

# Functional brain abnormalities in young adults at genetic risk for late-onset Alzheimer's dementia

Eric M. Reiman<sup>a,b,c,d,e</sup>, Kewei Chen<sup>a,d,f,g</sup>, Gene E. Alexander<sup>d,h</sup>, Richard J. Caselli<sup>d,i</sup>, Daniel Bandy<sup>a,d</sup>, David Osborne<sup>d,j</sup>, Ann M. Saunders<sup>k,l</sup>, and John Hardy<sup>m,n</sup>

<sup>a</sup>Positron Emission Tomography Center, Banner Good Samaritan Medical Center, Phoenix, AZ 85006; Departments of <sup>b</sup>Psychiatry and <sup>f</sup>Radiology, University of Arizona, Tucson, AZ 85724; Departments of <sup>g</sup>Mathematics and <sup>h</sup>Psychology, Arizona State University, Tempe, AZ 85287; Departments of <sup>i</sup>Neurology and <sup>j</sup>Psychology, Mayo Clinic, Scottsdale, AZ 85259; <sup>k</sup>Department of Medicine (Neurology), Joseph and Kathleen Bryan Alzheimer's Disease Research Center, Duke University, Durham, NC 27710; <sup>m</sup>Department of Neuroscience, Mayo Clinic, Jacksonville, FL 32224; <sup>n</sup>Translational Genomics Research Institute, Phoenix, AZ 85004; and <sup>d</sup>Arizona Alzheimer's Disease Consortium, 1111 East McDowell Road, Phoenix, AZ 85006

Edited by Marcus E. Raichle, Washington University School of Medicine, St. Louis, MO, and approved October 29, 2003 (received for review September 12, 2003)

Fluorodeoxyglucose positron emission tomography (PET) studies have found that patients with Alzheimer's dementia (AD) have abnormally low rates of cerebral glucose metabolism in posterior cingulate, parietal, temporal, and prefrontal cortex. We previously found that cognitively normal, late-middle-aged carriers of the apolipoprotein E  $\epsilon 4$  allele, a common susceptibility gene for late-onset Alzheimer's dementia, have abnormally low rates of glucose metabolism in the same brain regions as patients with probable AD. We now consider whether  $\epsilon 4$  carriers have these regional brain abnormalities as relatively young adults. Apolipoprotein E genotypes were established in normal volunteers 20–39 years of age. Clinical ratings, neuropsychological tests, magnetic resonance imaging, and PET were performed in 12  $\epsilon 4$  heterozygotes, all with the  $\epsilon 3/\epsilon 4$  genotype, and 15 noncarriers of the  $\epsilon 4$  allele, 12 of whom were individually matched for sex, age, and educational level. An automated algorithm was used to generate an aggregate surface-projection map that compared regional PET measurements in the two groups. The young adult  $\epsilon 4$  carriers and noncarriers did not differ significantly in their sex, age, educational level, clinical ratings, or neuropsychological test scores. Like previously studied patients with probable AD and late-middle-aged  $\epsilon 4$  carriers, the young  $\epsilon 4$  carriers had abnormally low rates of glucose metabolism bilaterally in the posterior cingulate, parietal, temporal, and prefrontal cortex. Carriers of a common Alzheimer's susceptibility gene have functional brain abnormalities in young adulthood, several decades before the possible onset of dementia.

apolipoprotein E | positron emission tomography | glucose metabolism | brain mapping | surrogate markers

Alzheimer's dementia (AD) afflicts  $\approx 10\%$  of those over the age of 65 and almost half of those over the age of 85 (1). To develop and test effective primary prevention therapies, it would be helpful to characterize brain changes associated with the susceptibility to AD as early as possible before the onset of cognitive impairment (2).

Fluorodeoxyglucose positron emission tomography (PET) studies have found that patients with AD have abnormally low cerebral metabolic rates for glucose (CMRgl) in posterior cingulate, parietal, temporal, and prefrontal cortex and a progressive decline in these rates over time (3–8). We (2, 9, 10) and others (11, 12) have been using PET to detect and track these functional brain abnormalities before the onset of dementia in carriers of the apolipoprotein E (APOE)  $\epsilon 4$  allele, a common Alzheimer's susceptibility gene associated with up to half of cases of late-onset AD (13–15). Previously, we found that cognitively normal 50- to 65-year-old  $\epsilon 4$  carriers have abnormally low CMRgl in each of the same regions as patients with probable AD and abnormal rates of regional CMRgl decline over time (2, 9, 10). We now consider whether  $\epsilon 4$  carriers have these regional brain abnormalities as relatively young adults, several decades before the possible onset of dementia. Our analysis of  $\epsilon 4$  carriers was restricted to those with the  $\epsilon 3/\epsilon 4$  genotype, found

in  $\approx 20\text{--}23\%$  of Caucasian populations (16) and  $\approx 11\text{--}36\%$  of different ethnic groups (17).

## Methods

**Subjects.** We used newspaper advertisements to recruit 135 20- to 39-year-old normal volunteers, who agreed that they would not be given information about their APOE genotype, provided their informed consent, and were studied under guidelines approved by human subjects committees at Banner Good Samaritan Medical Center and the Mayo Clinic (Rochester, MN). Venous blood samples were drawn, and APOE genotypes were characterized with analysis involving restriction fragment length polymorphisms (18).

Twelve  $\epsilon 4$  heterozygotes and 15 control subjects without the  $\epsilon 4$  allele satisfied our eligibility criteria and participated in our brain imaging studies. Thirteen of the control subjects had the  $\epsilon 3/\epsilon 3$  genotype, and 2 had the  $\epsilon 2/\epsilon 3$  genotype; 12 were individually matched to the  $\epsilon 4$  carriers for sex, age (within 3 years), and educational level (within 2 years), and 3 were matched to the  $\epsilon 4$  carriers for their mean age and educational level. Subjects denied an impairment in memory or other cognitive skills, did not satisfy criteria for a current psychiatric disorder, did not use centrally acting medications for at least 2 weeks before their PET session, and had a normal neurological examination. One subject in each group reported a history of hypertension. Investigators who were unaware of the subjects' APOE genotypes obtained data from medical and family histories, a neurologic examination, and a structured psychiatric interview (19). The subjects completed the Folstein modified Mini-Mental State Examination (20) and the Hamilton Depression Rating Scale (21), and all but one control subject completed a battery of neuropsychological tests (22).

**Brain Imaging.** Volumetric  $T_1$ -weighted magnetic resonance imaging and fluorodeoxyglucose PET were performed as described (2, 9, 23). PET was performed with the 951/31 ECAT scanner (Siemens, Knoxville, TN), a transmission scan, the i.v. injection of 10 mCi of  $^{18}\text{F}$ -fluorodeoxyglucose, and a 60-min dynamic sequence of emission scans as the subjects, who had fasted for at least 4 h, lay quietly in a darkened room with their eyes closed and directed forward. For whole brain measurements, CMRgl (mg/min per 100 g) was calculated by using the PET images, an image-derived radiotracer input function, plasma glucose levels, and a graphic

This paper was submitted directly (Track II) to the PNAS office.

Abbreviations: PET, positron emission tomography; AD, Alzheimer's dementia; CMRgl, cerebral metabolic rate for glucose; APOE, apolipoprotein E.

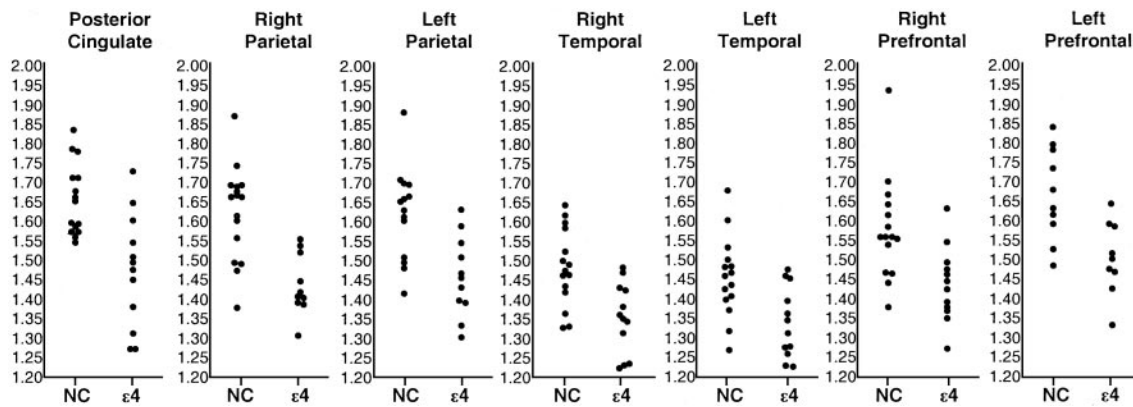
<sup>e</sup>To whom correspondence should be addressed. E-mail: eric.reiman@bannerhealth.com.

<sup>l</sup>Present address: GlaxoSmithKline R&D, Research Triangle Park, NC 27709.

<sup>n</sup>Present address: Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, MD 20892.

© 2003 by The National Academy of Sciences of the USA





**Fig. 2.** Regional/pontine CMRgl in young adult APOE  $\epsilon 4$  carriers and controls. Measurements were normalized to that in the pons, which appears to be the region least affected in patients with the probable AD (25), and extracted from the locations specified in Table 3, which were associated with the most significant differences between the  $\epsilon 4$  carriers ( $\epsilon 4$ ) and controls who do not carry the  $\epsilon 4$  allele (NC) ( $P < 0.001$ , uncorrected for multiple comparisons).

homozygotes (9) and heterozygotes (10), the group of young adult  $\epsilon 4$  carriers had abnormally low CMRgl in the posterior cingulate, parietal, temporal, and prefrontal cortex (Fig. 1 and Table 3). With the exception of the right temporal cortex, these abnormalities remained significant after correction for multiple comparisons (Table 3). Whereas  $\epsilon 4$  carriers may have lower CMRgl in other cortical regions ( $P < 0.001$ , uncorrected for multiple comparisons, Fig. 1), these additional metabolic reductions need to be confirmed in an independent comparison. For each of the locations specified in Table 3, the mean CMRgl was 8.0–10.5% lower in the  $\epsilon 4$  carriers than in the controls. These reductions were smaller than those previously reported in patients with probable AD (6), and there was considerable overlap between the  $\epsilon 4$  carrier and noncarrier groups' individual measurements (Fig. 2). The  $\epsilon 4$  carriers did not have abnormally high CMRgl, even at the more liberal significance level of  $P < 0.05$ , uncorrected for multiple comparisons.

In a post hoc analysis, the subjects' coregistered MRI and PET images, an automated brain mapping algorithm that permitted us to investigate the effects of partial volume averaging on PET measurements in each voxel (SPM99, Wellcome Department of Cognitive Neurology, University College, London) (28) and an algorithm intended to account for these effects (29) were used to generate and compare statistical brain maps of metabolic differences between the two subject groups with and without correction for individual differences in brain tissue volume. Because the pattern of metabolic differences between the two groups remained present after correction for partial volume averaging, and was not significantly different from the pattern identified without this correction, the functional brain abnormalities observed in

the young adult  $\epsilon 4$  carriers could not be solely attributed to alterations in the regional volume of brain tissue.

In another post hoc analysis, PET images from the 20- to 39-year-old subjects and previously studied 50- to 65-year-old subjects (10) were used to characterize and compare the effects of age on regional CMRgl in cognitively normal  $\epsilon 4$  heterozygotes and controls. Overall, the  $\epsilon 4$  carriers had significantly lower CMRgl than the controls in the posterior cingulate, parietal, temporal, and prefrontal cortex, and older age was significantly correlated with lower CMRgl in extensive areas of cerebral cortex, most prominently in the prefrontal and anterior cingulate cortex ( $P < 0.001$ , uncorrected for multiple comparisons). With possible exceptions in localized areas of the left prefrontal and posterior cingulate cortex ( $P < 0.005$ , uncorrected for multiple comparisons), the  $\epsilon 4$  carriers did not have significantly steeper age-related CMRgl declines than those in the  $\epsilon 4$  noncarriers. Thus, whereas 2-year follow-up studies have demonstrated abnormally high rates of regional CMRgl decline in older  $\epsilon 4$  carriers before the onset of cognitive impairment (2, 12), this cross-sectional comparison did not demonstrate abnormally steep posterior cingulate, parietal, or temporal CMRgl declines in  $\epsilon 4$  carriers between young adulthood (when the abnormalities are already present) and late middle age.

## Discussion

We previously found that cognitively normal late-middle-aged carriers of the APOE  $\epsilon 4$  allele, a common AD susceptibility gene, have functional brain abnormalities in the same brain regions as patients with probable AD. We now find that cognitively normal  $\epsilon 4$  carriers have functional brain abnormalities as relatively young adults, several decades before the

**Table 1. Characteristics of the young adult  $\epsilon 4$  carriers and control subjects**

Characteristic	Carriers, $n = 12$	Controls, $n = 15$	$P$ value
Age, yr	30.7 $\pm$ 5.4	31.2 $\pm$ 5.0	0.77*
Sex, female/male	9/3	12/3	0.28†
Handedness right, yes/no	12/0	14/1	>0.99†
Years of education	16.0 $\pm$ 1.7	16.1 $\pm$ 1.5	0.95*
Reported family history			
Any first-degree relative, yes/no	5/7	1/14	0.09†
Any first- or second-degree relative, yes/no	6/6	5/10	0.63†
Score on Hamilton Depression Scale	1.0 $\pm$ 1.9	1.1 $\pm$ 1.8	0.92*
Score on Mini-Mental State Examination	29.9 $\pm$ 0.3	29.9 $\pm$ 0.3	0.88*

Plus-minus values are means  $\pm$  SD.

\*The  $P$  value was calculated with an unpaired two-tailed  $t$  test.

†The  $P$  value was calculated with Pearson  $\chi^2$  two-tailed test, with continuity correction for  $n < 5$ .



**Table 2. Neuropsychological scores in young  $\epsilon 4$  heterozygotes and control subjects**

Test	$\epsilon 4$ Carriers, <i>n</i> = 12	Controls, <i>n</i> = 14	<i>P</i> value
	Mean ( $\pm$ SD) score		
AVLT			
Total learning	51.9 $\pm$ 6.7	52.6 $\pm$ 8.5	0.83
Short-term recall	11.2 $\pm$ 2.6	11.8 $\pm$ 2.5	0.54
Long-term recall	10.6 $\pm$ 3.1	11.7 $\pm$ 2.2	0.29
Complex figure test			
Copy	35.6 $\pm$ 1.0	35.4 $\pm$ 0.9	0.67
Recall	20.3 $\pm$ 5.6	21.0 $\pm$ 7.6	0.77
Boston naming test	54.4 $\pm$ 3.2	56.1 $\pm$ 2.7	0.15
WAIS-R			
Information	11.6 $\pm$ 1.5	11.0 $\pm$ 2.4	0.48
Digit span	10.8 $\pm$ 1.5	10.9 $\pm$ 3.3	0.93
Block design	11.3 $\pm$ 3.0	12.1 $\pm$ 2.5	0.46
Mental arithmetic	11.6 $\pm$ 2.2	11.2 $\pm$ 2.5	0.69
Similarities	11.6 $\pm$ 2.1	11.5 $\pm$ 2.1	0.92
Controlled oral word association test	43.3 $\pm$ 8.7	44.4 $\pm$ 10.0	0.79
WMS-R Orientation Subtest	13.9 $\pm$ 0.3	13.9 $\pm$ 0.3	0.91

AVLT, Auditory Verbal Learning Test; WAIS-R, Wechsler Adult Intelligence Scale-Revised; WMS-R, Wechsler Memory Scale-Revised. The *P* value was calculated with unpaired two-tailed *t* tests, uncorrected for multiple comparisons.

possible onset of dementia. These abnormalities were detected even though the young adult  $\epsilon 4$  carriers were restricted to persons with one copy of this allele, who tend to have a lower risk of AD and a later age of onset of dementia than those with two copies of this allele  $\epsilon 4$ , even though at least half of  $\epsilon 4$  heterozygotes are unlikely to develop AD, and even though the average age of onset of dementia in those  $\epsilon 4$  heterozygotes who develop the disorder appears to be in the seventh decade of life, about four decades older than the average age of subjects in this study (13–15).

In separate studies, we have found abnormally low CMRgl bilaterally in the posterior cingulate, parietal, temporal, and prefrontal cortex in patients with AD (8), cognitively normal  $\epsilon 4$  homozygotes and heterozygotes at 50–65 years of age (9, 10), and cognitively normal  $\epsilon 4$  heterozygotes at 20–39 years of age at the time of their baseline scans. As noted (9, 10), the CMRgl abnormalities could reflect reductions in the activity or density of terminal neuronal fields that innervate the implicated regions (30), the activity of synaptic glial cells (31), an impairment in glucose metabolism unrelated to local neuronal activity (32, 33), or a combination of these factors. As in patients with probable AD (29), the CMRgl abnormalities

in the young  $\epsilon 4$  carriers could not be solely attributed to alterations in brain volume.

The CMRgl reductions in our young adult  $\epsilon 4$  carriers may be the earliest brain abnormalities yet found in living persons at risk for late-onset AD. In postmortem studies, the characteristic histopathological features of AD include neuritic plaques, neurofibrillary tangles, and a loss of synapses. Whereas the type, density, and topographic distribution of this histopathology is likely to be extremely limited in normal young adults, postmortem studies have raised the possibility that the initial stage of AD histopathology [the selective involvement of neurofibrillary tangles in transentorhinal cortex (34, 35)] may be present several decades before the onset of dementia. In a study of young adults (mean age 38, range 22–46 years), this initial histopathological stage was observed in a higher frequency of persons with the  $\epsilon 3/\epsilon 4$  genotype than in controls with no copies of the  $\epsilon 4$  allele (36). Estimating the longitudinal progression of AD histopathology from cross-sectional data, a related study of persons 40–90 years of age suggested that this initial histopathological stage may precede AD by  $\approx$ 50 years (37). MRI studies have found significantly smaller hippocampal volumes in patients with AD and mild cognitive impair-

**Table 3. Location and magnitude of most significant reductions CMRgl**

Brain region	Atlas coordinates,* mm			Percent reduction	<i>T</i> value <sup>†</sup>	Corrected <i>P</i> value <sup>‡</sup>
	X	Y	Z			
Posterior cingulate	2	–32	25	10.5	3.77	0.0080
Right parietal	52	–53	47	9.8	3.41	0.048
Left parietal	–45	–50	52	10.0	3.63	0.049
Right temporal	65	–14	–7	8.6	3.48	0.033
Left temporal	–63	–37	7	8.0	3.05	0.13
Right prefrontal	45	42	29	8.9	3.12	0.010
Left prefrontal	–34	11	59	9.8	3.30	0.015

The reductions were identified by an automated search of posterior cingulate, parietal, temporal, and prefrontal cortex, regions previously found to be affected in patients with AD.

\*The coordinates were obtained from Talairach and Tournoux (25). *x* is the distance in mm to the right (+) or left (–) of the midline, *y* is the distance anterior (+) or posterior (–) to the anterior commissure, and *z* is the distance superior (+) or inferior (–) to a horizontal plane through the anterior and posterior commissures.

<sup>†</sup>*P* < 0.001 before correction for multiple comparisons.

<sup>‡</sup>Significance levels were corrected for multiple comparisons in each of the regions of interest previously associated with CMRgl reductions in the patients with probable AD. Details of the correction procedure, which included 20,000 Monte Carlo simulations, are available on request.

ment, correlations between reduced hippocampal volume and the severity of cognitive impairment, and progressive declines in hippocampal volume during the course of the illness (10, 23). Based on our study of late-middle-aged  $\epsilon 4$  homozygotes and controls and a review of the MRI literature, we suggested that the reduction in posterior cingulate CMRgl is apparent before the onset of memory decline in persons at risk for AD and that hippocampal volumes begin to decline some time later, in conjunction with the onset of memory decline and shortly before the onset of dementia (23).

The causal connections, if any, between the CMRgl abnormalities and the histopathological features of AD remain to be clarified. For instance, it is possible that the CMRgl abnormalities are attributable to AD histopathology. In a PET study of baboons, neurotoxic lesions of entorhinal and perirhinal cortex were associated with reduced rates of glucose metabolism in posterior cingulate, parietal, and temporal cortex (38), raising the possibility that the PET abnormalities in these regions reflect a reduction in the activity or density of projections arising in the vicinity of entorhinal cortex, the location of earliest and most extensive AD-related histopathology (34, 39). Still, it remains to be shown that the CMRgl abnormalities could be attributed to the very limited histopathology that may or may not be present in the cognitively normal young adult  $\epsilon 4$  carriers. Alternatively, it is possible that the CMRgl abnormalities found in young adult  $\epsilon 4$  carriers increase vulnerability to AD histopathology. Because there may be some correspondence between the cortical association areas preferentially affected metabolically and histopathologically in AD (40, 41), our findings raise the possibility that functional alterations provide a foothold for the subsequent onset of neuropathology in brain regions that are preferentially vulnerable to this disorder. If so, it may be possible to identify neurobiological processes that are involved in the predisposition to AD and precede the onset of previously known neuropathology, providing particularly early targets for a prevention therapy.

The functional brain abnormalities observed in the young adult  $\epsilon 4$  carriers could reflect a very early age-related decline in CMRgl or an abnormality in prenatal or early postnatal neurological development. In comparison with the other isoforms, the E4 isoform of APOE has been associated with higher cholesterol levels, increased aggregation of amyloid, less protection against amyloid-induced oxidative neurotoxicity, less efficient repair of neurons and synapses, less protection against the hyperphosphorylation of the microtubule-associated protein tau, the formation of neurofibrillary tangles, and a reduction in the outgrowth of neurons (13, 16, 42–45). Although any of these processes could have a role in the development of AD, some could have an additional role in neurological development.

Although it remains possible that the CMRgl abnormalities reflect aspects of the  $\epsilon 4$  allele unrelated to AD, PET studies suggest that these abnormalities are related to the development of this disorder. Although there may be some differences (46, 47), patients with probable AD had a similar pattern of reductions in regional CMRgl whether or not they had the  $\epsilon 4$  allele (48, 49). In patients with probable AD, the CMRgl abnormalities predicted the subsequent progression of dementia and the histopathological diagnosis of AD (7), were progressive (3–5, 8), and were correlated with dementia severity (6). In older  $\epsilon 4$  carriers who did not have dementia, CMRgl continued to decline in these and other brain regions and did so at higher rates than in control subjects who did not have this allele (2, 12). In those  $\epsilon 4$  carriers who had memory concerns, some of the metabolic abnormalities predicted a subsequent decline in memory (12). Our ongoing longitudinal PET study of late-middle-aged, cognitively normal  $\epsilon 4$  carriers, and studies in patients with mild cognitive impairment who have an increased annual risk of AD

(50), promise to clarify the extent to which the PET abnormalities are related to the risk of AD.

Although some studies do not detect a significant effect of early life factors (51, 52), it has been suggested that early life factors may contribute to the risk of late-onset AD (53–56). In a sample of nuns, those with lower measures of linguistic ability at a mean age of 22 years had lower cognitive function, a higher frequency of neuropathologically confirmed AD, and a higher density of neurofibrillary tangles in the hippocampus and cortex when they were assessed about six decades later (53). In a sample of Scottish residents, those with lower scores on a test of mental ability at the age of 11 had a higher frequency of late-onset dementia by the age of 75 (54). In several studies, lower educational levels were associated with an increased risk of AD (55, 56). The relationship between lower levels of intelligence and education and an increased risk of AD could be attributable to very early neurological alterations that lead to each of these conditions, differential cognitive reserve capacities in neuropathologically affected or unaffected brain regions (57), possible neuroprotective effects of cognitive stimulation (58), or a combination of these and other contributory factors.

In a post hoc cross-sectional analysis, PET images from our young adults and previously studied late-middle-aged subjects (10) were used to characterize and compare the effects of age on regional CMRgl in cognitively normal  $\epsilon 4$  heterozygotes and controls. Consistent with previous studies (9, 59–63), the late-middle-aged adults had lower CMRgl than young adults in extensive areas of the cerebral cortex, most prominently in the medial frontal lobe. In contrast to 2-year follow-up findings that regional CMRgl continues to decline at an abnormally high rate in older  $\epsilon 4$  carriers with and without memory concerns (2, 12), the CMRgl differences between young adulthood and late middle age were not significantly greater in the  $\epsilon 4$  carriers than in the controls. Although there may be other explanations to reconcile these findings, we have previously postulated that the rates of age-related CMRgl decline in different regions vary depending on a person's age, AD risk and stage, and their interactions (10). Longitudinal studies are needed to further characterize the course of CMRgl declines across the adult lifespan, the extent to which they are accelerated in  $\epsilon 4$  carriers and other persons at risk for AD, and the extent to which baseline and longitudinal CMRgl declines predict the subsequent onset of cognitive impairment and dementia.

This study has several limitations. For instance, because subjects in this study were selected by using newspaper advertisements, were mostly Caucasian and female, and had relatively high educational levels, additional studies are needed to determine whether our findings can be confirmed in young adult  $\epsilon 4$  carriers with other demographic features. Although CMRgl abnormalities have been reported in some of the same brain regions in relatives of patients with an early-onset, autosomal dominant form of AD (64) and a patient with mild cognitive impairment who subsequently developed AD (65), additional studies are needed to determine whether the same pattern of functional brain abnormalities is related to the risk of AD in persons with no copies of the  $\epsilon 4$  allele. Finally, whereas the  $\epsilon 4$  carriers and control subjects had significant differences in their regional PET measurements, there was considerable overlap between groups in their individual measurements. Neither genetic testing for APOE alleles nor PET are clinically indicated to predict a cognitively normal person's risk of AD. This information does not yet determine with sufficient accuracy whether or when a person might develop AD; this information may be associated with psychological and social risks; and established prevention therapies are not yet available (66).

If, as we believe, the functional brain abnormalities observed in young adult  $\epsilon 4$  carriers are associated with the risk of late-onset AD,

it may be possible to discover primary prevention therapies that target the contributing processes at an unusually early age, decades before the onset of cognitive decline, and perhaps at a more tractable stage of disease or level of susceptibility.

We thank Anita Prouty, Christine Burns, Sandra Yee-Benedetto, Heather Wheeler, Debra Intorcchia, Sandra Goodwin, Leslie Mullen, Susan Poulton, Dr. Wavrant-DeVrieze Fabienne, Louis Giordano, Alisa Domb, and Bernadette Romo for technical assistance; Dr. David Kuhl

for permission to use PET data from the University of Michigan, Ann Arbor; Dr. Satoshi Minoshima for permission to use image-analysis software; and Dr. Michael Lawson and Connie Boker for their encouragement. This study was supported by Alzheimer's Association Grants IIRG-98-068 (to E.M.R.) and IIRG-98-078 (to R.J.C.), the Arizona Alzheimer's Research Center (E.M.R. and R.J.C.), National Institute of Mental Health and National Institute on Aging Grants RO1 MH57899-01 and P30 AG19610-02 (to E.M.R.), the Banner Health Foundation, and the Mayo Clinic Foundation.

- Evans, D. A., Funkenstein, H. H., Albert, M. S., Scherr, P. A., Cook, N. R., Chown, M. J., Hebert, L. E., Hennekens, C. H. & Taylor, J. O. (1989) *J. Am. Med. Assoc.* **262**, 2551–2556.
- Reiman, E. M., Caselli, R. J., Chen, K., Alexander, G. E., Bandy, D. & Frost, J. (2001) *Proc. Natl. Acad. Sci. USA* **98**, 3334–3339.
- McGeer, E. G., Peppard, R. P., McGeer, P. L., Tuokko, H., Crockett, D., Parks, R., Akiyama, H., Calne, D. B., Beattie, B. L. & Harrop, R. (1990) *Can. J. Neurol. Sci.* **17**, 1–11.
- Smith, G. S., de Leon, M. J., George, A. E., Kluger, A., Volkow, N. D., McRae, T., Golomb, J., Ferris, S. H., Reisberg, B., Ciaravino, J., et al. (1992) *Arch. Neurol.* **49**, 1142–1150.
- Mielke, R., Herholz, K. & Grond, M. (1994) *Dementia* **5**, 36–41.
- Minoshima, S., Frey, K. A., Koeppe, R. A., Foster, N. L. & Kuhl, D. E. (1995) *J. Nucl. Med.* **36**, 1238–1248.
- Silverman, D. H., Small, G. W., Chang, C. Y., Lu, C. S., Kung De Aburto, M. A., Chen, W., Czernin, J., Rapoport, S. I., Pietrini, P., Alexander, G. E., et al. (2001) *J. Am. Med. Assoc.* **286**, 2120–2127.
- Alexander, G. E., Chen, K., Pietrini, P., Rapoport, S. & Reiman, E. M. (2002) *Am. J. Psychiatry* **159**, 738–745.
- Reiman, E. M., Caselli, R. J., Yun, L. S., Chen, K., Bandy, D., Minoshima, S., Thibodeau, S. N. & Osborne, D. (1996) *N. Engl. J. Med.* **334**, 752–758.
- Reiman, E. M., Caselli, R. J., Alexander, G. E. & Chen, K. (2001) *Clin. Neurosci. Res.* **1**, 194–206.
- Small, G. W., Mazziotta, J. C., Collins, M. T., Baxter, L. R., Phelps, M. E., Mandelkern, M. A., Kaplan, A., La Rue, A., Adamson, C. F., Chang, L., et al. (1995) *J. Am. Med. Assoc.* **273**, 942–947.
- Small, G. W., Ercoli, L. M., Silverman, D. H., Huang, S. C., Komo, S., Bookheimer, S. Y., Lavretsky, H., Miller, K., Siddarth, P., Rasgon, N. L., et al. (2000) *Proc. Natl. Acad. Sci. USA* **97**, 6037–6042.
- Strittmatter, W. J., Saunders, A. M., Schmechel, D., Pericak-Vance, M., Enghild, J., Salvesen, G. S. & Roses, A. D. (1993) *Proc. Natl. Acad. Sci. USA* **90**, 1977–1981.
- Corder, E. H., Saunders, A. M., Strittmatter, W. J., Schmechel, D. E., Gaskell, P. C., Small, G. W., Roses, A. D., Haines, J. L. & Pericak-Vance, M. A. (1993) *Science* **261**, 921–923.
- Saunders, A. M., Strittmatter, W. J., Schmechel, D., George-Hyslop, P. H., Pericak-Vance, M. A., Joo, S. H., Rosi, B. L., Gusella, J. F., Crapper-MacLachlan, D. R., Alberts, M. J., et al. (1993) *Neurology* **43**, 1467–1472.
- Mahley, R. W. (1988) *Science* **240**, 622–630.
- Hallman, D. M., Boerwinkle, E., Saha, N., Sandholzer, C., Menzel, H. J., Csazar, A. & Utermann, G. (1991) *Am. J. Hum. Genet.* **49**, 338–349.
- Hixson, J. E. & Vernier, D. T. (1990) *J. Lipid Res.* **31**, 545–548.
- Spitzer, R. L., Williams, J. B. W., Gibbon, M. & First, M. D. (1990) *User's Guide for the Structured Clinical Interview for DSM-III-R (SCID)* (Am. Psychiatric Press, Washington, DC).
- Folstein, M. F., Folstein, S. E. & McHugh, P. R. (1975) *J. Psychiatr. Res.* **12**, 189–198.
- Hamilton, M. (1960) *J. Neurol. Neurosurg. Psychiatry* **23**, 56–62.
- Lezak, M. D. (1983) *Neuropsychological Assessment* (Oxford Univ. Press, New York), 2nd Ed.
- Reiman, E. M., Uecker, A., Caselli, R. J., Lewis, S., Bandy, D., de Leon, M. J., De Santi, S., Convit, A., Osborne, D., Weaver, A., et al. (1998) *Ann. Neurol.* **44**, 288–291.
- Chen, K., Bandy, D., Reiman, E., Huang, S. C., Lawson, M., Feng, D., Yun, L. S. & Palant, A. (1998) *J. Cereb. Blood Flow Metab.* **18**, 716–723.
- Talairach, J. & Tournoux, P. (1988) *Coplanar Stereotaxic Atlas of the Human Brain* (Thieme, New York).
- Minoshima, S., Frey, K. A., Foster, N. L. & Kuhl, D. E. (1995) *J. Comput. Assist. Tomogr.* **19**, 541–547.
- Chen, K. C., Reiman, E. M., Bandy, D. & Alexander, G. E. (2003) in *Modeling and Control in Biomedical Systems*, eds. Feng, D. D. & Carson, E. R. (Elsevier, Hong Kong), pp. 11–15.
- Frackowiak, R. S. J., Friston, K. J., Frith, C. D., Dolan, R. J. & Mazziotta, J. C. (1997) *Human Brain Function* (Academic, San Diego).
- Ibanez, V., Pietrini, P., Alexander, G. E., Furey, M. L., Teichberg, D., Rajapakse, J. C., Rapoport, S. I., Schapiro, M. B. & Horwitz, B. (1998) *Neurology* **50**, 1585–1593.
- Schwartz, W. J., Smith, C. B., Davidsen, L., Savaki, H., Sokoloff, L., Mata, M., Fink, D. J. & Gainer, H. (1979) *Science* **205**, 723–725.
- Magistretti, P. J. & Pellerin, L. (1996) *Cereb. Cortex* **6**, 50–61.
- Piert, M., Koeppe, R. A., Giordani, B., Berent, S. & Kuhl, D. E. (1996) *J. Nucl. Med.* **37**, 201–208.
- Mark, R. J., Pang, Z., Geddes, J. W., Uchida, K. & Mattson, M. P. (1997) *J. Neurosci.* **17**, 1046–1054.
- Braak, H. & Braak, E. (1991) *Acta Neuropathol.* **82**, 239–259.
- Braak, H. & Braak, E. (1997) *Neurobiol. Aging* **18**, 351–357.
- Ghebremedhin, E., Schultz, C., Braak, E. & Braak, H. (1998) *Exp. Neurol.* **153**, 152–155.
- Ohm, T. G., Müller, H., Braak, H. & Bohl, J. (1994) *Neuroscience* **64**, 209–217.
- Meguro, K., Blaizot, X., Kondoh, Y., Le Mestric, C., Baron, J. C. & Chavoix, C. (1999) *Brain* **122**, 1519–1531.
- Hyman, B. T., Van Hoesen, G. W., Kromer, L. J. & Damasio, A. R. (1986) *Ann. Neurol.* **20**, 472–481.
- Shoghi-Jadid, K., Small, G. W., Agdeppa, E. D., Kepe, V., Ercoli, L. M., Siddarth, P., Read, S., Satyamurthy, N., Petric, A., Huang, S. C., et al. (2002) *Am. J. Geriatr. Psychiatry* **10**, 24–35.
- Mathis, C. A., Bacskai, B. J., Kajdasz, S. T., McLellan, M. E., Frosch, M. P., Hyman, B. T., Holt, D. P., Wang, Y., Huang, G. F., Debnath, M. L., et al. (2002) *Bioorg. Med. Chem. Lett.* **12**, 295–298.
- Wisniewski, T., Castano, E. M., Golabek, A., Vogel, T. & Frangione, B. (1994) *Am. J. Pathol.* **145**, 1030–1035.
- Miyata, M. & Smith, J. D. (1996) *Nat. Genet.* **14**, 55–61.
- Strittmatter, W. J., Weisgraber, K. H., Goedert, M., Saunders, A. M., Huang, D., Corder, E. H., Dong, L. M., Jakes, R., Alberts, M. J., Gilbert, J. R., et al. (1994) *Exp. Neurol.* **125**, 1663–1671.
- Nathan, B. P., Bellosa, S., Sanan, D. A., Weisgraber, K. H., Mahley, R. W. & Pitas, R. E. (1994) *Science* **264**, 850–852.
- Mielke, R., Zerres, K., Uhlhaas, S., Kessler, J. & Heiss, W. D. (1998) *Neurosci. Lett.* **254**, 49–52.
- Higuchi, M., Arai, H., Nakagawa, T., Higuchi, S., Muramatsu, T., Matsushita, S., Kosaka, Y., Itoh, M. & Sasaki, H. (1997) *NeuroReport* **8**, 2639–2643.
- Corder, E. H., Jelic, V., Basun, H., Lannfelt, L., Valind, S., Winblad, B. & Nordberg, A. (1997) *Arch. Neurol.* **54**, 273–277.
- Hirono, N., Mori, E., Yasuda, M., Ishii, K., Ikejiri, Y., Imamura, T., Shimomura, T., Hashimoto, M., Yamashita, H. & Sasaki, M. (1998) *Alzheimer Dis. Assoc. Disord.* **12**, 362–367.
- Petersen, R. C., Smith, G. E., Ivnik, R. J., Tangalos, E. G., Schaid, D. J., Thibodeau, S. N., Kokmen, E., Waring, S. C. & Kurland, L. T. (1995) *J. Am. Med. Assoc.* **273**, 1274–1278.
- Munoz, D. G., Ganapathy, G. R., Eliasziw, M. & Hachinski, V. (2000) *Arch. Neurol.* **57**, 85–89.
- Caselli, R. J., Hentz, J. G., Osborne, D., Graff-Radford, N. R., Barbieri, C. J., Alexander, G. E., Hall, G. R., Reiman, E. M., Hardy, J. & Saunders, A. M. (2002) *J. Am. Geriatr. Soc.* **50**, 49–54.
- Snowdon, D. A., Kemper, S. J., Mortimer, J. A., Greiner, L. H., Wekstein, D. R. & Markesbery, W. R. (1996) *J. Am. Med. Assoc.* **275**, 528–532.
- Whalley, L. J., Starr, J. M., Athawes, R., Hunter, D., Pattie, A. & Deary, I. J. (2000) *Neurology* **55**, 1455–1460.
- Evans, D. A., Beckett, L. A., Albert, M. S., Hebert, L. E., Scherr, P. A., Funkenstein, H. H. & Taylor, J. O. (1993) *Ann. Epidemiol.* **3**, 71–77.
- Stern, Y., Gurland, B., Tatemichi, T. K., Tang, M. X., Wilder, D. & Mayeux, R. (1994) *J. Am. Med. Assoc.* **271**, 1004–1010.
- Alexander, G. E., Furey, M. L., Grady, C. L., Pietrini, P., Brady, D. R., Mentis, M. J. & Schapiro, M. B. (1997) *Am. J. Psychiatry* **154**, 165–172.
- Kesslak, J. P., So, V., Choi, J., Cotman, C. W. & Gomez-Pinilla, F. (1998) *Behav. Neurosci.* **112**, 1012–1019.
- Kuhl, D. E., Metter, E. J., Riege, W. H. & Phelps, M. E. (1982) *J. Cereb. Blood Flow Metab.* **2**, 163–171.
- Salmon, E., Maquet, P., Sadzot, B., Degueldre, C., Lemaire, C. & Franck G. (1991) *Acta Neurol. Belg.* **91**, 288–295.
- Loessner, A., Alavi, A., Lewandowski, K. U., Mozley, D., Souder, E. & Gur, R. E. (1995) *J. Nucl. Med.* **36**, 1141–1149.
- Coffey, C. E., Wilkinson, W. E., Parashos, I. A., Soady, S. A., Sullivan, R. J., Patterson, L. J., Figiel, G. S., Webb, M. C., Spritzer, C. E. & Djang, W. T. (1992) *Neurology* **42**, 527–536.
- Terry, R. D., DeTeresa, R. & Hansen, L. A. (1976) *Arch. Psychiatr. Nervenkr.* **223**, 15–33.
- Kennedy, A. M., Frackowiak, R. S., Newman, S. K., Bloomfield, P. M., Seaward, J., Roques, P., Lewington, G., Cunningham, V. J. & Rossor, M. N. (1995) *Neurosci. Lett.* **186**, 17–20.
- Pietrini, P., Azari, N. P., Grady, C. L., Salerno, J. A., Gonzales-Aviles, A., Heston, L. L., Pettigrew, K. D., Horwitz, B., Haxby, J. V. & Schapiro, M. B. (1993) *Dementia* **4**, 94–101.
- American College of Medical Genetics, American Society of Human Genetics Working Group on ApoE and Alzheimer Disease (1995) *J. Am. Med. Assoc.* **274**, 1627–1629.