

ORIGINAL ARTICLE

Distribution of *Malassezia* Species on the Scalp in Korean Seborrheic Dermatitis Patients

Yang Won Lee, M.D., Hee Jin Byun, M.D.¹, Beom Joon Kim, M.D.², Dong Ha Kim, M.D.², Yun Young Lim, Ph.D.², Jin Woong Lee, M.D.², Myeung Nam Kim, M.D.², Donghak Kim, Ph.D.³, Young-Jin Chun, Ph.D.⁴, Seog Kyun Mun, M.D.⁵, Chan Woong Kim, M.D.⁶, Sung Eun Kim, M.D.⁶, Jae Sung Hwang, Ph.D.⁷

Department of Dermatology, Konkuk University College of Medicine, Seoul, ¹Department of Dermatology, Seoul National University College of Medicine, Seoul, ²Department of Dermatology, Chung-Ang University College of Medicine, Seoul, ³Department of Biological Sciences, Konkuk University, Seoul, ⁴College of Pharmacy, Chung-Ang University, Seoul, ⁵Department of Otorhinolaryngology-Head and Neck Surgery, Chung-Ang University College of Medicine, Seoul, ⁶Department of Emergency Medicine, Chung-Ang University College of Medicine, Seoul, ⁷Department of Genetic Engineering and Skin Biotechnology Center, Kyung Hee University, Seoul, Korea

Background: *Malassezia* species play an important role in the pathogenesis of seborrheic dermatitis. In particular, *M. restricta* and *M. globosa* are considered to be the predominant organisms in seborrheic dermatitis of Western countries. However, species distribution of *Malassezia* in seborrheic dermatitis has not been clearly determined yet in Asia. **Objective:** To identify the distribution of *Malassezia* species on the scalp of seborrheic dermatitis patients in Korea using 26S rDNA PCR-RFLP analysis. **Methods:** A total of 40 seborrheic dermatitis patients and 100 normal healthy volunteers were included in this study. For the identification of *Malassezia* species, the scalp scales of the subjects were analyzed by 26S rDNA PCR-RFLP analysis. **Results:** The most commonly identified *Malassezia* species were *M. restricta* in the seborrheic dermatitis patients, and *M. globosa* in the normal controls. In the seborrheic dermatitis group, *M. restricta* was identified in 47.5%, *M. globosa* in 27.5%, *M. furfur* in 7.5%, and *M. sympodialis* in 2.5% of patients. In the healthy control group, *M. globosa* was identified in 32.0%, *M. restricta* in 25.0%, *M. furfur* in 8.0%, *M. obtusa* in 6.0%, *M. slooffiae* in 6.0%, and *M. sympodialis* in 4.0% of subjects. **Conclusion:** *M. restricta* is considered to be the most im-

portant *Malassezia* species in Korean seborrheic dermatitis patients. (Ann Dermatol 23(2) 156~161, 2011)

-Keywords-

26S rDNA PCR-RFLP, *Malassezia*, Seborrheic dermatitis

INTRODUCTION

Malassezia species are lipophilic fungi which are regarded as part of the normal flora of human skin, and are found in 75~98% of healthy adults. These yeasts are the cause of pityriasis versicolor and *Malassezia* folliculitis, and appear to be involved in the pathogenesis of common skin disorders such as seborrheic dermatitis, psoriasis, and atopic dermatitis¹. In 1996, Guého et al.² classified *Malassezia* into seven species, *M. furfur*, *M. obtusa*, *M. globosa*, *M. slooffiae*, *M. sympodialis*, *M. pachydermatis*, and *M. restricta*, based on morphological and biochemical analysis. However, with the development of molecular methods such as polymerase chain reaction (PCR), an additional six species have been described which are *M. yamatoensis*, *M. nana*, *M. japonica*, *M. equine*, *M. caprae* and *M. dermatis*³. Among them, *M. restricta* and *M. globosa* are considered to be the most important pathogenic organisms in the development of seborrheic dermatitis⁴. However, some reports have also linked *M. furfur*, *M. sympodialis*, *M. obtusa*, and *M. slooffiae* with seborrheic dermatitis⁵.

The aim of this study was to identify *Malassezia* species on the scalp of 40 patients with seborrheic dermatitis in

Received August 10, 2010, Revised December 17, 2010, Accepted for publication December 17, 2010

*This study is supported by a grant of the Korea Healthcare Technology R&D Project, Ministry of Health & Welfare (A080065).

Corresponding author: Beom Joon Kim, M.D., Department of Dermatology, Chung-Ang University College of Medicine, 224-1 Heukseok-dong, Dongjak-gu, Seoul 156-755, Korea. Tel: 82-2-6299-1525, Fax: 82-2-823-1049, E-mail: beomjoon@unitel.co.kr

Korea using 26S rDNA polymerase chain reaction restriction fragment length polymorphism (PCR-RFLP). We also compared the results of seborrheic dermatitis patients with those of 100 normal healthy volunteers.

MATERIALS AND METHODS

Standard strains

Six reference strains of *Malassezia* species, which are known to be involved in the pathogenesis of seborrheic dermatitis, were used to determine the 26S rDNA PCR-RFLP pattern of *Malassezia* yeasts in this study^{4,5}. The standard strains were purchased from Centralbureau Boor Schimmelcultures (CBS, Utrecht, Netherlands), Biological Resource Center and included *M. furfur* (CBS 1878), *M. restricta* (CBS 7877), *M. obtusa* (CBS 7876), *M. sympodialis* (CBS 7222), *M. slooffiae* (CBS 7956) and *M. globosa* (CBS 7966).

Subjects

A total of 140 subjects, including 40 seborrheic dermatitis (with scalp involvement) patients and 100 healthy volunteers as controls, were enrolled in this study (Table 1). Seborrheic dermatitis on the lesions of the scalp was clinically diagnosed, with seborrhea, greasy, yellowish, pityriasiform scaling related to irregular sharply delimited erythematous plaques. Some seborrheic dermatitis patients had pronounced seborrhea and inflamed erythematous folliculitis. The patients without scalp lesions or with only dandruff on the scalp lesions were excluded. We included only those patients who had not used any topical or oral treatment during the previous 2 months. No concomitant diseases were registered. The mean age was significantly higher in the seborrheic dermatitis group (42.5, range 25~54) than in the control group (35.7, range 22~66) ($p=0.001$). The proportion of males was significantly higher in the seborrheic dermatitis group (65.0%) than in the control group (60.0%) ($p=0.045$).

Sample preparation

Scalp scales were collected by swabbing in both the seborrheic dermatitis and normal control group. A sterile

cotton swab was moistened in 100 μ l of the detergent fluid containing 0.1% Triton X-100 in 0.075 mol phosphate buffer (pH 7.9), and was rubbed five times against a round area of 3 cm diameter on the scalp skin. Then, only the fiber-tipped part of the swab was cut and placed in 900 μ l of the detergent fluid. This was placed into a shaker for 30 seconds to evenly disperse the *Malassezia* yeasts in the solution. Approximately 100 μ l of sampled skin was mixed with 900 μ l of the detergent. From this, 100 μ l was taken and applied evenly on Leeming & Notman culture medium, and incubated at 34°C for 14 days.

Genomic DNA isolation

Genomic DNA isolation was performed by the glass beads method. The obtained *Malassezia* yeasts from the patients were suspended in 400 μ l lysis buffer (100 mM Tris-HCl pH 8.0, 1.0% SDS, 2.0% Triton X-100, 10 mM EDTA, 100 mM NaCl). A total of 400 μ l P/C/I (phenol: chloroform: isoamyl alcohol=25:24:1, v/v) and 400 mg glass beads (diameter 0.5 mm) were added. This mixture was vortexed for 10 minutes and centrifuged at 15,000 rpm for 10 minutes. Total DNA was precipitated from the aqueous fraction with isopropanol and centrifuged at 15,000 rpm for 15 minutes. The DNA pellet was washed in 70% ethanol and resuspended in sterile water.

26S rDNA PCR

Primers were designed to amplify the 26S rDNA region of all six species of *Malassezia*. The sequences were forward, 5'-TAACAAGGATTCCCCTAGTA-3' and reverse, 5'-ATTA-CGCCAGCATCCTAAG-3'. The conditions of the early stage of reaction were as follows: pre-denaturation at 95°C for 14 minutes, denaturation at 94°C for 45 seconds, annealing at 55°C for 45 seconds, extension of 35 cycles at 72°C for 1 minute, and final extension at 72°C for 7 minutes. Amplified DNA was confirmed by electrophoresis on a 1.5% agarose gel stained with ethidium bromide (0.5 μ g/ml) by using 1 \times TAE migrating buffer (pH 8.0, 40 mM Tri-acetate 1 mM EDTA).

Table 1. Age and sex distribution of subjects

Age groups	Seborrheic dermatitis group			Control group		
	Male (%)	Female (%)	Total (%)	Male (%)	Female (%)	Total (%)
20~29	6 (60)	4 (40)	10 (100)	22 (55)	18 (45)	40 (100)
30~39	13 (65)	7 (35)	20 (100)	23 (58)	17 (43)	40 (100)
40~	7 (70)	3 (30)	10 (100)	10 (50)	10 (50)	20 (100)
Total	26 (65)	14 (35)	40 (100)	60 (60)	40 (40)	100 (100)

RFLP analysis

The amplified 26S rDNA was purified using the Accu-Prep PCR purification kit (Bioneer, Daejeon, Korea). Two restriction enzymes, Hha I (Takara Biomedicals, Otsu, Japan) and BstF51 (SibEnzyme, Novosibirsk, Russia) were used. The restriction enzyme reaction was performed with 10 × PCR buffer, 10 U of the restriction enzyme, and 7.5 μl of the PCR products. After the reaction, the RFLP pattern was analyzed with DNA fragments in gel electrophoresis.

Statistic analysis

Chi-squared test was conducted for the data obtained from 26S rDNA PCR-RFLP analysis to statistically determine the difference in the frequency and distribution of *Malassezia*

species in the seborrheic dermatitis and control group.

RESULTS

Confirmation of six standard strains by 26S rDNA PCR-RFLP analysis

Using colony PCR to amplify the 26S rDNA of *Malassezia* standard strains, 580 bp PCR bands were confirmed in all of the six standard strains including *M. furfur*, *M. globosa*, *M. restricta*, *M. obtusa*, *M. sympodialis*, and *M. slooffiae* (Fig. 1). All the six standard strains showed different RFLP patterns using the restriction enzyme *Hha1*. With the restriction enzyme *BstF51*, *M. globosa* and *M. obtusa* showed the same RFLP pattern, whereas the other four strains showed different patterns (Fig. 2). Therefore, we confirmed that these six standard strains can be identified and differentiated by 26S rDNA PCR-RFLP analysis.

Malassezia species identified in the seborreic dermatitis group vs. the control group

Malassezia yeasts of the seborrheic dermatitis group were cultured in 34 out of 40 specimens, (identification rate 85%). In the seborrheic dermatitis group, *M. restricta* (47.5%) was most commonly recovered, and *M. globosa* (27.5%) was the second most commonly recovered species. *M. furfur* and *M. sympodialis* were identified in 7.5% and 2.5% of the seborrheic dermatitis patients, respectively. *M. obtusa* and *M. slooffiae* were not recovered in any of the seborrheic dermatitis patients.

Malassezia yeasts of the control group were cultured in 81 out of 100 specimens, and thus the identification rate was 81%. In the control group, *M. globosa* (32.0%) was most commonly recovered, and *M. restricta* (25.0%) was the second most commonly recovered species. The complete set of results for the recovered species is shown in Table 2. Only the most abundant species for each individual was counted, not accounting for the combined infection

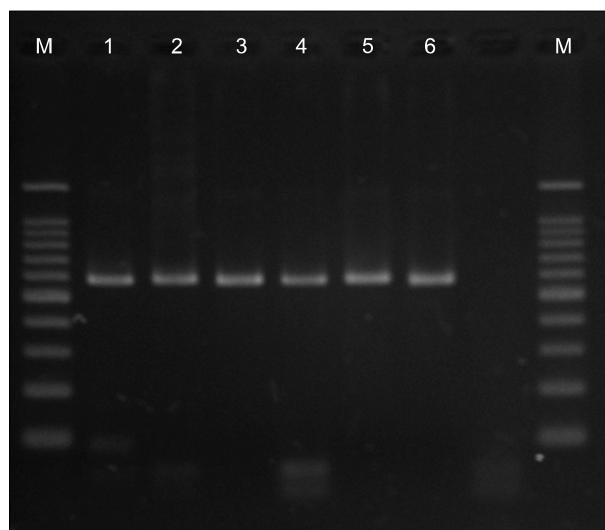


Fig. 1. 26S rDNA PCR in the six *Malassezia* standard strains. The size of PCR products were almost 580 bp. Lane M: marker, lane 1: *M. furfur*, lane 2: *M. globosa*, lane 3: *M. restricta*, lane 4: *M. obtusa*, lane 5: *M. sympodialis*, lane 6: *M. slooffiae*.

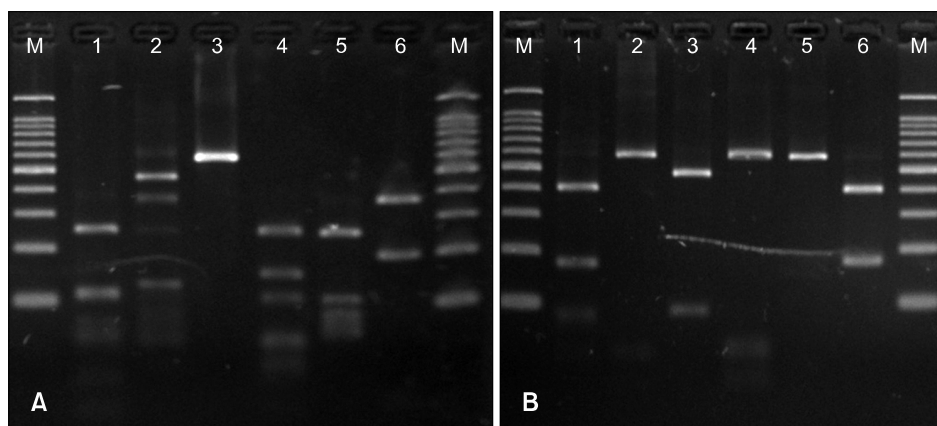


Fig. 2. 26S rDNA PCR-RFLP pattern in the six *Malassezia* standard strains. 26S rDNA PCR products were digested with two restriction enzymes. (A) *Hha1*; all the six standard strains showed different RFLP patterns. (B) *BstF51*; *M. globosa* and *M. obtusa* showed the same RFLP patterns. The other four strains all showed different patterns. Lane M: marker, lane 1: *M. furfur*, lane 2: *M. globosa*, lane 3: *M. restricta*, lane 4: *M. obtusa*, lane 5: *M. sympodialis*, lane 6: *M. slooffiae*.

Table 2. *Malassezia* species identified in the seborrheic dermatitis and control group

<i>Malassezia</i> species*	Seborrheic dermatitis No. (%)	Healthy control No. (%)
<i>M. restricta</i>	19 (47.5)	25 (25.0)
<i>M. globosa</i>	11 (27.5)	32 (32.0)
<i>M. furfur</i>	3 (7.5)	8 (8.0)
<i>M. obtusa</i>	0 (0)	6 (6.0)
<i>M. sympodialis</i>	1 (2.5)	4 (4.0)
<i>M. slooffiae</i>	0 (0)	6 (6.0)
No growth	6 (15.0)	19 (19.0)
Total	40 (100)	100 (100)

*Only the most abundant species in each individual was counted, not accounting for the combined infection of two or more species.

of two or more species.

In comparison to the seborrheic dermatitis group with the control group, the prevalence of *M. restricta* was significantly higher in the seborrheic dermatitis group (47.5%) than in the control group (25.0%) ($p < 0.05$). The other five species, *M. furfur*, *M. globosa*, *M. obtusa*, *M. sympodialis*, and *M. slooffiae*, did not show the significant differences in prevalence between the seborrheic dermatitis group and the control group.

The prevalence of co-colonization with two or more *Malassezia* species

The prevalence of co-colonization of two or more *Malassezia* species was 15.0% in the seborrheic dermatitis group, and slightly higher in the control group, at 21.0%. This difference was not statistically significant ($p = 0.339$). The recovered species are shown in Table 3. Co-colonization of *M. restricta* with *M. globosa* was the most common in both groups.

Differences in the distribution of *Malassezia* species according to age and sex

The identified *Malassezia* species did not show significant differences among the different age groups (20~29, 30~39, ≥ 40) both in the seborrheic dermatitis and control group. According to sex, *Malassezia* species neither showed significant differences in either the seborrheic dermatitis or control group.

DISCUSSION

Recently, various molecular biological methods have been applied for the identification of *Malassezia* species, such as nested PCR, real-time PCR, RFLP, and sequencing analysis⁶⁻¹³. 26S rDNA PCR-RFLP is one such method that uses 26S rDNA containing highly conserved sequences

Table 3. Co-colonization of *Malassezia* species in the seborrheic dermatitis and control group

<i>Malassezia</i> species	Seborrheic dermatitis No. (%)	Healthy control No. (%)
<i>M. restricta</i> + <i>M. globosa</i>	2 (5.0)	8 (8.0)
<i>M. restricta</i> + <i>M. obtusa</i>	2 (5.0)	6 (6.0)
<i>M. restricta</i> + <i>M. slooffiae</i>	1 (3.0)	0 (0)
<i>M. restricta</i> + <i>M. sympodialis</i>	1 (3.0)	4 (4.0)
<i>M. globosa</i> + <i>M. furfur</i>	0 (0)	3 (3.0)
Total	6 (15.0)	21 (21.0)

yet enough sequence variation for specific interspecies identification. This method requires only PCR and one or two restriction enzymes. Therefore, it is less demanding than most the other molecular biological approaches¹³. There are five reports of the successful use of 26S rDNA PCR-RFLP for the identification of *Malassezia* species¹³⁻¹⁷. In this study, we also confirmed the accuracy and effectiveness of this method.

Seborrheic dermatitis is a chronic skin disease affecting the seborrheic areas of the body including the scalp, face, and upper trunk. It manifests as erythema, scales, and follicular papules. In seborrheic dermatitis patients, *Malassezia* yeasts show a significantly higher count and density on the lesional skin than on the normal skin of the same patient and on the skin of healthy controls⁴. The yeast density is considered the basic difference between normal skin and skin with seborrheic dermatitis. However, this does not correlate positively with the severity of the disease^{4,18}. In previous reports, the species distribution of *Malassezia* in seborrheic dermatitis showed geographical peculiarities. In Japan and USA, *M. globosa* was most frequently isolated in seborrheic dermatitis, whereas *M. restricta* was the second most frequently isolated. In Eastern Europe, *M. obtusa* was the most frequently recovered species⁴. As for the species distribution of *Malassezia* in Korean seborrheic dermatitis patients, there has been only one report by Lim et al.¹⁹. In this report, *M. restricta* was most commonly identified, in eight of 10 patients, and *M. globosa* was recovered in only one patient. In our study, 34 Korean seborrheic dermatitis patients were investigated, and *M. restricta* was also most commonly identified (55.9%), showing a significantly higher recovery rate compared to the normal controls (30.9%). However, Jang et al.¹³ recently reported *M. restricta* was identified in 56.8% of healthy Korean scalp skin samples. Their results also showed a higher prevalence of *M. restricta* in older age groups, in contrast to a higher prevalence of *M. globosa* in younger age groups. Because our seborrheic dermatitis patients had a

higher age than the healthy controls, we cannot completely exclude that the age difference between the two groups may act as a confounding factor.

Age may affect the prevalence of *Malassezia* infection, because *Malassezia* species are lipophilic fungi, and therefore, the growth of *Malassezia* greatly depends on the sebum content which differs according to age. The recovery rate of *Malassezia* is known to be highest in the twenties when sebaceous glands show greatest activity²⁰. However, there have not been many reports regarding the age-dependent difference of *Malassezia* species in seborrheic dermatitis. Jang et al.¹³ reported the results of healthy individuals, not of seborrheic dermatitis patients, which showed predominance of *M. globosa* in the younger age groups and *M. restricta* in older age groups. However, in another study with 770 healthy Japanese, *M. restricta* predominated in younger ages, whereas *M. globosa* predominated in older ages, which is inverse to the Korean report^{13,20}. We can confirm that the distribution of *Malassezia* species is greatly influenced by geographical locations or ethnic groups. In our study, identified *Malassezia* species did not differ significantly according to age groups. This could be due to the relatively small number of the subjects and the narrow range of the participants' age in our study.

As for the gender-dependent difference of *Malassezia* infection in seborrheic dermatitis, there have been no reports yet. In our study, there was no gender-dependent difference in the distribution of *Malassezia* species in the seborrheic dermatitis or control group. As for healthy skin, there have been a few reports regarding the gender-dependent difference of *Malassezia* distribution^{13,20}. In a Japanese study, *M. globosa* to *M. restricta* ratio changed noticeably after 16 years in males, and 23 years in females²⁰.

In our study, 15.0% of seborrheic dermatitis patients showed co-colonization of two or more *Malassezia* species. This was similar to the previous report by Gaitanis et al.²¹, which showed that 18% of Greek seborrheic dermatitis patients were co-infected with *M. globosa*, the most commonly identified species in Greeks, together with other *Malassezia* species²¹. However, as a higher prevalence of co-colonization was found in our healthy volunteers (21.0%), co-colonization of two or more *Malassezia* species may not be important in the development of seborrheic dermatitis.

In our study, only the six reference strains of *Malassezia* species, which have been reported to be related to seborrheic dermatitis, were used, and the newly reported species, including *M. yamatoensis*, *M. nana*, *M. japonica*, *M. equine*, *M. caprae* and *M. dermatis*, were not included.

As there have been a few reports regarding the roles of these new species in seborrheic dermatitis, further studies will be important.

This study is a culture-based study on data gathered from Korean subjects to examine distribution of *Malassezia* yeast. Culture-based techniques are expected to be useful in the clinical assessment of *Malassezia* species associated with other fungal infections. However, this conventional method was complex in culture conditions and had a lower chance of successful culture. The assessment of additional types of clinical samples is indicated to determine the broad applicability of the approach. Recently, non-culture based techniques which avoided the culture process and directly extracted fungal DNA and differentiated *Malassezia* species was introduced^{22,23}. Further studies are required to evaluate this new technique and compare the differences in the rate of successful identification on the scalp in seborrheic dermatitis patients.

In conclusion, *M. restricta* appears to be the most important *Malassezia* species in Korean seborrheic dermatitis patients. Age and gender do not seem to greatly influence the distribution of *Malassezia* species. This was a pioneer study which first investigated more than ten Korean seborrheic dermatitis patients. Further studies including a larger numbers of patients will be needed to confirm our results.

REFERENCES

1. Janik MP, Heffernan MP. Yeast infections: candidiasis, pityriasis versicolor. In: Wolff K, Goldsmith LA, Katz SI, Gilchrest BA, Paller AS, Leffell DJ, editors. Fitzpatrick's dermatology in general medicine. 7th ed. New York: McGraw-Hill, 2008:1822-1830.
2. Guého E, Midgley G, Guillot J. The genus *Malassezia* with description of four new species. Antonie Van Leeuwenhoek 1996;69:337-355.
3. Sugita T, Tajima M, Ito T, Saito M, Tsuboi R, Nishikawa A. Antifungal activities of tacrolimus and azole agents against the eleven currently accepted *Malassezia* species. J Clin Microbiol 2005;43:2824-2829.
4. Zisova LG. *Malassezia* species and seborrheic dermatitis. Folia Med (Plovdiv) 2009;51:23-33.
5. Gupta AK, Batra R, Bluhm R, Boekhout T, Dawson TL Jr. Skin diseases associated with *Malassezia* species. J Am Acad Dermatol 2004;51:785-798.
6. Morishita N, Sei Y, Sugita T. Molecular analysis of *Malassezia* microflora from patients with pityriasis versicolor. Mycopathologia 2006;161:61-65.
7. Sugita T, Tajima M, Tsuboku H, Tsuboi R, Nishikawa A. Quantitative analysis of cutaneous *Malassezia* in atopic dermatitis patients using real-time PCR. Microbiol Immunol 2006;50:549-552.
8. Gupta AK, Boekhout T, Theelen B, Summerbell R, Batra R.

- Identification and typing of *Malassezia* species by amplified fragment length polymorphism and sequence analyses of the internal transcribed spacer and large-subunit regions of ribosomal DNA. *J Clin Microbiol* 2004;42:4253-4260.
9. Gandra RF, Simão RC, Matsumoto FE, da Silva BC, Ruiz LS, da Silva EG, et al. Genotyping by RAPD-PCR analyses of *Malassezia furfur* strains from pityriasis versicolor and seborrheic dermatitis patients. *Mycopathologia* 2006;162: 273-280.
 10. Guillot J, Deville M, Berthelemy M, Provost F, Guého E. A single PCR-restriction endonuclease analysis for rapid identification of *Malassezia* species. *Lett Appl Microbiol* 2000; 31:400-403.
 11. Makimura K, Tamura Y, Kudo M, Uchida K, Saito H, Yamaguchi H. Species identification and strain typing of *Malassezia* species stock strains and clinical isolates based on the DNA sequences of nuclear ribosomal internal transcribed spacer 1 regions. *J Med Microbiol* 2000;49:29-35.
 12. Sugita T, Kodama M, Saito M, Ito T, Kato Y, Tsuboi R, et al. Sequence diversity of the intergenic spacer region of the rRNA gene of *Malassezia globosa* colonizing the skin of patients with atopic dermatitis and healthy individuals. *J Clin Microbiol* 2003;41:3022-3027.
 13. Jang SJ, Lim SH, Ko JH, Oh BH, Kim SM, Song YC, et al. The investigation on the distribution of *Malassezia* yeasts on the normal Korean skin by 26S rDNA PCR-RFLP. *Ann Dermatol* 2009;21:18-26.
 14. Lee YW, Kim SM, Oh BH, Lim SH, Choe YB, Ahn KJ. Isolation of 19 strains of *Malassezia dermatis* from healthy human skin in Korea. *J Dermatol* 2008;35:772-777.
 15. González A, Sierra R, Cárdenas ME, Grajales A, Restrepo S, Cepero de Garcia MC, et al. Physiological and molecular characterization of atypical isolates of *Malassezia furfur*. *J Clin Microbiol* 2009;47:48-53.
 16. Zomorodian K, Mirhendi H, Tarazooie B, Zeraati H, Hallaji Z, Balighi K. Distribution of *Malassezia* species in patients with psoriasis and healthy individuals in Tehran, Iran. *J Cutan Pathol* 2008;35:1027-1031.
 17. Mirhendi H, Makimura K, Zomorodian K, Yamada T, Sugita T, Yamaguchi H. A simple PCR-RFLP method for identification and differentiation of 11 *Malassezia* species. *J Microbiol Methods* 2005;61:281-284.
 18. Clift DC, Dodd HJ, Kirby JD, Midgley G, Noble WC. Seborrheic dermatitis and malignancy. An investigation of the skin flora. *Acta Derm Venereol* 1988;68:48-52.
 19. Lim SW, Shin MG, Lim JY, Yun SJ, Kim SJ, Lee SC, et al. Nested PCR for detection of *Malassezia* species from patient skin scales and clinical strains. *Korean J Dermatol* 2008;46:446-452.
 20. Sugita T, Suzuki M, Goto S, Nishikawa A, Hiruma M, Yamazaki T, et al. Quantitative analysis of the cutaneous *Malassezia* microbiota in 770 healthy Japanese by age and gender using a real-time PCR assay. *Med Mycol* 2010;48: 229-233.
 21. Gaitanis G, Velegaki A, Alexopoulos EC, Chasapi V, Tsigonia A, Katsambas A. Distribution of *Malassezia* species in pityriasis versicolor and seborrheic dermatitis in Greece. Typing of the major pityriasis versicolor isolate *M. globosa*. *Br J Dermatol* 2006;154:854-859.
 22. Tajima M, Sugita T, Nishikawa A, Tsuboi R. Molecular analysis of *Malassezia* microflora in seborrheic dermatitis patients: comparison with other diseases and healthy subjects. *J Invest Dermatol* 2008;128:345-351.
 23. Takahata Y, Sugita T, Kato H, Nishikawa A, Hiruma M, Muto M. Cutaneous *Malassezia* flora in atopic dermatitis differs between adults and children. *Br J Dermatol* 2007;157:1178-1182.