

## Video Article

# Non-radioactive *in situ* Hybridization Protocol Applicable for Norway Spruce and a Range of Plant Species

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## Abstract

The high-throughput expression analysis technologies available today give scientists an overflow of expression profiles but their resolution in terms of tissue specific expression is limited because of problems in dissecting individual tissues. Expression data needs to be confirmed and complemented with expression patterns using e.g. *in situ* hybridization, a technique used to localize cell specific mRNA expression. The *in situ* hybridization method is laborious, time-consuming and often requires extensive optimization depending on species and tissue. *In situ* experiments are relatively more difficult to perform in woody species such as the conifer Norway spruce (*Picea abies*). Here we present a modified DIG *in situ* hybridization protocol, which is fast and applicable on a wide range of plant species including *P. abies*. With just a few adjustments, including altered RNase treatment and proteinase K concentration, we could use the protocol to study tissue specific expression of homologous genes in male reproductive organs of one gymnosperm and two angiosperm species; *P. abies*, *Arabidopsis thaliana* and *Brassica napus*. The protocol worked equally well for the species and genes studied. *AtAP3* and *BnAP3* were observed in second and third whorl floral organs in *A. thaliana* and *B. napus* and DAL13 in microsporophylls of male cones from *P. abies*. For *P. abies* the proteinase K concentration, used to permeabilize the tissues, had to be increased to 3 g/ml instead of 1 g/ml, possibly due to more compact tissues and higher levels of phenolics and polysaccharides. For all species the RNase treatment was removed due to reduced signal strength without a corresponding increase in specificity. By comparing tissue specific expression patterns of homologous genes from both flowering plants and a coniferous tree we demonstrate that the DIG *in situ* protocol presented here, with only minute adjustments, can be applied to a wide range of plant species. Hence, the protocol avoids both extensive species specific optimization and the laborious use of radioactively labeled probes in favor of DIG labeled probes. We have chosen to illustrate the technically demanding steps of the protocol in our film.

Anna Karlgren and Jenny Carlsson contributed equally to this study.

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## Video Link

The video component of this article can be found at <http://www.jove.com/video/1205/>

## Protocol

This non-radioactive mRNA *in situ* hybridization protocol is optimized for Norway spruce tissues and described in detail. It is based on an Arabidopsis/rapeseed *in situ* hybridization protocol optimized in our groups, which in turn is based on protocols from the Meyerowitz and Irish labs and optimized by Vivian Irish, Cindy Lincoln and Jeff Long among others<sup>1-4</sup>.

## PROCEDURE

### 1. FIXATION AND EMBEDDING

To provide RNA retention tissues need to be fixated and embedded in wax before an *in situ* experiment is performed. The fixation and embedding is a critical step since poorly fixed materials might give a low or undetectable signal even though the mRNA is highly abundant. The tissues are dehydrated, cleared and embedded in wax in gradual changes to avoid tissue damage. To further avoid shrinkage and swelling of the cells 0.85% NaCl is added to the first ethanol steps in the dehydration of Norway spruce tissues. It is possible to leave out the NaCl in the ethanol (e.g. it is left out in the Arabidopsis/rapeseed protocol). The dehydration step during embedding may be performed on ice to keep RNase activity at minimum or at +4°C. In this protocol the 4% paraformaldehyde fix solution contains 0.25% glutaraldehyde, which is supposed to give a better RNA retention even though that some argue that it probably makes no difference (e.g. it is left out in the Arabidopsis/rapeseed protocol).

Most likely any standard fixation and embedding protocol can be used. As seen below the Norway spruce embedding differs slightly from the Arabidopsis/rapeseed protocol.

**1.1 Day 1 Collecting plant material and paraformaldehyde fixation**

1. Collect the plant material of interest and dissect it if necessary. Keep the tissues on ice and place them in ice-cold fix solution (see recipes below) directly or as soon as possible. *N.B.! Keep on ice all the time!*
2. Apply vacuum to the samples until the paraformaldehyde starts to bubble. Hold the vacuum for up to three hours (e.g. 2 x 15 min for Arabidopsis and rapeseed floral tissues or one hour for Norway spruce male cones) and release the vacuum slowly. The tissues are supposed to sink after the vacuum infiltration of the fixative, but for certain tissues containing a lot of air it is not possible (e.g. Arabidopsis flowers). A piece of gauze can be used to make the tissues stay in the fixative.
3. Replace the fixative and incubate at +4°C overnight (~12-16 hours) gently shaking.

**Option 1**

*N.B.! Norway spruce embedding version below (steps 4-8). Make sure scintillation tubes or glass vials are used, at least in step 5.*

**1.2 Day 2 Dehydration N.B.! Norway spruce embedding version.**

4.	0.85% NaCl	30 min	on ice
	50% EtOH + 0.85% NaCl	90 min	on ice
	70% EtOH + 0.85% NaCl	90 min	on ice

*Pause! It is possible to stop here and store the tissues in 70% EtOH + 0.85% NaCl at +4°C for several months.*

	85% EtOH + 0.85% NaCl	90 min	+4°C
	95% EtOH (+ eosin)	90 min	+4°C

*N.B.! Eosin is added to stain the tissues pink making them easier to detect during embedding and sectioning but it may be excluded.*

	100% EtOH (+ eosin)	90 min	+4°C
	100% EtOH (+ eosin)	overnight	+4°C

**1.3 Day 3 Dehydration and embedding N.B.! Norway spruce embedding version.**

5.	100 % EtOH (+ eosin)	2 h	room temperature
	50% EtOH + 50% HistoClear II	1 h	room temperature
	100% HistoClear II	1 h	room temperature
	100% HistoClear II	1 h	room temperature
	50% HistoClear II + 50% Histowax	overnight	+40-50°C

*N.B.! In the last step the concentration of Histowax is not crucial, just pour in some wax chips.*

**1.4 Day 4 Embedding N.B.! Norway spruce embedding version.**

6.	100 % melted Histowax	+60°C	(Change morning and evening)
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**1.5 Day 5 Embedding N.B.! Norway spruce embedding version.**

7.	100 % melted Histowax	+60°C	(Change morning and evening)
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**1.6 Day 6 Embedding N.B.! Norway spruce embedding version.**

8.	100 % melted Histowax	+60°C	(Change morning and evening)
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*N.B.! It is ok if the wax occasionally is changed once a day, but then it takes two days to complete one day of changes. The wax change can also continue for two extra days without a problem. This is recommended for large samples since it helps the infiltration of the wax into the center of these tissues.*

**Option 2**

*N.B.! Arabidopsis/rapeseed embedding version below (steps 4-8).*

**1.2 Day 2 Dehydration N.B.! Arabidopsis/rapeseed embedding version.**

4. *N.B.! All steps at +4°C and gently shaking, if not else is stated.*

1x PBS	30 min
1x PBS	30 min
30% EtOH	60 min
40% EtOH	60 min
50% EtOH	60 min
60% EtOH	60 min
70% EtOH	60 min

*Pause! It is possible to stop here and store the tissues in 70% EtOH at +4°C for several months.*

85% EtOH	60 min
95% EtOH (+ eosin)	overnight

*N.B.! Eosin is added to stain the tissues pink making them easier to detect during embedding and sectioning but it may be excluded.*

**1.3 Day 3 Dehydration and embedding N.B.! Arabidopsis/rapeseed embedding version.**

5. *N.B.! All steps at room temperature and gently shaking, if not else is stated.*

100% EtOH (+ eosin)	30 min
100% EtOH (+ eosin)	30 min
100% EtOH (+ eosin)	60 min
100% EtOH (+ eosin)	60 min

*N.B.! Transfer the tissue to scintillation vials or other (glass) vials.*

75% EtOH + 25% HistoClear II	30 min
50% EtOH + 50% HistoClear II	30 min
25% EtOH + 75% HistoClear II	30 min
100% HistoClear II	60 min
100% HistoClear II	60 min
100% HistoClear II + ¼ volumes Histowax chip	overnight (no shaking)

*N.B.! In the last step the concentration of Histowax is not crucial, just pour in some wax chips.*

**1.4 Day 4 Embedding N.B.! Arabidopsis/rapeseed embedding version.**

6. Place the vials with tissues at +42°C until the wax chips melt completely. Next add ¼ volume of wax chips and let them melt completely. Move to +60°C and leave the vials for several hours. Finally, replace wax/HistoClear II with freshly melted wax and incubate over night at +60°C.

**1.5 Day 5 Embedding N.B.! Arabidopsis/rapeseed embedding version.**

7.	100 % melted Histowax	+60°C	(Change morning and evening)
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**1.6 Day 6 Embedding N.B.! Arabidopsis/rapeseed embedding version.**

8.	100 % melted Histowax	+60°C	(Change morning and evening)
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*N.B.! In the Arabidopsis/rapeseed protocol (compared to the Norway spruce protocol) there is a Day 7 Embedding in the same way as in day 5-6, i.e. 100 % melted Histowax at +60°C and change morning and evening*

*N.B.! It is ok if the wax occasionally is changed once a day, but then it takes two days to complete one day of changes. The wax change can also continue for two extra days without a problem. This is recommended for large samples since it helps the infiltration of the wax into the center of these tissues.*

**1.7 Day 7 Embedding (Day 8 in the Arabidopsis/rapeseed protocol) (Film! Technical details are shown in the film)**

9. Preheat petri dishes and/or molds at +60°C. Turn on a hot plate (+60°C) and make sure melted Histowax is available.
  10. Pour the tissues and melted Histowax into a petri dish (or a mold) placed on the hot plate. Add more Histowax so that the tissues are covered. Heat a pair of tweezers and distribute the tissues evenly in the dish. Fill the petri dish (or mold). If the wax hardens before the tissues are in the right position, heat it up again or remove the hardened wax with the heated pair of tweezers.
  11. Let the wax block harden at room temperature before placing it at +4°C. It is better to make several wax blocks instead of embedding the tissues too tightly since the wax block might crack when pieces are cut out during sectioning.
- Pause! Store at +4°C. The tissues are stable for years.*

*N.B.! Work RNase free from this step and onwards. Use gloves, DEPC-treat MilliQ-water, buffers, glassware etc., autoclave buffers and bake glassware etc. when appropriate.*

## 2. SECTIONING ( *Film! Technical details are shown in the film* )

12. Turn on two hot plates (+60°C and +42°C) and make sure melted Histowax is available.
13. Heat a scalpel and carefully cut out a wax block of tissue from the petri dish (it might crack if one is too violent), or remove it from the mold. When a single wax block has been separated, shape the piece into the right size and shape to facilitate the slicing in the microtome.
14. The wax piece should have an extra wax "heel" underneath the tissue so it can be attached to the block holder. Heat the block holder and the heel on the hot plate (+60°C) and fuse them together. It might be necessary to put on a drop of Histowax II to make the fusion stable. Let it harden at +4°C.
15. Cut the wax block into 6-8 µm thick sections connected in a long ribbon using a microtome.  
*N.B.! This will be easier if the wax block is cold, i.e. store the wax block at +4°C and bring it from the fridge when starting the cutting. It can also be beneficial to keep the knife cold.*
16. Study the ribbons of sections in a magnifying glass and mark the ones to be used.
17. Put RNase-free MilliQ-water on the slides (Probe-On Plus from Fisher Biotechnology, which are pre-cleaned and charged). Cut the ribbon into smaller pieces and put them ("back" or "shiny" side down) on the slides. The slides with ribbons should be placed on a +42°C plate. The ribbon should flatten out (this is important!) in the water. Let the sections sit for a few minutes and then use a Kimwipe/tissue paper to drain of the water.
18. Leave the slides on the plate for 1-2 hours.
19. Incubate the slides at +42°C overnight so that the tissues adhere to slides.  
*Pause! It is recommended to use the sections as soon as possible, but they can be stored dry in closed boxes with desiccant up to several days at +4°C.*

## 3. THE MAKING OF PROBES

*In situ* hybridization detects expression using a labeled probe specific for a certain mRNA. The probe, which is most often a piece of complementary RNA, can be tagged with digoxigenin, DIG. DIG is a small molecule, which can be attached to uridine and thereby incorporated in the RNA probe and then detected using DIG-specific antibodies.

The designing and making of the probe is the most critical step in a RNA *in situ* hybridization experiment. Care should be taken to make sure the probe has a high specificity and is labeled with high quality UTPs. Sometimes the hybridization temperature and/or reaction length have to be adjusted for specific probes. As always, care should be taken to avoid RNase contamination.

Before labeling, isolate a template according your standard protocols, use e.g. cDNA clones, PCR-fragments, and linearize the template. Sense and antisense fragments are isolated and amplified from the template using gene specific primers. For PCR amplified fragments do not use enzymes that produce 3' overhangs. To all fragments a T7 (or SP6 or T3) overhang is added using primers carrying the T7-sequence or using a vector with a T7-promoter.

The sense (mRNA or (-) strand) probe have the same sequence as the target mRNA and will not hybridize to the target mRNA, while the antisense (anti-mRNA or (+) strand) probe have the complementary sequence and thus hybridize to the target giving the signal of interest. The sense probe is used as a negative control and no signal will be obtained if the experiment worked properly. A positive control is also needed, e.g. a probe that detects a house-keeping gene. The majority of the slides should be hybridized with antisense probes, while only a few slides should be hybridized to the different control probes.

### 3.1 Probe labeling using *In vitro* transcription

Reagents from the DIG RNA Labeling Kit (SP6/T7; 11175025910, Roche Applied Science, Mannheim, Germany) or similar are used when labeling the probes. The 20 µl reaction described here should yield ~10 µg of DIG-labeled RNA.

20. Make an *in vitro* transcription to incorporate labeled nucleotides. Add the following reagents to a tube (keep on ice):

Ingredient	volume/amount
10 x NTP labeling mixture	2 µl
10 x Transcription buffer	2 µl
Protector RNase inhibitor	1 µl (20U)
RNA T7 polymerase (SP6 or T3)	2 µl
Template DNA (500-1000 ng)	x µl
RNase- free MilliQ-water	y µl, to a final volume of 18 µl

21. Mix the NTP labeling mixture, transcription buffer, RNase inhibitor ( *N.B.! if the probe is short, e.g. <500bp, you may increase the RNase inhibitor concentration to 40U* ), polymerase, template DNA and water and then briefly centrifuge.
22. Incubate the reaction mix at +37°C for 2 hours ( *N.B.! if the probe is short, e.g. <500bp, increase the reaction time to 4-6 hours* ).
23. Add 2 µl DNase I and incubate at +37°C for 15 min.
24. Stop the reaction by adding 2 µl 0.2 M EDTA (pH 8.0). Check the product on gel.
25. Precipitate the probe by adding 2.5 µl of 4 M LiCl and 75 µl of >99.5% ethanol. Mix well and incubate at -20°C overnight ( *N.B.! Do not use tRNA as carrier. Instead use glycogen or acrylamide according to the manufacturers* ).

26. Centrifuge (13 000 rpm) for 30 minutes at +4°C and remove the supernatant. Wash the pellet (at least 10 min 13 000 rpm) two times in 70% ethanol (~150-300 µl).
27. Remove the supernatant and let the pellet dry. Resuspend the probe in 30 µl RNase-free water for 1-2 hours on ice.  
*Pause! The probe can be stored at -20°C for more than a year.*  
*N.B.! Save 0.5 µl samples from step 21 (before the in vitro transcription), step 23 (before the DNase treatment), step 24 (after the DNase treatment, but before the precipitation) and 1 µl sample from step 27 (when the probe has been resuspended). Run the samples on a non-denaturing 1% TBE gel. If the DNase reaction has worked properly the template-band will disappear. The in vitro transcription should generate a thick RNA band of approximately the correct size. If you see multiple bands, denature the samples at +80°C for 5 minutes followed by quenching on ice before loading. If the recovery from step 27 looks bad it might be so because the probe was not resuspended well. Incubate the RNA at +55°C 5-10 minutes to get it into solution.*

### 3.2 Hydrolysis of long probes

*N.B.! If your probe is larger than 150 bp it should be reduced to 50-150 bp to give a high signal, due to better penetration. The probe may be chemically degraded in a carbonate buffer (pH 10.2). If the probe is of the correct size skip the hydrolysis step, but remember to add one volume formamide, i.e. 30 µl, to the probe.*

28. Prepare 0.2 M sodium bicarbonate (NaHCO<sub>3</sub>) and 0.2 M sodium carbonate (Na<sub>2</sub>CO<sub>3</sub>). Both should be fresh.
29. Test the two buffers by mixing 5 ml H<sub>2</sub>O, 2 ml 0.2 M NaHCO<sub>3</sub> and 3 ml 0.2 M Na<sub>2</sub>CO<sub>3</sub>. The pH should be ~10.2.
30. Calculate the incubation time:

Incubation time (in minutes) = $(L_0 - L_f) / (K * L_0 * L_f)$
L <sub>0</sub> = starting length of the probe in kb
L <sub>f</sub> = final length of the probe in kb = e.g. 0.100 kb
K = rate constant for hydrolysis = 0.11 kb <sup>-1</sup> min <sup>-1</sup>

31. Hydrolyze half of your probe, save the rest for later. Dilute probe to 50 µl with RNase free water and add 20 µl 0.2 M NaHCO<sub>3</sub> (add first) and 30 µl 0.2 M Na<sub>2</sub>CO<sub>3</sub>. Mix and incubate at +60°C for the calculated incubation time.
32. Stop the reaction by adding 10 µl of 1 M NaOAc pH 4.7.
33. Precipitate the probe by adding 10 µl 4 M LiCl<sub>2</sub> and 300 µl ethanol (*N.B.! Use glycogen or acrylamide as carrier, if needed*). Incubate at -20°C overnight.
34. Centrifuge (13 000 rpm) for 30 minutes at +4°C and remove the supernatant. Wash the pellet two times in 70% ethanol (~300 µl). Centrifuge (13 000 rpm) at least 10 min in each wash at +4°C.
35. Remove the supernatant and let the pellet dry. Resuspend the probe in 30 µl RNase-free MilliQ-water. Add one volume formamide, i.e. 30 µl, to the probe.  
*Pause! The probe can be stored at -20°C for more than a year.*  
*N.B.! Take 2 µl of the unhydrolyzed probe and 2 µl of the hydrolyzed probe (before the formamide is added) and check on a non-denaturing 1% TBE gel. There should be a difference in size between the two samples. The probes might need to be denatured at +80°C for 5 minutes followed by quenching on ice before loading.*

## 4. IN SITU HYBRIDIZATION ( Film! Technical details are shown in the film)

Make sure that all solutions are RNase free! It is not necessary to DEPC-treat everything, but use at least autoclaved MilliQ-water. Make sure that all glassware are heated at +200°C overnight or at least for 5 hours. Treat plastic containers and stir bars with 0.1 M NaOH with agitation overnight. Rinse the plastic containers several times with autoclaved MilliQ-water (~500 ml/container). It is crucial to rinse the containers carefully to avoid traces of NaOH that might disturb the hybridization. DEPC-treat all stock solutions, except the Tris-buffers. Check all solutions etc. before starting the experiment to ensure that everything is available. Until step 61 (except steps 51-52) we keep the slides in a rack, which is easily moved between containers.

### 4.1 In situ section pretreatment

36. Deparaffinize/dehydrate sections:

2x 10 min Histoclear II (use glass containers)
2x 1-2 min 100% EtOH
1-2 min 95% EtOH
1-2 min 90% EtOH
1-2 min 80% EtOH
1-2 min 60% EtOH
1-2 min 30% EtOH
1-2 min H <sub>2</sub> O

37. Wash the slides for 15-20 min in 2x SSC at room temp (*N.B.! Prepare the paraformaldehyde used in step 42 during the incubation time*).
38. Treat the tissues for 30 min with proteinase K at +37°C with gentle agitation (e.g. 1 µg/ml for Arabidopsis/rapeseed and 3 µg/ml for Norway spruce). Mix and preheat to +37°C 100 mM Tris pH 8 and 50 mM EDTA. Add proteinase K immediately before treating the slides. *N.B.! The*

*proteinase K treatment needs to be optimized depending on tissue, plant species and age. ( N.B.! Prepare the triethanolamine used in step 45 during the incubation time).*

39. Treat the slides for 2 min with 2mg/ml glycine in 1x PBS at room temperature. ( N.B.! Mix the glycine with PBS the day before using it to ensure that the glycine is properly dissolved).
40. Wash the slides for 2 minutes in 1x PBS at room temperature.
41. Wash the slides for 2 minutes in 1x PBS at room temperature.
42. Incubate the slides for 10 minutes with 4% (w/v) paraformaldehyde (made fresh) in 1x PBS pH 7 at room temperature. Use glass container.
43. Wash the slides for 5 minutes in 1x PBS at room temperature.
44. Wash the slides for 5 minutes in 1x PBS at room temperature. (Dispense acetic anhydride into the solution in step 45).
45. Incubate the slides for 10 minutes in 0.1M triethanolamine (made fresh, pH 8) and acetic anhydride at room temperature.
46. Wash the slides for 5 minutes in 1x PBS at room temperature.
47. Wash the slides for 5 minutes in 1x PBS at room temperature.
48. Dehydrate:

30 sec 30% EtOH
30 sec 60% EtOH
30 sec 80% EtOH
30 sec 90% EtOH
30 sec 95% EtOH
2x 30 sec 100% EtOH

*Pause! It is possible to stop here and store the slides in a container with a small amount of EtOH in the bottom at +4°C for several hours.*

#### 4.2 In situ hybridization ( Film! Technical details are shown in the film)

Decide which slides to use with which probes, do not forget to use both sense and antisense probes as well as a positive control. (N.B.! ProbeOn Plus slides from Fisher Biotechnology have a white frosting that allows them to be sandwiched into pairs during the hybridization/detection steps of the protocol.) The same probe with the same concentration should be used on a specific slide pair. Determine how much hybridization solution to make based on the total number of slide pairs. Preheat the Dextran sulfate in +60°C. The hybridization solution is very viscous from the Dextran sulfate. Put the hybridization solution in +60°C and it will be easier to handle. Air-dry the slides on clean paper towels or Kimwipe/ tissue papers before the probe is applied. The slides must be completely dry. For each pair of slides add the probe. It is important to test different probe concentrations (e.g. 0.5, 2 and 4 µl) to find the optimal concentration before the actual experiment is performed.

Hybridization solution	5 slide pairs	10 slide pairs	15 slide pairs	20 slide pairs
10x <i>in situ</i> salts	100 µl	200 µl	300 µl	400 µl
deionized formamide	400 µl	800 µl	1200 µl	1600 µl
50 % Dextran sulfate (+60 °C)	200 µl	400 µl	600 µl	800 µl
50x Denhardt's solution	20 µl	40 µl	60 µl	80 µl
tRNA (100mg/ml)	10 µl	20 µl	30 µl	40 µl
H <sub>2</sub> O (DEPC treated)	70 µl	140 µl	210 µl	280 µl
Total volume	800 µl	1600 µl	2400 µl	3200 µl

49. Dilute the probe in RNase-free MilliQ-water to a final volume of 20 µl. Add one volume (i.e. 20 µl) formamide to the probe giving a total volume of 40 µl. Heat the probe to 80°C for 2 minutes. Place on ice for 2 min. Spin down. Keep the probe on ice. This probe-mixture is enough for one pair of slides. Multiply according the total number of slide pairs for the specific probe.
50. Add 160 µl of hybridization solution for each pair of slides giving a final volume of 200 µl (i.e. 160 µl hybridization solution + 40 µl probe) for each slide pair. Mix without generating bubbles.
51. Apply the probe to one slide and slowly fuse the two slides together. (N.B.! Sandwiching can only be done with Probe-On Plus slides, or equivalent slides. If slides without frosting are used use cover slips instead of sandwiching.)
52. Elevate slides above wet paper towels in a plastic container (which seals tightly) using plastic pipettes. Hybridize +50-55°C overnight (12-16 hours).

#### 4.3 In situ post hybridization (Film! Technical details are shown in the film)

53. Prepare the *in situ* post hybridization by warming ~2 L 0.2x SSC (+55°C) and ~2 L 1x NTE (+37°C) overnight. More SSC and NTE are needed if the RNase treatment (steps 57-59 below) is performed.
54. Dip each slide pair into warm 0.2x SSC (see above) to separate and rinse them. Then place them in a rack in a container with warm 0.2x SSC.
55. Wash the slides for 60 min in 0.2x SSC with gentle agitation at +55°C. ( N.B.! Prepare the Boehringer block solution used in step 63 during the incubation time.)
56. Repeat the wash in 0.2x SSC for 60 min. ( N.B.! If the RNase step is performed let the RNase thaw during this incubation, if not continue to step 60.)
57. Optional: Wash the slides for 5 min in warm 1x NTE (see above) at +37°C with gentle agitation.
58. Optional: Repeat the wash in warm 1x NTE for 5 min at +37°C with gentle agtation.

59. Optional: Treat the slides with RNase (20 µg/ml RNase A in 1x NTE) for 30 min at +37°C with gentle agitation. Then wash for 5 min in 1x NTE at +37°C with gentle agitation. Repeat the 1x NTE wash.
60. Wash the slides for 60 min in 0.2x SSC at +55°C with gentle agitation. ( *N.B.!* Prepare the block solution used in steps 64-65 and 67 during the incubation time.)
61. Wash the slides for 5 min in 1x PBS at room temp (*Pause!* You may store slides in 1XPBS at +4°C overnight if you want to continue the next day.)
62. Place the slides on the bottom of a large plastic container with the sample side up.
63. Incubate the slides with 1% Boehringer block in 100 mM Tris pH 7.5, 150 mM NaCl for 45 min at room temperature with gentle agitation. Just cover the slides with the blocking solution. Pour off the blocking solution while the slides still are in the plastic container or place the slides in a new container. When pouring off the solution the slides usually adhere to the plastic container, but make sure that they do not fall out off the container.
64. Replace the Boehringer block solution with 1.0% BSA in 100 mM Tris pH 7.5, 150 mM NaCl, 0.3% Triton X-100. Perform the incubation as in step 63.
65. Dilute the anti-dig antibodies (1:1250, i.e. 8 µl antibodies in 10 ml BSA) in the BSA/Tris/NaCl/Triton solution from step 64. Make a puddle of antibody solution in a plastic weigh dish. Sandwich slides together to allow capillary forces to pull up the solution. Drain on Kimwipe/tissue paper and repeat, try to avoid bubbles.
66. Elevate slides above wet paper towels in plastic container and allow the slides to sit at room temp for 2 hours.
67. Drain slides on Kimwipe/tissue paper and separate. Place on bottom of plastic container as in step 63. Wash 4x in the BSA/Tris/NaCl/Triton solution. 15 min each time, gentle agitation at room temperature.
68. 10 min 100 mM Tris pH 9.5, 100 mM NaCl, 50 mM MgCl<sub>2</sub>. Place on bottom of plastic container as in step 63.
69. Dip slide in Tris pH 9.5/NaCl/MgCl<sub>2</sub> solution to ensure all of detergent is washed off.
70. Make sandwiches and draw up Western blue solution as in 65. Repeat once.
71. Place slides in plastic container above wet paper towels in total darkness (wrap in tin foil) for 1-5 days at room temperature. Check after 6, 12 and 24 hours then once every day.
72. Drain slides, rinse in 1xTE to stop reaction. Put on cover slips before the slides are examined in microscope. The slides can be saved in 1x TE for several months but take photos as soon as possible.

## RECIPES/STOCK SOLUTIONS

*N.B.!* Use RNase-free MilliQ-water as much as possible, e.g. DEPC-treat MilliQ-water (Add 0.1% DEPC. Leave overnight. Autoclave (20 minutes at 15 psi, i.e. 1.05 kg/cm<sup>2</sup>, on liquid cycle) or use at least autoclaved MilliQ-water. When preparing buffers and stock solutions it is also possible to dissolve RNase-free salts etc. in DEPC-treated water, instead of DEPC-treating the buffers and stock solutions themselves. If you do not DEPC-treat water, buffers and stock-solutions, at least autoclave or filter sterilize them.

*Pause!* Store buffers and stock-solutions at room temperature, if not else is stated.

*N.B.!* You can find many of the buffers as well as hints on how to make them in *Molecular Cloning* (Sambrook, J., and D.W. Russell. 2001. *Molecular Cloning A Laboratory Manual*. 3 ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, USA).

### Acetic anhydride in 0.1M triethanolamine (pH 8, volume: 800 ml).

Ingredient	volume/amount	final concentration
Triethanolamine	10.4 ml	0.1 M triethanolamine
MilliQ-water	786.4 ml	final volume 800 ml
HCl	3.2 ml	giving pH 8.0
Acetic anhydride	4.8 ml	0.6%

- To make 0.1 M triethanolamine buffer, pH 8.0, dilute 10.4 ml of triethanolamine in 786.4 ml of H<sub>2</sub>O and add 3.2 ml of HCl to bring the pH to 8.0 (check with pH indicator).
- Elevate slide rack with broken 10 ml plastic pipettes or similar in a container of triethanolamine with a stir bar in the bottom.
- Dispense 4.8 ml acetic anhydride into the triethanolamine a few minutes before putting slides in so that it is mixed well.

*N.B.!* Make the 0.1M triethanolamine fresh and add acetic anhydride just before incubation.

*N.B.!* Use glass container. Triethanolamine buffer has to be used because acetic anhydride is unstable in water.

### Agarose (1%) gel in 1 x TBE (volume: 100 ml)

Ingredient	volume/amount	final concentration
Agarose	1 g	1%
1 x TBE	up to 100 ml	

Mix agarose with 1 x TBE. Heat/boil until the agarose has melted. Cast a gel. Run the gel in a 1 x TBE buffer.

### Dextran sulfate

Dilute 5 g Dextran sulfate to 10 ml with DEPC-treated MilliQ-water (i.e. 50% (w/v)) and dissolve in 60°C.

*Pause!* Store at -20°C.

**0.5 M EDTA pH8.0 (titriplex III, volume 500 ml)**

Ingredient	volume/amount	final concentration
EDTA (372.24 g/mol)	93.06 g	0.5 M EDTA
MilliQ-water	up to 500 ml	
NaOH	~ 10 g pellets	giving pH 8.0
DEPC	0.5 ml	0.1% DEPC

Dissolve EDTA in water (happens at pH 8.0), adjust to a final volume of 500 ml. Add 0.1% DEPC. Leave over night. Autoclave.

**Fix solution/fixative (steps 1-3, 42) - 4% (w/v) paraformaldehyde in 1x PBS, pH 7.0 (650 ml)**

Ingredient	volume/amount	final concentration
MilliQ-water	585 ml	
10x PBS	65 ml	1x PBS
10M NaOH	520 µl	giving pH ~11
Paraformaldehyde	26 g	4%
Concentrated HCl	449 µl	giving pH ~7

- Mix water, PBS and NaOH and heat to +60-70°C (the temperature and the high pH will make it easier to dissolve the paraformaldehyde).
- Add (in fume hood) paraformaldehyde.
- When the solution has cleared place it on ice and adjust pH to 7.0 by adding 449 µl concentrated HCl.

N.B.! Make fresh.

N.B.! In the spruce protocol glutaraldehyde was added to a final concentration of 0.25%, i.e. 6.5 ml 25% glutaraldehyde to the total volume of 650 ml or 10 ml 25% glutaraldehyde to the total volume of 1000 ml (remember to reduce the water with an equal volume).

N.B.! Add a few drops of Tween 20 and/or Triton X-100, after the pH is adjusted, to reduce surface tension to facilitate the fixation in steps 1-3. DO NOT USE IN STEP 42!

N.B.! When fixing plant tissues a smaller volume (e.g. 100 - 200 ml) of fixative is generally needed.

**10x in situ salts (volume 100 ml)**

Ingredient	volume/amount	final concentration
5 M NaCl	60 ml	3M NaCl
1 M Tris-Cl pH 8.0	10 ml	0.100 M Tris
1 M Na Phosphate pH 6.8	10 ml	0.100 M Na Phosphate
0.5 M EDTA	10 ml	0.050 M EDTA
MilliQ-water	up to 100 ml	

Mix a Na Phosphate buffer with pH 6.8 (46.3 ml 1 M Na<sub>2</sub>HPO<sub>4</sub> + 53.7 ml 1 M NaH<sub>2</sub>PO<sub>4</sub>). Mix NaCl, Tris, Na phosphate buffer, EDTA and water. Autoclave. *Pause! Store at -20°C.*

**4 M LiCl<sub>2</sub> (lithium chloride, volume: 100 ml)**

Ingredient	volume/amount	final concentration
LiCl <sub>2</sub> (42.39 g/mol)	16.956 g	4M LiCl <sub>2</sub>
MilliQ-water	up to 100 ml	
DEPC	0.1 ml	0.1% DEPC

Dissolve LiCl<sub>2</sub> in water, adjust to a final volume of 100 ml. Add 0.1% DEPC. Leave over night. Autoclave. *Pause! Store at +4°C.*

**1 M MgCl<sub>2</sub> · H<sub>2</sub>O (magnesium chloride, volume: 100 ml)**

Ingredient	volume/amount	final concentration
MgCl <sub>2</sub> · H <sub>2</sub> O (203.30 g/mol)	20.33 g	1 M MgCl <sub>2</sub> · H <sub>2</sub> O
MilliQ-water	up to 100 ml	
DEPC	0.1 ml	0.1% DEPC

Dissolve MgCl<sub>2</sub> in water, adjust to a final volume of 100 ml. Add 0.1% DEPC. Leave over night. Autoclave.



*N.B.! MgCl<sub>2</sub> is very hygroscopic. Do not store opened bottles for long periods of time.*

**5 M NaCl (sodium chloride, volume: 1 l)**

Ingredient	volume/amount	final concentration
NaCl (58.44 g/mol)	292.2 g	5 M NaCl
MilliQ-water	up to 1l	
DEPC	1 ml	0.1% DEPC

Dissolve NaCl in water, adjust to a final volume of 1 l. Add 0.1% DEPC. Leave over night. Autoclave.

**1 M NaOAc pH 4.7 (sodium acetate, volume: 100 ml)**

Ingredient	volume/amount	final concentration
NaOAc (82.03 g/mol)	0.8203 g	1 M NaOAc
MilliQ-water	up to 100 ml	
Acetic acid		giving pH 4.7
DEPC	0.1 ml	0.1% DEPC

Dissolve NaOAc in water. Adjust the pH to 4.7. Adjust to a final volume of 100 ml. Add 0.1% DEPC. Leave over night. Autoclave.

**0.2 M Na<sub>2</sub>CO<sub>3</sub> pH11.4 (sodium carbonate, volume: 10 ml)**

Ingredient	volume/amount	final concentration
Na <sub>2</sub> CO <sub>3</sub>	0.21198 g	0.2 M Na <sub>2</sub> CO <sub>3</sub> pH=11.4
MilliQ-water	up to 10 ml	

Dissolve Na<sub>2</sub>CO<sub>3</sub> in water; adjust to a final volume of 10 ml. The pH should be 11.4.  
*N.B.! Make fresh.*

**0.2 M NaHCO<sub>3</sub> pH8.2 (sodium bicarbonate, volume: 10 ml)**

Ingredient	volume/amount	final concentration
NaHCO <sub>3</sub>	0.16802 g	0.2 M NaHCO <sub>3</sub> : pH=8.2
MilliQ-water	up to 10 ml	

Dissolve NaHCO<sub>3</sub> in water; adjust to a final volume of 10 ml. The pH should be 8.2.  
*N.B.! Make fresh.*

**1 M Na<sub>2</sub>HPO<sub>4</sub> · 2 H<sub>2</sub>O (volume 500 ml)**

Ingredient	volume/amount	final concentration
Na <sub>2</sub> HPO <sub>4</sub> · 2 H <sub>2</sub> O (177.99 g/mol)	88.995 g	1 M Na <sub>2</sub> HPO <sub>4</sub>
MilliQ-water	up to 500 ml	
DEPC	0.5 ml	0.1% DEPC

Dissolve Na<sub>2</sub>HPO<sub>4</sub> in water, adjust to a final volume of 500 ml. Add 0.1% DEPC. Leave over night. Autoclave.

**1 M NaH<sub>2</sub>PO<sub>4</sub> · H<sub>2</sub>O (volume 500 ml)**

Ingredient	volume/amount	final concentration
NaH <sub>2</sub> PO <sub>4</sub> · H <sub>2</sub> O (137.99 g/mol)	68.995 g	1 M NaH <sub>2</sub> PO <sub>4</sub>
MilliQ-water	up to 500 ml	
DEPC	0.5 ml	0.1% DEPC

Dissolve NaH<sub>2</sub>PO<sub>4</sub> in water, adjust to a final volume of 500 ml. Add 0.1% DEPC. Leave over night. Autoclave.

**5x NTE (total volume: 1l)**

Ingredient	volume/amount	final concentration
5 M NaCl	500 ml	2.5 M NaCl
1 M Tris-Cl pH 8	25 ml	50 mM Tris-Cl pH 8
0.5 M EDTA	5 ml	5 mM EDTA
MilliQ-water	470 ml	final volume 1l

Mix NaCl, Tris, EDTA and water. Do not DEPC-treat. Autoclave.

**10 x PBS (phosphate-buffered saline) pH 7.0 (total volume: 1l)**

Ingredient	volume/amount	final concentration
5 M NaCl	260 ml	1.3 M NaCl
1 M Na <sub>2</sub> HPO <sub>4</sub>	70 ml	70 mM Na <sub>2</sub> HPO <sub>4</sub>
1 M NaH <sub>2</sub> PO <sub>4</sub>	30 ml	30 mM NaH <sub>2</sub> PO <sub>4</sub>
MilliQ-water	640 ml	final volume 1l
DEPC	1 ml	0.1% DEPC

Mix NaCl, Na<sub>2</sub>HPO<sub>4</sub>, NaH<sub>2</sub>PO<sub>4</sub> and water. Adjust the pH to pH 7.0 with HCl. Add 0.1% DEPC. Leave over night. Autoclave.

**20x SSC pH 7.0 (total volume: 1l)**

Ingredient	volume/amount	final concentration
NaCl (58.44 g/mol)	175.32 g	3 M NaCl
Na Citrate (294.1 g/mol)	88.23 g	0.300 M Na Citrate
MilliQ-water	up to 1 l	
HCl		giving pH 7.5
DEPC	1 ml	0.1% DEPC

Dissolve NaCl and Na Citrate in ~ 800 ml water. Adjust the pH to pH 7.0 with HCl. Adjust the volume to 1 l with water. Add 0.1% DEPC. Leave over night. Autoclave.

**5 x TBE (volume: 1l)**

Ingredient	volume/amount	final concentration
Tris (121.14 g/mol)	54 g	~ 0.45 M Tris
Boric acid (61.83 g/mol)	27.5 g	~ 0.45 M boric acid
EDTA (372.24 g/mol)	3.7 g	~ 0.01 M EDTA
MilliQ-water	up to 1l	

Mix Tris, boric acid, EDTA and water. The pH should be ~ 8.3. Dilute to 1 x TBE before use.

**10 x TE pH 8.0 (Tris EDTA, volume: 1l)**

Ingredient	volume/amount	final concentration
1 M Tris-Cl, pH 8.0	100 ml	0.100 M Tris-Cl
0.5 M EDTA pH8.0	100 ml	0.050 M EDTA
MilliQ-water	up to 1l	

Mix Tris, EDTA and water. Do not DEPC-treat. Autoclave.

*N.B.! Tris should not be DEPC-treated! Use RNase-free powder and dilute with DEPC-treated/RNase-free MilliQ- water.*

1 M Tris-HCl pH 7.5 - 9.5 (volume 500 ml)

Ingredient	volume/amount	final concentration
Tris (121.14 g/mol)	60.57 g	1 M Tris
MilliQ-water	up to 500 ml	
HCl		giving pH 7.5
HCl		giving pH 8.0
NaOH		giving pH 9.5

Dissolve Tris in DEPC-treated water. Adjust to the desired pH. Adjust to a final volume of 500 ml. Autoclave.  
*N.B.! Tris should not be DEPC-treated! Use RNase-free powder and dilute with DEPC-treated/RNase-free MilliQ- water.*  
*N.B.! In Molecular Cloning (see above) there is a table describing how much concentrate HCl is needed to achieve the correct pH.*

MATERIAL AND METHODS

The *in situ* protocol is presented above and in the film. Here additional information of the plant material and probes used in this specific example are described.

Plant material and experimental design

Male cones from *Picea abies* [L.] Karst. (Norway spruce) were collected in Uppsala, Sweden, autumn 2007. Eight cones were sectioned and sections from each cone were used in each treatment. Three proteinase K concentrations were tested; 1, 3 and 5 µg/µl and each concentration were treated with or without RNase. Slides not treated with RNase were left in PBS during RNase treatment. *Arabidopsis thaliana* [L.] Heynh. and *Brassica napus* L., were grown under controlled conditions in a culture chamber with 22°C/18°C day/night temperatures and a photoperiod of 16 h. Young inflorescences containing floral buds, stages 0-10 (stages according to Smyth<sup>5</sup>) were studied.

cRNA probes

Total RNA was isolated from *P. abies* male cones as previously described<sup>6</sup> and from *A. thaliana* and *B. napus* using TRIzol (Gibco BRL, Frederick, Maryland, USA) and the Qiagen RNeasy Plant Mini Kit (Qiagen, Hilden, Germany), respectively, in accordance with manufacturers' recommendations. The cDNA was synthesized from 0.5-1 µg total RNA, depending on species, using Superscript III Reverse Transcriptase (Invitrogen, Carlsbad, California, USA) following the manufacturer's instructions. *AtAP3* and *BnAP3* sense and antisense fragments and *P. abies* sense fragments were isolated and amplified using gene specific primers (Table 1). *DAL13* antisense fragments were isolated and amplified using primers as in 7. A T7 overhang was added to fragments from *A. thaliana* and *P. abies* using modified reverse primers carrying the T7-sequence (Table 1). A 500 bp fragment was isolated using *BnAP3* specific primers (Table 1) and cloned into a pGEM-T vector with a T7-promotor. For the *P. abies* fragments all PCR reactions were performed in a final volume of 20 µl containing 0.2 U Phusion High-Fidelity DNA Polymerase (Finnzymes, Espoo, Finland), 1x Phusion HF buffer, 200 µM of each dNTP, 0.3 µM of each primer and 25-50 ng cDNA and a standard PCR program was used with annealing temperature 60-55°C (touch-down -1°C/cycle) followed by 55°C. Sense and antisense cRNA probes for all three species were synthesized with *in vitro* transcription using the DIG RNA Labeling Kit (SP6/T7; Roche Applied Science, Mannheim, Germany) according to the manufacturer's recommendations and long probes were digested to approximately 100-150 bp fragments using a Na<sub>2</sub>CO<sub>3</sub> buffer according to<sup>3</sup>.

RESULTS

Here in the result section we describe three different examples of typical results from *in situ* experiments.

The genetic mechanism that regulates reproductive development in gymnosperms and angiosperms by specifying male and female organ identity appears to be evolutionarily conserved<sup>7-9</sup> despite that the two seed plant lineages separated 285 million years ago<sup>10</sup>. In *A. thaliana* the floral homeotic genes *APETALA3* (*AP3*<sup>11</sup>) and *PISTILLATA* (*PI*<sup>12</sup>) are necessary and sufficient to specify male reproductive development within the context of a flower, and this function appears to be conserved within the angiosperm lineage<sup>13</sup>. In gymnosperms, which lack flowers and have their reproductive organs arranged in separate male and female cones (Fig 1A-C), genes homologous to *AP3* and *PI* are specifically expressed in the pollen bearing organs of the male cone, the microsporophylls<sup>7,14</sup>. Hence, a similar set of homologous genes in both angiosperms and gymnosperms defines the pollen-bearing organs.

Gene specific probes directed against *AtAP3* and *BnAP3*, respectively, gave signals from stage 3 in young floral buds in a region corresponding to whorl two and three in *A. thaliana* and *B. napus*. In later staged buds the signals were restricted to developing petals and stamens (Fig. 2A and B). These results are in agreement with previous studies<sup>11,15,16</sup>. Probes directed towards the *AP3* homologue in *P. abies*, *DAL13*, produced signals in the microsporophylls of *P. abies* male cones both before and after apical meristem termination (Fig. 2C and D), as expected from previous studies<sup>7,14</sup>. Sense probes gave no signal above background (Fig. 3A-B and data not shown). Taken together, these results demonstrate expression of *A. thaliana* *AP3* and its orthologs in *B. napus* and *P. abies* in developing male reproductive organs.

**Figure 1.** *A. thaliana* and *B. napus* flowers and pollen producing male cones of the conifer *P. abies* Pictures show inflorescences with floral buds and open flowers of *A.thaliana* and *B.napus* in **A** and **B** respectively. Note that angiosperm flowers apart from the sterile perianth harbors both

male and female reproductive organs; stamens and carpels. Shown in **C** is a twig of the gymnosperm *P. abies* with green vegetative needles and red reproductive male cones.

**Figure 2.** Expression of *A. thaliana* *AP3* and homologous genes in *B. napus* and *P. abies*. Micrographs show longitudinal sections of *A. thaliana* and *B. napus* inflorescences in **A** and **B**, respectively and *P. abies* male cones in **C** and **D**. Sections hybridized with antisense probes directed against *AtAP3* in **A** and *BnAP3* in **B** show signal in second and third whorl floral organs. Numbers indicates floral stages according to Smyth<sup>5</sup>. Antisense probe directed towards the *DAL13* gene gave signal in the pollen bearing microsporophylls of *P. abies* male cones, as shown in **C-D**. Examples of microsporophylls are indicated by arrows. Arrowheads in A-D point to hybridization signal, which appears as purple. In **C** and **D** phenolic compounds give brownish unspecific color to individual cells in the central pith. s, sepal; p, petal; st, stamen; c, carpel; ms, microsporophyll; pi, pith; pp, pre-pollen cells. Bar: 100  $\mu$ m.

**Figure 3.** *DAL13* expression in male cones from *P. abies*. All Micrographs (**A-H**), are showing longitudinal sections of male cones after apical meristem termination. Arrowheads point to cell types in which *DAL13* is expressed. Sections in **A** and **B** are hybridized with a sense control probe to determine background staining. Micrographs **C** to **H** are hybridized with a *DAL13* antisense probe. Sections from experiments with and without RNase treatment are shown in **D**, **F**, **H** and **C**, **E**, **G** respectively. Micrographs from experiments with 1  $\mu$ g/ml proteinase K are shown in **C** and **D**, 3  $\mu$ g/ml in **E** and **F**, and 5  $\mu$ g/ml in **G** and **H**. Bar: 100  $\mu$ m.

## Discussion

Two aspects of the *P. abies* protocol were optimized compared to the *A. thaliana/B. napus* protocol, RNase treatment and proteinase K concentration. The RNase removes background signals and thus increases the signal specificity<sup>17</sup>. The proteinase K treatment is needed to permeabilize the tissues so the probe can enter and hybridize to the RNA pool of interest. The *DAL13* antisense probe gave a higher signal without RNase-treatment (Fig. 3C, E, G) compared to sections treated with RNase for 30 min (Figure 2D, F and H). Since the RNase-treatment did not enhance the signal it was removed from the protocol. The proteinase K treatment were tested at three different concentrations; 1, 3 and 5  $\mu$ g/ml. In the case of *P. abies* a low or no signal was observed using 1  $\mu$ g/ml proteinase K (Fig. 3C-D). A strong signal was obtained with both 3  $\mu$ g/ml (Fig. 3E-F) and 5  $\mu$ g/ml (Fig. 3G-H) proteinase K. Since no apparent difference in signal strength was found when using 5  $\mu$ g/ml proteinase K compared to 3  $\mu$ g/ml, we chose to use 3  $\mu$ g/ml in our protocol. It is likely that the need of a higher proteinase K concentration in the *P. abies* male cones compared to the *A. thaliana* and *B. napus* floral buds is due to more compact tissue with higher levels of phenolics and polysaccharides.

During the fixation and embedding some differences between the *A. thaliana/B. napus* protocol and the *P. abies* protocol are evident. The tissue of interest is fixed to provide RNA retention and this is a critical step since poorly fixed materials might give a low or undetectable signal even though the mRNA is highly abundant. The tissue is dehydrated, cleared and embedded in wax in gradual changes to avoid tissue damage, and in some protocols 0.85% NaCl is added to the ethanol in the dehydration to further avoid shrinkage and swelling of the cells<sup>3</sup>. The dehydration step during embedding can be performed on ice to keep RNase activity at minimum. In the *P. abies* protocol the 4% paraformaldehyde fix solution contains 0.25% glutaraldehyde, which is supposed to give a better RNA retention<sup>17</sup> even though that some argue that it probably make no difference<sup>3</sup>. After sectioning the material is fixed onto slides and pretreated (i.e. dewaxed, rehydrated, permeabilized, treated to reduce non-specific binding of the probe and finally dehydrated) before the hybridization.

In the protocol presented here the whole detection procedure can be performed in ordinary laboratories, the signal develops usually within 1-3 days and the ratio between signal over background can continuously be monitored. The DIG-labeled probes usually give a more distinct signal as compared to radioactive labeled probes, are more stable and can be reused in consecutive experiments. On the other hand, the sensitivity is reduced compared to radioactive probes<sup>17</sup>. *In situ* hybridization detects expression using a labeled probe specific for a certain mRNA. Radio-labeling is a robust and sensitive labeling method but it requires handling of radioactivity and it takes several weeks before the results can be detected. Antigen-labeled probes on the other hand, are more stable, less hazardous and require exposure to the detection medium for a few days only<sup>17</sup>. Thus, antigen-labeling methods are currently dominating. The most critical step regardless of protocol is the probe design. Care should be taken to make sure the probe has a high specificity and is labeled with high quality UTPs. Sometimes the hybridization temperature and/or reaction length have to be adjusted for specific probes.

Our modified protocol based on DIG-labeled probes will facilitate localization of mRNA expression simultaneously in species ranging from *A. thaliana* to *P. abies*, and should with minor adjustments be usable in other plant species. The protocol has, beside male cones, also been tested on different stages of vegetative shoots and in both zygotic and somatic embryos from *P. abies* with good results (unpublished results; Karlgren *et al.*).

## Disclosures

The authors have nothing to disclose.

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