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MicroRNA Processing and Binding Site Polymorphisms are not Replicated in the Ovarian Cancer Association Consortium

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Abstract

Background—Single nucleotide polymorphisms (SNPs) in microRNA-related genes have been associated with epithelial ovarian cancer (EOC) risk in two reports, yet associated alleles may be inconsistent across studies.

Methods—We conducted a pooled analysis of previously-identified SNPs by combining genotype data from 3,973 invasive EOC cases and 3,276 controls from the Ovarian Cancer Association Consortium. We also conducted imputation to obtain dense coverage of genes and comparable genotype data for all studies. In total, 226 SNPs within 15 kilobases of 4 miRNA biogenesis genes (*DDX20, DROSHA, GEMIN4,* and *XPO5)* and 23 SNPs located within putative miRNA binding sites of 6 genes (*CAV1, COL18A1, E2F2, IL1R1, KRAS,* and *UGT2A3*) were genotyped or imputed and analyzed in the entire dataset.

Results—After adjustment for European ancestry, no overall association was observed between any of the analyzed SNPs and EOC risk.

Conclusions—Common variants in these evaluated genes do not appear to be strongly associated with EOC risk.

Impact—This analysis suggests earlier associations between EOC risk and SNPs in these genes may have been chance findings, possibly confounded by population admixture. To more adequately evaluate the relationship between genetic variants and cancer risk, large sample sizes are needed, adjustment for population stratification should be performed, and use of imputed SNP data should be considered.

Keywords

miRNA processing; binding sites; inherited susceptibility; ovarian cancer; genetic variants

Introduction

MicroRNAs (miRNAs) are short, non-coding RNAs that regulate translation (1). SNPs in precursor and mature miRNAs, their processing machinery, or in miRNA binding sites of target genes have been implicated in cancer risk (2). Liang et al. (3) analyzed 238 SNPs from 8 miRNA processing genes and 138 genes containing potential miRNA binding sites in 339 EOC cases and 349 controls self-reported to be Caucasian, and identified associations between EOC risk and 13 SNPs from 4 processing genes (*DDX20, DROSHA/RNASEN, GEMIN4, XPO5)* and 7 binding site genes (*ATG4A, CAV1, COL18A1, E2F2, IL1R1, KRAS,* and *UGT2A3)*. We (4) genotyped 318 SNPs in 18 miRNA processing genes in 2,172 EOC

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cases and 3,052 controls of European ancestry, and identified 6 SNPs from 4 genes (*DROSHA, FMR1, LIN28, LIN28B*) as significantly associated with EOC risk. Here we conducted a pooled analysis of variants reported as risk-associated by Liang et al (3) in 3,973 cases and 3,276 controls from the international Ovarian Cancer Association Consortium (OCAC) (5). We imputed SNPs to expand coverage of genes and regions, totaling 249 SNPs from 10 of the 11 highlighted genes (3).

Material and Methods

Participating OCAC studies were from North America (US-CAN), the United Kingdom (UK), and Poland (POL). Study characteristics have been reported (4) and are summarized in Table 1. Briefly, cases had pathologically-confirmed primary invasive EOC. Controls had at least one ovary intact when interviewed. All studies collected data on disease status, selfreported ethnicity, and histologic subtype. Subjects with <80% European ancestry were excluded (4), and the first two principal components (PCs) representing European ancestry were estimated for all SNPs with call rates >99% using Golden Helix SVS PCA function, algorithmically equivalent to EigenSTRAT. The protocol was approved by the institutional review board at each site, and all participants provided written informed consent. Pooled data included 3,973 cases (51% serous) and 3,276 controls.

SNP genotyping and quality control have been described (4, 6). SNP imputation was carried out within studies (US-CAN, UK, POL) with MACH version 1.0.16 using CEU phased data from HapMap release 22 (genome build 36). We imputed data for 186 SNPs that span 15 kb upstream and downstream of each miRNA processing gene or reside in a putative miRNA binding site in the 3′ UTR of target genes as predicted by SNPInfo (7) and/or PolymiRTS (8); the remaining 63 SNPs were directly genotyped.

Study-specific odds ratios (OR) and 95% confidence intervals (CI) were estimated using unconditional logistic regression. Log-additive genetic models were fit for each SNP, modeling the number of copies of the minor allele. For imputed SNPs, we used expected counts of minor alleles obtained from MACH. Study-specific estimates were adjusted for age at diagnosis/interview (US-CAN, POL), component study sites (US-CAN), and the first two PCs (US-CAN, UK, POL). Allele frequencies across studies were similar, suggesting low genetic heterogeneity between populations and appropriateness for combining data. Pooled estimates were adjusted for a) study (US-CAN, UK, POL) and b) study and the first two PCs. We used PLINK for statistical analysis (10).

Results

Two hundred twenty-six SNPs were evaluated within or near miRNA processing genes *DDX20* (n=17), *DROSHA* (n=179), *GEMIN4* (n=11), and *XPO5* (n=19). Table 2 displays association results for the 6 processing SNPs (or their tagSNPs) identified by Liang et al. (3); none were risk-associated. Of all other miRNA processing SNPs evaluated, only 3 *DROSHA* SNPs were associated with risk (*P*<0.05) when accounting for study site only, but none retained statistical significance after further adjustment for ancestry (See Supplemental Table 1).

There were 23 SNPs predicted to disrupt miRNA binding within 6 of the 7 candidate genes (3). We did not evaluate SNPs within *ATG4A* because neither genotype nor imputed data were available for SNPs within the 3′ UTR. Table 2 shows results from the 6 binding site SNPs (or their tagSNPs) identified by Liang et al. (3). To minimize redundancy due to tagSNPs, results from 21 of the 23 binding site SNPs evaluated are displayed in Supplemental Table 1. Only one previously-identified binding site SNP, *CAV1* rs9920 (3),

and two imputed *CAV1* SNPs (rs1049314 and rs8713) were associated with risk in the pooled, study site-adjusted analysis (Table 2; Supplemental Table 1). However, none of these *CAV1* SNPs were risk-associated after further adjustment for ancestry.

Study-specific estimates were generally similar across studies, and results did not change appreciably when considering a dominant genetic model or serous-only histology (data not shown).

Discussion

We did not detect consistent associations between the majority of previously-identified polymorphisms (3) and EOC risk. Although we did identify associations between EOC risk and 3 SNPs flanking the 3′UTR of *DROSHA* and 3 SNPs in miRNA binding sites of *CAV1,* none retained statistical significance after controlling for European ancestry. Consistent with recent large-scale (11) but not smaller studies (3, 12), we did not identify associations between EOC risk and SNPs in miRNA binding sites of *KRAS*.

Several explanations exist for not replicating the findings presented by Liang et al. (3). First, our analysis suggests their results may be confounded by population admixture, underscoring the importance of estimating population stratification rather than relying on self-reported ancestry in genetic association studies. Due to their relatively small sample size (3), chance is an alternate explanation for their findings. Our pooled sample had at least 90% statistical power to detect a SNP with a minor allele frequency of 0.09 and a log-additive OR of 1.2. This analysis highlights the importance of having large studies and/or combining genotype data from multiple studies to increase statistical power to detect true associations, and demonstrates the utility of population stratification and imputation.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Totals represent the number of non-Hispanic white Europeans passing genotyping quality control criteria and meeting study site-specific inclusion/exclusion criteria. *1*Totals represent the number of non-Hispanic white Europeans passing genotyping quality control criteria and meeting study site-specific inclusion/exclusion criteria.

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2 Cases from NEC that were evaluated as part of this investigation represent postmenopausal advanced papillary serous carcinomas; 26 of these cases were ascertained as part of a hospital-based pre-*2*Cases from NEC that were evaluated as part of this investigation represent postmenopausal advanced papillary serous carcinomas; 26 of these cases were ascertained as part of a hospital-based preoperative study operative study

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Table 2

Association between selected miRNA processing and miRNA binding site SNPs and epithelial ovarian cancer risk in a pooled analysis. Association between selected miRNA processing and miRNA binding site SNPs and epithelial ovarian cancer risk in a pooled analysis.

Abbreviations: US-CAN=United States-Canada; UK=United Kingdom; POL=Poland; maj=major; min=minor; miR=miRNA; UTR= untranslated region; ns=non- synonymous SNP; ss=synonymous SNP; 5 ξ λ λ á OR (CI) =odds ratio (confidence interval); MAF=minor allele frequency among all controls; all P-values are two-sided. *P*-values are two-sided. OR (CI) =odds ratio (confidence interval); MAF=minor allele frequency among all controls; all

 $a_{\text{The major allele represents the most frequently-occurring allele and serves as the reference allele during modeling.}$ *a*The major allele represents the most frequently-occurring allele and serves as the reference allele during modeling.

 b SNP location derived from Illumina annotation files. HapMap2 data (http://hapmap.ncbi.nlm.nih.gov/), and dbSNP (http://www.ncbi.nlm.nih.gov/projects/SNP/). SNPinfo http://snpinfo.niehs.nih.gov/ and *b*SNP location derived from Illumina annotation files, HapMap2 data [\(http://hapmap.ncbi.nlm.nih.gov/](http://hapmap.ncbi.nlm.nih.gov/)), and dbSNP ([http://www.ncbi.nlm.nih.gov/projects/SNP/\)](http://www.ncbi.nlm.nih.gov/projects/SNP/). SNPinfo<http://snpinfo.niehs.nih.gov/> and the PolymiRTS database (http://compbio.uthsc.edu/miRSNP) were used to predict miRNAs whose binding activity may be altered due to the SNP location. the PolymiRTS database (<http://compbio.uthsc.edu/miRSNP>) were used to predict miRNAs whose binding activity may be altered due to the SNP location.

Genotype data was imputed for all participants using MACH version 1.0.16 using phased data from HapMap release 22 (genome build 36) derived from individuals with European ancestry (CEU). *c*Genotype data was imputed for all participants using MACH version 1.0.16 using phased data from HapMap release 22 (genome build 36) derived from individuals with European ancestry (CEU).

 d Pooled OR and 95% CI estimated using a log-additive model adjusted for study (US-CAN, UK, POL) *d*Pooled OR and 95% CI estimated using a log-additive model adjusted for study (US-CAN, UK, POL)

Pooled OR and 95% CI estimated using a log-additive model adjusted for study and the first two principal components representing European ancestry *e*Pooled OR and 95% CI estimated using a log-additive model adjusted for study and the first two principal components representing European ancestry

 $f_{\rm{DDX20~rs19714~is~in~linkage}}$ disequilibrium (LD) (r² =0.90) with rs197383 identified by Liang et al. 2 =0.90) with rs197383 identified by Liang et al. *fDDX20* rs19714 is in linkage disequilibrium (LD) (r

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 8DROSHA rs9292427 is in LD (r^2 =0.98) with rs4867329 identified by Liang et al. 2 =0.98) with rs4867329 identified by Liang et al. gDROSHA rs9292427 is in LD (r

 h SNP deviates from Hardy Weinberg Equilibrium among all controls with PHWE values of 0.020 for rs607613, 0.040 for rs615435, 0.013 for rs2740349, 0.004 for rs3732133, and 0.034 for rs17147016, *h*SNP deviates from Hardy Weinberg Equilibrium among all controls with PHWE values of 0.020 for rs607613, 0.040 for rs615435, 0.013 for rs2740349, 0.004 for rs3732133, and 0.034 for rs17147016, respectively. respectively.

iGEMIN4 SNP pair in LD (r iGEMIN4 SNP pair in LD $(r^2$ =1)

jE2F2 SNP pair in LD (r j_{E2F2} SNP pair in LD (r² =0.97) k KRAS rs13096 is in LD (r^2 =1) with rs10771184 identified by Liang et al. $2 = 1$) with rs10771184 identified by Liang et al. *kKRAS* rs13096 is in LD (r