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## Requirements for mammalian carboxylesterase inhibition by substituted ethane-1,2-diones

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### Abstract

Carboxylesterases (CE) are ubiquitous enzymes found in both human and animal tissues and are responsible for the metabolism of xenobiotics. This includes numerous natural products, as well as a many clinically used drugs. Hence the activity of these agents is likely dependent upon the levels and location of CE expression. We have recently identified benzil is a potent inhibitor of mammalian CEs, and in this study, we have assessed the ability of analogues of this compound to inhibit these enzymes. Three different classes of molecules were assayed: One containing different atoms vicinal to the carbonyl carbon atom and the benzene ring [PhXC(O)C(O)XPh, where X = CH<sub>2</sub>, CHBr, N, S, or O]; a second containing a panel of alkyl 1,2-diones demonstrating increasing alkyl chain length; and a third consisting of a series of 1-phenyl-2-alkyl-1,2-diones. In general, with the former series of molecules, heteroatoms resulted in either loss of inhibitory potency (when X = N), or conversion of the compounds into substrates for the enzymes (when X = S or O). However, the inclusion of a brominated methylene atom resulted in potent CE inhibition. Subsequent analysis with the alkyl diones [RC(O)C(O)R, where R ranged from CH<sub>3</sub> to C<sub>8</sub>H<sub>17</sub>] and 1-phenyl-2-alkyl-1,2-diones [PhC(O)C(O)R where R ranged from CH<sub>3</sub> to C<sub>6</sub>H<sub>13</sub>], demonstrated that the potency of enzyme inhibition directly correlated with the hydrophobicity (clogP) of the molecules. We conclude from these studies that that the inhibitory power of these 1,2-dione derivatives depends primarily upon the hydrophobicity of the R group, but also on the electrophilicity of the carbonyl group.

### 1. Introduction

Carboxylesterases (CE<sup>1</sup>) are enzymes found in a wide range of organisms, from humans to bacteria [1]. CEs are known to be involved in the hydrolysis of ester-containing xenobiotics [1] using a catalytic serine within a Ser-His-Glu triad to initiate hydrolysis of the molecule.

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<sup>1</sup>Abbreviations: CE - carboxylesterase CPT-11 - Irinotecan, 7-ethyl-10-[4-(1-piperidino)-1-piperidino]carbonyloxycamptothecin; hAChE - human acetylcholinesterase; hBChE - human butyrylcholinesterase; hBr3 - human brain carboxylesterase; hCE1 - human carboxylesterase 1; hiCE - hCE2, human intestinal carboxylesterase; HPLC - high performance liquid chromatography; 4-MUA - 4-methylumbelliferone acetate; o-NPA - nitrophenyl acetate; rCE - rabbit liver carboxylesterase; SN-38 - 7-ethyl-10-hydroxycamptothecin.

The products that result from this reaction are the respective alcohol and carboxylic acid [2, 3]. Two major CEs exist in humans, human liver CE (hCE1; CES1) and human intestinal CE (hiCE; CES2) [2, 3] with hCE1 being primarily expressed in the liver, while hiCE is found in both the liver and the small intestine. A third human CE, hBr3 (CES3), has been described, but very little is known about the levels of expression and/or the substrate specificity of this enzyme [4]. While the exact role of these proteins in mammals is unclear, and endogenous substrates have not been definitively identified, the patterns of expression are consistent with them playing a protective role. Furthermore, since the carboxylic ester chemotype is present in numerous agents including natural products, pesticides and clinically used drugs, de facto they are substrates for these enzymes [5–10]. Hence drug hydrolysis, which can result in either activation or inactivation of the molecule, is dependent upon the levels of CE expressed in exposed tissues and the substrate specificity of the protein.

One such chemotherapeutic agent that is metabolized by CEs is the anticancer drug irinotecan (CPT-11, 7-ethyl-10-[4-(1-piperidino)-1-piperidino]carbonyloxycamptothecin [6, 9, 11]). CPT-11 is a carbamate-derived prodrug that is hydrolyzed by hiCE into its active metabolite, SN-38 (7-ethyl-10-hydroxycamptothecin) [12, 13]. The latter is a potent topoisomerase I poison which exerts its toxicity at low nanomolar concentrations. We have previously demonstrated the efficient activation of CPT-11 by a rabbit liver CE (rCE) and used this enzyme to modulate tumor cells sensitivity to this drug [14–16]. The development of clinical approaches using this technology is currently underway. However, the toxicity of CPT-11 (delayed diarrhea) is in part due to high levels of hiCE that are expressed in the intestine [6, 17]. Therefore, identifying specific hiCE inhibitors which could be used in combination with CPT-11 to ameliorate the delayed diarrhea, may have clinical utility [2, 3]. Previously, we determined that small molecules containing the ethane 1,2-dione moiety were potent inhibitors of CEs [18–21]. These compounds demonstrated no activity toward human acetyl- or butyrylcholinesterase, and one class, the benzils, inhibited hiCE intracellularly and modulated cellular response to CPT-11 [22]. Preliminary studies indicated that the planarity of the ethane-1,2-dione group could determine specificity of enzyme inhibition and that inhibitor potency was increased when phenyl groups were present within the molecule. In this study, we sought to determine the chemical requirements for inhibition of CEs by ethane 1,2-diones and to assess whether nucleophilic attack by the serine O<sub>γ</sub> atom within the active site is the mechanism by which enzyme inhibition occurs. This included evaluating the role of the atoms immediately adjacent to the carbonyl groups towards inhibitor potency and to assess the necessity for the inclusion of the phenyl rings. Our results indicate that the atoms bonded to the carbonyl groups in the 1,2-diones play a major role towards inhibitor potency, both by moderating carbonyl electrophilicity, and compound hydrophobicity. Indeed, molecules with clogP > 2.75 were much more potent inhibitors of mammalian CEs than more hydrophilic compounds (clogP < 2.75). Furthermore, aromaticity within the molecule is not a requirement for enzyme inhibition.

## 2. Materials and Methods

### 2.1 Chemicals and General Chemistry

All solvents and starting materials were purchased from Sigma Aldrich (St. Louis, MO) and were ACS grade or better. Benzil (**1**), butane-2,3-dione (**8**), hexane-3,4-dione (**9**), and 1-phenyl-1,2-propanedione (**17**) were obtained from Sigma Aldrich. Diphenyl ethanedioate (**4**), 1,4-diphenylbutane-2,3-dione (**5**), 1,4-dibromo-1,4-diphenylbutane-2,3-dione (**6**) and 1-bromo-1,4-diphenylbutane-2,3-dione (**7**) were purchased from MDPI (Basel, Switzerland). Octane-4,5-dione (**10**) was obtained from the Developmental Therapeutics Program at NIH and 1,2-dicyclohexylethane-1,2-dione (**16**) was synthesized as previously described [20, 23]. The structures of the compounds described in this paper are illustrated in Table 1.

For compounds synthesized as part of these studies, melting points were determined using a Mel-temp (Barnstead International, Dubuque, IA) and purity and structures were assessed by TLC, NMR, HRMS and total C, H, N, O, S analysis (as appropriate). Complete physical data is presented as Supplementary Information.

## 2.2 Enzymes

Pure hCE1, hiCE, and rabbit liver CE (rCE) were obtained by purification from baculovirus infected *Spodoptera frugiperda* cells as previously described [24]. Human acetylcholinesterase (hAChE) and butyrylcholinesterase (hBChE) were obtained from Sigma Biochemicals (St. Louis, MO).

## 2.3 Synthesis of oxanilide (*N, N'*-diphenylethanedi- amide (2))

Oxanilide was synthesized from aniline and oxalyl chloride as previously described [25]. Physical parameters were consistent with that reported previously. Identity was confirmed by melting point, NMR, and elemental analysis.

## 2.4 Synthesis of *S*<sup>1</sup>, *S*<sup>2</sup>-diphenyl ethanebis(thioate) (3)

The named compound was synthesized by condensation of thiophenol with oxalyl chloride in dichloromethane [26]. Identity was confirmed by melting point, NMR, and elemental analysis.

## 2.5 Synthesis of alkyl diones and phenyl alkyl diones

Alkyl diones (**11–15**) were synthesized by oxidation of the corresponding alkyne using potassium permanganate in an acetone/water phase transfer system [27]. Routinely, 10.8 mM 1-phenyl-1-alkyne dissolved in 420 ml acetone was added to 240 ml aqueous 32 mM NaHCO<sub>3</sub> and 22 mM MgSO<sub>4</sub>. Potassium permanganate (6.65 g, 42 mmol) was added in one portion and the reaction stirred for 4 hours at rt. Small portions of NaNO<sub>2</sub> and 10% H<sub>2</sub>SO<sub>4</sub> were added until the reaction was clear (~5 g and 50 ml, respectively) and solid NaCl was then added to achieve saturation. Samples were worked up using standard approaches and compound identity was confirmed by <sup>1</sup>H NMR, elemental analysis (Microlab Inc., Norcross, GA), and HRMS (UIUC School of Chemical Sciences, Urbana, IL). Typically yields were greater than 70% and compound physical parameters were consistent with those previously reported.

## 2.6 cLogP calculations

cLogP values were calculated using ChemSilico Predict v2.0 software (ChemSilico LLC, Tewksbury, MA).

## 2.7 Determination of *K*<sub>i</sub> values using *o*-nitrophenyl acetate (*o*-NPA) as a substrate

Inhibition of CEs was assessed using a multiwell plate spectrophotometric assay with 3 mM *o*-NPA as a substrate [21, 28]. Inhibitors (100 μM) and substrate were aliquoted into wells of a 96-well plate and enzyme was added. The rate of change in absorbance at 420 nm was recorded at 15 s intervals for 5 min and data was compared to wells without inhibitor. Data was expressed as the yield of *o*-nitrophenol per minute per milligram of protein. Compounds demonstrating greater than 50% reduction of CE activity were re-screened with dilutions ranging from 1 pM to 100 μM. Data was fitted to the following equation to determine the mode of inhibition [29]:

$$i = \frac{[I]\{[s](1 - \beta) + Ks(\alpha - \beta)\}_-}{[I]\{[s] + \alpha Ks\} + Ki\{\alpha[s] + \alpha Ks\}}$$

where  $i$  is the fractional inhibition,  $[I]$  is the inhibitor concentration,  $[s]$  is the substrate concentration,  $\alpha$  is the change in affinity of the substrate for enzyme,  $\beta$  is the change in the rate of enzyme substrate complex decomposition,  $K_s$  is the dissociation constant for the enzyme substrate complex, and  $K_i$  is the inhibitor constant. Examination of the curve fits where  $\alpha$  ranged from 0 to  $\infty$  and  $\beta$  ranged from 0 to 1 was performed using GraphPad Prism software and Perl data language [21, 28]. Curves were generated and analyzed using Akaike's information criteria [30, 31]. Using the equation generated by Prism considered to be the best fit for the datasets,  $K_i$  values were computed [21, 28].

## 2.8 Inhibition of acetylcholinesterase and butyrylcholinesterase

Inhibition of hAChE and hBChE by compounds was determined as previously described using 1 mM of either acetylthiocholine (AcTCh) or butyrylthiocholine (BuTCh), respectively, as substrates [32–35].

## 2.9 Irreversible enzyme inhibition assays

To assess irreversible enzyme inhibition by selected compounds, inhibitors were added to hiCE at a final concentration of 10  $\mu$ M in Hepes pH 7.4 and incubated on ice for 40 mins. At this time, samples were diluted 1:500 and enzyme activity was determined using o-NPA as a substrate.

## 2.10 Enzyme inhibition assays and determination of $K_i$ values using CPT-11 as a substrate

CPT-11 (20  $\mu$ M) and inhibitor concentrations ranging from 0–100  $\mu$ M were pre-incubated for 2 min at 37°C in 50 mM HEPES pH 7.4. Purified hiCE (25 U) was then added and the sample was incubated at 37°C for 5 min. The reaction was terminated by addition of 100  $\mu$ l of acid methanol and the amount of SN-38 in the sample was determined using high performance liquid chromatography [14, 36].  $K_i$  values were then determined using Prism software as described earlier.

## 2.11 Determination of ability of compounds to act as substrates

Compounds **2**, **3**, or **4** (200  $\mu$ M) were incubated with enzyme for varying time periods (1, 5, or 20 min) at either rt or 37°C in a total volume of 200  $\mu$ l of 50 mM Hepes pH 7.4 and reactions were terminated by addition of acetonitrile. Samples were then evaluated by reverse-phase high-performance liquid chromatography and products were identified by UV detection. These results were compared to samples without enzyme and to samples with marker compounds consisting of aniline, thiophenol, or phenol.

## 2.12 Intracellular CE inhibition assay

Intracellular CE inhibition was assessed using the substrate 4-methylumbelliferone acetate (4-MUA) as previously described [22]. Briefly,  $10^6$  cells were suspended in PBS with stirring in a cuvette and were pre-incubated with inhibitor (10  $\mu$ M) for 1 hour. 4-MUA was then added (100  $\mu$ M) and fluorescence was determined using a Hitachi F-2000 fluorimeter, with an excitation wavelength of 365 nm and an emission wavelength of 460 nm. Data was collected over 2 minutes and the percentage of inhibition of product formation as compared to cell not treated with inhibitor was determined by assessing the difference in fluorescence intensity at 30 s.

## 2.13 Growth inhibition assay

Growth inhibition assays using inhibitors and CPT-11 were undertaken as previously described [15, 22, 37]. Briefly, cells were plated in 6-well plates (50,000 cells/well) and allowed to adhere overnight. Following pre-incubation with inhibitor (10  $\mu$ M) for 1 hour, cells were treated for 2 hours with both inhibitor and varying concentrations of CPT-11 (0–

100  $\mu\text{M}$ ). Cells were then washed and allowed to grow in drug-free medium for 4 days (~3 cell doublings), prior to harvesting and counting using a Coulter Z2 counter (Beckman Coulter, Fullerton, CA). Routinely, all concentrations were conducted in triplicate and the percentage of surviving cells was determined for each treatment condition and compared to control cells not exposed to inhibitor. Survival curves were generated and  $\text{IC}_{50}$  values were calculated using Prism software (GraphPad, San Diego, CA).

## 2.14 Molecular dynamics simulations

Molecular dynamics simulations to evaluate the dihedral angle of the 1,2-dione bond [(O)CC(O)] were performed as previously described [19]. Briefly, analyses were run using the MM3 module of Alchemy (Tripos Inc., St. Louis, MO) with angle measurements recorded every 10 fs, over a total time frame of 15 ps, using a time step of 1 fs.

## 2.15 Computational chemistry

All computational analyses were performed using Gaussian 03 software (Gaussian, Wallingford, CT). Molecules were constructed using Gauss-View and geometry optimization was performed using the density functional theory method using the B3LYP hybrid functional [38] with the 6-31G(d, p) basis set [39]. Charge distribution was then visualized using Molden (Centre for Molecular and Biomolecular Informatics, Radboud University Nijmegen Medical Centre, Nijmegen, Netherlands).

## 3. Results

### 3.1 Analysis of CE inhibition by 1,4-diphenylbutane-2,3-dione analogues

To assess the role of the atom vicinal to the carbonyl carbon atoms in the molecule towards enzyme inhibition, we synthesized a series of 1,4-diphenylbutane-2,3-dione analogues containing a variety of different moieties in the  $\alpha$ -position. In general, symmetrical compounds were used to ensure that any inhibitory effects would not be as a result of competitive inhibition by two different sites of nucleophilic attack by the protein.

#### 3.1.1 Oxanilide, $S^1$ , $S^2$ -diphenyl ethanebis(thioate) and diphenyl ethanedioate

—Oxanilide (**2**) was not an inhibitor of hCE1, hiCE or rCE (Table 1) and also lacked activity towards the cholinesterases. Similarly, analogues containing either sulfur (**3**;  $S^1$ ,  $S^2$ -diphenyl ethanebis(thioate)) or oxygen (**4**; diphenyl ethanedioate) yielded molecules that were ineffective at enzyme inhibition (Table 1). However, unlike oxanilide, the latter two compounds underwent spontaneous hydrolysis in aqueous solution liberating either thiophenol or phenol, respectively (data not shown). Furthermore, this reaction was not enhanced by the addition of CE. While detailed kinetics were not determined, greater than 40% of each compound was hydrolyzed within 5 mins of addition to buffered aqueous (pH 7.4) solution. While this was expected for **3** (as it is an activated thioester), it was unclear whether compound **4** would be subject to hydrolysis. Our results confirm that the ester is also labile under these conditions.

#### 3.1.2 1,4-Diphenylbutane-2,3-dione, 1-bromo-1,4-diphenylbutane-2,3-dione and 1,4-dibromo-1,4-diphenylbutane-2,3-dione

—Insertion of a carbon atom in the  $\alpha$ -position (**5**; 1,4-diphenylbutane-2,3-dione) resulted in a molecule that was effective at CE inhibition, but the potency was ~200- to 480-fold weaker than that seen with benzil (Table 1). The mode of enzyme inhibition was deemed to be partially competitive (i.e., indicating that while the small molecule structurally resembled a CE substrate, they were unable to reduce substrate hydrolysis to 0%, even at infinite inhibitor concentrations). The brominated derivatives, 1,4-dibromo-1,4-diphenylbutane-2,3-dione (compound **6**) and 1-bromo-1,4-diphenylbutane-2,3-dione (compound **7**), were considerably more active, with the dibromo

analogue ~30- to 5000-fold more potent inhibitor than **5**. These results suggest that the activation of the carbonyl group was a major effector in improving inhibitor function. While compound **7** was generally more potent than **5**, dramatic improvements in the inhibition constants for this molecule were only observed with hiCE (Table 1).

It is possible the bromine analogues may change the mechanism of enzyme inhibition by allowing nucleophilic attack on the methylene atom rather than the carbonyl carbon. This would then result in irreversible inhibition of the protein. Therefore, we evaluated whether compound **6** could inactivate hiCE following incubation with the protein. As indicated in Table 1 however, no irreversible inhibition of the enzyme was observed with this analogue.

**3.1.3 Flexibility of the (O)C-C(O) bond**—In previous studies, we have seen a correlation between the flexibility of rotation around the 1,2-dione bond and the inhibitory potency of the molecules. Typically, compounds that adopted a more rigid structure tended to be more potent CE inhibitors [19]. Therefore using molecular dynamics approaches, we evaluated the rotation of this bond for benzil and compound **5**. As indicated in Figure 1, the 1,2-dione in benzil rapidly adopted a stable conformation with a dihedral angle of ~ 230°. In contrast, with 1,4-diphenylbutane-2,3-dione, rotation around the dione continued to oscillate over the entire course of the dynamics run. Since compound **5** is a poorer inhibitor than benzil, these results are consistent with those seen previously for a panel of heteroaromatic ethane-1,2-diones, where increased oscillation was observed with weaker molecule potency [19].

### 3.2 Synthesis and enzyme inhibition by alkyl-1,2-diones

Since the active sites of the mammalian CEs are highly hydrophobic [2], we hypothesized that small molecules with larger clogP values would be more potent enzyme inhibitors. Therefore, we obtained, or generated, a series of alkyl-1,2-diones ranging from butane-2,3-dione to octadecane-9,10-dione. Synthesis was accomplished by oxidation of the corresponding alkyne using potassium permanganate in an acetone/water phase transfer system. Yields were routinely greater than 70% and most compounds were pale yellow crystalline solids. Chemical and physical properties for these compounds are presented as supplementary information.

CE inhibition by the alkyl-1,2-diones was assessed for both hCE1 and hiCE, as well as rCE, using *o*-NPA as a substrate (Table 2). As can be seen, as the compounds become larger, the molecules become more potent inhibitors. Since this is likely due to the increase in the hydrophobicity of the compounds, we calculated the clogP of the alkyl diones using commercially available software. As indicated in Table 2, both inhibitor potency and the clogP values increased with increasing chain length. As an example, compound **15** containing eight carbon atoms in the alkyl group, was a very potent inhibitor of the mammalian CEs (0.84 nM and 5.7 nM for hCE1 and hiCE, respectively) and demonstrated a clogP value of 6.03. As before, the mode of enzyme inhibition by these compounds was found to be partially competitive.

### 3.3 Carboxylesterase inhibition by 1-phenyl-2-alkyl-1,2-diones

Having observed potent inhibition by the alkyl-1,2-diones, we next evaluated a series of phenyl alkyl derivatives. These were chosen since our original data demonstrated that benzil (**1**) was an excellent inhibitor and that potency, and potentially selectivity, might be afforded by the alkyl chain. Therefore, we synthesized a series of 1-phenyl-1,2-alkyldiones using similar methodology to that described for the alkyl derivatives. Similar yields (~70%) were obtained and all physical parameters were consistent with their structure (see Supporting Information).

As indicated in Table 3, potent enzyme inhibition was observed when the alkyl chain exceeded 4 carbon atoms. For example, 1-phenyl-1,2-octanedione (**22**), the most potent compound synthesized, yielded  $K_i$  values of 2.2 nM, 28.7 nM and 28.1 nM for hCE1, hiCE and rCE, respectively. In general, these molecules did not generate inhibition constants as low as that seen for the alkyl-1,2-diones, however these molecules were clearly effective CE inhibitors. Again, the mode of enzyme inhibition was determined to be partially competitive, and inhibitor potency appeared to correlate with the clogP of the compounds.

### 3.4 Correlation between clogP and inhibitory potency for the 1,2-diones

Since we saw a trend towards increased inhibitory potency with higher clogP values for both the alkyl-1,2-diones, and 1-phenyl-1,2-alkyldiones, we evaluated whether correlations could be determined for these parameters. As indicated in Figure 2, linear correlations could be described for both individual datasets (panel A – alkyl-1,2-diones; panel B - 1-phenyl-2-alkyl-1,2-diones), as well as the combined values for all compounds (panel C). The linear regression analyses yielded  $r^2$  values ranging from 0.72 to 0.99, depending upon both the enzyme and the class of compounds. Overall, these data indicate that the clogP and  $K_i$  values for all of the CEs and inhibitors that were evaluated appear to be related, with increasing clogP resulting in more potent enzyme inhibition.

### 3.5 Modulation of CPT-11 metabolism and intracellular carboxylesterase inhibition

Having identified several inhibitors that could potentially inhibit hiCE, we sought to determine the  $K_i$  values for this enzyme when using CPT-11 as a substrate. This was undertaken only with compounds that demonstrated reasonable efficacy at inhibiting o-NPA hydrolysis (compounds **11–15** and **18–22**). As indicated in Table 4, the majority of these molecules were effective at inhibiting hiCE-mediated CPT-11 metabolism, with compound **15** (octadecane-9,10-dione) yielding the lowest  $K_i$  value (~24 nM). Having established that these inhibitors could modulate CPT-11 hydrolysis in vitro, we assessed the ability of these compounds to inhibit hiCE intracellularly. This was determined using 4-MUA as a substrate in a fluorometric assay and would provide a measure of the cell permeability of the molecules [22]. As shown in Table 4, all of the inhibitors were effective at inhibiting hiCE within U373MG cells, with the most potent inhibition observed with compound **22** (1-phenyl-2-octane-1,2-dione).

To assess whether intracellular inhibition could modulate CPT-11 cytotoxicity, we undertook drug survival assays in cells expressing hiCE, following exposure to selected inhibitors. As indicated in Table 5, both benzil and 1-phenyl-2-octane-1,2-dione reduced the toxicity of CPT-11 to U373MGhiCE cells by ~6- and 3-fold respectively. Interestingly, compound **13** (tetradecane-7,8-dione) was ineffective in this assay, despite demonstrating good in vitro biochemical properties (Tables 2 and 4), and inhibiting hiCE by ~91% in the fluorometric assay.

## 4. Discussion

Past publications have demonstrated the importance of the 1,2-dione chemotype within molecules for the selective inhibition of CE [19–21]. Specifically, benzil was found to be an excellent inhibitor of hiCE, hCE1 and rCE. We previously hypothesized that the mechanism of enzyme inhibition by these compounds was due to abortive nucleophilic attack on one of the carbonyl carbon atoms of the molecule by the catalytic serine [21]. Since the tetrahedral intermediate that would be formed from this reaction would contain a C-C bond (rather than the C-O bond in the ester), we postulated that the enzyme would be unable to cleave this bond. Hence, an equilibrium would be established between the free inhibitor and the enzyme-inhibitor complex, resulting in reversible inhibition. We reasoned that the simplest

way to test this hypothesis would be to generate compounds containing either deactivated (e.g. oxanilide) or activated (e.g., diphenyl ethanedioate, 1,4-dibromo-1,4-diphenylbutane-2,3-dione) carbonyl groups, and then assess their activity towards the CEs. Therefore the studies presented here detail the interaction of the mammalian CEs with small molecules containing either highly electrophilic, or essentially inactivated, carbonyl carbon atoms.

For oxanilide (**2**), where the  $\alpha$ -atom with respect to the carbonyl carbon is nitrogen, no enzyme inhibition was observed (Table 1). This is likely due to the decreased electrophilicity of the carbonyl. This renders the molecule less susceptible to nucleophilic attack by the enzyme and as a consequence, oxanilide is unable to inhibit the mammalian CEs. This would suggest that activated electrophiles, such as the carbonyl groups present in compounds **3** and **4**, would be effective CE inhibitors. However, both analogues were both subject to spontaneous hydrolysis by water and this reaction was not enhanced by the addition of enzyme (data not shown). Since oxygen and sulfur are both electron withdrawing in nature, and each carbonyl group is further activated by the adjacent one, the 1,2-dione in these compounds is highly reactive, to the extent that even poor nucleophiles such as water, can readily hydrolyze these molecules.

1,4-Diphenylbutane-2,3-dione (**5**) was a much poorer inhibitor of the CEs than benzil (Table 1) indicating that the proximity of the benzene ring is important towards biological activity. In benzil, the phenyl group is directly attached to this moiety and it would be anticipated that due to resonance effects, this would reduce the electrophilicity of the carbonyl group. Furthermore with compound **5**, such an effect would be negated by the methylene atom, and inductive effects from the phenyl ring would actually activate this group. However, the biological data (Table 1) clearly demonstrate that this is not the case. This suggests that other parameters play a role in inhibitor potency, rather than just the electrophilicity of the carbonyl carbon atom.

We have previously demonstrated the flexibility of the 1,2-dione bond is an important parameter with respect to compound potency [19]. For a series of heteroaromatic ethane-1,2-diones, we observed that the ability of the dione bond to adopt a relatively stable conformation correlated with biological activity. In addition, if the final dihedral angle that the molecule adopted was too close to  $230^\circ$ , it could be predicted that the compound would be a potent CE inhibitor. As indicated in Figure 1, the final dihedral angle for benzil was  $\sim 230^\circ$ , whereas for compound **5**, the 1,2-dione was still oscillating and never achieved a stable conformation at the end of the dynamics simulations. The exact reason why this parameter correlates with inhibitor potency is not clear, but may be due to the alignment of the small molecule in a configuration such that nucleophilic attack by the catalytic serine is facile.

Furthermore, as indicated in Figure 3, the stability of the conformation adopted by benzil is presumably due to conjugation of the  $\pi$  electrons between the phenyl ring and the adjacent carbonyl group. This accounts for the high frequency  $1660\text{cm}^{-1}$  carbonyl IR stretch seen with this compound [20]. However, no conjugation occurs between the carbonyl group themselves. With molecule **5**, no conjugation is observed between the phenyl group and the carbonyl moiety (the IR carbonyl wave number is  $1709\text{cm}^{-1}$  [40]) and hence the rotation around the carbon – carbon bonds within the butane moiety is less restricted. These observations are consistent with the molecular dynamics simulations presented in Figure 1. Also, as can be seen in Figure 3, the electron distribution on the aromatic rings, coincident with the side of attack by a nucleophile in benzil, is reduced as compared to compound **5**. Hence, this may make the carbonyl carbon atom in 1,4-diphenylbutane-2,3-dione less prone to interaction with the serine  $O_\gamma$  atom due to increased repulsion from the charge localized on the benzene rings. Alternatively, interaction of benzil with the enzyme may result in a



conformational change in the former that results in increased affinity for the protein. Since both the molecular dynamics simulations and the computational approaches (Figures 1 and 3, respectively), indicate that benzil and compound **5** adopt different poses, it is possible that reaction of these molecules with the CEs, would result in a significant change in the energetics of the system. Since the biological data indicates that the interaction of benzil with these proteins is clearly favored, as compared to **5**, we are currently undertaking very detailed computational analyses of the proposed enzyme-inhibitor complexes to assess whether significant changes in small molecule conformation can be used to determine and/or predict inhibitor potency.

Activation of the carbonyl carbon by the addition of a bromine atom on the vicinal carbon atom (compounds **6** and **7**) resulted in decreased  $K_i$  values for enzyme inhibition. Due to the electron withdrawing properties of the halogen, and the fact that bromine is a relatively good leaving group, it is possible that nucleophilic attack by the  $O_\gamma$  within the catalytic serine could occur either at the carbonyl, or the methylene, carbon atom. The latter would result in covalent bond formation and irreversible inhibition of the enzyme. However, as indicated in Table 1, no loss of enzyme activity was observed following dilution of the inhibitor from the reaction. These results imply that interaction of the protein with the inhibitor occurred at the carbonyl carbon and not at the vicinal carbon atom. Additionally, since the kinetic analyses indicated that the mode of enzyme inhibition was partially competitive, this infers that these inhibitors bind to the same site within the CE, and that they structurally resemble the substrate. This would argue that a similar mechanism of interaction likely occurs between the enzyme and the inhibitor/substrate. Again, these data support the hypothesis that the catalytic serine undergoes nucleophilic attack of the carbonyl carbon atom. Furthermore, the halogens also increased the hydrophobicity of the molecules (Table 1), likely allowing for a further increase inhibitor potency. Since the active site gorges of CE are very hydrophobic [2, 21], compounds demonstrating greater clogP values would preferentially localize within this domain. Evidence to this effect has been noticed with a series of isatin analogues [18]. Overall, these results suggest that the electrophilicity of the carbonyl carbon atom plays a role in determining the potency of inhibition towards CEs.

To further examine the effect of small molecule hydrophobicity on enzyme inhibition, we synthesized a series of alkyl-1,2-diones and evaluated their inhibitory potency against the CEs. We reasoned that by simply increasing the size of the alkyl group, we could effect an increase in the clogP of the compounds, without significantly altering the electrostatic configuration of the molecule. As indicated in Table 2, increasing the length of the alkyl side chains resulted in dramatically reduced  $K_i$  values for the inhibitors with the CEs. This correlated well with the increasing clogP values of the compounds (Figure 2). This phenomenon was observed for all three mammalian CEs, suggesting that the environment within the active site was similar for each protein. As indicated above, since the active sites of these enzymes are highly hydrophobic, this increase in potency is likely due to preferred localization of the longer chain compounds within this domain. Indeed, since good correlation coefficients were observed between the  $K_i$  values for enzyme inhibition and the clogP values of these molecules (Figure 2), this suggests that this parameter is an important factor in determining inhibitor potency. Based upon our observations with the 1,4-diphenylbutane-2,3-dione analogues and the alkyl-1,2-diones, we hypothesized that compounds containing a phenyl group and alkyl chain on either side of the dione moiety should yield very potent CE inhibitors. As displayed in Table 3, we saw selective enzyme inhibition by such molecules, with inhibitory potency correlating with the hydrophobicity of the molecule (Figure 2). These results are in agreement with those described for the alkyl-1,2-diones and further reinforce the importance of hydrophobicity towards enzyme inhibition.

Having identified the necessary properties for these molecules to effect CE inhibition *in vitro*, we evaluated the ability of selected compounds to modulate enzyme activity in cells and to determine whether these inhibitors could alter CPT-11 cytotoxicity in cells expressing hiCE. We confirmed that the majority of the alkyl and phenylalkyl derivatives were cell permeable and were effective at inhibition hydrolysis of CPT-11 by hiCE. Furthermore, in cell viability studies, benzil and compound **22** were effective at modulating the toxicity of this drug. Surprisingly, tetradecane-7,8-dione (**13**) was ineffective at reducing CPT-11 toxicity in cells expressing hiCE. Attempts to identify a mechanism for this lack of activity (e.g. the use of serum-free culture media to eliminate protein binding, the use of glass culture dishes to minimize interaction with the plasticware, etc) have so far been unsuccessful. Potentially, these compounds maybe subjected to metabolic processes that may inactivate them, but as yet we have not identified a specific mechanism for the lack of activity of this molecule. However it is clear that compounds of this type can be used to modulate intracellular levels of CE activity and can be used as biochemical tools to evaluate enzyme function.

## 5. Conclusions

Overall, our studies have delineated the roles of selected atoms within the 1,2-dione CE-specific inhibitors and demonstrated that the electrophilicity of the carbonyl group, the conformation that the molecules adopt, and the hydrophobicity of these compounds play major roles in determining inhibitor potency. Our data further support the hypothesis that nucleophilic attack by the protein on the carbonyl carbon atom represents the most likely mechanism of enzyme inhibition. This has yielded several series of small molecules that demonstrate potency and selectivity towards CE, and elucidated the chemical requirements for the design of further compounds. These studies are currently underway.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

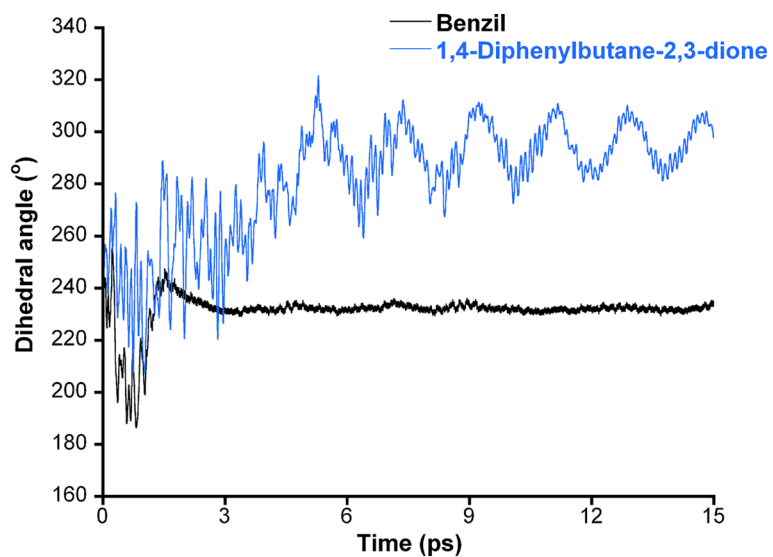
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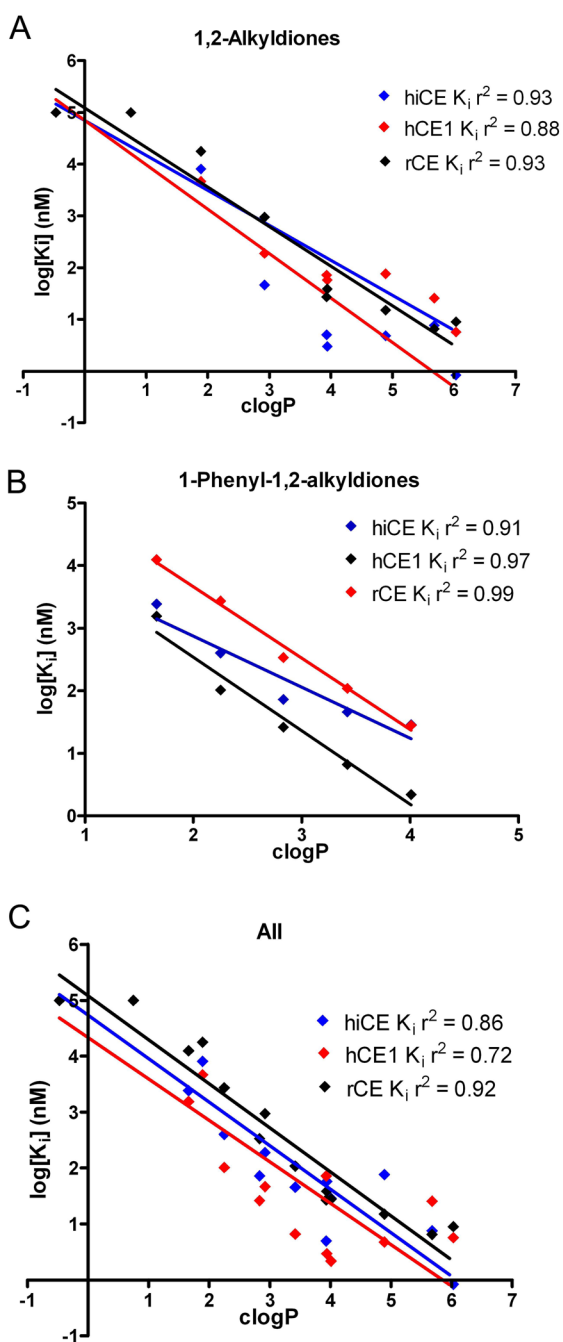
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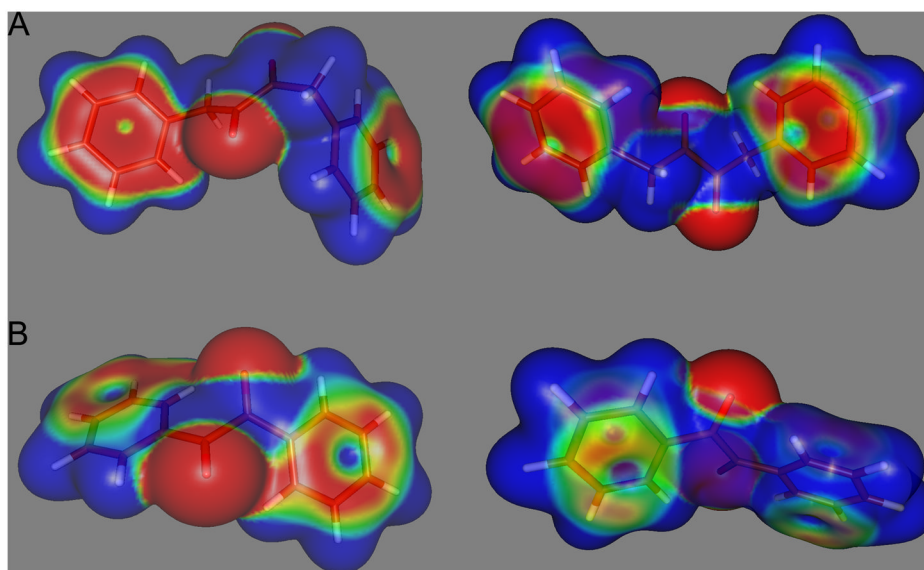
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**Figure 1.** Plot of the dihedral angle for the 1,2-dione group [(O)CC(O)] versus time for benzil (black line) and 1,4-diphenylbutane-2,3-dione (**5**; blue line) for the molecular dynamics simulations.

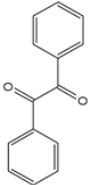
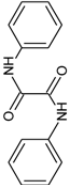
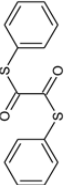
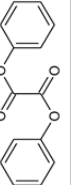
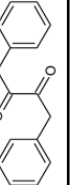
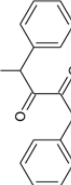
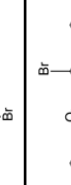


**Figure 2.** Graphs demonstrating the correlation between  $\text{clogP}$  and  $K_i$  values for the alkyl 1,2-diones (A), the 1-phenyl-1,2-alkyldiones (B) and composite results for both classes of compounds (C). Data are shown for the three different enzymes hiCE (blue), hCE1 (red) and rCE (black) and the corresponding linear correlation coefficient is indicated on the graph.



**Figure 3.** Charge distribution patterns in 1,4-diphenylbutane-2,3-dione (**5**; A) and benzil (**1**; B). Images are oriented with the oxygen atoms projecting either towards (left panel) or away (right panel) from the viewer. Red indicates relative negative charge and blue relative positive charge.

**Table 1**  
Structures, enzyme inhibition constants and kinetic parameters for the aromatic ethane-1,2-diones.

ID	Name	Structure	clogP	$K_i$ (nM)					hiCE irreversible inhibition (%)
				hCEI	hiCE	rCE	hAChE	hBChE	
1	Benzil		3.02	$15 \pm 1.9^a$	$45 \pm 3.2^a$	$103 \pm 19^a$	$>100,000$	$>100,000$	$<8$
2	Oxamiliide		2.24	$>100,000$	$>100,000$	$>100,000$	$>100,000$	$>100,000$	ND
3	<i>S</i> <sup>1</sup> , <i>S</i> <sup>2</sup> -Diphenyl ethane bis(thioate)		3.62	$>100,000$	$>100,000$	$>100,000$	$>100,000$	$>100,000$	ND
4	Diphenyl ethanedioate		3.39	$>100,000$	$>100,000$	$>100,000$	$>100,000$	$>100,000$	ND
5	1,4-Diphenylbutane-2,3-dione		3.19	$3,080 \pm 230$	$21,610 \pm 2,270$	$>100,000$	$>100,000$	$>100,000$	ND
6	1,4-Dibromo-1,4-diphenyl butane-2,3-dione		4.07	$107 \pm 21$	$334 \pm 25$	$20.5 \pm 3.3$	$>100,000$	$>100,000$	$<5$
7	1-Bromo-1,4-diphenyl butane-2,3-dione		3.71	$2,630 \pm 70$	$267 \pm 38$	$3,680 \pm 320$	$>100,000$	$>100,000$	ND

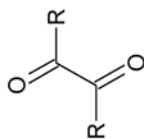
<sup>a</sup>Data taken from Wadkins et al. [21]



Table 2

Structure, logP and  $K_i$  values for enzyme inhibition for alkyl 1,2-diones.

ID	Name	R	clogP	$K_i$ (nM)					
				hCE1	hiCE	rCE	hAChE	hBChE	
8	Butane-2,3-dione	CH <sub>3</sub>	-0.47	>100,000	>100,000	>100,000	>100,000	>100,000	>100,000
9	Hexane 3,4-dione	C <sub>2</sub> H <sub>5</sub>	0.75	>100,000	>100,000	>100,000	>100,000	>100,000	>100,000
10	Octane-4,5-dione	C <sub>3</sub> H <sub>7</sub>	1.89	8,060 ± 330	4,670 ± 260	17,780 ± 1,290	>100,000	>100,000	>100,000
11	Decane-5,6-dione	C <sub>4</sub> H <sub>9</sub>	2.92	46.6 ± 9.3	189 ± 15	944 ± 75	>100,000	>100,000	>100,000
12	Dodecane-6,7-dione	C <sub>5</sub> H <sub>11</sub>	3.94	3.0 ± 0.4	57.7 ± 2.2	38.6 ± 1.6	>100,000	>100,000	>100,000
13	Tetradecane-7,8-dione	C <sub>6</sub> H <sub>13</sub>	4.89	4.8 ± 0.3	76.4 ± 9.4	15.1 ± 0.9	>100,000	>100,000	>100,000
14	Hexadecane-8,9-dione	C <sub>7</sub> H <sub>15</sub>	5.68	7.6 ± 1.6	25.6 ± 2.3	6.5 ± 1.9	>100,000	>100,000	>100,000
15	Octadecane-9,10-dione	C <sub>8</sub> H <sub>17</sub>	6.03	0.84 ± 0.28	5.7 ± 1.4	9.0 ± 3.8	>100,000	>100,000	>100,000
16	1,2-Dicyclohexylethane-1,2-dione	Cyc. C <sub>6</sub> H <sub>11</sub>	3.93 <sup>a</sup>	5.0 ± 0.7 <sup>a</sup>	72 ± 7.0 <sup>a</sup>	27 ± 7.7	>100,000	>100,000	>100,000

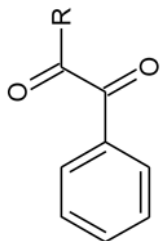


<sup>a</sup>Data taken from Hyatt et al [20].

**Table 3**

Structure, logP and  $K_i$  values for enzyme inhibition for 1-phenyl-alkyl-1,2-diones.

ID	Name	R	clogP	$K_i$ (nM)					
				hCE1	hiCE	rCE	hAChE	hBChE	
17	Phenyl-1,2-propanedione	CH <sub>3</sub>	1.07	5,270 ± 1,730	1,840 ± 260	4,930 ± 1,320	>100,000	>100,000	
18	Phenyl-1,2-butanedione	C <sub>2</sub> H <sub>5</sub>	1.66	1,550 ± 380	2,420 ± 150	12,500 ± 1,980	>100,000	>100,000	
19	Phenyl-1,2-pentanedione	C <sub>3</sub> H <sub>7</sub>	2.25	102 ± 12	401 ± 21	2,710 ± 170	>100,000	>100,000	
20	Phenyl-1,2-hexanedione	C <sub>4</sub> H <sub>9</sub>	2.83	26.1 ± 2.2	72.7 ± 5.1	337 ± 22	>100,000	>100,000	
21	Phenyl-1,2-heptanedione	C <sub>5</sub> H <sub>11</sub>	3.42	6.65 ± 0.22	45.5 ± 6.1	110 ± 10	>100,000	>100,000	
22	Phenyl-1,2-octanedione	C <sub>6</sub> H <sub>13</sub>	4.01	2.19 ± 0.08	28.7 ± 2.3	28.1 ± 1.8	>100,000	>100,000	



**Table 4**

$K_i$  values for the inhibition of CPT-11 hydrolysis by mammalian CEs with alkyl- and phenyl alkyl-1,2-diones and intracellular enzyme inhibition using 4-MUA as a substrate.

ID	Alkyl chain length (carbon atoms)	hiCE (CPT-11) (nM)	Intracellular hiCE inhibition (%) <sup>a</sup>
11	4	140 ± 18.0	85.0
12	5	193 ± 26.8	90.0
13	6	271 ± 42.7	91.2
14	7	66.8 ± 9.75	92.9
15	8	24.5 ± 2.35	80.4
18	2	2,010 ± 189	70.3
19	3	495 ± 39	86.1
20	4	168 ± 16.6	93.0
21	5	98.1 ± 20.6	93.6
22	6	130 ± 11.0	95.1
Benzil (1)	NA	175 ± 29 <sup>b</sup>	95.1 <sup>c</sup>

<sup>a</sup>Data represent the results obtained from a representative experiment.

<sup>b</sup>Taken from Wadkins et al. [21]

<sup>c</sup>Taken from Hyatt et al. [22]

**Table 5**Modulation of IC<sub>50</sub> values for CPT-11 in U373MG cells expressing hiCE by selected 1,2-dione inhibitors

Compound	IC <sub>50</sub> (μM) <sup>a</sup>
CPT-11	3.6
Benzil ( <b>1</b> )	>100
CPT-11 + Benzil ( <b>1</b> )	22.9
<b>13</b>	>100
CPT-11 + <b>13</b>	4.0
<b>22</b>	>100
CPT-11 + <b>22</b>	10.2

<sup>a</sup>Data are representative values from several experiments.