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# Antineutrophil Cytoplasmic Antibodies, Autoimmune Neutropenia, and Vasculitis

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# Abstract

**Objectives**—Reports of an association between antineutrophil cytoplasmic antibodies (ANCA) and autoimmune neutropenia have rarely included cases of proven vasculitis. A case of ANCA-associated vasculitis (AAV) with recurrent neutropenia is described and relevant literature on the association between ANCA, neutropenia, and vasculitis is reviewed.

**Methods**—Longitudinal clinical assessments and laboratory findings are described in a patient with AAV and recurrent episodes of profound neutropenia from December 2008 – October 2010. A PubMed database search of the medical literature was performed for papers published from 1960 through October 2010 to identify all reported cases of ANCA and neutropenia.

**Results**—A 49 year-old man developed recurrent neutropenia, periodic fevers, arthritis, biopsyproven cutaneous vasculitis, sensorineural hearing loss, epididymitis, and positive tests for ANCA with specificity for antibodies to both proteinase 3 and myeloperoxidase. Antineutrophil membrane antibodies were detected during an acute neutropenic phase and were not detectable in a post-recovery sample, whereas ANCA titers did not seem to correlate with neutropenia. An association between ANCA and neutropenia has been reported in 74 cases from 24 studies in the context of drug/toxin exposure, underlying autoimmune disease, or chronic neutropenia without underlying autoimmune disease. In these cases, the presence of atypical ANCA patterns and other antibodies were common; however, vasculitis was uncommon and when it occurred was usually limited to the skin and in cases of underlying toxin exposure.

**Conclusions**—ANCA is associated with autoimmune neutropenia, but systemic vasculitis rarely occurs in association with ANCA and neutropenia. The interaction between neutrophils and ANCA may provide insight into understanding both autoimmune neutropenia and AAV.

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#### Keywords

vasculitis; neutropenia; antineutrophil cytoplasmic antibody (ANCA)

# INTRODUCTION

Autoimmune neutropenia is defined as a circulating absolute neutrophil count (ANC) of less than 1500/µL due to an immune-mediated process. Primary autoimmune neutropenia occurs in the absence of any other detectable pathology and is usually a benign condition seen in newborns. Secondary autoimmune neutropenia occurs in cases of underlying malignancy, infection, toxic exposure, or autoimmune disease including Felty's syndrome, systemic lupus erythematosis, and large granular lymphocyte syndrome (1). Antineutrophil membrane antibodies, directed against antigens located on the neutrophil cell surface, are recognized causes of both primary and secondary autoimmune neutropenia (1). In contrast, despite neutrophil antigens being the antigenic target of antineutrophil cytoplasmic antibodies (ANCA), neutropenia is not typically associated with ANCA or ANCA-associated diseases.

A case of recurrent neutropenia in a patient with ANCA who subsequently developed ANCA-associated vasculitis (AAV) is described, and the existing medical literature on the potential association between ANCA, autoimmune neutropenia, and vasculitis is reviewed.

# METHODS

Longitudinal clinical and laboratory assessments were performed in a case of AAV and recurrent neutropenia. Relevant information was excerpted from medical records and contemporaneous interviews with the patient. Serial assessments for ANCA and antineutrophil membrane antibodies were performed during periods of neutropenia and recovery. Testing was performed for both cytoplasmic (c) and perinuclear (p) ANCA by indirect immunofluorescence, and enyzme-linked immunosorbent assays (ELISA) were used to test for antibodies to proteinase 3 (PR3) and myeloperoxidase (MPO) (2). Granulocyte-reactive antibodies directed against neutrophil membrane antigens were measured using a modified indirect granulocyte immunofluorescence test (GIFT) (3).

A PubMed search of the medical literature for 1960 through October 2010 was performed using the following subject heading terms and keywords: [leukopenia OR leucopenia OR neutropenia OR agranulocytosis] and [systemic vasculitis OR antineutrophil cytoplasmic antibodies OR antineutrophil cytoplasmic antibody vasculitis OR ANCA OR vasculitis]. Bibliographies of identified reports and review articles were hand searched for additional references. Only pertinent literature, primarily in the English language, was included. Articles describing neutropenia as a side effect of therapy in cases of AAV with medications known to cause neutropenia (e.g. cyclophosphamide, rituximab, azathioprine) were excluded from further review.

#### RESULTS

A case of AAV and recurrent neutropenia with disease manifestations including periodic fevers, arthritis, biopsy-proven cutaneous vasculitis, sensorineural hearing loss, epididymitis, and positive tests for antineutrophil membrane antibodies and ANCA with specificity for antibodies to both PR3 and MPO is described.

## CASE REPORT

In June 2008, a previously healthy 49 year-old man developed a distinctive pattern of highspiking fevers (temperature >39°C), arthralgias, and fatigue. On day 1 of his typical symptom cycle, he had progressive fatigue. On day 2, he had fevers, shaking chills, and symmetric arthralgias. On day 3, these symptoms were most severe. On day 4, he gradually recovered. On days 5–7, he felt well.

The patient was hospitalized in July 2008 for evaluation of fever and found to be neutropenic with a total white blood cell count of  $2300/\mu$ L and an absolute neutrophil count (ANC) of  $0/\mu$ L. A bone marrow biopsy revealed normal marrow precursors, maturation arrest of myeloid elements, and no malignancy. The neutropenia spontaneously resolved and no explanation for the fevers was identified.

For the next several months, the patient had recurrent fevers, arthralgias, neutropenia, and he always recovered spontaneously. His symptoms usually improved with acetaminophen, and he never received colony stimulating factors for treatment of neutropenia. In August 2008 while neutropenic, an arthrocentesis performed on a painful left ankle yielded inflammatory synovial fluid (15,000 white blood cells; 73% polymorphonuclear leukocytes, 11% histiocytes, 6% lymphocytes) with no crystals detected. In November 2008, his ANC again trended down to  $0/\mu$ L before spontaneous recovery.

The patient's past medical history was notable for longstanding hypertension that was wellcontrolled with diltiazem. He took no other medications or supplements. There was no family history of autoimmune diseases or periodic fever syndromes. He lived in Massachusetts, USA and had not recently traveled out of state. He worked in a managerial position. He had a 30-year history of tobacco use but had quit smoking cigarettes several years ago. He reported limited alcohol intake and had no history of illicit drug use, including cocaine.

In December 2008, he was again hospitalized for evaluation of fever and arthralgias. At admission, he was febrile ( $103^{\circ}$  F) and tachycardic. The conjunctivae were injected. Fingers, wrists, elbows, and shoulders were tender without effusions. There was no splenomegaly or lymphadenopathy. Cutaneous, sinonasal, cardiopulmonary, gastrointestinal, neurologic, and genitourinary exams were unremarkable. The WBC at admission was 7500/µL (55% polymorphonuclear leukocytes, 36% lymphocytes, 7% monocytes, 2% eosinophils). ANC was 4125/µL but subsequently fell to 0/µL on hospital day 7 and spontaneously normalized by day 14. Tests for autoimmune and infectious diseases, and for genetic periodic fever syndromes, were negative (Table 1). A bone marrow biopsy during a period of absolute neutropenia again revealed maturation arrest of myeloid elements. Flow cytometry of marrow and peripheral blood did not demonstrate a clonal lymphocyte population.

A diagnosis of vasculitis was considered. Testing for ANCA was positive for moderate titers of antibodies to both PR3 and MPO with a perinuclear immunofluorescence staining pattern. Magnetic resonance imaging of the chest and abdomen, computerized tomography of sinuses, urinalysis, and ophthalmologic exam, revealed no evidence of vasculitis.

In January 2009, he presented with fevers, arthralgias, and purple macules on his right leg. Skin biopsy was consistent with leukocytoclastic vasculitis with perivascular lymphocytic and neutrophilic infiltrate with leukocytoclasia and extravasated erythrocytes. Treatment for AAV was initiated with high-dose glucocorticoids and oral methotrexate (20mg/week) with rapid resolution of the rash, fevers, and arthralgias.

In May 2009 the patient presented with acute right-sided hearing loss. An audiogram, when compared to a baseline audiogram obtained in March 2009, demonstrated a new right-sided 10–15 decibel sensorineural hearing loss across all frequencies. Treatment included reinitiation of high-dose glucocorticoids, discontinuation of methotrexate, and initiation of oral cyclophosphamide (1mg/kg/day). A subsequent audiogram (June 2009) showed a return to baseline hearing threshold. During treatment with cyclophosphamide, he had only one episode of mild transient neutropenia. In October 2009 the cyclophosphamide was discontinued and azathioprine was started. In late October 2009 the patient developed pancreatitis which resolved when azathioprine was discontinued.

In December 2009 mycophenolate mofetil (2000mg/day) was started. Two weeks later, the patient had severe bilateral testicular pain; ANC was  $0.1 \text{K/}\mu\text{L}$ . Scrotal ultrasound showed bilateral enlargement of the epididymes with hyperemia and cyst formation consistent with epididymitis. He was treated for 3 days with broad-spectrum antibiotics without improvement. Prednisone 60mg daily was then started with resolution of symptoms and recovery of neutrophil count within several days. Mycophenolate mofetil was discontinued, and he was treated with rituximab (375mg/m<sup>2</sup>; weekly infusions × 4 doses) (4).

As of October 2010, no further episodes of neutropenia or active manifestations of vasculitis with the exception of arthralgias requiring low doses of prednisone (5–10mg daily) have been noted. ANCA titers for PR3 and MPO have remained persistently positive in low titers. He was treated with a second course of rituximab ( $375mg/m^2$ ; weekly infusions × 4 doses) six months after initial treatment for maintenance of disease remission.

Serial ANCA measurements from December 2008 – December 2009 confirmed the persistence of ANCA with dual specificity to PR3 and MPO. A summary timeline of events, treatment strategies, neutrophil counts, and ANCA titers is provided (Figure 1).

To test for other potential autoimmune causes of neutropenia, granulocyte-reactive antibodies directed against neutrophil membrane antigens were measured using a modified indirect granulocyte immunofluorescence test (GIFT). IgM antineutrophil membrane antibodies were detectable during an acute neutropenic phase and were not detectable in the second post-recovery sample (Figure 2).

#### LITERATURE REVIEW

The literature regarding a possible association between ANCA and neutropenia is mostly limited to case reports and small case series. The search identified 24 articles that described an association between ANCA and neutropenia in a total of 74 patients. Cases reported to date can be categorized into three different clinical scenarios: cases of drug/toxin exposure, cases that occurred in the context of active autoimmune disease, and cases of chronic neutropenia without underlying autoimmune disease. Patient-level data are summarized in Table 2. Among all included studies, female prevalence was 79% and mean patient age was 42 years (range 8–70 years). Reported ANCA immunofluorescence patterns were PANCA 38%, atypical-ANCA 37%, and C-ANCA 25%. ELISA testing demonstrated ANCA specificity to lactoferrin 22%, MPO 18%, unidentifiable antigen 15%, multiple antigens 5%, PR3 4%, and human neutrophil elastase 0.1%; testing for antigen specificity was not described in 35% of cases.

Ten of 74 patients had suspected vasculitis (biopsy-proven in 6 cases). Skin lesions with leukocytoclastic vasculitis was the most commonly reported manifestation of vasculitis (8/10 suspected cases and 6/6 biopsy-proven cases) and all biopsy-proven cases were described in the setting of an underlying toxic exposure.

Antibodies other than ANCA were frequently described. Testing for antibodies directed to neutrophil surface membrane antigens can be technically challenging and is not usually performed in routine clinical practice (1). Antineutrophil membrane antibodies were detected in 8 of 16 cases in which testing occurred and were detected in at least one case in each of the three clinical categories. Antinuclear antibodies were also often present (33 of 46 cases in which such testing was reported) in all three clinical categories.

#### **Drug-induced ANCA and neutropenia**

Twenty-four cases of ANCA and neutropenia in the setting of a suspected identifiable medication or toxin were detected. Cessation of the suspected agent typically resulted in improvement in symptoms and laboratory abnormalities. Despite an extensive list of medications known to be associated with neturopenia(5), relatively few of these medications were associated with both ANCA and neutropenia.

Propylthiouracil (PTU) was a commonly implicated medication. P-ANCA with specificity for MPO was frequently reported. In a series of 56 patients with Grave's disease treated with PTU, MPO-ANCA was detected in 21 cases (38%) and neutropenia occurred in 3 of these cases (5%) (6). Akamizu et al. conducted *in vitro* cytotoxicity tests on serum from a patient who developed neutropenia and ANCA while being treated with PTU and demonstrated that ANCA lysed neutrophils via a complement-dependent mechanism but not by antibody-dependent cell-mediated cytotoxicity (7). Antineutrophil membrane antibodies and biopsy-proven vasculitis have been reported in cases of PTU exposure (8–10). Methimazole has also been implicated as causing development of ANCA and neutropenia (11).

Cases of lupus-like syndromes with overlapping features of systemic vasculitis have been described in association with minocycline and hydralazine. Ahmed et al. report an 18 year old patient who developed moderate neutropenia, c-ANCA with specificity to PR3, high-titer ANA, and constitutional symptoms while taking minocycline for acne (12). Sangala et al. described a patient with SLE taking hydralazine who developed biopsy-proven, pauci-immune glomerulonephritis and pancytopenia (13). The neutropenia was attributed to a lupus-like syndrome and resolved with cessation of hydralazine. Testing for antineutrophil membrane antibodies was not performed in either case.

Recently, an association with neutropenia and ANCA has been reported in users of cocaine adulterated with levamisole. Levamisole was developed as an antihelminth medication and is known to have immunostimulating effects with production of autoantibodies (14). Knowles et al. describe 60 cases of severe neutropenia associated with cocaine tainted with levamisole (15). Four of 5 cases tested for ANCA were positive (2 for c-ANCA; 2 for p-ANCA), and an additional case had detectable antineutrophil membrane antibodies. An overlap of clinical features seems to define cases of exposure to levamisole/cocaine with findings including: severe neutropenia; ANCA production with antibodies to PR3, MPO, and/or human neutrophil elastase; purpura with a predilection for the earlobes; antiphospholipid antibodies; and necrotic skin lesions with a mixed pathologic pattern of leukocytocalstic vasculitis and microthrombus (16–19). Antineutrophil membrane antibodies were present in one of two cases of ANCA and neutropenia in which testing occurred (18). Independent of cocaine exposure, a case of ANCA and neutropenia has been described in a child being treated with levamisole as adjuvant therapy for nephrotic syndrome (20).

#### Underlying autoimmune disease with ANCA and neutropenia

ANCA and neutropenia has been reported in association with other active autoimmune diseases including Felty's syndrome, autoimmune liver diseases, and Sjögren's syndrome.

Felty's syndrome, a clinical triad of rheumatoid arthritis, neutropenia, and splenomegaly, has been associated with ANCA. Juby et al. report 33% prevalence of ANCA in a series of 32 patients with Felty's syndrome with severe neutropenia (21). Immunofluorescence staining showed either p-ANCA or an atypical pattern; however, testing specific ANCA antigens was not performed. Coremans et al. detected ANCA in 23 of 30 (77%) patients with Felty's syndrome (22). Specificity to lactoferrin was detected in 50% of patients with Felty's syndrome compared to 4% of a comparison group of patients with rheumatoid arthritis without Felty's syndrome. A high frequency of extra-articular disease has also been observed in Felty's syndrome with up to 28% prevalence of vasculitis reported in one series (23).

ANCA and neutropenia has also been reported in the context of other autoimmune diseases. Autoimmune liver diseases, including autoimmune hepatitis and primary sclerosis cholangitis, have been associated with non-specific ANCA, presence of antineutrophil membrane antibodies, and severe neutropenia in the absence of vasculitis (24–26). ANCA and severe neutropenia has also been described in three cases of Sjögren's syndrome, and in one case, cutaneous vasculitis and antineutrophil membrane antibodies were reported (27–28).

#### ANCA and chronic neutropenia without underlying autoimmune disease

Detection of ANCA has been reported in 9 cases of chronic neutropenia where no other infectious, toxic, or autoimmune etiology was identified. In a series of 9 patients, Coppo et al. described an association with ANCA and non-cyclic neutropenia in 7 patients without an identifiable underlying autoimmune disease (27). ANCA immunofluorescence patterns were mostly atypical with an unidentifiable antigen on ELISA testing, although PR3 was detected in one patient. Antineutrophil membrane antibodies were detected in 3 of 7 cases. One patient in the series developed leukocytoclastic vasculitis, and many of these patients had additional findings suggestive of a potential underlying autoimmune mechanism including hemolytic anemia, thrombocytopenia, and other circulating autoantibodies. Rodrigues et al. demonstrated ANCA in the serie of two patients with cyclic neutropenia and no other underlying autoimmune disease; these ANCA reacted with a novel 60kDa protein but not to PR3 or MPO (29).

#### DISCUSSION

This case and the existing literature support an association between ANCA and neutropenia. This association can occur in cases of drug exposure, in association with other active autoimmune diseases, and in cases of chronic neutropenia without underlying autoimmune disease. Reported ANCA immunoflouresence patterns were varied with approximately equal proportions of atypical and perinuclear patterns and fewer cytoplasmic patterns. Antibodies to either PR3 or MPO, the two antigens used to diagnose and categorize AAV (30), were commonly observed in cases of ANCA-associated neutropenia, yet few of these cases developed clinical evidence of vasculitis. When vasculitis did occur, it was usually limited to skin involvement in the setting of underlying exposure to a toxin. Whether a causal relationship exists between ANCA and neutropenia is unclear, and the presence of additional antibodies, especially antineutrophil membrane antibodies, complicates issues of causality.

The present case is unique because neutropenia was associated with systemic vasculitis and not only cutaneous disease, and occurred without evidence of an underlying toxic exposure. This patient was not taking any medications known to cause drug-induced vasculitis and repeated urine toxicology screening tests were negative for cocaine. The manifestations of

inflammatory arthritis (31), biopsy-proven cutaneous vasculitis (32), sensorineural hearing loss responsive to immunosuppressants (33), and epididymitis (34), combined with the ANCA test results, provide compelling evidence for a diagnosis of AAV with multi-organ involvement. It is notable that this patient did not develop sinonasal, pulmonary, or renal disease, which are usual organ systems affected in the commonly definable subsets of AAV [granulomatosis with polyangiitis (Wegener's), microscopic polyangiitis, and Churg Straus syndrome].

This case and the existing literature raise interesting questions about the pathophysiology of neutropenia in association with ANCA and antineutrophil membrane antibodies. Potential underlying mechanisms for autoimmune neutropenia include cellular or humoral suppression of granulopoesis, antibody-mediated sequestration or margination of neutrophils, and peripheral destruction of neutrophils (35). Evidence supporting one or more of these hypotheses was present in the current case and many of the previously reported cases. Maturation arrest of myeloid precursors was seen on bone marrow biopsy in the present case and several other cases (12, 24–25, 27–28) suggesting possible cellular or immune-mediated central marrow suppression. Cases in which splenomegaly was found suggest the potential for neutrophil sequestration (9, 20–22, 27, 36). Evidence for peripheral destruction of neutrophils was supported by the presence of antibodies to neutrophil antigens and by the findings of hypercellularity on bone marrow biopsy (7, 13, 37).

Antineutrophil membrane antibodies, directed against neutrophil cell surface antigens, are known to cause neutropenia via complement-mediated cytotoxicity (38) and phagocytosis (39). Whether ANCA are directly causal for neutropenia is unclear. In the present case, unlike in most other cases of ANCA and neutropenia reported in the literature, both ANCA and antineutrophil membrane antibodies were measured longitudinally in periods of severe neutropenia and recovery. ANCA and antineutrophil membrane antibodies were detected in the patient's serum during periods of severe neutropenia. IgM antineutrophil membrane antibodies were detected during a period of absolute neutropenia but were no longer detectable during neutrophil recovery. In contrast, ANCA titers did not seem to correlate with neutrophil counts. These data suggest that antineutrophil membrane antibodies, rather than ANCA, were causal for neutropenia but do not exclude the possibility that the underlying etiology was multi-factorial and that ANCA may have been contributory.

The observation that the patient in current report had several weeks of recurrent fever and neutropenia before developing features of systemic vasculitis supports a hypothesis that autoimmune neutropenia may induce the formation of ANCA. Lysis of neutrophils by antineutrophil membrane antibodies and/or ANCA antigen migration to the neutrophil cell surface promoted by endogenous pyrogens (40–41), may expose the immune system to cytoplasmic antigens including PR3 and MPO (41–42) in an immunogenic environment, leading to the generation of ANCA and subsequent systemic vasculitis (43). It should be noted, however, that the pathogenicity of ANCA in AAV is somewhat controversial (44). Only a few of the patients reported in the literature with ANCA and neutropenia developed vasculitis despite developing antibodies to PR3 and MPO, suggesting that factors in addition to ANCA may be necessary in the pathogenesis of AAV. Alternatively vasculitis may be under-recognized when it occurs in association with neutropenia because the clinical features of vasculitis may be atypical in this setting, as evidenced by the present case.

Neutrophils are felt to play a primary role in the acute injury of AAV (45). In the present case, symptoms of vasculitis occurred both in periods of absolute neutropenia and in times of neutrophil recovery. The paradox of vasculitis in the setting of absolute neutropenia may be explained by the fact that circulating levels of neutrophils may not necessarily reflect neutrophil burden in other tissues. For example, the currently-described patient developed an

Treating a patient with severe neutropenia with medications that can both cause neutropenia and increase the risk of infection is a major clinical challenge but was justified in this case given the suspected autoimmune etiology of the neutropenia. The patient experienced fewer neutropenic episodes following the initiation of treatment with immunosuppressive therapy. Repeated dosing of rituximab was particularly efficacious in achieving sustained disease remission compared to other immunosuppressant agents.

ANCA is associated with autoimmune neutropenia but causality has not been established. ANCA testing should be considered in cases of unexplained neutropenia, and a positive ANCA should increase clinical suspicion for possible underlying exposures to toxins or autoimmune diseases. Although clinically-apparent vasculitis appears to be uncommon in ANCA-associated neutropenia, cutaneous and systemic manifestations have been reported. Further investigation into the interaction between neutrophils and ANCA may provide important insights into the pathophysiology underlying autoimmune neutropenia and AAV.

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Events 1, 3, and 4 (open arrows): fever, arthralgias, and absolute neutropenia. Events 2, 5, 6, and 7 (closed arrows): manifestations of ANCA-associated vasculitis.

Event 2: inflammatory ankle effusion and neutropenia.

Event 5: cutaneous leukocytoclastic vasculitis, fevers, and arthralgias.

Event 6: sensorineural hearing loss.

Event 7: epididymitis, fever, and absolute neutropenia.

Anti-proteinase 3 titers ranged from 359–1689 units (normal< 2.8 units).

Anti-myeloperoxidase titers ranged from 250–18452 units (normal< 20 units).

ANC = absolute neutrophil count; ANCA = antineutrophil cytoplasmic antibodies.



Figure 2. IgM antineutrophil membrane antibodies at time of absolute neutropenia (December 2008) and the subsequent two month period during which neutropenia resolved Antineutrophil antibody results are expressed as ratios of median fluorescence intensity (RFI) of patient serum-sensitized target cells over median fluorescence intensity of cells incubated in normal control serum. A ratio of  $\geq 2.5$  is considered positive for IgM neutrophil reactive antibodies (above red dashed line). Neutropenia is defined as an absolute neutrophil count < 1500/UL (below blue dashed line). Antibody testing was done courtesy of the Blood Center of Wisconsin Platelet and Neutrophil Immunology Laboratory.

#### Table 1

#### Selected laboratory tests in current case

| Test   | Value                    | Normal Range        |
|--|--------------------------|---------------------|
| Tests for autoimmune diseases                        |                          |                     |
| Antinuclear antibody                                 | Negative                 | Negative            |
| Rheumatoid factor (IU/mL)                            | <20                      | <20                 |
| Complement component 3 (C3) (g/L)                    | 0.92                     | 0.80-1.70           |
| Complement component 4 (C4) (g/L)                    | 0.15                     | 0.16-0.40           |
| C-reactive protein ( <i>mg/L</i> )                   | 153                      | <10                 |
| Erythrocyte sedimentation rate (mm/hr)               | 98                       | 0–15                |
| IgG ( <i>g/L</i> )                                   | 17.20                    | 7.00–16.00          |
| IgA ( <i>g/L</i> )                                   | 9.21                     | 0.70-4.00           |
| IgM ( <i>g/L</i> )                                   | 23.60                    | 0.46-3.04           |
| Serum immunofixation                                 | No monoclonal bands      | No monoclonal bands |
| Tests for infectious diseases                        |                          |                     |
| Blood cultures (bacterial, fungal)                   | Negative                 | Negative            |
| Human immunodeficiency virus (HIV)                   | Non-reactive             | Non-reactive        |
| Rapid plasma reagin (RPR)                            | Non-reactive             | Non-reactive        |
| Anti-Steptolysin O titer (IU/ml)                     | <25                      | <125                |
| Hepatitis B surface antigen                          | Negative                 | Negative            |
| Hepatitis C antibody                                 | Negative                 | Negative            |
| Parvovrius B19 (PCR)                                 | Negative                 | Negative            |
| Babesiosis (PCR)                                     | Negative                 | Negative            |
| Ehrlichiosis (PCR)                                   | Negative                 | Negative            |
| Lyme Disease (Antibody, PCR)                         | Negative                 | Negative            |
| Tests for genetic diseases                           |                          |                     |
| Familial Mediterranean Fever (FMF)                   | No MEFV gene mutations   | No mutations        |
| TNF Recepter-1 Associated Periodic Syndromes (TRAPS) | No TNFR1 gene mutations  | No mutations        |
| Cryopyrin-Associated Periodic Syndromes (CAPS)       | No CIAS 1 gene mutations | No mutations        |

Table 2

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Literature review of association between ANCA and neutropenia

| Reference              | Attributed<br>Cause | Subject<br>Age, Sex  | Degree of<br>Neutropenia | ANCA (IF)                | ANCA<br>(ELISA) | Signs of<br>Vasculitis                               | Antineutrophil<br>Membrane<br>Antibodies | ANA | Other                          |
|------------------------|---------------------|----------------------|--------------------------|--------------------------|-----------------|--|--|-----|--------------------------------|
| Suspected Drug-Induced |                     |                      |                          |                          |                 |  |  |     |                                |
|                        |                     | 46,F                 | ND                       | p-ANCA                   | MPO             | Purpura  | ΩN                                       | ΟN  |                                |
| Sera 2000 (6)          | Propylthiouriacil   | 28,F                 | ND                       | p-ANCA                   | MPO             | None   | ŊŊ                                       | QN  |                                |
|                        |                     | 33,F                 | ND                       | p-ANCA                   | MPO             | None   | ND                                       | ΟN  |                                |
| Noh 2001 (45)          | Propylthiouriacil   | 22,F                 | Mild                     | p-ANCA                   | MPO             | None   | ND                                       | +   |                                |
| Akamizu 2002 (7)       | Propylthiouriacil   | 45,F                 | Severe                   | p-ANCA                   | MPO,PR3         | None   | QN                                       | +   | Hypercellular marrow           |
| Yamada 2002 (8)        | Propylthiouriacil   | 13,F                 | ND                       | p-ANCA                   | MPO             | Hematuria  | +  | ŊŊ  |                                |
| Farah 2006 (10)        | Propylthiouriacil   | 70,F                 | Severe                   | p-ANCA                   | QN              | Skin-LCV *   | QN                                       | QN  |                                |
| Finucane 2008 (37)     | Propylthiouriacil   | 31,F                 | Moderate                 | c-ANCA                   | PR3             | None   | QN                                       | Q   | Hypercellular marrow           |
| Ozkok 2009 (9)         | Propylthiouriacil   | 36,F                 | Moderate                 | p-ANCA                   | MPO             | Skin-LCV*  | QN                                       | I   | Splenomegaly Plasmacytosis     |
| Kawachi 1995 (11)      | Methimazole         | 24,F                 | Moderate                 | p-ANCA                   | MPO             | LCV*   | QN                                       | +   |                                |
| Sangala 2010 (13)      | Hydralazine         | 55,F                 | Severe                   | p-ANCA                   | MPO             | Pauci-immune GN <sup>*</sup> ; Skin-LCV <sup>*</sup> | QN                                       | +   | Hypercellular marrow           |
| Ahmed 2008 (12)        | Minocycline         | 18,M                 | Mild                     | c-ANCA                   | PR3             | None   | ND                                       | +   | Maturation arrest              |
| Knowles 2009 (15)      | Cocaine/Le vamisole | 4 Cases <sup>#</sup> | Severe                   | p-ANCA (2)<br>c-ANCA (2) | DN              | QN   | QN                                       | ΟN  |                                |
| Bradford 2010 (17)     | Cocaine/Levamisole  | 39,F                 | Mild                     | a-ANCA                   | HNE             | Skin-LCV *   | ΟN                                       | +   | LAC                            |
|                        |                     | 49,F                 | Severe                   | a-ANCA                   | MPO, PR3, HNE   | None   | ND                                       | +   |                                |
|                        |                     | 45,F                 | Severe                   |                          |                 | None   | I  |     | Plasmacytosis                  |
| Czuchlewski 2010 (18)  | Cocaine/Levamisole  | 37,M<br>35,F         | Se vere<br>Se vere       | QN                       | DN              | None<br>None   | + QN                                     | Ŋ   | Plasmacytosis<br>Plasmacytosis |
|                        |                     | 57,F                 | Severe                   | p-ANCA                   | QN              | None   | - AN                                     | Ð   | ACA                            |
| Walsh 2010 (16)        | Cocaine/Levamisole  | 22,F                 | Moderate                 | p-ANCA                   | ŊŊ              | Skin-LCV*  | QN                                       | QN  | ACA                            |
|                        |                     |                      |                          |                          |                 |  |  |     |                                |

| Reference                  | Attributed<br>Cause            | Subject<br>Age, Sex | Degree of<br>Neutropenia | ANCA (IF)                 | ANCA<br>(ELISA)                                   | Signs of<br>Vasculitis                                   | Antineurrophi<br>Membrane<br>Antibodies | ANA             | Other                   |
|----------------------------|--------------------------------|---------------------|--------------------------|---------------------------|---|--|---|-----------------|-------------------------|
| arbano 1999 (20)           | Levamisole                     | 8,M                 | Ŋ                        | p-ANCA                    | ŊŊ  | None   | ŊŊ                                      | 1               | ACA, Splenomegaly       |
| derlying Autoimmune        | ? Disease                      |                     |                          |                           |   |  |   |                 |                         |
| <b>Juby 1992</b> (21)      | Felty's Syndrome               | 11 cases#           | Mild-Severe              | p-ANCA (4)<br>c-ANCA (7)  | QN  | Î  | QN                                      | QN              | Splenomegaly (7)        |
| <b>Coremans 1993</b> (22)  | Felty's Syndrome               | 23 cases#           | Mild-Severe              | a-ANCA (22)<br>p-ANCA (1) | Lactoferrin (15)<br>MPO (4)<br>Not identified (4) | ΩN   | QN                                      | + (17)<br>- (6) |                         |
| Wehbe 2010 (24)            | Autoimmune hepatitis           | 49,F                | Severe                   | a-ANCA                    | ND  | None   | +                                       | ΟN              | Maturation arrest       |
| Cuadrado 2009 (25)         | Autoimmune hepatitis           | 68,F                | Severe                   | p-ANCA                    | ΩN  | None   | -                                       | +               | Maturation arrest       |
| <b>Hanawa 2010</b> (26)    | Primary Sclerosing Cholangitis | 19,M                | Severe                   | c-ANCA                    | ΠN  | None   | +                                       | +               |                         |
| <b>Jani 2002</b> (36)      | Ulcerative Colitis             | 61,F                | Moderate                 | p-ANCA                    | MPO,PR3   | None   | -                                       | +               | Splenomegaly            |
| Krishnan 1997 (28)         | Sjöogren's Syndrome            | 52,F                | Severe                   | QN                        | ŊŊ  | Skin-LCV   | +                                       | +               | Maturation arrest       |
| <b>Coppo 2004</b> (27)     | Sjöogren's Syndrome            | 62,F<br>54.F        | Severe                   | p-ANCA<br>p-ANCA          | MPO<br>Not Identified                             | None<br>None   | 1 1                                     | + +             | Maturation arrest, LGLS |
|                            |                                | ****                | 200                      | LOUT 4                    |   | 0101   |   | -               |                         |
| Present Report             | ANCA-Associated Vasculitis     | 49,M                | Severe                   | p-ANCA                    | MPO,PR3   | Skin-LCV $^{*}$ SN hearing loss, arthritis, epididymitis | +                                       | 1               | Maturation arrest       |
| o Underlying Autoimm       | une Disease                    |                     |                          |                           |   |  |   |                 |                         |
|                            |                                | 16,M                | Severe                   | c-ANCA                    | Not identified                                    | None   | +                                       | I               | Maturation arrest       |
|                            |                                | 46,F                | Mild                     | c-ANCA                    | PR3   | None   | I                                       | +               |                         |
|                            | :                              | 56,F                | Severe                   | p-ANCA                    | MPO,HNE   | Skin-LCV   | I                                       | I               | Splenomegaly            |
| Coppo 2004 (27)            | Idiopathic                     | 60,M                | Severe                   | a-ANCA                    | Not identified                                    | None   | + -                                     | ı -             | Splenomegaly            |
|                            |                                | 1/1C                | PIEM                     | C-ANCA                    | Not identified                                    | None   | + 1                                     | + 1             |                         |
|                            |                                | 67,F                | Severe                   | p-ANCA                    | Lactoferrin                                       | None   | I                                       | +               | Maturation arrest       |
| <b>todrigues 2009</b> (29) | Cyclic Neutropenia             | 38,M<br>53,F        | Severe                   | c-ANCA (2)                | 60kDA Protein                                     | None   | QN                                      | QN              | LGLS (1)                |

Degree of neutropenia: mild - 1000-1500/mm<sup>3</sup>; moderate- 500-1000/mm<sup>3</sup>; severe < 500/mm<sup>3</sup>.

cardiolipin antibodies.

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 $\#_{\rm II}$  Indicates subject level data on age and sex not provided.

\* Indicates biopsy-proven finding