

# Nanomaterials in controlled drug release

Xin-Jun Cai · Ying-Ying Xu

Received: 24 March 2011 / Accepted: 3 June 2011 / Published online: 1 July 2011  
© Springer Science+Business Media B.V. 2011

**Abstract** In past years with the advances of chemistry and material sciences, the development of nanotechnology brought generations of nanomaterials with specific biomedical properties. These include the nanoparticle-based drug delivery, nanosized drugs, and nanomaterials for tissue engineering. The present article focuses on the use of nanomaterials in controlled drug release. The applications of nanomaterials with nano-enabled drug release characteristics brought many benefits when compared to the traditional (bulk) materials. We discuss the current advances and propose some future directions for the technology development.

**Keywords** Drug release · Nanomaterial · Nanoparticle · Gel · Colloid · Drug delivery · Self-assembly

Nanomaterials include all kinds of materials with at least one of the dimensions in the scale of 100 nm (Suri et al. 2007). In recent decade, nanomaterials expanded greatly due to the fast development of techniques in material science, and most bulk

materials could be processed in order to acquire nano-features. Generally, nanomaterials include fullerene, nanoparticles, colloids, and nano-gels, which have different applications in various areas. In the present article, we would like to focus on nanoparticle and nano-gel-based control of drug release.

## Nanoparticles

In past decade, drug delivery made with nanoparticles or nanovesicles has brought great commercial benefits in the industry, with 24 products exceeding 5 billion US dollar sales (Huynh et al. 2009; Jatariu et al. 2009; Kateb et al. 2010; Provenzale and Silva 2009).

### Variety of nanoparticles

The idea of nanoparticle emerged in 1970s as the delivery substrate for anticancer drugs and vaccines. Various nanoparticles since then were designed and developed, including poly(ethylene oxide)-poly(L-lactic acid)/poly(b-benzyl-L-aspartate), poly(lactide-co-glycolide)-[(propylene oxide)-poly(ethylene oxide)], polyphosphazene derivatives, poly(ethylene glycol)-coated nanospheres, azidothymidin (AZT)/dideoxycytidine (ddc) nanoparticles, poly(isobutylcynoacrylate) nanocapsules, poly(g-benzyl-L-glutamate)/poly(ethylene oxide), chitosan-poly(ethylene oxide) nanoparticles, methotrexate-o-carboxymethylate chitosan, solid lipid nanoparticles (SLNs), and so on (Blasi et al.

---

X.-J. Cai · Y.-Y. Xu (✉)  
Department of pharmacy, Integrated Chinese and Western  
Medicine Hospital of Zhejiang Province, 310003  
Hangzhou, Zhejiang, China  
e-mail: zjtemcxj@163.com

X.-J. Cai  
e-mail: xinjun\_cai@163.com

2009; Caruso et al. 2010; Chekhonin et al. 2009; Fonseca et al. 2009; Gilmore et al. 2008; Graf et al. 2009; Huynh et al. 2009; Jatariu et al. 2009; Kaur et al. 2008; Khalil and Mainardes 2009; Ljubimova et al. 2008; Mistry et al. 2009b; Modi et al. 2010; Namdeo et al. 2008; Shilpi et al. 2007; Tosi et al. 2008; Wong et al. 2010). Further to these polymer nanoparticles, there are metal-based, lipid-based, and biological nanoparticles. These different surface modifications could result in different interacting efficiency between the nanoparticles and cells, suggesting the diversity of applications (Table 1).

### Drug release from nanoparticles

There are several basic approaches to control drug release from nanoparticles. First, the different affinities of nanoparticle surface molecules to different cells and tissues lead varies targeting efficiency, suggesting a targeted drug delivery effect. Therefore, the local drug concentration could be significantly higher in the regions of interest, especially when the nanoparticles were further coated with antigen-recognized antibodies. Secondly, the biodegradation of nanoparticles would vary due to the different natures of the materials, the diverse biodegrading activity of enzymes among tissues, and the co-applied adjuvants. Thirdly, it would be ideal to design multi-layer nanoparticles with adjustable degrading curve, with a controlled drug release (Denora et al. 2009; Huynh et al. 2009; Jatariu et al. 2009). Adjuvants could be further combined to regulate the release of specific drugs from the nanoparticle when needed.

### Administration routes of nanoparticles

Many routes of administration of nanoparticles are available, such as oral delivery, inhalation, transdermal,

implantation, and injection. Inhalation was generally used to treat diseases relevant to respiratory systems, while the olfactory nerve could also mediate the direct drug delivery from the nasal epithelium into the brain, without crossing the blood–brain barrier (Al-Ghanaem et al. 2010; Ali et al. 2010; Jogani et al. 2008; Kumar et al. 2009; Mistry et al. 2009a; Nochi et al. 2010; Patel et al. 2009). The idea originated from the observation that some viral infections can propagate into the brain through the olfactory nerve, and some metal dusts were found in the olfactory bulb of mine workers (Charlton et al. 2007; Oberdörster et al. 2004). Now, the nasal drug delivery was recognized as a routine route of drug administration, even for some diseases of the brain (Betbeder et al. 2000; Fisher and Ho 2002; Patel et al. 2009). This requires the small sized nanoparticles to be transported by the olfactory nerve; and more important, any potential biohazards could lead to serious effects of the central nervous system functions. The application requires cautious evaluation before any drug could be adopted clinically for nasal-brain delivery.

### Virus-based drug delivery

More arguments suggest that virus-based drug delivery could be categorized as a biological nanoparticle system. The viruses are with the natural infection routes and could have self-proliferation. Therefore, the utilization of any viral-based drug delivery should consider following aspects: selection of the proper virus to use; engineering of the virus to remove hazardous effects; packaging of the drug/gene sequences into the virus/viral protein-coated nanoparticle; activity and proliferation control following targeting. The main groups of viral vector include enveloped and non-enveloped groups. Enveloped viral vectors include retrovirus (RNA), lentivirus (RNA) and HSV-1 (dsDNA); non-enveloped viral

**Table 1** Nanoparticles for controlled drug release and targeted delivery

Types	Source	Bio-compatibility	Controlled release	Targeting efficiency
Polymer	Chemical synthesis	Could be improved with surface modification	Yes	Depends
Metal	Mechanical	Low	Less easier	Low
Lipid	Chemical synthesis	Low	Yes	Low
Biological	Chemical synthesis or biological	High	Depends	Varies

vectors include AAV (ssDNA) and adenovirus (dsDNA) (Bok 2004; Fisher and Ho 2002; Fonseca et al. 2009; Heilbronn and Weger 2010; Poeschla 2003; Sleeper et al. 2009). These viral vectors are of different sizes, varied tissue inflammation reactions, different genome integration abilities, and diverse transfection efficiencies. One of the most widely used systems is the AAV-based drug delivery (Alexander and Hauswirth 2008; Buch et al. 2008; Grieger and Samulski 2005; Martin et al. 2002; Surace and Auricchio 2003, 2008), which is characterized by numerous successes in past decade.

### Nano-gel

Microgels are known as intra-molecularly crosslinked macromolecules (ICM), which are formed through polymerization of precursors. Conceivably, the nanogels are polymerized from nano-scale precursors. Nano-level gels and colloids were widely adopted for drug delivery and tissue engineering. In some cases the nanogels contain nanoparticles, and are considered as a manner of nanoparticle delivery. Here we focus on nanofiber-formed nanogels, especially on nanoscaffolds. One of the most important nanogels is the self-assembly nanoscaffolds hydrogel.

Molecular self-assembly is a strategy in nanofabrication. The ideal molecules were designed and produced; under certain conditions, these molecules could aggregate into desired structures due to the molecular forces and shape-complementarity. The known weak interacting forces included Van der Waals, capillary,  $\pi$ - $\pi$ , and hydrogen bonds. And the individual molecules would follow the order during self-assembly processes.

One example would be the self-assembly peptide nanofiber scaffolds. Because peptides are synthesized from amino acids, the degradation of the peptide-based scaffolds would lead to no immune response and side effects (Kopecek and Yang 2009; Zhao et al. 2010). Moreover, these reported self-assembly peptide nanofiber scaffolds are with high water content, mimicking the natural extracellular matrix, and have been shown to be useful in 3D cell culture, tissue repair, and regeneration (Collier et al. 2010; Ellis-Behnke et al. 2006a, b; Guo et al. 2007; Kopecek and Yang 2009; Ling et al. 2011; Zhao et al. 2010). One interesting application is to engineer the peptide

molecules with side chains of pharmaceutical effects (Zhao et al. 2010), and the hydrogel during biological degradation will guide the drug release reactions. The other way is to mix drug particles, including nanoparticles, with the nanogels before administration. The attachment/interaction of drug particles to specific chains and bases in the nanogels could elongate the drug effect for a long period, and the drugs could be retained in the desired regions to treat.

Other than the peptide-based nanogels, many other types of nanomaterials in gel form were developed, including the fabrication of some microgels into nanoscale, such as the nano-collagen fibers. The acquirement of nano-scale characteristic could bring special characteristics to these bulk materials and remains to be one hot research topic currently.

### Summary

In conclusion, the newly emerging nanotechnology has largely facilitated the drug delivery processes in many aspects of pharmacology. In the area of cytotechnology, nanoparticles and nanogels provide excellent testing tools to examine the efficiency and effectiveness of drugs, as well as potential toxicity in a high throughput manner. In the future, there will be more nanomaterials available for such applications.

### Reference

- Alexander JJ, Hauswirth WW (2008) Adeno-associated viral vectors and the retina. *Adv Exp Med Biol* 613:121–128
- Al-Ghananeem AM, Saeed H, Florence R, Yokel RA, Malkawi AH (2010) Intranasal drug delivery of didanosine-loaded chitosan nanoparticles for brain targeting; an attractive route against infections caused by aids viruses. *J Drug Target* 18:381–388
- Ali J, Ali M, Baboota S, Sahani JK, Ramassamy C, Dao L, Bhavna (2010) Potential of nanoparticulate drug delivery systems by intranasal administration. *Curr Pharm Des* 16(14):1644–1653
- Betbeder D, Spérandio S, Latapie JP, de Nadai J, Etienne A, Zajac JM, Francés B (2000) Biovector nanoparticles improve antinociceptive efficacy of nasal morphine. *Pharm Res* 17:743–748
- Blasi P, Schoubben A, Giovagnoli S, Rossi C, Ricci M (2009) Lipid nanoparticles for drug delivery to the brain: in vivo veritas. *J Biomed Nanotechnol* 5:344–350
- Bok D (2004) Gene therapy of retinal dystrophies: achievements, challenges and prospects. *Novartis Found Symp* 255:4–12 discussion -6, 177–178

- Buch PK, Bainbridge JW, Ali RR (2008) AAV-mediated gene therapy for retinal disorders: from mouse to man. *Gene Ther* 15:849–857
- Caruso G, Raudino G, Caffo M, Alafaci C, Granata F, Lucerna S, Salpietro FM, Tomasello F (2010) Nanotechnology platforms in diagnosis and treatment of primary brain tumors. *Recent Pat Nanotechnol* 4(2):119–124
- Charlton S, Jones NS, Davis SS, Illum L (2007) Distribution and clearance of bioadhesive formulations from the olfactory region in man: effect of polymer type and nasal delivery device. *Eur J Pharm Sci* 30:295–302
- Chekhonin VP, Baklaushev VP, Iusubalieva GM (2009) Prospects for targeted therapy for gliomas. *Vestn Ross Akad Med Nauk* (4):30–42
- Collier JH, Rudra JS, Gasiorowski JZ, Jung JP (2010) Multi-component extracellular matrices based on peptide self-assembly. *Chem Soc Rev* 39:3413–3424
- Denora N, Trapani A, Laquintana V, Lopodota A, Trapani G (2009) Recent advances in medicinal chemistry and pharmaceutical technology—strategies for drug delivery to the brain. *Curr Top Med Chem* 9:182–196
- Ellis-Behnke RG, Liang YX, Tay DK, Kau PW, Schneider GE, Zhang S, Wu W, So KF (2006a) Nano hemostat solution: immediate hemostasis at the nanoscale. *Nanomedicine* 2:207–215
- Ellis-Behnke RG, Liang YX, You SW, Tay DK, Zhang S, So KF, Schneider GE (2006b) Nano neuro knitting: peptide nanofiber scaffold for brain repair and axon regeneration with functional return of vision. *Proc Natl Acad Sci USA* 103:5054–5059
- Fisher RS, Ho J (2002) Potential new methods for antiepileptic drug delivery. *CNS Drugs* 16:579–593
- Fonseca SB, Pereira MP, Kelley SO (2009) Recent advances in the use of cell-penetrating peptides for medical and biological applications. *Adv Drug Deliv Rev* 61:953–964
- Gilmore JL, Yi X, Quan L, Kabanov AV (2008) Novel nanomaterials for clinical neuroscience. *J Neuroimmune Pharmacol* 3:83–94
- Graf A, McDowell A, Rades T (2009) Poly(alkylcyanoacrylate) nanoparticles for enhanced delivery of therapeutics—is there real potential? *Expert Opin Drug Deliv* 6:371–387
- Grieger JC, Samulski RJ (2005) Adeno-associated virus as a gene therapy vector: vector development, production and clinical applications. *Adv Biochem Eng Biotechnol* 99:119–145
- Guo J, Su H, Zeng Y, Liang YX, Wong WM, Ellis-Behnke RG, So KF, Wu W (2007) Reknitting the injured spinal cord by self-assembling peptide nanofiber scaffold. *Nanomedicine* 3:311–321
- Heilbronn R, Weger S (2010) Viral vectors for gene transfer: current status of gene therapeutics. *Handb Exp Pharmacol* (197):143–170
- Huynh NT, Passirani C, Saulnier P, Benoit JP (2009) Lipid nanocapsules: a new platform for nanomedicine. *Int J Pharm* 379:201–209
- Jatariu A, Peptu C, Popa M, Indrei A (2009) Micro- and nanoparticles—medical applications. *Rev Med Chir Soc Med Nat Iasi* 113:1160–1169
- Jogani VV, Shah PJ, Mishra P, Mishra AK, Misra AR (2008) Intranasal mucoadhesive microemulsion of tacrine to improve brain targeting. *Alzheimer Dis Assoc Disord* 22:116–124
- Kateb B, Chiu K, Black KL, Yamamoto V, Khalsa B, Ljubimova JY, Ding H, Patil R, Portilla-Arias JA, Modo M, Moore DF, Farahani K, Okun MS, Prakash N, Neman J, Ahdoot D, Grundfest W, Nikzad S, Heiss JD (2010) Nanoplatfoms for constructing new approaches to cancer treatment, imaging, and drug delivery: what should be the policy? *Neuroimage* 54(Suppl 1):S106–S124
- Kaur IP, Bhandari R, Bhandari S, Kakkar V (2008) Potential of solid lipid nanoparticles in brain targeting. *J Control Release* 127:97–109
- Khalil NM, Mainardes RM (2009) Colloidal polymeric nanoparticles and brain drug delivery. *Curr Drug Deliv* 6:261–273
- Kopecek J, Yang J (2009) Peptide-directed self-assembly of hydrogels. *Acta Biomater* 5:805–816
- Kumar M, Misra A, Pathak K (2009) Formulation and characterization of nanoemulsion of olanzapine for intranasal delivery. *PDA J Pharm Sci Technol* 63:501–511
- Ling PM, Cheung SW, Tay DK, Ellis-Behnke RG (2011) Using self-assembled nanomaterials to inhibit the formation of metastatic cancer stem cell colonies in vitro. *Cell Transplant* 20:127–131
- Ljubimova JY, Fujita M, Ljubimov AV, Torchilin VP, Black KL, Holler E (2008) Poly(malic acid) nanoconjugates containing various antibodies and oligonucleotides for multitargeting drug delivery. *Nanomedicine (Lond)* 3:247–265
- Martin KR, Klein RL, Quigley HA (2002) Gene delivery to the eye using adeno-associated viral vectors. *Methods* 28:267–275
- Mistry A, Glud SZ, Kjems J, Randel J, Howard KA, Stolnik S, Illum L (2009a) Effect of physicochemical properties on intranasal nanoparticle transit into murine olfactory epithelium. *J Drug Target* 17:543–552
- Mistry A, Stolnik S, Illum L (2009b) Nanoparticles for direct nose-to-brain delivery of drugs. *Int J Pharm* 379:146–157
- Modi G, Pillay V, Choonara YE (2010) Advances in the treatment of neurodegenerative disorders employing nanotechnology. *Ann N Y Acad Sci* 1184:154–172
- Namdeo M, Saxena S, Tankhiwale R, Bajpai M, Mohan YM, Bajpai SK (2008) Magnetic nanoparticles for drug delivery applications. *J Nanosci Nanotechnol* 8:3247–3271
- Nochi T, Yuki Y, Takahashi H, Sawada S, Mejima M, Kohda T, Harada N, Kong IG, Sato A, Kataoka N, Tokuhara D, Kurokawa S, Takahashi Y, Tsukada H, Kozaki S, Akiyoshi K, Kiyono H (2010) Nanogel antigenic protein-delivery system for adjuvant-free intranasal vaccines. *Nat Mater* 9:572–578
- Oberdörster G, Sharp Z, Atudorei V, Elder A, Gelein R, Kreyling W, Cox C (2004) Translocation of inhaled ultrafine particles to the brain. *Inhal Toxicol* 16:437–445
- Patel MM, Goyal BR, Bhadada SV, Bhatt JS, Amin AF (2009) Getting into the brain: approaches to enhance brain drug delivery. *CNS Drugs* 23:35–58
- Poeschla EM (2003) Non-primate lentiviral vectors. *Curr Opin Mol Ther* 5:529–540
- Provenzale JM, Silva GA (2009) Uses of nanoparticles for central nervous system imaging and therapy. *AJNR Am J Neuroradiol* 30:1293–1301
- Shilpi S, Jain A, Gupta Y, Jain SK (2007) Colloidosomes: an emerging vesicular system in drug delivery. *Crit Rev Ther Drug Carrier Syst* 24:361–391

- Sleeper MM, Bish LT, Sweeney HL (2009) Gene therapy in large animal models of human cardiovascular genetic disease. *ILAR J* 50:199–205
- Surace EM, Auricchio A (2003) Adeno-associated viral vectors for retinal gene transfer. *Prog Retin Eye Res* 22:705–719
- Surace EM, Auricchio A (2008) Versatility of AAV vectors for retinal gene transfer. *Vision Res* 48:353–359
- Suri SS, Fenniri H, Singh B (2007) Nanotechnology-based drug delivery systems. *J Occup Med Toxicol* 2:16
- Tosi G, Costantino L, Ruozi B, Forni F, Vandelli MA (2008) Polymeric nanoparticles for the drug delivery to the central nervous system. *Expert Opin Drug Deliv* 5:155–174
- Wong HL, Chattopadhyay N, Wu XY, Bendayan R (2010) Nanotechnology applications for improved delivery of antiretroviral drugs to the brain. *Adv Drug Deliv Rev* 62:503–517
- Zhao X, Pan F, Xu H, Yaseen M, Shan H, Hauser CA, Zhang S, Lu JR (2010) Molecular self-assembly and applications of designer peptide amphiphiles. *Chem Soc Rev* 39:3480–3498