# p53, Oxidative Stress, and Aging

Dongping Liu and Yang Xu

## **Abstract**

Mammalian aging is associated with elevated levels of oxidative damage of DNA, proteins, and lipids as a result of unbalanced prooxidant and antioxidant activities. Accumulating evidence indicates that oxidative stress is a major physiological inducer of aging. p53, the guardian of the genome that is important for cellular responses to oxidative stresses, might be a key coordinator of oxidative stress and aging. In response to low levels of oxidative stresses, p53 exhibits antioxidant activities to eliminate oxidative stress and ensure cell survival; in response to high levels of oxidative stresses, p53 exhibits prooxidative activities that further increase the levels of stresses, leading to cell death. p53 accomplishes these context-dependent roles by regulating the expression of a panel of genes involved in cellular responses to oxidative stresses and by modulating other pathways important for oxidative stress responses. The mechanism that switches p53 function from antioxidant to prooxidant remains unclear, but could account for the findings that increased p53 activities have been linked to both accelerated aging and increased life span in mice. Therefore, a balance of p53 antioxidant and prooxidant activities in response to oxidative stresses could be important for longevity by suppressing the accumulation of oxidative stresses and DNA damage. Antioxid. Redox Signal. 15, 1669–1678.

# p53 Is a Critical Tumor Suppressor

THE CRITICAL TUMOR SUPPRESSOR P53 plays important roles in cell-cycle arrest, apoptosis, senescence, or differentiation in response to various genotoxic and cellular stresses, including oxidative stress (73, 102, 133). As a guardian of the genome, the inactivation of wild-type p53 function by direct gene mutation or disruption of pathways important for p53 activation is a prerequisite for the development of most human cancers (35, 92, 127). As a transcription factor, p53 consists of two N-terminal transactivation domains, a core DNA-binding domain and a C-terminal oligomerization domain (55, 92). Because of its potent activity in inducing apoptosis and senescence, the p53 stability and activity are tightly regulated by posttranslational mechanisms (47, 51, 129). In the absence of stresses, p53 is inactive and unstable because of its interaction with Mdm2 and MdmX, which inactivate p53 and ubiquitinate p53 for proteasome-dependent degradation (Fig. 1). In response to stresses, p53 is modified posttranslationally through phosphorylation, acetylation, methylation, and sumoylation at various sites, disrupting the interaction between p53 and its negative regulators, leading to the activation and stabilization of p53 (68, 85, 104).

As a transcription factor, p53 can directly regulate the expression of hundreds of genes, products of which mediate various p53-dependent functions (Fig. 2) (43, 53, 69). For example, p21 and  $14-3-3\sigma$  are responsible for p53-dependent cell-cycle arrest (30, 31, 50); p53 can also induce embryonic stem (ES) cell differentiation by suppressing the expression of Nanog, which is required for the self-renewal of ES cells (64). In response to high levels of DNA damage, p53 induces apoptosis and senescence by upregulating apoptotic genes such as Noxa and Puma (66, 71). These functions of p53 prevent the passage of DNA damage to the daughter cells and thus maintain genomic stability. In response o oxidative stresses, p53 activates the transcription of a number of genes involved in regulating oxidative stress, such as Sestrin, glutathione peroxidase (GPX), aldehyde dehydrogenase (ALDH), and tumor protein 53–induced nuclear protein 1(TP53INP1) (14, 16, 115, 130). p53 can also regulate the cellular oxidative stress levels by modulating glycolysis through inducing the expression of TIGAR (TP53-induced glycolysis and apoptosis regulator) and suppressing the expression of phosphoglycerate mutase (PGM) (9, 58).

# p53 and Aging

Recent studies have functionally linked p53 to aging in various organisms (Fig. 3). The p53 orthologue in Caenorhabditis elegans, Cep-1, is involved in negatively regulating the life span of the worm, because the reduced expression of Cep-1 results in increased longevity (4). Expression of dominant-negative versions of Drosophila melanogaster p53 (Dmp53) in adult neurons extends the life span and increases the genotoxic stress

Section of Molecular Biology, Division of Biological Sciences, University of California, San Diego, La Jolla, California.



FIG. 1. Activation of p53 in response to DNA damage and oxidative stresses. In the absence of stresses, the negative regulators of p53, such as Mdm2/MdmX, suppress p53 activity and induce its degradation. In response to DNA damage and oxidative stress, p53 and its negative regulators are posttranslationally modified, leading to p53 activation by disrupting the interaction between p53 and its negative regulators.

resistance in the fly (8). Because the expression of the dominantnegative Dmp53 does not further increase the life span of flies that are calorie restricted, these findings suggest that p53 is involved in mediating the calorie-restricted life span in flies. However, mutagenesis studies in C. elegans show that certain mutations extending the life span increase activities of p53 and cancer resistance (94). Therefore, increased p53 activities are associated with both accelerated aging and increased life span in C. elegans.

A similarly complicated scenario is also observed when studying the roles of p53 in mammalian aging. One mouse model, in which the N-terminus of p53 is truncated, exhibits increased p53 activities and accelerated aging (119). However, because of the large deletion of the genomic DNA upstream of

p53 that contains 24 genes (40), it remains unclear whether any of these deleted genes is responsible for these aging phenotypes. The potential involvement of N-terminus– truncated p53 in aging is further supported by the overexpression of the N-terminus–deleted p53 isoform p44 in mice, leading to accelerated aging (72). This study suggests that p44 modulates the life span by inhibiting the PTEN and IGF signal pathways (39, 75, 110). To link p53 to aging in humans, a recent study shows that polymorphism of p53 at codon 72 (arginineto-proline substitution) reduces p53 activities, correlating with increased life span but also with higher cancer risk in older individuals (120). Therefore, it has been suggested that p53 might suppress cancer at the cost of longevity.

The notion that increased p53 activity induces aging in mice is challenged by recent studies of mouse models with increased p53 activities. For example, mice with a hypomorphic mutation in Mdm2 exhibit increased p53 activity but normal life span (78). In addition, mice with an additional copy of p53 and ARF exhibit an enhanced expression of antioxidant activity and decreased levels of endogenous oxidative stresses, correlating with increased life span (74). Therefore, the increased antioxidant activity of p53 in these transgenic mice prevents the accumulation of oxidative stresses to the high levels required to induce p53-dependent apoptosis and senescence, thus delaying aging in these mice. In summary, the functions of p53 in aging are complex and could be context dependent. In this context, mild and transient activation of p53 in response to a low dosage of oxidative stress could protect cells from oxidative damage. In contrast, persistent activation of p53 in response to high levels of oxidative stresses can result in cell death and organismal aging. In further support of this notion, persistent activation of p53 depletes adult stem cells primarily through p53-dependent apoptosis (64).

### Oxidative Stress and Aging

The free radical hypothesis remains the most well-established theory on the mechanism of aging (46). The increased



FIG. 2. p53 target genes are mediators of various p53-dependent functions in response to DNA damage and oxidative stresses.

Organism	Genetic modification	Impact on p53 activity	Impact on lifespan
C.elegans	Reduced expression	Reduced	Extended
Drosophila	Dominant negative	Reduced	Extended
Mouse	N-terminus deleted	Increased	Shortened
	p44	Increased	Shorteded
	p53T21S23D knock-in	Increased	Shortened
	Transgenic Arf or p53	Increased	No impact
	Transgenic Arf and p53	Increased	Extended
	Hypomorphic Mdm2	Increased	No impact

FIG. 3. Summary of the modulation of p53 effects on the lifespan of various organisms.

ROS production and a decreased antioxidant capacity are thought to contribute to the aging process by oxidative modification of different macromolecules, such as lipids, proteins, and genomic DNA (12, 20, 25, 62, 63, 65, 96, 109, 117). In the context of DNA, oxidative damage to mitochondrial and nuclear DNA is significantly increased in different tissues in old rats and mice (20, 45, 61, 67, 76, 82, 116). Levels of lipidperoxidation products are also increased with aging (44, 83, 87, 97, 108, 113, 119, 123). In addition, aging-related oxidative modification of different proteins causes changes in protein structure, enzyme activities, transcriptional activities, and signal-transduction pathways (32, 70, 103, 111, 112, 124), leading to age-related diseases. In summary, the levels of oxidative damage are increased during aging in various organisms, including C. elegans (11, 52, 121), flies (3, 64), and mice (22, 74, 79).

Free radicals are physiologic byproducts of metabolism and are rapidly eliminated by various antioxidant enzymes in cells (23). For example, the antioxidant enzymes, including superoxide dismutase (SOD), catalase, and peroxiredoxins, convert superoxide to hydrogen peroxide and eventually to water (5, 19, 99). SODs catalyze the breakdown of the superoxide anion into oxygen and hydrogen peroxide. Mice lacking SOD2 develop neurologic defects and die soon after birth because of excessive mitochondrial production of ROS (77); mice lacking SOD1 are viable but have numerous pathologies and a reduced life span (98). Catalase converts hydrogen peroxide into water and oxygen (19, 132). Humans and mice deficient in catalase can still efficiently remove  $H_2O_2$ , implying that other enzymes are also involved in this reaction (72, 88). Peroxiredoxins catalyze the reduction of hydrogen peroxide, organic peroxide, and peroxynitrite (99). These enzymes can be divided into there classes: typical 2-cysteine peroxiredoxins, atypical 2-cysteine peroxiredoxins, and 1-cysteine peroxiredoxins (128). Mice lacking peroxiredoxins 1 and 2 have a shortened life span (55, 86). Together, these findings underscore the importance of antioxidant enzymes in preventing aging processes. In further support of this notion, a diet rich in the building-block nutrients of antioxidant enzymes, including cofactors for SOD (manganese, zinc, and copper), show beneficial effects on delaying aging (1, 24, 49, 59, 81, 106).

In further support of the notion that oxidative stress is an inducer of aging, treatment with antioxidants can increase the life spans of various organisms and has a beneficial impact on aging-related diseases (6, 29, 38, 57, 114, 119). A low dose of dietary supplement with antioxidants partially mimics the effects of caloric restriction and delays aging in mice (6), and long-term treatment with free radical scavenging Schisandrin B, a dibenzocyclooctadiene derivative isolated from the fruit of Schisandra chinensis, delays aging-related functional impairment in various organs and improves the survival rate of aging mice (114). A dietary supplement of cysteine, which is required for the synthesis of the primary antioxidant glutathione, has clear benefits in delaying some aspects of aging (29). However, clinical trials have also found no significant beneficial effects of supplementation with antioxidant vitamin E, indicating that not all antioxidants have antiaging activities (55, 107, 125).

#### p53 and Oxidative Stress

ROS levels have a significant impact on cell growth, survival and development, and tumorigenesis (17). p53 plays key and complex roles in cellular responses to oxidative stresses (84, 100). In response to low levels of oxidative stresses, p53 plays primarily antioxidant roles. In this context, a number of p53 target genes, including Sestrin, glutathione peroxidase (GPX), and aldehyde dehydrogenase (ALDH), are involved in reducing oxidative stresses (Fig. 4). For example, Sestrin protects the cells from hydrogen peroxide–induced damage by generation of peroxiredoxins (14). GPX is a primary antioxidant enzyme that scavenges hydrogen peroxide or organic hydroperoxides (115). Aldehyde dehydrogenase (ALDH) also contributes to the antioxidant function of p53 (130).

p53 can also reduce the intracellular levels of ROS by regulating cellular metabolism. In this context, p53 induces the expression of TIGAR (TP53-induced glycolysis and apoptosis regulator), which slows glycolysis and promotes the production of NAPDH to decrease ROS levels (9). In addition, p53 suppresses the expression of phosphoglycerate mutase (PGM), leading to a decrease of pyruvate required for oxidative respiration in mitochondria and thus reduced ROS production (10, 74).

In response to high levels of oxidative stress, p53 exhibits prooxidative activities by turning on prooxidative genes such as PIG3 and proline oxidase (27, 95). Overexpression of these genes leads to higher levels of oxidative stress. In addition, p53 induces the expression of BAX and PUMA, which induce apoptosis through the release of cytochrome  $c$  from mitochondria (66, 71). The prooxidative activities of p53 also include the inhibition of the expression of antioxidant genes, leading to increased cellular oxidative stresses to induce apoptosis. For example, p53 could repress the expression of SOD2 and Nrf2, resulting in sensitivity to oxidative stress or inducing apoptosis (28, 34, 91). Interestingly, p53-induced upregulation of MnSOD and GPX, but not catalase, increases oxidative stress and apoptosis (54), suggesting that the balance of antioxidant enzyme and oxidative stress is important for cell survival. In summary, p53 plays important but context-dependent roles in regulating cellular oxidative stresses, and the levels of oxidative-stress damage dictate whether the p53 behavior is that of a protector or a killer (100).



# p53 Interacts with Other Pathways Involved in Oxidative Stress and Aging

In addition to its direct regulation of genes involved in oxidative stresses, p53 also interact with other pathways that are involved in aging and oxidative stresses, which are summarized here (Fig. 5).

#### Sirt1

The Sirt1 gene encodes the NAD-dependent histone deacetylase, which is important for the longevity in yeast and mammalian species by calorie restriction (42, 60, 64, 122). Sirt1 can deacetylate and inactivate p53, leading to impaired cell growth arrest and apoptosis in response to oxidative stresses (101). In addition, the expression of a dominant-negative version of Sirt1 increases the cellular sensitivity to oxidative stress, further indicating its antioxidant roles in cellular responses to oxidative stresses. However, the roles of Sirt1 in suppressing p53 in response to oxidative stresses remain to be fully established. In contrast to the prediction that Sirt1 deficiency would increase p53 activity, recent studies indicate that deficiency of Sirt1 extends the replicative capacity of mouse embryonic fibroblasts (MEFs) under the conditions of chronic oxidative stress due to the inefficient activation of p53 (21). However, the physiological relevance of replicative senescence in aging is not clear, because it primarily reflects a cell-culture phenomenon in the presence of nonphysiologically high levels of oxygen. Because Sirt1 is an NAD-dependent deacetylase, and NAD levels are regulated by cellular metabolism and levels of ROS, these findings implicate a complex functional interaction of p53, Sirt1, oxidative stresses, and aging.

# p66Shc

p66Shc, a downstream target of p53, is indispensable for p53-dependent elevation of intracellular oxidative stresses and apoptosis (118). p66Shc is a splice variant of  $p52Shc/$ p46Shc, a cytoplasmic signal transducer involved in the transmission of mitogenic signal from activated receptors to FIG. 4. Context-dependent roles of p53 in cellular responses to oxidative stresses by turning on distinct target genes. At basal or low levels of oxidative stress, p53 regulates the expression of Sestrin, GPX, ALDH, TP53INP1, SOD2, TIGAR, and PGM to eliminate ROS, and therefore, promotes cellular survival. In response to high levels of oxidative stress, p53 induces the expression of prooxidative genes and suppresses the expression of antioxidant genes to increase ROS levels and promote apoptosis. Unbalanced antioxidants can also induce ROS to promote cell death.

Ras (93). However, p66Shc is not involved in regulating Ras signal but instead is involved in inducing apoptosis in response to oxidative stresses (80). The important role of p66Shc in oxidative stresses and aging is indicated by the findings that ablation of p66Shc enhances cellular resistance to apoptosis induced by oxidative stresses and extends the life span of p66Shc-deficient mice (79). In this context, cytochrome c release after oxidative signals is impaired in p66Shc-deficient cells (90). Therefore, p66Shc functionally links p53 to oxidative stress response and aging.

#### FoxO

Forkhead box O (FoxO) transcription factors are important mediators of the PI3K/Akt signaling pathway and regulate the cellular responses to oxidative stresses and the life span (56, 105). p53 negatively regulates the activities of FoxO by inducing the expression of serum- and glucocorticoid-inducible kinase (SGK), a negative regulator of FoxO and PTEN (37). In addition, Sirt1 can deacetylate FoxO3 and FoxO4, thus



FIG. 5. Functional interaction between p53 and other pathways important for oxidative stress response and aging.

attenuating FoxO-induced apoptosis and cell-cycle arrest (41). Therefore, the balance of the functional interaction among Sirt1, FoxO, and p53 might play important roles in regulating oxidative stresses and aging.

#### $APE/Ref1$

The expression of APE/Ref1 is decreased in senescent human bone marrow–derived mesenchymal stem cells (hBMSCs) with increased endogenous ROS levels. Overexpression of APE1/Ref-1 suppresses superoxide production and decreases senescence in hBMSCs (48). In addition, aging mice have an impaired induction of APE in response to oxidative damage (15). The activities of  $APE/Ref1$  are negatively regulated by p53 (131), implicating another pathway for p53 to modulate oxidative stresses and aging.

# Caveolin-1

Expression of Caveolin-1 is induced in fibroblasts undergoing oxidative stress–induced senescence, and the antioxidant prevents the senescence and upregulation of Caveolin-1 (36, 126). Overexpression of Caveolin-1 in MEF induces the premature senescence through a p53-p21–dependent pathway, suggesting that Caveolin-1 could activate p53 dependent premature senescence after oxidative stresses (36). In this context, Caveolin-1 binds to Mdm2 and disrupts the binding of Mdm2 to p53, leading to the activation of p53 in response to oxidative stresses. The activation of p53 and induction of premature senescence are compromised in the Caveolin-1–null MEFs, confirming that Caveolin-1 is an upstream activator of p53 in response to oxidative stresses (7).

#### FoxM1C-Bmi1 pathway

Bmi1 is a negative regulator of the  $Ink4a/Arf$  and p53; FoxM1C induces the expression of Bmi1 to prevent the oxidative stress–induced cellular senescence by inhibiting the expression of p53 (13, 18, 33, 89). Bmi1 is important to repress the prooxidant activities of p53 in neurons and to suppress oxidative stress–induced apoptosis and premature aging-like phenotypes (18). In addition, targeted depletion of Bmi1 sensitizes tumor cells to p53-mediated apoptosis in response to radiation therapy (2).

#### Bach1

For transcription factors, the recruitment of co-activators or co-repressors to p53 target promoters is critical for p53 dependent transcription. Bach1 is induced by oxidative stresses and forms a complex with p53, histone deacetylase 1, and nuclear co-repressor N-coR, promoting histone deacetylation and suppression of certain p53 target genes (26). In this context, Bach1 inhibits oxidative stress–induced cellular senescence by disrupting p53-dependent gene expression (26).

# **Conclusion**

The accumulation of oxidative stress and oxidative damage is a major inducer of aging. Many pathways involved in cellular responses to oxidative stresses regulate the aging process and the life spans of various organisms. p53 plays important but context-dependent roles in cellular responses to low or high levels of oxidative stresses. In response to low levels of oxidative stresses, p53 exhibits antioxidant activities and promotes cellular survival; in response to high levels of oxidative stresses, p53 exhibits prooxidative activities to induce cellular apoptosis. Both functions of p53 can prevent the accumulation of oxidative damage in cells and thus maintain genomic stability. p53 accomplishes these functions by direct transcriptional regulation of genes involved in oxidativestress responses or modulating other pathways important in oxidative-stress responses.

Consistent with its context-specific roles in oxidative-stress responses, the roles of p53 in aging appear to be complex as well. In this context, increased p53 activities can accelerate aging in some transgenic mouse models but not in others (72, 74, 78, 119). In addition, the increase of the gene dosage of ARF and p53 does not promote aging but increases the life span of transgenic mice (74). Therefore, the roles of p53 in aging could also be context dependent. The accumulation of oxidative stresses in old mice could turn on the apoptotic or senescent roles of p53, thus promoting the aging process. However, increased dosages of p53 and ARF could ensure more efficient elimination of oxidative stress and thus prevent the accumulation of oxidative stresses to high levels in old mice. In support of this notion, a significant reduction of DNA damage occurs in old transgenic mice with additional copies of p53 and ARF (74). p53 primarily plays a protective role to increase the life span in these transgenic mice. Therefore, further elucidation of the mechanism that governs the context-dependent roles of p53 in oxidative-stress responses and the functional outcomes of the interaction between p53 and other pathways involved in cellular responses to oxidative stresses will shed light on its role in aging.

#### Acknowledgment

This work is supported by an NIH grant to YX (R01 CA94254).

#### References

- 1. Airede AK. Copper, zinc and superoxide dismutase activities in premature infants: a review. East Afr Med J 70: 441– 444, 1993.
- 2. Alajez NM, Shi W, Hui AB, Yue S, Ng R, Lo KW, Bastianutto C, O'Sullivan B, Gullane P, and Liu FF. Targeted depletion of BMI1 sensitizes tumor cells to P53-mediated apoptosis in response to radiation therapy. Cell Death Differ 16: 1469–1479, 2009.
- 3. Arking R, Buck S, Berrios A, Dwyer S, and Baker GT 3rd. Elevated paraquat resistance can be used as a bioassay for longevity in a genetically based long-lived strain of Drosophila. Dev Genet 12: 362–370, 1991.
- 4. Arum O and Johnson TE. Reduced expression of the Caenorhabditis elegans p53 ortholog cep-1 results in increased longevity. J Gerontol A Biol Sci Med Sci 62: 951–959, 2007.
- 5. Bannister JV, Bannister WH, and Rotilio G. Aspects of the structure, function, and applications of superoxide dismutase. CRC Crit Rev Biochem 22: 111–180, 1987.
- 6. Barger JL, Kayo T, Vann JM, Arias EB, Wang J, Hacker TA, Wang Y, Raederstorff D, Morrow JD, Leeuwenburgh C, Allison DB, Saupe KW, Cartee GD, Weindruch R, and Prolla TA. A low dose of dietary resveratrol partially mimics caloric restriction and retards aging parameters in mice. PLoS One 3: e2264, 2008.
- 7. Bartholomew JN, Volonte D, and Galbiati F. Caveolin-1 regulates the antagonistic pleiotropic properties of cellular senescence through a novel Mdm2/p53-mediated pathway. Cancer Res 69: 2878–2886, 2009.
- 8. Bauer JH, Poon PC, Glatt-Deeley H, Abrams JM, and Helfand SL. Neuronal expression of p53 dominant-negative proteins in adult Drosophila melanogaster extends life span. Curr Biol 15: 2063–2068, 2005.
- 9. Bensaad K, Tsuruta A, Selak MA, Vidal MN, Nakano K, Bartrons R, Gottlieb E, and Vousden KH. TIGAR, a p53 inducible regulator of glycolysis and apoptosis. Cell 126: 107–120, 2006.
- 10. Bensaad K and Vousden KH. p53: new roles in metabolism. Trends Cell Biol 17: 286–291, 2007.
- 11. Berdichevsky A, Viswanathan M, Horvitz HR, and Guarente L. C. elegans SIR-2.1 interacts with 14-3-3 proteins to activate DAF-16 and extend life span. Cell 125: 1165–1177, 2006.
- 12. Bergamini E, Bizzarri R, Cavallini G, Cerbai B, Chiellini E, Donati A, Gori Z, Manfrini A, Parentini I, Signori F, and Tamburini I. Ageing and oxidative stress: a role for dolichol in the antioxidant machinery of cell membranes? J Alzheimers Dis 6: 129–135, 2004.
- 13. Bruggeman SW, Valk-Lingbeek ME, van der Stoop PP, Jacobs JJ, Kieboom K, Tanger E, Hulsman D, Leung C, Arsenijevic Y, Marino S, and van Lohuizen M. Ink4a and Arf differentially affect cell proliferation and neural stem cell self-renewal in Bmi1-deficient mice. Genes Dev 19: 1438–1443, 2005.
- 14. Budanov AV, Sablina AA, Feinstein E, Koonin EV, and Chumakov PM. Regeneration of peroxiredoxins by p53 regulated sestrins, homologs of bacterial AhpD. Science 304: 596–600, 2004.
- 15. Cabelof DC, Raffoul JJ, Ge Y, Van Remmen H, Matherly LH, and Heydari AR. Age-related loss of the DNA repair response following exposure to oxidative stress. J Gerontol A Biol Sci Med Sci 61: 427–434, 2006.
- 16. Cano CE, Gommeaux J, Pietri S, Culcasi M, Garcia S, Seux M, Barelier S, Vasseur S, Spoto RP, Pebusque MJ, Dusetti NJ, Iovanna JL, and Carrier A. Tumor protein 53-induced nuclear protein 1 is a major mediator of p53 antioxidant function. Cancer Res 69: 219–226, 2009.
- 17. Chao C, Hergenhahn M, Kaeser MD, Wu Z, Saito S, Iggo R, Hollstein M, Appella E, anmd Xu Y. Cell type- and promoter-specific roles of Ser18 phosphorylation in regulating p53 responses. J Biol Chem 278: 41028–41033, 2003.
- 18. Chatoo W, Abdouh M, David J, Champagne MP, Ferreira J, Rodier F, and Bernier G. The polycomb group gene Bmi1 regulates antioxidant defenses in neurons by repressing p53 pro-oxidant activity. J Neurosci 29: 529–542, 2009.
- 19. Chelikani P, Fita I, and Loewen PC. Diversity of structures and properties among catalases. Cell Mol Life Sci 61: 192– 208, 2004.
- 20. Chen JH, Hales CN, and Ozanne SE. DNA damage, cellular senescence and organismal ageing: causal or correlative? Nucleic Acids Res 35: 7417–7428, 2007.
- 21. Chua KF, Mostoslavsky R, Lombard DB, Pang WW, Saito S, Franco S, Kaushal D, Cheng HL, Fischer MR, Stokes N, Murphy MM, Appella E, and Alt FW. Mammalian SIRT1 limits replicative life span in response to chronic genotoxic stress. Cell Metab 2: 67–76, 2005.
- 22. Collins AR, Lyon CJ, Xia X, Liu JZ, Tangirala RK, Yin F, Boyadjian R, Bikineyeva A, Pratico D, Harrison DG, and

Hsueh WA. Age-accelerated atherosclerosis correlates with failure to upregulate antioxidant genes. Circ Res 104: e42– e54, 2009.

- 23. Davies KJ. Oxidative stress: the paradox of aerobic life. Biochem Soc Symp 61: 131, 1995.
- 24. Davis CD and Feng Y. Dietary copper, manganese and iron affect the formation of aberrant crypts in colon of rats administered 3,2'-dimethyl-4-aminobiphenyl. J Nutr 129: 1060–1067, 1999.
- 25. De Bont R and van Larebeke N. Endogenous DNA damage in humans: a review of quantitative data. Mutagenesis 19: 169–185, 2004.
- 26. Dohi Y, Ikura T, Hoshikawa Y, Katoh Y, Ota K, Nakanome A, Muto A, Omura S, Ohta T, Ito A, Yoshida M, Noda T, and Igarashi K. Bach1 inhibits oxidative stress-induced cellular senescence by impeding p53 function on chromatin. Nat Struct Mol Biol 15: 1246–1254, 2008.
- 27. Donald SP, Sun XY, Hu CA, Yu J, Mei JM, Valle D, and Phang JM. Proline oxidase, encoded by p53-induced gene-6, catalyzes the generation of proline-dependent reactive oxygen species. Cancer Res 61: 1810–1815, 2001.
- 28. Drane P, Bravard A, Bouvard V, and May E. Reciprocal downregulation of p53 and SOD2 gene expression-implication in p53 mediated apoptosis. Oncogene 20: 430–439, 2001.
- 29. Droge W. Oxidative stress and ageing: is ageing a cysteine deficiency syndrome? Phil Trans R Soc Lond B Biol Sci 360: 2355–2372, 2005.
- 30. el-Deiry WS, Harper JW, O'Connor PM, Velculescu VE, Canman CE, Jackman J, Pietenpol JA, Burrell M, Hill DE, and Wang Y, et al.  $WAF1/CIP1$  is induced in p53-mediated G1 arrest and apoptosis. Cancer Res 54: 1169–1174, 1994.
- 31. el-Deiry WS, Tokino T, Velculescu VE, Levy DB, Parsons R, Trent JM, Lin D, Mercer WE, Kinzler KW, and Vogelstein B. WAF1, a potential mediator of p53 tumor suppression. Cell 75: 817–825, 1993.
- 32. Ethen CM, Reilly C, Feng X, Olsen TW, and Ferrington DA. Age-related macular degeneration and retinal protein modification by 4-hydroxy-2-nonenal. Invest Ophthalmol Vis Sci 48: 3469–3479, 2007.
- 33. Fan C, He L, Kapoor A, Gillis A, Rybak AP, Cutz JC, and Tang D. Bmi1 promotes prostate tumorigenesis via inhibiting p16(INK4A) and p14(ARF) expression. Biochim Biophys Acta 1782: 642–648, 2008.
- 34. Faraonio R, Vergara P, Di Marzo D, Pierantoni MG, Napolitano M, Russo T, and Cimino F. p53 suppresses the Nrf2-dependent transcription of antioxidant response genes. J Biol Chem 281: 39776–39784, 2006.
- 35. Freeman J, Schmidt S, Scharer E, and Iggo R. Mutation of conserved domain II alters the sequence specificity of DNA binding by the p53 protein. EMBO J 13: 5393–5400, 1994.
- 36. Galbiati F, Volonte D, Liu J, Capozza F, Frank PG, Zhu L, Pestell RG, Lisanti MP. Caveolin-1 expression negatively regulates cell cycle progression by inducing  $G(0)/G(1)$  arrest via a  $p53/p21(WAF1/Cip1)$ -dependent mechanism. Mol Biol Cell 12: 2229–2244, 2001.
- 37. Garinis GA, van der Horst GTJ, Vijg JHJ, and Hoeijmakers J. DNA damage and ageing: new-age ideas for an age-old problem. Nat Cell Biol 10: 1241–1247, 2008.
- 38. Gaziano JM. Vitamin E and cardiovascular disease: observational studies. Ann N Y Acad Sci 1031: 280–291, 2004.
- Gems D and Partridge L. Insulin/IGF signalling and ageing: seeing the bigger picture. Curr Opin Genet Dev 11: 287– 292, 2001.
- 40. Gentry A and Venkatachalam S. Complicating the role of p53 in aging. Aging Cell 4: 157–160, 2005.
- 41. Giannakou ME and Partridge L. The interaction between FOXO and SIRT1: tipping the balance towards survival. Trends Cell Biol 14: 408–412, 2004.
- 42. Guarente L. Sir2 links chromatin silencing, metabolism, and aging. Genes Dev 14: 1021–1026, 2000.
- 43. Gudkov A. Microarray analysis of p53-mediated transcription: multi-thousand piece puzzle or invitation to collective thinking. Cancer Biol Ther 2: 444–445, 2003.
- 44. Gupta A, Hasan M, Chander R, and Kapoor NK. Agerelated elevation of lipid peroxidation products: diminution of superoxide dismutase activity in the central nervous system of rats. Gerontology 37: 305–309, 1991.
- 45. Hamilton ML, Van Remmen H, Drake JA, Yang H, Guo ZM, Kewitt K, Walter CA, and Richardson A. Does oxidative damage to DNA increase with age? Proc Natl Acad Sci U S A 98: 10469–10474, 2001.
- 46. Harman D. Aging: a theory based on free radical and radiation chemistry. J Gerontol 11: 298–300, 1956.
- 47. Haupt Y, Maya R, Kazaz A, and Oren M. Mdm2 promotes the rapid degradation of p53. Nature 387: 296–299, 1997.
- 48. Heo JY, Jing K, Song KS, Seo KS, Park JH, Kim JS, Jung YJ, Hur GM, Jo DY, Kweon GR, Yoon WH, Lim K, Hwang BD, Jeon BH, and Park JI. Downregulation of APE1/Ref-1 is involved in the senescence of mesenchymal stem cells. Stem Cells 27: 1455–1462, 2009.
- 49. Hercberg S, Galan P, Preziosi P, Bertrais S, Mennen L, Malvy D, Roussel AM, Favier A, and Briancon S. The SU.VI.MAX Study: a randomized, placebo-controlled trial of the health effects of antioxidant vitamins and minerals. Arch Intern Med 164: 2335–2342, 2004.
- 50. Hermeking H, Lengauer C, Polyak K, He TC, Zhang L, Thiagalingam S, Kinzler KW, and Vogelstein B. 14-3-3 sigma is a p53-regulated inhibitor of  $G2/M$  progression. Mol Cell 1: 3–11, 1997.
- 51. Honda R, Tanaka H, and Yasuda H. Oncoprotein MDM2 is a ubiquitin ligase E3 for tumor suppressor p53. FEBS Lett 420: 25–27, 1997.
- 52. Honda Y and Honda S. Oxidative stress and life span determination in the nematode Caenorhabditis elegans. Ann N Y Acad Sci 959: 466–474, 2002.
- 53. Huang J, Logsdon N, Schmieg FI, and Simmons DT. p53 mediated transcription induces resistance of DNA to UV inactivation. Oncogene 17: 401–411, 1998.
- 54. Hussain SP, Amstad P, He P, Robles A, Lupold S, Kaneko I, Ichimiya M, Sengupta S, Mechanic L, Okamura S, Hofseth LJ, Moake M, Nagashima M, Forrester KS, and Harris CC. p53-induced up-regulation of MnSOD and GPx but not catalase increases oxidative stress and apoptosis. Cancer Res 64: 2350–2356, 2004.
- 55. Jeffers JR, Parganas E, Lee Y, Yang C, Wang J, Brennan J, MacLean KH, Han J, Chittenden T, Ihle JN, McKinnon PJ, Cleveland JL, and Zambetti GP. Puma is an essential mediator of p53-dependent and -independent apoptotic pathways. Cancer Cell 4: 321–328, 2003.
- 56. Katic M and Kahn CR. The role of insulin and IGF-1 signaling in longevity. Cell Mol Life Sci 62: 320–343, 2005.
- 57. Kim J, Takahashi M, Shimizu T, Shirasawa T, Kajita M, Kanayama A, and Miyamoto Y. Effects of a potent antioxidant, platinum nanoparticle, on the lifespan of Caenorhabditis elegans. Mech Ageing Dev 129: 322–331, 2008.
- 58. Kondoh H, Lleonart ME, Gil J, Wang J, Degan P, Peters G, Martinez D, Carnero A, and Beach D. Glycolytic enzymes can modulate cellular life span. Cancer Res 65: 177–185, 2005.
- 59. Lamb DJ, Tickner ML, Hourani SM, and Ferns GA. Dietary copper supplements modulate aortic superoxide dismutase, nitric oxide and atherosclerosis. Int J Exp Pathol 86: 247–255, 2005.
- 60. Langley E, Pearson M, Faretta M, Bauer UM, Frye RA, Minucci S, Pelicci PG, and Kouzarides T. Human SIR2 deacetylates  $p53$  and antagonizes  $PML/p53$ -induced cellular senescence. EMBO J 21: 2383–2396, 2002.
- 61. Lemon JA, Rollo CD, and Boreham DR. Elevated DNA damage in a mouse model of oxidative stress: impacts of ionizing radiation and a protective dietary supplement. Mutagenesis 23: 473–482, 2008.
- 62. Lenaz G. Role of mitochondria in oxidative stress and ageing. Biochim Biophys Acta 1366: 53–67, 1998.
- 63. Lenaz G, Bovina C, Formiggini G, and Parenti Castelli G. Mitochondria, oxidative stress, and antioxidant defences. Acta Biochim Pol 46: 1–21, 1999.
- 64. Liu DP, Ou L, Clemenson Jr GD, Chao C, Lutske ME, Zambetti GP, Gage FH, and Xu Y. Puma is required for p53-induced depletion of adult stem cells. Nat Cell Biol 12: 993–998, 2010.
- 65. Linnane AW, Marzuki S, Ozawa T, and Tanaka M. Mitochondrial DNA mutations as an important contributor to ageing and degenerative diseases. Lancet 1: 642–645, 1989.
- 66. Liu Z, Lu H, Shi H, Du Y, Yu J, Gu S, Chen X, Liu KJ, and Hu CA. PUMA overexpression induces reactive oxygen species generation and proteasome-mediated stathmin degradation in colorectal cancer cells. Cancer Res 65: 1647– 1654, 2005.
- 67. Lopez-Torres M and Barja G. Calorie restriction, oxidative stress and longevity. Rev Esp Geriatr Gerontol 43: 252–260, 2008.
- 68. Lu H and Levine AJ. Human TAFII31 protein is a transcriptional coactivator of the p53 protein. Proc Natl Acad Sci USA 92: 5154–5158, 1995.
- 69. Lu H, Lin J, Chen J, and Levine AJ. The regulation of p53 mediated transcription and the roles of hTAFII31 and mdm-2. Harvey Lect 90: 81–93, 1994.
- 70. Machado A, Ayala A, Gordillo E, Revilla E, and Santa Maria C. Relationship between enzymatic activity loss and post-translational protein modification in aging. Arch Gerontol Geriatr 12: 187–197, 1991.
- 71. Macip S, Igarashi M, Berggren P, Yu J, Lee SW, and Aaronson SA. Influence of induced reactive oxygen species in p53-mediated cell fate decisions. Mol Cell Biol 23: 8576– 8585, 2003.
- 72. Maier B, Gluba W, Bernier B, Turner T, Mohammad K, Guise T, Sutherland A, Thorner M, and Scrable H. Modulation of mammalian life span by the short isoform of p53. Genes Dev 18: 306–319, 2004.
- 73. Mansur CP. The regulation and function of the p53 tumor suppressor. Adv Dermatol 13: 121–166, 1997.
- 74. Matheu A, Maraver A, Klatt P, Flores I, Garcia-Cao I, Borras C, Flores JM, Vina J, Blasco MA, and Serrano M. Delayed ageing through damage protection by the  $Arf/p53$ pathway. Nature 448: 375–379, 2007.
- 75. Mattson MP, Maudsley S, and Martin B. A neural signaling triumvirate that influences ageing and age-related disease: insulin/IGF-1, BDNF and serotonin. Ageing Res Rev 3: 445– 464, 2004.
- 76. Meissner C. Mutations of mitochondrial DNA: cause or consequence of the ageing process? Z Gerontol Geriatr 40: 325–333, 2007.
- 77. Melov S, Schneider JA, Day BJ, Hinerfeld D, Coskun P, Mirra SS, Crapo JD, and Wallace DC. A novel neurological phenotype in mice lacking mitochondrial manganese superoxide dismutase. Nat Genet 18: 159–163, 1998.
- 78. Mendrysa SM, O'Leary KA, McElwee MK, Michalowski J, Eisenman RN, Powell DA, and Perry ME. Tumor suppression and normal aging in mice with constitutively high p53 activity. Genes Dev 20: 16–21, 2006.
- 79. Migliaccio E, Giorgio M, Mele S, Pelicci G, Reboldi P, Pandolfi PP, Lanfrancone L, and Pelicci PG. The p66shc adaptor protein controls oxidative stress response and life span in mammals. Nature 402: 309–313, 1999.
- 80. Migliaccio E, Mele S, Salcini AE, Pelicci G, Lai KM, Superti-Furga G, Pawson T, Di Fiore PP, Lanfrancone L, and Pelicci PG. Opposite effects of the p52shc/p46shc and p66shc splicing isoforms on the EGF receptor-MAP kinase-fos signalling pathway. EMBO J 16: 706–716, 1997.
- 81. Mocchegiani E, Malavolta M, Muti E, Costarelli L, Cipriano C, Piacenza F, Tesei S, Giacconi R, and Lattanzio F. Zinc, metallothioneins and longevity: interrelationships with niacin and selenium. Curr Pharm Des 14: 2719–2732, 2008.
- 82. Montaner B, O'Donovan P, Reelfs O, Perrett CM, Zhang X, Xu YZ, Ren X, Macpherson P, Frith D, and Karran P. Reactive oxygen-mediated damage to a human DNA replication and repair protein. EMBO Rep 8: 1074–1079, 2007.
- 83. Montine TJ, Neely MD, Quinn JF, Beal MF, Markesbery WR, Roberts LJ, and Morrow JD. Lipid peroxidation in aging brain and Alzheimer's disease. Free Radic Biol Med 33: 620–626, 2002.
- 84. Nakamizo A, Amano T, Zhang W, Zhang XQ, Ramdas L, Liu TJ, Bekele BN, Shono T, Sasaki T, Benedict WF, Sawaya R, and Lang FF. Phosphorylation of Thr18 and Ser20 of p53 in Ad-p53-induced apoptosis. Neurol Oncol 10: 275–291, 2008.
- 85. Neilsen PM, Cheney KM, Li CW, Chen JD, Cawrse JE, Schulz RB, Powell JA, Kumar R, and Callen DF. Identification of ANKRD11 as a p53 coactivator. J Cell Sci 121: 3541–3552, 2008.
- 86. Neumann CA, Krause DS, Carman CV, Das S, Dubey DP, Abraham JL, Bronson RT, Fujiwara Y, Orkin SH, and Van Etten RA. Essential role for the peroxiredoxin Prdx1 in erythrocyte antioxidant defence and tumour suppression. Nature 424: 561–565, 2003.
- 87. Nowak M, Swietochowska E, Wielkoszynski T, Marek B, Karpe J, Gorski J, Glogowska-Szelag J, Kos-Kudla B, and Ostrowska Z. Changes in blood antioxidants and several lipid peroxidation products in women with age-related macular degeneration. Eur J Ophthalmol 13: 281–286, 2003.
- 88. Ogata M. Acatalasemia. Hum Genet 86: 331–340, 1991.
- 89. Oguro H, Iwama A, Morita Y, Kamijo T, van Lohuizen M, and Nakauchi H. Differential impact of Ink4a and Arf on hematopoietic stem cells and their bone marrow microenvironment in Bmi1-deficient mice. J Exp Med 203: 2247– 2253, 2006.
- 90. Orsini F, Migliaccio E, Moroni M, Contursi C, Raker VA, Piccini D, Martin-Padura I, Pelliccia G, Trinei M, Bono M,

Puri C, Tacchetti C, Ferrini M, Mannucci R, Nicoletti I, Lanfrancone L, Giorgio M, and Pelicci PG. The life span determinant p66Shc localizes to mitochondria where it associates with mitochondrial heat shock protein 70 and regulates trans-membrane potential. J Biol Chem 279: 25689– 25695, 2004.

- 91. Pani G, Bedogni B, Anzevino R, Colavitti R, Palazzotti B, Borrello S, and Galeotti T. Deregulated manganese superoxide dismutase expression and resistance to oxidative injury in p53-deficient cells. Cancer Res 60: 4654–4660, 2000.
- 92. Pavletich NP, Chambers KA, and Pabo CO. The DNAbinding domain of p53 contains the four conserved regions and the major mutation hot spots. Genes Dev 7: 2556–2564, 1993.
- 93. Pelicci G, Lanfrancone L, Grignani F, McGlade J, Cavallo F, Forni G, Nicoletti I, Grignani F, Pawson T, and Pelicci PG. A novel transforming protein (SHC) with an SH2 domain is implicated in mitogenic signal transduction. Cell 70: 93–104, 1992.
- 94. Pinkston JM, Garigan D, Hansen M, and Kenyon C. Mutations that increase the life span of C. elegans inhibit tumor growth. Science 313: 971–975, 2006.
- 95. Polyak K, Xia Y, Zweier JL, Kinzler KW, and Vogelstein B. A model for p53-induced apoptosis. Nature 389: 300–305, 1997.
- 96. Poulsen HE. Oxidative DNA modifications. Exp Toxicol Pathol 57 suppl 1: 161–169, 2005.
- 97. Pratico D. Lipid peroxidation and the aging process. Sci Aging Knowledge Environ 50: 1–4 , 2002.
- 98. Reaume AG, Elliott JL, Hoffman EK, Kowall NW, Ferrante RJ, Siwek DF, Wilcox HM, Flood DG, Beal MF, Brown RH Jr, Scott RW, and Snider WD. Motor neurons in  $Cu/Zn$ superoxide dismutase-deficient mice develop normally but exhibit enhanced cell death after axonal injury. Nat Genet 13: 43–47, 1996.
- 99. Rhee SG, Chae HZ, and Kim K. Peroxiredoxins: a historical overview and speculative preview of novel mechanisms and emerging concepts in cell signaling. Free Radic Biol Med 38: 1543–1552, 2005.
- 100. Sablina AA, Budanov AV, Ilyinskaya GV, Agapova LS, Kravchenko JE, and Chumakov PM. The antioxidant function of the p53 tumor suppressor. Nat Med 11: 1306– 1313, 2005.
- 101. Sakaguchi K, Herrera JE, Saito S, Miki T, Bustin M, Vassilev A, Anderson CW, and Appella E. DNA damage activates p53 through a phosphorylation-acetylation cascade. Genes Dev 12: 2831–2841, 1998.
- 102. Sandor J, Ambrus T, and Ember I. The function of the p53 gene suppressor in carcinogenesis. Orv Hetil 136: 1875– 1883, 1995.
- 103. Schoneich C. Protein modification in aging: an update. Exp Gerontol 41: 807–812, 2006.
- 104. Scolnick DM, Chehab NH, Stavridi ES, Lien MC, Caruso L, Moran E, Berger SL, and Halazonetis TD. CREB-binding protein and p300/CBP-associated factor are transcriptional coactivators of the p53 tumor suppressor protein. Cancer Res 57: 3693–3696, 1997.
- 105. Sedding DG. FoxO transcription factors in oxidative stress response and ageing: a new fork on the way to longevity? Biol Chem 389: 279–283, 2008.
- 106. Selenius M, Rundlof AK, Olm E, Fernandes AP, and Bjornstedt M. Selenium and the selenoprotein thioredoxin reductase in the prevention, treatment and

diagnostics of cancer. Antioxid Redox Signal 12: 867–880, 2009.

- 107. Sesso HD, Buring JE, Christen WG, Kurth T, Belanger C, MacFadyen J, Bubes V, Manson JE, Glynn RJ, and Gaziano JM. Vitamins E and C in the prevention of cardiovascular disease in men: the Physicians' Health Study II randomized controlled trial. JAMA 300: 2123–2133, 2008.
- 108. Shinohara R, Mano T, Nagasaka A, Hayashi R, Uchimura K, Nakano I, Watanabe F, Tsugawa T, Makino M, Kakizawa H, Nagata M, Iwase K, Ishizuki Y, and Itoh M. Lipid peroxidation levels in rat cardiac muscle are affected by age and thyroid status. J Endocrinol 164: 97– 102, 2000.
- 109. Siedlak SL, Casadesus G, Webber KM, Pappolla MA, Atwood CS, Smith MA, and Perry G. Chronic antioxidant therapy reduces oxidative stress in a mouse model of Alzheimer's disease. Free Radic Res 43: 156–164, 2009.
- 110. Sonntag WE, Lynch C, Thornton P, Khan A, Bennett S, and Ingram R. The effects of growth hormone and IGF-1 deficiency on cerebrovascular and brain ageing. J Anat 197: 575–585, 2000.
- 111. Soskic V, Groebe K, and Schrattenholz A. Nonenzymatic posttranslational protein modifications in ageing. Exp Gerontol 43: 247–257, 2008.
- 112. Stadtman ER. Protein modification in aging. J Gerontol 43: B112–B120, 1988.
- 113. Stewart RR and Bewley JD. Lipid peroxidation associated with accelerated aging of soybean axes. Plant Physiol 65: 245–248, 1980.
- 114. Takubo K, Ohmura M, Azuma M, Nagamatsu G, Yamada W, Arai F, Hirao A, and Suda T. Stem cell defects in ATMdeficient undifferentiated spermatogonia through DNA damage-induced cell-cycle arrest. Cell Stem Cell 2: 170–182, 2008.
- 115. Tan M, Li S, Swaroop M, Guan K, Oberley LW, and Sun Y. Transcriptional activation of the human glutathione peroxidase promoter by p53. J Biol Chem 274: 12061–12066, 1999.
- 116. Terzioglu M and Larsson NG. Mitochondrial dysfunction in mammalian ageing. Novartis Found Symp 287: 197–208, 2007.
- 117. Toescu EC, Myronova N, and Verkhratsky A. Age-related structural and functional changes of brain mitochondria. Cell Calcium 28: 329–338, 2000.
- 118. Trinei M, Giorgio M, Cicalese A, Barozzi S, Ventura A, Migliaccio E, Milia E, Padura IM, Raker VA, Maccarana M, Petronilli V, Minucci S, Bernardi P, Lanfrancone L, and Pelicci PG. A p53-p66Shc signalling pathway controls intracellular redox status, levels of oxidation-damaged DNA and oxidative stress-induced apoptosis. Oncogene 21: 3872– 3878, 2002.
- 119. Tyner SD, Venkatachalam S, Choi J, Jones S, Ghebranious N, Igelmann H, Lu X, Soron G, Cooper B, Brayton C, Hee Park S, Thompson T, Karsenty G, Bradley A, and Donehower LA. p53 mutant mice that display early ageingassociated phenotypes. Nature 415: 45–53, 2002.
- 120. van Heemst D, Mooijaart SP, Beekman M, Schreuder J, de Craen AJ, Brandt BW, Slagboom PE, and Westendorp RG. Variation in the human TP53 gene affects old age survival and cancer mortality. Exp Gerontol 40: 11-15, 2005.
- 121. Vanfleteren JR and Braeckman BP. Mechanisms of life span determination in Caenorhabditis elegans. Neurobiol Aging 20: 487–502, 1999.
- 122. Vaziri H, Dessain SK, Ng Eaton E, Imai SI, Frye RA, Pandita TK, Guarente L, and Weinberg RA. hSIR2(SIRT1) functions as an NAD-dependent p53 deacetylase. Cell 107: 149–159, 2001.
- 123. Videla LA, Fernandez V, and Valenzuela A. Age-dependent changes in rat liver lipid peroxidation and glutathione content induced by acute ethanol ingestion. Cell Biochem Funct 5: 273–280, 1987.
- 124. Viner RI, Ferrington DA, Williams TD, Bigelow DJ, and Schoneich C. Protein modification during biological aging: selective tyrosine nitration of the SERCA2a isoform of the sarcoplasmic reticulum Ca2+-ATPase in skeletal muscle. Biochem J 340: 657–669, 1999.
- 125. Vivekananthan DP, Penn MS, Sapp SK, Hsu A, and Topol EJ. Use of antioxidant vitamins for the prevention of cardiovascular disease: meta-analysis of randomised trials. Lancet 361: 2017–2023, 2003.
- 126. Volonte D, Zhang K, Lisanti MP, and Galbiati F. Expression of caveolin-1 induces premature cellular senescence in primary cultures of murine fibroblasts. Mol Biol Cell 13: 2502–2517, 2002.
- 127. Walker DR, Bond JP, Tarone RE, Harris CC, Makalowski W, Boguski MS, and Greenblatt MS. Evolutionary conservation and somatic mutation hotspot maps of p53: correlation with p53 protein structural and functional features. Oncogene 18: 211–218, 1999.
- 128. Wood ZA, Schroder E, Robin Harris J, and Poole LB. Structure, mechanism and regulation of peroxiredoxins. Trends Biochem Sci 28: 32–40, 2003.
- 129. Xu Y. Regulation of p53 responses by post-translational modifications. Cell Death Differ 10: 400–403, 2003.
- 130. Yoon KA, Nakamura Y, and Arakawa H. Identification of ALDH4 as a p53-inducible gene and its protective role in cellular stresses. J Hum Genet 49: 134–140, 2004.
- 131. Zaky A, Busso C, Izumi T, Chattopadhyay R, Bassiouny A, Mitra S, and Bhakat KK. Regulation of the human APendonuclease ( $APE1/Ref-1$ ) expression by the tumor suppressor p53 in response to DNA damage. Nucleic Acids Res 36: 1555–1566, 2008.
- 132. Zamocky M and Koller F. Understanding the structure and function of catalases: clues from molecular evolution and in vitro mutagenesis. Prog Biophys Mol Biol 72: 19–66, 1999.
- 133. Zhu D, Wu J, Spee C, Ryan SJ, and Hinton DR. BMP4 mediates oxidative stress-induced retinal pigment epithelial cell senescence and is overexpressed in age-related macular degeneration. J Biol Chem 284: 9529–9539, 2009.

Address correspondence to: Dr. Yang Xu Bonner Hall 3430 UCSD 0322 9500 Gilman Drive La Jolla, CA 92093-0322

E-mail: yangxu@ucsd.edu

Date of first submission to ARS Central, September 13, 2010; date of final revised submission, October 14, 2010; date of acceptance, November 4, 2010.

#### Abbreviations Used ALDH = aldehyde dehydrogenase  $APE/Ref1 = a$ purinic/apyrimidinic endonuclease/ redox factor-1  $BAI1 = brain-specific angiogenesis inhibitor 1$ Dmp53 = Drosophila melanogaster p53  $ES$  cell = embryonic stem cell  $FoxO =$  for khead box  $O$  $GPX = glutathione peroxidase$  $GST = glutathione S-transferase$ hBMSCs = human bone marrow-derived mesenchymal stem cells  $MEF = mouse$  embryonic fibroblast NQO1 = NAD(P)H dehydrogenase [quinone] 1  $Nrf1 = NF-E2$ -related factor 2  $\mathrm{PGM} = \mathrm{phosphoglycerate}$  mutase  $PIG3 = p53$ -inducible gene 3 Puma  $=$  p53 upregulated modulator of apoptosis  $ROS = reactive$  oxygen species  $SGK =$  serum- and glucocorticoid-inducible kinase  $SOD = superoxide$  dismutase  $TIGAR = TP53$ -induced glycolysis and apoptosis regulator  $TP53INP1 =$ tumor protein 53-induced nuclear protein 1

 $TSP1 =$ thrombospondin-1