study is the small number of patients. The observation of ^{99m}Tc -annexin V myocardial uptake in three out of six patients might be explained by the random distribution and the episodic nature of the apoptotic process. Second, no cardiac biopsies were obtained. Therefore, a validation of the ^{99m}Tc -annexin V myocardial uptake with histology was not possible.

Recently modification of the highly specific ARVC/D Task Force criteria (published in 1994) has been proposed [25]. The revision incorporates new knowledge and technology to improve especially the sensitivity of the Task Force criteria without changing the high specificity. However, at the time of patient inclusion the 1994 criteria were used [12]. As expected, because of the relatively unchanged specificity, reevaluation of the patients included in our study according to the new criteria did not change the clinical diagnosis in any patient.

Conclusion

Apoptosis may be detected noninvasively in ARVC/D, and this may lead to a better understanding of the role of apoptosis in the pathophysiology of ARVC/D. Whether it will allow monitoring of the disease course or the response to various treatments aimed at counteracting disease progression remains to be studied.

Acknowledgments Dr. H.L. Tan was supported by the Royal Netherlands Academy of Arts and Sciences (KNAW) and the Netherlands Organization for Scientific Research (NWO, ZonMW-Vici 918.86.616).

Conflicts of interest None.

Open Access This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

References

1. Asimaki A, Tandri H, Huang H, Halushka MK, Gautam S, Basso C, et al. A new diagnostic test for arrhythmogenic right ventricular cardiomyopathy. *N Engl J Med*. 2009;360:1075–84.
2. Marcus FI, Zareba W, Calkins H, Towbin JA, Basso C, Bluemke DA, et al. Arrhythmogenic right ventricular cardiomyopathy/dysplasia clinical presentation and diagnostic evaluation: results from the North American Multidisciplinary Study. *Heart Rhythm*. 2009;6:984–92.
3. Basso C, Thiene G, Corrado D, Angelini A, Nava A, Valente M. Arrhythmogenic right ventricular cardiomyopathy. Dysplasia, dystrophy, or myocarditis? *Circulation*. 1996;94:983–91.
4. van Heerde WL, Robert-Offerman S, Dumont E, Hofstra L, Doevendans PA, Smits JF, et al. Markers of apoptosis in cardiovascular tissues: focus on Annexin V. *Cardiovasc Res*. 2000;45:549–59.
5. Blankenberg FG, Katsikis PD, Tait JF, Davis RE, Naumovski L, Ohtsuki K, et al. In vivo detection and imaging of phosphatidylserine expression during programmed cell death. *Proc Natl Acad Sci U S A*. 1998;95:6349–54.
6. Hofstra L, Liem IH, Dumont EA, Boersma HH, van Heerde WL, Doevendans PA, et al. Visualisation of cell death in vivo in patients with acute myocardial infarction. *Lancet*. 2000;356:209–12.
7. Hofstra L, Dumont EA, Thimister PW, Heidendal GA, DeBruine AP, Elenbaas TW, et al. In vivo detection of apoptosis in an intracardiac tumor. *JAMA*. 2001;285:1841–2.
8. Kietselaer BL, Reutelingsperger CP, Boersma HH, Heidendal GA, Liem IH, Crijns HJ, et al. Noninvasive detection of programmed cell loss with ^{99m}Tc -labeled annexin A5 in heart failure. *J Nucl Med*. 2007;48:562–7.
9. Narula J, Strauss HW. Invited commentary: P.S.* I love you: implications of phosphatidyl serine (PS) reversal in acute ischemic syndromes. *J Nucl Med*. 2003;44:397–9.
10. Tokita N, Hasegawa S, Maruyama K, Izumi T, Blankenberg FG, Tait JF, et al. ^{99m}Tc -Hynic-annexin V imaging to evaluate inflammation and apoptosis in rats with autoimmune myocarditis. *Eur J Nucl Med Mol Imaging*. 2003;30:232–8.
11. Campian ME, Verberne HJ, Hardziyenka M, de Bruin K, Selwaness M, van den Hoff MJ, et al. Serial noninvasive assessment of apoptosis during right ventricular disease progression in rats. *J Nucl Med*. 2009;50:1371–7.
12. McKenna WJ, Thiene G, Nava A, Fontaliran F, Blomstrom-Lundqvist C, Fontaine G, et al. Diagnosis of arrhythmogenic right ventricular dysplasia/cardiomyopathy. Task Force of the Working Group Myocardial and Pericardial Disease of the European Society of Cardiology and of the Scientific Council on Cardiomyopathies of the International Society and Federation of Cardiology. *Br Heart J*. 1994;71:215–8.
13. Awad MM, Calkins H, Judge DP. Mechanisms of disease: molecular genetics of arrhythmogenic right ventricular dysplasia/cardiomyopathy. *Nat Clin Pract Cardiovasc Med*. 2008;5:258–67.
14. Merner ND, Hodgkinson KA, Haywood AF, Connors S, French VM, Drenckhahn JD, et al. Arrhythmogenic right ventricular cardiomyopathy type 5 is a fully penetrant, lethal arrhythmic disorder caused by a missense mutation in the TMEM43 gene. *Am J Hum Genet*. 2008;82:809–21.

15. Mallat Z, Tedgui A, Fontaliran F, Frank R, Durigon M, Fontaine G. Evidence of apoptosis in arrhythmogenic right ventricular dysplasia. *N Engl J Med*. 1996;335:1190–6.
16. James TN. Normal and abnormal consequences of apoptosis in the human heart. *Annu Rev Physiol*. 1998;60:309–25.
17. Valente M, Calabrese F, Thiene G, Angelini A, Basso C, Nava A, et al. In vivo evidence of apoptosis in arrhythmogenic right ventricular cardiomyopathy. *Am J Pathol*. 1998;152:479–84.
18. Laufer EM, Reutelingsperger CP, Narula J, Hofstra L. Annexin A5: an imaging biomarker of cardiovascular risk. *Basic Res Cardiol*. 2008;103:95–104.
19. Hirt UA, Leist M. Rapid, noninflammatory and PS-dependent phagocytic clearance of necrotic cells. *Cell Death Differ*. 2003;10:1156–64.
20. Brouckaert G, Kalai M, Krysko DV, Saelens X, Vercammen D, Ndlovu M, et al. Phagocytosis of necrotic cells by macrophages is phosphatidylserine dependent and does not induce inflammatory cytokine production. *Mol Biol Cell*. 2004;15:1089–100.
21. Campian ME, Verberne HJ, Hardziyenka M, de Groot EA, van Moerkerken AF, van Eck-Smit BL, et al. Assessment of inflammation in patients with arrhythmogenic right ventricular cardiomyopathy/dysplasia. *Eur J Nucl Med Mol Imaging*. 2010;37:2079–85.
22. Tabib A, Loire R, Chalabreysse L, Meyronnet D, Miras A, Malicier D, et al. Circumstances of death and gross and microscopic observations in a series of 200 cases of sudden death associated with arrhythmogenic right ventricular cardiomyopathy and/or dysplasia. *Circulation*. 2003;108:3000–5.
23. Basso C, Ronco F, Marcus F, Abudurehman A, Rizzo S, Frigo AC, et al. Quantitative assessment of endomyocardial biopsy in arrhythmogenic right ventricular cardiomyopathy/dysplasia: an in vitro validation of diagnostic criteria. *Eur Heart J*. 2008;29:2760–71.
24. Calabrese F, Basso C, Carturan E, Valente M, Thiene G. Arrhythmogenic right ventricular cardiomyopathy/dysplasia: is there a role for viruses? *Cardiovasc Pathol*. 2006;15:11–7.
25. Marcus FI, McKenna WJ, Sherrill D, Basso C, Bauce B, Bluemke DA, et al. Diagnosis of arrhythmogenic right ventricular cardiomyopathy/dysplasia: proposed modification of the task force criteria. *Circulation*. 2010;121:1533–41.