

NIH Public Access

Author Manuscript

Science. Author manuscript; available in PMC 2012 February 5.

Published in final edited form as:

Science. 2011 August 5; 333(6043): 753-755. doi:10.1126/science.1206883.

Extending the Carbon Chain: Hydrocarbon Formation Catalyzed by Vanadium/Molybdenum Nitrogenases*

Yilin Hu**,†, **Chi Chung Lee****, and **Markus W. Ribbe**†

Department of Molecular Biology and Biochemistry, University of California, Irvine, CA 92697-3900

Abstract

Vanadium-dependent nitrogenase was previously shown in a small-scale reaction to catalyze reductive catenation of CO to C_2H_4 , C_2H_6 , C_3H_6 and C_3H_8 . Here, we report the identification of additional hydrocarbon products, α -C₄H₈, *n*-C₄H₁₀ and CH₄, in a scaled-up reaction featuring 20 milligrams of vanadium-iron protein, the catalytic component of vanadium nitrogenase. Additionally, we show that the more common molybdenum-dependent nitrogenase can generate the same hydrocarbons from CO, although $CH₄$ was not detected. The identification of CO as a substrate for both molybdenum- and vanadium-nitrogenases strengthens the hypothesis that COreduction is an evolutionary relic of the function of the nitrogenase family; moreover, the comparison between the CO-reducing capacities of the two nitrogenases suggests that the identity of heterometal at the active cofactor site impacts the efficiency and product distribution of this reaction.

> Nitrogenase enzymes catalyze the reduction of dinitrogen (N_2) to ammonia (NH_3) , a key step in the global nitrogen cycle (1). The molybdenum (Mo)- and vanadium (V)-dependent nitrogenases are two homologous members of this enzyme family, distinguished primarily by the heterometals (Mo or V) at their respective cofactor centers (FeMoco or FeVco) (2). Both nitrogenases are composed of a specific reductase (*nifH*- or *vnfH-*encoded Fe protein) and a catalytic component (*nifDK*-encoded MoFe protein or *vnfDGK-*encoded VFe protein). Additionally, both enzymes presumably form a functional complex between the two component proteins during catalysis (3), in which electrons are transferred in an adenosine triphosphate (ATP)-dependent process from the reductase to the cofactor site of the catalytic component, where substrate reduction eventually occurs.

> The homology between Mo- and V-nitrogenases renders their substrate profiles highly similar (4); yet, the two nitrogenases react differently with carbon monoxide (CO). In a small-scale reaction (5, 6), V nitrogenase catalyzes reductive catenation of CO to ethylene (C_2H_4) , ethane (C_2H_6) and propane (C_3H_8) ; in contrast, these products have not been detected in the same reaction catalyzed by the wild-type Mo nitrogenase, although a very recent report has indicated that the Val^{q70} -substituted Mo nitrogenase variants can catalyze the formation of low molecular-weight hydrocarbon products from CO (7–9). The ability of V nitrogenase to catalyze hydrocarbon formation from CO is somewhat analogous to the capacities of late transition metal catalysts in the industrial Fischer-Tropsch process (10). However, the enzymatic reaction uses H^+ —instead of H_2 —for hydrocarbon formation (11),

^{*}This manuscript has been accepted for publication in Science. This version has not undergone final editing. Please refer to the complete version of record at <http://www.sciencemag.org/>. The manuscript may not be reproduced or used in any manner that does not fall within the fair use provisions of the Copyright Act without the prior, written permission of AAAS.

[†]To whom correspondence should be addressed. yilinh@uci.edu (Y.H.) or mribbe@uci.edu (M.W.R.).

These authors contributed equally to this work.

and it requires the participation of both components of the enzyme, the hydrolysis of ATP, and the supplies of electron sources (7). Thus, V nitrogenase-based hydrocarbon formation likely involves the reductive protonation of CO, a process that parallels the nitrogenasebased NH₃ formation pathway, which involves the reductive protonation of N_2 .

A fourth hydrocarbon product of V nitrogenase catalysis—propylene (C_3H_6) —can be detected in the standard reaction upon the substitution of H_2O by D_2O (11). One possible explanation for this effect is a stalling of the reaction by D_2O and the consequent accumulation of intermediates that are normally turned over quickly into other products. This observation leads to the speculation that other hydrocarbon products may have remained unidentified due to their minor yields. In this scenario, a sufficiently scaled-up reaction would allow us to identify these products either directly in H_2O or following the substitution of H_2O by D_2O .

Indeed, when we scaled up the reaction by 130-fold, C_3H_6 , along with C_2H_4 , C_2H_6 and C_3H_8 , was readily detected in the H₂O-based reactions using either ¹²CO (Fig. S1B and C, *traces 1*) or 13CO (Fig. S1B and C, *traces 2*) as the substrate (12). Moreover, we identified three additional products in the large-scale, H₂O-based assays. Gas chromatography-mass spectroscopy (GC-MS) analyses revealed the identities of these products as methane (CH_4) , α-butylene (C₄H₈) and *n*-butane (C₄H₁₀), which displayed *m/z* ratios of 16.032, 56.064 and 58.080, respectively, in the reaction of 12CO (Fig. S1A and D, *traces 1*) and the expected mass shifts of $+1$, $+4$ and $+4$, respectively, on substituting ¹³CO for ¹²CO (Fig. S1A and D, *traces 2*). This complete set of products was also detected in the D_2O -based reactions with the mass shifts expected for the D/H substitution (Fig. S1A-D, *traces 3* and *4*).

Unexpectedly, when V nitrogenase was replaced by Mo nitrogenase in the scaled-up reaction, the same two- and three-carbon products were detected in the H_2O -based reaction (Fig. S2B and C). Thus, like its V counterpart, Mo nitrogenase used CO and H^+ as the carbon and hydrogen sources for hydrocarbon formation. Furthermore, the deuterium effect documented for C₃H₆ in the small-scale V nitrogenase reaction (11) was duplicated for α- C_4H_8 and $n-C_4H_{10}$ in the scaled-up Mo nitrogenase reaction. These four-carbon products, which were not detected in the H2O-based reaction (Fig. S2D, *traces 1* and *2*), could be clearly identified in the D₂O-based reaction (Fig. S2D, *traces 3* and 4). In contrast, the onecarbon product (CH₄) remained unidentified even upon the substitution of H₂O by D₂O (Fig. S2A, *traces 3* and *4*), suggesting that CH4 (if any) was formed at an extremely low yield in the Mo nitrogenase-catalyzed reaction (13).

The observation of CO-reduction by Mo nitrogenase comes as a big surprise, as CO has never been shown to be a substrate of Mo nitrogenase; instead, it has long been regarded as a potent inhibitor for the reduction of all known substrates of Mo nitrogenase except $H^+(1)$. The presence of CO suppresses the ability of Mo nitrogenase to turn over substrates other than $H⁺$ and, consequently, the electrons which are normally used for the reduction of these substrates are routed toward the concomitant reduction of H^+ . The fact that CO inhibits the V nitrogenase-catalyzed H⁺-reduction by \sim 76% suggests a re-routing of electrons from H⁺reduction to CO-reduction, an argument supported by the formation of hydrocarbon products from CO in this reaction $(4, 7)$; in contrast, CO exerts a minimal impact on the H⁺reduction by Mo nitrogenase, indicating that an almost untraceable quantity of electrons (14) are diverted from H^+ -reduction to CO-reduction, thereby limiting the ability of Mo nitrogenase to generate hydrocarbons from CO.

Consistent with this hypothesis, quantitative GC analyses show that Mo nitrogenase (Fig. 1B) is considerably less efficient than its V counterpart (Fig. 1A) in hydrocarbon formation, and the total specific activity of product formation by the former is only \sim 0.1% and \sim 2%,

respectively, of that by the latter in H_2O - and D_2O -based reactions (Fig. 1C). The disparate CO-reducing activities of V- and Mo-nitrogenase parallel the differential capacities of synthetic V- and Mo-compounds to reductively couple two CO moieties into functionalized acetylene ligands (15), suggesting that V (a group VB transition metal) is superior to Mo (a group VIB transition metal) in this particular type of reaction. The substitution of H_2O by D2O, however, has a much more dramatic effect on the reaction catalyzed by Mo nitrogenase than that catalyzed by its V counterpart, leading to an increase in the specific activity of total product formation by ~21-fold in the case of the former (Fig. 1C, Mo, *2 vs. 1*) and by only ~12% in the case of the latter (Fig. 1C, V, 2 vs. 1). In both cases, the increase in hydrocarbon formation is accompanied by a \sim 30% decrease in D₂ evolution, suggesting a possible shift of electrons from D^+ -reduction toward CO-reduction (16).

A closer examination of the product distributions of V- and Mo-nitrogenases reveals additional features that set the reactions catalyzed by the two nitrogenases apart (Fig. 2; Table S1). Compared to V nitrogenase (Fig. 2A, 1), Mo nitrogenase (Fig. 2B, 1) displays a much lower C_2H_4/C_2H_6 ratio in its product profile; moreover, Mo nitrogenase generates a lower percentage of two-carbon products, and rather seems to shift the H_2O -based reaction toward the formation of C_3H_6 and C_3H_8 . In the presence of D_2O , however, Mo nitrogenase shifts the product distribution back toward two-carbon products; additionally, there is an appreciable increase in the C_2H_4/C_2H_6 ratio of its product profile (Fig. 2B, 2). Contrary to Mo nitrogenase, V nitrogenase is less affected by the deuterium effect; nevertheless, it shows a small decrease in the C_2H_4/C_2H_6 ratio of its product profile and a slight shift toward the formation of three- and four-carbon products—an opposite trend compared to its Mo counterpart—in the D₂O-based reaction (Fig. 2A, 2). Strikingly, these opposite changes in D_2O render a closer resemblance between the product profiles of V- and Mo-nitrogenases (Fig. 2A *vs*. B, *2*), pointing to possible similarities between the CO-reducing mechanisms of these two nitrogenases.

One common trait shared by the reactions catalyzed by V- and Mo-nitrogenases is the overwhelmingly predominant formation/release of C_2H_4 (Fig. 1 and 2). The fact that CH₄ the one-carbon product that could result directly from the reductive protonation of CO—is observed in such lower abundance than C_2H_4 suggests that the formation of a C-C bond between CO derivatives is relatively rapid. The observation that C_2H_6 —the two-carbon product that could originate from the reductive protonation of C_2H_4 —drops sharply in quantity relative to C_2H_4 indicates that the release of C_2H_4 is favored over the further reduction of this product. Furthermore, the huge discrepancy between the amounts of twoand three-/four-carbon products points to a potential competition between the formation of two-carbon products (particularly C_2H_4) and the extension of the carbon chain from the same two-carbon intermediate. The preferred C_2H_4 formation catalyzed by nitrogenases likely reflects a dominant impact of kinetic factors that guide the overall product distribution, resulting in a drastically decreased yield of hydrocarbon formation beyond twocarbon-length.

A second conserved feature between the reactions catalyzed by the two nitrogenases is the formation of products in the following rank: $CH_2=CH_2 \rightarrow CH_3=CH_3=CH_2=CH_2$ $CH_3 > CH_2=CH_2-CH_3 > CH_3-CH_2-CH_2-CH_3 > CH_2=CH_2-CH_2-CH_3$ (Fig. 1 and 2). As such, except for the two-carbon products, both nitrogenases display a preference toward the formation of saturated, three- or four-carbon alkanes rather than their corresponding, unsaturated alkenes, suggesting a possible mechanistic switch following the formation of the first C-C bond. One plausible account for the reaction beyond the two-carbon stage involves the formation of alkene prior to a fast protonation step that converts alkene to alkane. Such a hypothesis is supported by two lines of evidence. First, Mo nitrogenase displays a product profile of higher three- and four-carbon alkene/alkane ratios than its V counterpart (Table

Science. Author manuscript; available in PMC 2012 February 5.

S1), which could be explained by its limited access to electrons for CO-reduction that slows down the coupled electron/proton addition to the unsaturated C=C bonds of alkenes. Second, both V- and Mo-nitrogenases show increased three- and four-carbon alkene/alkane ratios in the presence of D_2O (Table S1), which may reflect a stalling effect of deuterium on the protonation step (16).

The discrepancy between the CO-reducing capacities of Mo- and V-nitrogenases suggests a prevailing impact of heterometal chemistry on the efficiency of the reaction; at the same time, the merging of the product profiles of the two nitrogenases in $D₂O$ points to a critical role of coupled electron/proton tunneling in the mechanism of this particular reaction. The discovery that Mo nitrogenase is capable of utilizing CO as a substrate—albeit at a considerably lower rate than its V counterpart—more broadly supports the hypothesis that CO-reduction is an evolutionary relic of the function of this enzyme family. The parallelism between the nitrogenase-catalyzed CO- and N_2 -reduction—demonstrated further by the ability of V nitrogenase to form CH_4 , an analogous product of NH_3 —strengthens the theory that the ancestral nitrogenase may represent an evolutionary link between carbon and nitrogen cycles on earth (7). In a practical vein, the industrial Fischer-Tropsch process uses H2—a costly syngas component—for CO-reduction and shows a tendency toward excessive formation of CH_4 —a low-value product (17) whereas the nitrogenase-based reaction utilizes H+ for hydrocarbon formation and favors the formation of C-C bond over the cleavage of C-O bond. These features make nitrogenase an attractive template for cost-efficient hydrocarbon formation that is directed toward C-C coupling and carbon chain extension. Borrowing a trick or two from this ancient enzyme family, perhaps an efficient strategy could be developed in the future for controlled fuel production from CO?

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We thank Prof. D. C. Rees and Dr. N. Dalleska of Caltech (Pasadena) for help on the GC-MS analysis. This work was supported by Herman Frasch Foundation grant 617-HF07 (M.W.R.) and National Institutes of Health grant GM 67626 (M.W.R.).

References and Notes

- 1. Burgess BK, Lowe DJ. Chem Rev. 1996; 96:2983. [PubMed: 11848849]
- 2. Hales B. Avd Inorg Biochem. 1990; 8:165.
- 3. Schindelin H, Kisker C, Schlessman JL, Howard JB, Rees DC. Nature. 1997; 387:370. [PubMed: 9163420]
- 4. Lee CC, Hu Y, Ribbe MW. Proc Natl Acad Sci U S A. 2009; 106:9209. [PubMed: 19478062]
- 5. A small-scale nitrogenase reaction typically features 0.15 mg VFe or MoFe protein, the catalytic component of V- or Mo-nitrogenase. Such an assay condition was established empirically some 30 years ago (*6*) and has since been used as the conventional scale of *in vitro* nitrogenase assays in the field.
- 6. Burgess BK, Jacobs DB, Stiefel EI. Biochim Biophys Acta. 1980; 614:196. [PubMed: 6930977]
- 7. Lee CC, Hu Y, Ribbe MW. Science. 2010; 329:642. [PubMed: 20689010]
- 8. Based on the retention time on a gas chromatograph column, the low molecular-weight hydrocarbon products formed by the Val^{q70} -substituted Mo nitrogenase variants have been assigned as methane, ethylene, ethane, propylene and propane (*9*).
- 9. Yang ZY, Dean DR, Seefeldt LC. J Biol Chem. 2011; 286:19417. [PubMed: 21454640]
- 10. Rofer-DePoorter CK. Chem Rev. 1981; 81:447.

Hu et al. Page 5

- 11. Lee CC, Hu Y, Ribbe MW. Angew Chem Int Ed Engl. 2011; 50:5545. [PubMed: 21538750]
- 12. Materials and methods are detailed in supporting material online.
- 13. Given the detection threshold of CH_4 (0.0007 nmol/nmol protein/min) and the activity of V nitrogenase in CH4 production (0.12 nmol/nmol protein/min), the Mo nitrogenase is at least 170 fold less active than its V counterpart in catalyzing the formation of $CH₄$ from CO.
- 14. Only 0.04% of electrons can be traced in the hydrocarbon products formed in the Mo nitrogenasecatalyzed reaction.
- 15. Carnahan EC, Protasiewicz JD, Lippard SJ. Acc Chem Res. 1993; 26:90.
- 16. The effect of deuterium on nitrogenase-catalyzed CO-reduction is likely multifaceted. Apart from the inverse kinetic isotope effects (*i.e.*, $k_H/k_D < 1$) of D₂O that favor the formation of deuterated products (18 , 19), other solvent effects of D_2O , such as those affecting the protein conformation, the interaction between proteins, and the network of hydrogen bonds, have been observed (*20–23*).
- 17. Khodadadi AA, Hudgins RR, Silveston PL. Can J Chem Engin. 1996; 74:695.
- 18. Kurtz KA, Fitzpatrick PF. J Am Chem Soc. 1997; 119:1155.
- 19. Churchill DG, Janak KE, Wittenberg JS, Parkin G. J Am Chem Soc. 2003; 125:1403. [PubMed: 12553844]
- 20. Karsten WE, Lai CJ, Cook PF. J Am Chem Soc. 1995; 117:5914.
- 21. Morgan TV, et al. Biochemistry. 1990; 29:3077. [PubMed: 2159783]
- 22. Cassman M. Arch Biochem Biophys. 1974; 165:60. [PubMed: 4374134]
- 23. Sheu SY, Schlag EW, Selzle HL, Yang DY. J Phys Chem A. 2008; 112:797. [PubMed: 18193847]
- 24. Glasoe PK, Long FA. J Phys Chem. 1960; 64:188.
- 25. Gavini N, Burgess BK. J Biol Chem. 1992; 267:21179. [PubMed: 1400428]

Fig. 1.

Specific activities for individual product formation catalyzed by (**A**) V- and (**B**) Monitrogenase and (**C**) comparison of total activities for hydrocarbon formation by the two nitrogenases in the presence of (1) H₂O or (2) D₂O. Legend: ① CH₄; ② C₂H₄; ③ C₂H₆; ④ C_3H_6 ; \textcircled{S} C_3H_8 ; \textcircled{S} α -C₄H₈; \textcircled{S} *n*-C₄H₁₀. Expanded scales of α -C₄H₈ and *n*-C₄H₁₀ formation are shown as insets. Specific activities were determined by quantitative GC analyses and calculated based on the formation of products (see Fig. S3) in the first 10 min of reaction (linear range). Data are presented as mean \pm SD ($N = 5$).

Fig. 2.

Distributions of hydrocarbons formed by (**A**) V- and (**B**) Mo-nitrogenases in the presence of (1) H_2O or (2) D_2O . The total amounts of hydrocarbons detected in V (A)- and Mo (B)based reactions were set as 100%, respectively, and the percentages of individual products were determined accordingly for each nitrogenase. The alkene/alkane ratios of V- and Monitrogenases are summarized in Table S1.