

# Compounds Which Serve as the Sole Source of Carbon or Nitrogen for *Salmonella typhimurium* LT-2

DAVID GUTNICK,<sup>1</sup> JOSEPH M. CALVO, TADEUSZ KLOPOTOWSKI, AND BRUCE N. AMES

National Institutes of Health, Bethesda, Maryland 20014, Department of Biochemistry and Molecular Biology, Cornell University, Ithaca, New York 14850, Institute of Biochemistry and Biophysics, Polish Academy of Sciences, Warsaw 12, Poland, and Department of Biochemistry, University of California, Berkeley, California 94720

Received for publication 7 August 1969

About 600 compounds were screened as possible carbon or nitrogen sources for *Salmonella typhimurium* LT-2. About 100 utilizable compounds were found.

In an attempt to provide new genetic markers in the chromosome of *Salmonella typhimurium*, a search for compounds which can serve as a sole carbon source or as a sole nitrogen source for this organism was undertaken. About 100 utilizable compounds were found. Mapping of the genes concerned with the utilization of these compounds should add considerable detail to the known genetic map (5) on which 200 genes are already localized.

These compounds will also be useful in the investigation of strains carrying deletion mutations if a gene involved in the utilization of one of these compounds is deleted. For example, certain strains, in which part of the leucine operon is deleted, cannot utilize arabinose as a sole source of carbon because the deletion extends into the nearby arabinose operon (2), while a strain with a leucine deletion extending in the other direction cannot utilize a number of other carbon sources (Calvo et al., unpublished data). In contrast, we have determined that *his-520*, in which a large deletion extends through the histidine operon and lipopolysaccharide (rfb) regions, grows on all the listed sources of carbon and nitrogen.

The list of compounds should also be useful for taxonomic studies comparing related enteric bacteria. Such studies often make use of smaller lists of compounds (1, 4).

## MATERIALS AND METHODS

Compounds were tested for their ability to support the growth of *S. typhimurium* strain LT-2 by a simple auxanographic technique. Minimal salts-agar was prepared by dissolving the following in a liter of distilled

water: K<sub>2</sub>SO<sub>4</sub>, 1.0 g; K<sub>2</sub>HPO<sub>4</sub>, 13.5 g; KH<sub>2</sub>PO<sub>4</sub>, 4.7 g; MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.1 g; agar, 15 g. For testing possible carbon sources, plates containing 30 ml of sterile minimal agar were overlaid with 2 ml of soft agar (0.6% in 0.5% NaCl solution) containing 0.1 ml of an overnight nutrient broth culture of bacteria and 0.1 ml of 0.1 M NH<sub>4</sub>Cl. For testing possible nitrogen sources, the 0.1 M NH<sub>4</sub>Cl was replaced by 0.2 ml of 50% glucose. When testing the amino acids as nitrogen sources, an additional set of plates was used containing 0.1% citrate as a sole carbon source.

Approximately 2 mg of each compound was placed on the surface of the solidified agar, and growth of the bacteria in the agar was recorded after incubation for 2 days at 37 C. Because the compound diffuses from the original point of deposition, a concentration gradient is established which allows the organism to grow even if certain concentrations are inhibitory. For example, compounds such as lactic acid, tested without prior neutralization, resulted in an inner zone of inhibition surrounded by a region of profuse growth. As controls, compounds were placed on similar plates lacking bacteria. No more than four compounds were tested on a single plate, and those compounds which gave positive results were each retested on an individual plate.

All chemicals were obtained from commercial sources; their purity was not determined, although several independent batches were tried whenever a positive response was obtained. This auxanographic technique was found to be simple, sensitive, and reliable.

## RESULTS AND DISCUSSION

We have found about 100 utilizable carbon (Table 1) and nitrogen (Table 2) sources out of some 600 compounds that were screened. We first tested all of the natural compounds that seemed likely to be utilizable; this yielded most of our 100 compounds. Further screening yielded

<sup>1</sup> Present address: Department of Microbiology, Tel Aviv University, Ramat Aviv, Israel.

TABLE 1. Compounds which serve as sole carbon sources for *Salmonella typhimurium* LT-2

Acetic acid	L-Fucose	Guanosine	Oleic acid
N-acetyl-D-glucosamine	Fumaric acid	Inosine	Oxaloacetic acid
N-acetyl-D-mannosamine	Galactaric acid (mucic acid)	meso-Inositol	3-Phosphoglyceric acid
Adenosine <sup>a</sup>	Galactitol (dulcitol)	DL-Isocitric acid	L-Proline <sup>a</sup>
L-Alanine <sup>a</sup>	D-Galactose	$\alpha$ -Ketoglutaric acid	Propionic acid
L-Arabinose	D-Glucaric acid (saccharic acid)	L-Lactic acid	Pyruvic acid
DL-Citramalic acid	D-Gluconolactone	Lauric acid	L-Rhamnose
Citric acid	D-Gluconic acid	L-Lyxose	D-Ribose
L-Cysteine <sup>a</sup>	D-Glucosamine	L-Malic acid	D-Serine <sup>a</sup>
Cytidine	D-Glucose	D-Maltose	L-Serine <sup>a</sup>
2-Deoxyadenosine	D-Glucose-6-phosphate	D-Mannitol	D-Sorbitol
2-Deoxycytidine	D-Glucuronic acid	D-Mannose	Succinic acid
2-Deoxyguanosine	D-Glucuronolactone	D-Mannosamine	meso-Tartaric acid <sup>b</sup>
2-Deoxy-D-ribose	DL-glyceric acid	Melibiose	Thymidine
Deoxyuridine	Glycerol	L-Methionyl-L-alanine <sup>a</sup>	D-Trehalose
Dihydroxyacetone	$\alpha$ -Glycerophosphate	6-Methylaminopurine riboside	Tricarballic acid
D-Erythrose <sup>b</sup>	Glycolic acid	$\alpha$ -Methyl-D-galactoside	Tridecanoic acid
D-Fructose		Myristic acid	Uridine
D-Fructose-6-phosphate			D-Xylose

<sup>a</sup> Can serve as the sole source of both carbon and nitrogen.

<sup>b</sup> The bacteria will mutate to use this compound.

TABLE 2. Compounds which serve as sole nitrogen sources for *Salmonella typhimurium*

Adenosine <sup>a</sup>	Deoxycytidine
L-Alanine <sup>a</sup>	D-Glutamic acid
Ammonium chloride	L-Glutamic acid
L-Arginine	L-Glutamine
D-Asparagine	Glutathione (reduced)
L-Asparagine	Glycine
D-Aspartic acid	L-Homoserine
L-Aspartic acid	DL- $\beta$ -Hydroxy glutamic acid
L-Cysteine <sup>a</sup>	L-Methionyl-L-alanine <sup>a</sup>
L-Cystine	L-Proline <sup>a</sup>
Cytidine	D-Serine <sup>a</sup>
Cytosine	L-Serine <sup>a</sup>
Deoxyadenosine	

<sup>a</sup> Can serve as the sole source of both carbon and nitrogen.

few new utilizable compounds; therefore, we feel that the list is probably fairly exhaustive in its present form. Table 3 lists the compounds which are not utilizable as sole carbon or nitrogen sources.

The utilization by *Escherichia coli* of a number of carbon sources related to the Krebs cycle has recently been reviewed, with particular emphasis on regulation (3). The pattern of the utilization of fatty acids is of some interest, as there are detailed studies of this in *E. coli* (7). The pattern is fairly similar; the C<sub>4</sub> to C<sub>10</sub> fatty acids are not utilized in either of the two organisms.

In comparison with organisms such as *Pseudomonas* (6), *S. typhimurium* is somewhat limited in the range of compounds that can be utilized as a sole carbon source for growth. Thus, phosphoryl-

ated compounds are, in general, poorly utilized. Few of the fatty acids are utilized, amides and ester linkages are generally not hydrolyzed, aromatic rings are not degraded to useful carbon sources, few of the naturally occurring amino acids serve as carbon sources, large molecules are not degraded, and few of the wide variety of available sugars and sugar derivatives are utilized. Some known metabolic intermediates on the list of nonutilizable compounds are presumably not utilized because they do not get into the cells, e.g., *cis*-aconitic acid and ribose-5-phosphate, 6-phosphogluconate, 3-phosphoglycerate, and glucose-1-phosphate.

As noted in a footnote to Table 1, several compounds in the table only support the growth of mutant derivatives of the parent strain. It is likely that by using more bacteria in the inoculum or by employing mutagens, other mutants would be identified which utilized some of the compounds in Table 3. Further study of such clones might provide information on interesting pathways or uptake systems in *Salmonella*.

It is of interest to make a rough estimate of the percentage of the *Salmonella* genome used for the capacity to utilize these various carbon and nitrogen sources. An estimate is made as follows. There are about 4,000 genes in a *Salmonella* (or *E. coli*) if one takes the average size gene to be about 1,000 bases in length. If one allows a permease and about two enzymes for the utilization of each of the compounds involved, that would account for 300 genes. In addition, there may be some utilizable compounds that we have not tested; we allow 100 genes for this. Thus, approximately

TABLE 3. Compounds which do not serve either as a sole carbon or a sole nitrogen source for *Salmonella typhimurium*<sup>a</sup>

Acetamide	$\delta$ -Amino- <i>n</i> -valeric acid	D-Chondrosamine (galactosamine)	Formimino glycine
Acetone oxime	Amygdalin	Cinnamic acid ( <i>trans</i> )	Formyl alanine
<i>N</i> -acetyl-D-alanine	Aniline	Citraconate, sodium	Formyl glycine
<i>N</i> -acetyl-L-alanine	L-Anserine	L-Citrulline	Formyl-D-phenylalanine
Acetyl-D-alloisoleucine	Anthranilic acid	Creatine	Formyl-L-phenylalanine
<i>N</i> -acetyl-S-benzyl-DL-cysteine	Arabinic acid	Creatinine	D-Fructose-1, 6-diphosphate
Acetyl-dihydro-phenylalanine ethyl ester	D-Arabinose	Cratonic acid	D-Fucose
<i>N</i> -acetyl-galactosamine	D-Arabitol	Cyanate, potassium	2-Furan pyruvic acid oxime
<i>N</i> -acetyl-glycine	L-Arabitol	DL(+)-Cystathionine, <i>allo</i>	$\beta$ -D-Galactose-1-phosphate
<i>N</i> -acetyl-L-leucine	D-Araboascorbic acid	Cysteic acid	$\alpha$ -D-Galacturonic acid
$\epsilon$ -Acetyl-L-lysine	D-Arabonate, calcium	3-Deoxyglucose	Gelatin
<i>N</i> -acetyl-D-methionine	Arcaïne sulfate	2-Deoxy-D-glucose	Gentiobiose
<i>N</i> -acetyl-DL-methionine	D-Arginine	$\alpha, \beta$ -Diacetyl-diaminopropionic acid	<i>d</i> -Glucoscorbic acid
1-Acetyl-3-methyl urea	DL-Arterenol	3, 5-Diamino benzoic acid	Glucoheptonic acid
<i>N</i> -acetyl-D-phenylalanine	L-Ascorbic acid	2, 4-Diaminobutyric acid	D- <i>gluco</i> -Heptulose
Acetyl-DL-phenylalanyl glycine	L-Aspartic diethyl ester	4, 5-Diamino-2, 6-dihydroxypyrimidine	L- <i>gluco</i> -Heptulose
Acetylene dicarboxylic acid	Azelaic acid	$\alpha, \epsilon$ -Diaminopimelic acid	$\alpha$ -D-Glucose pentaacetate
Acetyl-DL-proline	L-Azetidine-2-carboxylic acid	2, 6-Diaminopurine	$\beta$ -D-Glucose-1-phosphate
Acetyl-DL-tyrosine	Barbital, sodium	L-Dihydroorotic acid	L-Glutamic acid dimethyl ester
<i>cis</i> -Aconitic acid	Barbituric acid	Dihydroourocnic acid	L-Glutamic acid- $\alpha$ -ethyl ester
Adenine	Benzamide	Dihydroxynorephedrine·HCl	Glutaric acid
Adenosine-3, 5-cyclic phosphate	Benzidine	Dihydroxyphenylalanine (DOPA)	DL-Glyceraldehyde
Adenosine-5-phosphate	Benzilic acid	$\beta, \beta$ -Dimethyl acrylic acid	D- <i>glycero</i> -D- <i>allo</i> -heptose
Adipamide	Benzoic acid	5, 5-Dimethyl-1, 3-cyclohexane dione	D- <i>glycero</i> -D- <i>galacto</i> -heptose
Adipic acid	Benzoin	3, 3-Dimethyl glutaric acid	D- <i>glycero</i> -L- <i>galacto</i> -heptose
Adonitol	Benzoin acetate	3, 5-Dinitro salicylic acid	D- <i>glycero</i> -D- <i>gluco</i> -heptitol
Agmatine sulfate	DL- <i>N</i> -benzoyl-2-alanine	Diphenylamine	D- <i>glycero</i> -D- <i>gluco</i> -heptose
$\beta$ -Alanine	Benzoyl glycine	Dipicolinic acid	L- <i>glycero</i> -D- <i>gulo</i> -heptitol
D-Alanine	<i>S</i> -benzyl cysteine	Egg albumin	D- <i>glycero</i> -D- <i>gulo</i> -heptose
DL- $\alpha$ -Alanine ethyl ester	Benzyl glycinate	<i>d</i> -Epinephrine	D- <i>glycero</i> -D- <i>ido</i> -heptose
DL-Alanylglycylglycine	Betaine·HCl	Erythritol	$\beta$ -Glycerophosphoric acid
Allantoin	Bicarbonat, sodium	<i>erythro</i> -L- <i>galacto</i> -octitol	Glycine amide
Allantolactone	Bicine	D- <i>erythro</i> -L- <i>gluco</i> -octitol	Glycine ethyl ester
D-Allose	Biotin	D- <i>erythro</i> - <i>talo</i> -octitol	Glycine methyl ester
D-Altrose	Biuret	D- <i>erythro</i> -L- <i>talo</i> -octose	Glycocyamine
DL- $\alpha$ -Aminoadipic acid	Brucine	Esculin	Glycogen
<i>p</i> -Aminobenzoic acid	Bufotenine mono oxalate hydrate	Ethanolamine	Glycyl-D-methionine
$\alpha$ -Aminobutyric acid	<i>n</i> -Butyl-DL-malate	Ethanolamine phosphate	Glycylglycine
DL- $\beta$ -Aminobutyric acid	<i>n</i> -Butyramide	L-Ethionine	Glyoxylic acid
$\epsilon$ -Amino- <i>n</i> -carpoic acid	<i>n</i> -Butyric acid	Ethyl acetate	Guanidine
$\omega$ -Aminocaprylic acid	D-Butyrolin	Ethyl anthranilate	$\alpha$ -Guanidobutyric acid
1-Aminocyclopentane carboxylic acid	Caffeine	Ethyl lactate	Guanido-DL-phenylalanine
$\alpha$ -Aminodecanoic acid	<i>n</i> -Caproic acid	Ethyl levulinate	Guanidopipelic acid
Aminoguanidine sulfate	Carbamyl glutamic acid	<i>N</i> -ethylmaleimide	$\beta$ -Guanidopropionic acid
Aminoguanidine bicarbonate	Carbamyl phosphate	Ethylamine·HCl	Guanine
$\alpha$ -Aminolauric acid	<i>N</i> -carboboxy glucosamine	Ethylene glycol	D-Gulonolactone
$\delta$ -Aminolevulinic acid	Casein	Ethyl itaconate	L-Gulonolactone
2-Amino-2-methyl-1, 3-propanediol	Catechol	5-Fluorouracil deoxy-ribose	Hemin
DL-2-Amino phenylacetic acid	D-Cellobiose	Folic acid	
2-Aminopurine	Cellotriose	Formate, sodium	
2-Aminopyridine	Celtrobose		
	L- $\alpha$ -Chloro- $\beta$ -phenylpropionyl glycine		
	Cholesterol		
	Cholic acid		
	Choline chloride		

TABLE 3—Continued

<i>n</i> -Heptylic acid	Maleic acid	D-Norvaline	Serotonin
1-Hexadecanol	D-Malic acid	Nucleinate, sodium	Sorbic acid
Hexamethylenetetramine	Malonamide	<i>n</i> -Octanoic acid	D-Sorbose
Hippuric acid	Malonic acid	Orcinol	L-Sorbose
Histamine diphosphate	DL-Mandelic acid	DL-Ornithine·HCl	Spermidine
L-Histidine	L-Mannitol	Orotic acid	Spermine
L-Homocitrulline	D-Mannoheptulose	Oxalic acid	Stearamide
L-Homocysteic acid	D-Mannuronic acid	Palmitic acid	Stearanilide
Hydrazine sulfate	D-Melezitose	D-Pantothenic acid	Stearic acid
<i>p</i> -Hydrazinobenzoic acid	Menthol glucuronide	D-Pantoyl lactone	Succinamide
Hydrocinnamic acid	Mercaptosuccinic acid	Perseitol	Succinimide
<i>trans</i> - $\beta$ -Hydromuconic acid	<i>meso</i> -Allitol	Phenethyl- $\beta$ -D-galactoside	Sucrose
<i>p</i> -Hydroxybenzoic acid	<i>meso-glycero-gulo</i> -Heptitol	Phenol	Sulfanilate, sodium
$\beta$ -Hydroxybutyric acid	L(+) <i>meso</i> Lanthionine	D-Phenylalanine	D-Tagaturonic acid
D(-)- $\beta$ -Hydroxybutyric acid	D-Methionine	L-Phenylalanine	D-Taloheptulose
DL-Hydroxylysine	L-Methionine	Phenylalanine methyl ester	D-Talose
Hydroxyproline	DL-Methionine sulfone	Phenylalanyl phenylalanine	Tannic acid
Hydroxy-D-proline, <i>allo</i>	DL-Methionine sulfoxide	Phenylalanine	D-Tartaric acid
Hydroxypyruvic acid	$\alpha$ -Methyl- <i>N</i> -acetyl mannosamine furanoside	Phenylalanine	L-Tartaric acid
8-Hydroxyquinoline	Methyl amine·HCl	L-Phenylephrine	Taurine
Hypoxanthine	Methyl albumin	DL-C-phenyl glycine	Taurocholic acid
L-Iduronic acid·barium	<i>S</i> -methyl-L-cysteine	DL- $\alpha$ -Phenyllactic acid	2,4,5,6-Tetra-aminopyrimidine sulfate
Imidazole	$\beta$ -Methyl-D-galactoside	6-Phosphogluconic acid	Tetrahydroxysuccinic acid
Indole	$\alpha$ -Methyl-D-glucoside	Phosphoribosyl pyrophosphate	Tetramethylammonium bromide
Indole-3-acetic acid	$\beta$ -Methyl-D-glucoside	DL- <i>O</i> -phosphoserine	Thiamine
3-(2-Amino-ethyl) indole hydrochloride	DL- $\alpha$ -Methyl glutamic acid	DL- <i>O</i> -phosphothreonine	Thioglycolate, sodium
Inosine-5-phosphate	Methyl itaconate	Phthalic acid	Thiosemicarbazide
Inulin	$\alpha$ -Methyl-D-mannoside	Phthalimide	Thiourea
3-Iodopropionic acid	Methyl- $\alpha$ -L-rhamnoside	$\alpha$ -Picolinic acid	Threamine
Isatin	DL- $\alpha$ -Methyl serine	Picric acid	D- <i>threo</i> -L- <i>gulo</i> -Octose
Isobutyric acid	DL- <i>O</i> -methyl serine	2,6-Piperidine dicarboxylic acid	D-Threonine
Isoglutamine	Methyl- $\beta$ -thiogalactoside	Piperonal	L-Threonine
DL-Isoleucamine·HCl	$\alpha$ -Methyl-D-xyloside	Polygalacturonic acid	L-Threonine, <i>allo</i>
D-Isoleucine, <i>allo</i>	$\beta$ -Methyl-D-xyloside	Prolamine·HCl	Thymine
L-Isoleucine	Mevalonic acid	D-Proline	2,4,5-Triamino-6-hydroxy pyrimidine sulfate
L-Isoleucine benzyl ester	2,3-Monoacetone-D-xylulofuranose	L-Proline methyl ester	Tripalmitin
L-Isoleucine ethyl ester	<i>trans trans</i> Muconic acid	Propionamide	Tristearin
L-Isoleucine methyl ester	<i>trans trans</i> Muconic acid	Protocatechuic acid	Tryptamine
L-Isoleucyl-L-isoleucine	$\alpha$ -Naphthol	Putrescine	D-Tryptophan
DL- <i>N</i> -isopropyl arterenol	$\beta$ -Naphthyl alanine	4( <i>p</i> -Nitrobenzyl) pyridine	L-Tryptophan
Itaconic acid	Nicotinamide	Pyridoxine	L-Tryptophan ethyl ester
5-Keto-D-gluconate	Nicotinamide adenine dinucleotide	Pyrocatechol	D-Tyrosine
$\beta$ -Ketoglutaric acid	Nicotinamide adenine dinucleotide phosphate	Pyruvoyl glycine	L-Tyrosine
Kojic acid	Nicotinic acid	DL-Pyruvoyl phenylalanine	L-Tyrosine- <i>O</i> -sulfate
D-Lactic acid	Nitrate, sodium	Quercetin	Uracil
Lactose	Nitrite, sodium	D-Raffinose	Urea
Lecithin	<i>p</i> -Nitrobenzoic acid	Resorcinol	Uric acid
L-Leucinamide	<i>p</i> -Nitrobenzoyl-L-glutamic acid	D-Rhamnose	Valeric acid
D-Leucinamide	Nitroguanidophenylalanine	Riboflavin	D-Valine
Leucinamine	$\beta$ -Nitroguanidopropionic acid	D-Ribonolactone	L-Valine
D-Leucine	4-Nitroimidazole	D-Ribose-5-phosphate	DL-Valine ethyl ester
L-Leucine	Norleucamine·HCl	Salicin	Vanillic acid
L-Leucine ethyl ester	DL-Norleucine	Salicylaldehyde	Vanillin
L-Leucine methyl ester	DL-Normetanephine	Salicylamide	Vanillyl alcohol
D-Leucyl-L-isoleucine	Norvalamine·HCl	Salicylic acid	Xanthine
Levulinic acid		Sedoheptulose anhydride	Xanthosine
L-Lysine		Semicarbazide·HCl	Xanthurenic acid
L-Lysine methyl ester		DL-Serine ethyl ester	Xylitol
D-Lyxonic acid			L-Xylose
D-Lyxose			

\* Additional compounds *not* serving as carbon sources are listed in Table 2, and additional compounds *not* serving as nitrogen sources are listed in Table 1.

10% (400/4,000) of the genome is responsible for the capacity to utilize these compounds.

#### ACKNOWLEDGMENTS

We thank Anne Liggett for technical assistance and V. Ginsburg for samples of many carbohydrates.

This research was supported by Atomic Energy Commission contract AT(1101) 34, no. 156, to B.N.A., by Public Health Service grant 5R01 AM 12092-02 to B.N.A., and by National Science Foundation grant GB-3557 to J.M.C.

#### LITERATURE CITED

1. Kauffman, F. 1951. Enterobacteriaceae. Ejnar Munksgaard, Copenhagen.
2. Kessler, D. P., and E. Englesberg. 1969. Arabinose-leucine deletion mutants of *Escherichia coli* B/r. *J. Bacteriol.* **98**: 1159-1169.
3. Kornberg, H. L. 1967. The regulation of anaplerotic enzymes in *E. coli*. *Bull. Soc. Chim. Biol.* **49**:1479-1490.
4. Krieg, R. E., and W. R. Lockhart. 1966. Classification of enterobacteria based on overall similarity. *J. Bacteriol.* **92**: 1275-1280.
5. Sanderson, K. E. 1967. Revised linkage map of *Salmonella typhimurium*. *Bacteriol. Rev.* **31**:354-372.
6. Stanier, R. Y., N. J. Palleroni, and M. Doudoroff. 1966. The aerobic pseudomonads: a taxonomic study. *J. Gen. Microbiol.* **43**:159-271.
7. Weeks, G., M. Shapiro, R. O. Burns, and S. J. Wakil. 1969. Control of fatty acid metabolism. I. Induction of the enzymes of fatty acid oxidation in *Escherichia coli*. *J. Bacteriol.* **97**:827-836.