



Published in final edited form as:

Neurobiol Aging. 2009 May ; 30(5): 739–751. doi:10.1016/j.neurobiolaging.2007.08.022.

Prominent Phenotypic Variability Associated with Mutations in *Progranulin*

Brendan J. Kelley, MD^{a,k}, Wael Haidar, MD^{a,k}, Bradley F. Boeve, MD^{a,k}, Matt Baker, BSc^g, Neill R. Graff-Radford, MBChB^e, Thomas Krefft, MD^h, Andrew R. Frank, MD^{a,k}, Clifford R. Jack Jr, MD^c, Maria Shiung, BA^c, David S. Knopman, MD^{a,k}, Keith A. Josephs, MST, MD^a, Sotirios A. Parashos, MDⁱ, Rosa Rademakers, PhD^g, Mike Hutton, PhD^g, Stuart Pickering-Brown, PhD^j, Jennifer Adamson, BS^g, Karen M. Kuntz, RN^k, Dennis W. Dickson, MD^f, Joseph E. Parisi, MD^b, Glenn E. Smith, PhD^{d,k}, Robert J. Ivnik, PhD^{d,k}, and Ronald C. Petersen, PhD, MD^{a,k}

^aDepartment of Neurology, Mayo Clinic, Rochester, Minnesota 55905 USA

^bDepartment of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, Minnesota 55905 USA

^cDepartment of Radiology, Mayo Clinic, Rochester, Minnesota 55905 USA

^dDepartment of Psychiatry and Psychology, Mayo Clinic, Rochester, Minnesota 55905 USA

^eDepartment of Neurology, Mayo Clinic, Jacksonville, Florida 32224 USA

^fDepartment of Neuropathology Laboratory, Mayo Clinic, Jacksonville, Florida 32224 USA

^gDepartment of Neurogenetics Laboratory, Mayo Clinic, Jacksonville, Florida 32224 USA

^hNeurology Clinic, Slidell, LA 70458 USA

ⁱStruthers Parkinson's Center, Golden Valley, Minnesota 55427 USA

^jCentre for Clinical Neurosciences, University of Manchester, Salford, M6 8HD, UK

^kRobert H. and Clarice Smith and Abigail Van Buren Alzheimer's Disease Research Program of the Mayo Foundation

Abstract

Mutations in progranulin (*PGRN*) are associated with frontotemporal dementia with or without parkinsonism. We describe the prominent phenotypic variability within and among eight kindreds evaluated at Mayo Clinic Rochester and/or Mayo Clinic Jacksonville in whom mutations in *PGRN* were found. All available clinical, genetic, neuroimaging and neuropathologic data was reviewed. Age of onset ranged from 49 to 88 years and disease duration ranged from 1 to 14 years. Clinical diagnoses included frontotemporal dementia (FTD), primary progressive aphasia, FTD with parkinsonism, parkinsonism, corticobasal syndrome, Alzheimer's disease, amnesic mild cognitive impairment, and others. One kindred exhibited maximal right cerebral hemispheric atrophy in all four affected individuals, while another had maximal left hemisphere involvement in all three of the affected. Neuropathologic examination of 13 subjects revealed frontotemporal lobar degeneration with ubiquitin-positive inclusions plus neuronal intranuclear inclusions in all cases. Age of onset, clinical phenotypes and MRI findings associated with most *PGRN* mutations varied

Correspondence to: Bradley F. Boeve, M.D. Mayo Clinic Department of Neurology 200 First Street SW Rochester, MN 55905
Phone: 507-538-1038 Fax: 507-538-6012 bboeve@mayo.edu.

Disclosure The authors report no potential conflicts of interest.

significantly both within and among kindreds. Some kindreds with *PGRN* mutations exhibited lateralized topography of degeneration across all affected individuals.

Keywords

Frontotemporal dementia; FTDP-17; Progranulin; PGRN; MRI

1. Introduction

In 1892, Arnold Pick first described behavioral and language abnormalities in association with frontotemporal lobar degeneration. Subsequently, many familial cases of frontotemporal dementia with parkinsonism were linked to chromosome 17 (FTDP-17) (Foster et al. 1997). Some were associated with mutations in the microtubule associated protein tau (*MAPT*), (Hutton et al. 1998; Ingram and Spillantini 2002) while others were not (Kertesz et al. 2000; Rademakers et al. 2002; Mackenzie et al. 2006b). Many cases of FTDP-17 did not exhibit immunostaining for tau on pathologic examination (Kertesz et al. 2000; Rosso et al. 2001; Savioz et al. 2003; Mackenzie et al. 2006b; van der Zee et al. 2006), and mutations in *MAPT* were absent in all of these. We recently reported several FTDP-17 kindreds having frontotemporal lobar degeneration with ubiquitin-positive inclusions (FTLD-U) neuropathology in association with mutations in the gene encoding *Progranulin* (*PGRN*) (Baker et al. 2006; Boeve et al. 2006; Gass et al. 2006; Mackenzie et al. 2006b). The eight families described here extend the phenotypic and MRI findings associated with mutations in *PGRN* and highlight the variability in presentation, clinical course and neuroimaging findings within and among kindreds.

2. Methods

2.1 Subjects

Mutations in *PGRN* were identified in eight families whose probands were evaluated at Mayo Clinic Rochester (n=6) or Mayo Clinic Jacksonville (n=2). At least one affected individual from each family was enrolled in the Mayo Clinic Alzheimer Disease Research Center, a Mayo Foundation Institutional Review Board-approved program. All available clinical records and neuroimaging studies on affected members of these kindreds were reviewed. Genetic analyses, MRI scans, and autopsies were performed after subjects or appropriate proxies provided written consent.

2.2 Clinical Evaluations

Age of onset was the age at which the subject first demonstrated behavioral or personality change, memory loss, motor changes, or other neurological changes as noted by themselves, family, friends or colleagues. All neurobehavioral clinical data (Members of the Department of Neurology 1998) were reviewed.

2.3 Laboratory Analyses

DNA was extracted from peripheral blood leucocytes and sequence analysis of *MAPT* and *PGRN* from patient genomic DNA was performed as previously described (Hutton et al. 1998; Baker et al. 2006).

2.4 Neuroimaging Examinations

MRI was performed using a GE scanner at 1.5 Tesla, and images of the brain were obtained in the sagittal (T1-weighted), axial [proton-density, T2-weighted, and fluid attenuation inversion recovery (FLAIR)], and coronal (T1-weighted and FLAIR) planes.

2.5 Neuropathologic Examination

Sections of neocortex, hippocampus, thalamus, basal ganglia, midbrain, pons, medulla and cerebellum were stained with hematoxylin and eosin and thioflavin-S fluorescent microscopy and immunocytochemistry for phospho-tau. Sections of cortex and hippocampus were stained with Bielschowsky and Luxol fast blue, and immunostained for ubiquitin, neurofilament, A β 40, and A β 42. Midbrain and amygdala sections were studied with alpha-synuclein immunostains. Immunohistochemistry for TDP-43 was applied in selected cases (1:8000; Proteintech Group, Chicago, IL).

Neuropathological diagnoses were based on established guidelines (McKhann et al. 2001). A diagnosis of frontotemporal lobar degeneration with ubiquitin-only immunoreactive neuronal changes (FTLD-U) was made if there was neuronal loss and gliosis affecting frontal or temporal lobe, plus ubiquitin-positive; tau, alpha-synuclein and neurofilament negative intraneuronal inclusions or neurites in frontal or temporal neocortex or hippocampal dentate granule cells in the absence of any evidence of motor neuron degeneration (Josephs et al. 2006a). In cases with multiple pathologies, TDP-43 immunohistochemistry was used to confirm FTLD-U.

3. Results

3.1 Family Descriptions

Pedigrees are shown in Figure 1. Clinical, radiologic, and neuropathologic findings are summarized in Table 1, with additional clinical details presented in Table 2. Representative MRI scans are shown in Figure 2, and Figure 3 is a schematic drawing localizing the mutations in *PGRN*. Descriptions of Kindred 7 (Mesulam et al. 2007), two members of Kindred 1 (Boeve et al. 2002) and the proband of Kindred 5 (Boeve et al. 2006) have previously been reported.

3.2 Demographics

Symptom onset among 38 individuals ranged from 49 to 88 (mean 64.5 ± 9.6 years). Of 44 individuals reported to be affected, 25 were female. Among the 18 subjects (7 male) with clinical evaluations, age at onset ranged from 49 to 80 years (mean 61.1 ± 8.1 years). In Kindred 2, mean age of symptomatic onset was 75.8 ± 5.0 years in generation II and 60.7 ± 5.5 years in generation III. In Kindred 3, mean age of symptomatic onset was 70.9 ± 10.4 years in generation III and 52 ± 2.8 years in generations IV and V.

Disease duration ranged from 1 year to 14 years, with a mean of 6.6 ± 3.2 years (31 individuals). Two individuals had accidental deaths within 1 year of onset. Excluding these, the range was 2 to 14 years (mean 7.0 ± 3.0 years). Among the 13 subjects clinically evaluated who subsequently died, mean duration was 7.15 ± 2.9 years (range 2 to 13 years).

3.3 Clinical Features

As Table 2 indicates, personality change was an initial or early symptom (developing within one year of onset) in 10 of the 18 patients evaluated clinically (55%), and later developed in another 7 patients (cumulative 94%). Hyperoral behaviors developed in 8 of the 12 patients from whom this history was collected. Executive dysfunction (evidenced on

neuropsychological testing) was an initial or early feature in 7 of 18 patients for whom this data was available (39%). It later developed in all but one of these patients (94.1%). Eleven of the 18 patients evaluated clinically (61%) had personality changes and/or executive dysfunction features within the first year of symptomatic disease. Initial memory impairment was present in 8 of the 18 patients seen clinically (44%), developing prior to personality change or executive dysfunction in 2 unrelated patients. Memory impairment was prominent enough in six members of Kindred 2 that their initial diagnoses were AD or amnesic MCI, and AD remained the final diagnosis in four individuals. Language impairment eventually developed in 16 of the 18 individuals for whom this data was available, and was an initial or early symptom in 8 (44%).

Parkinsonism was seen in 11 of 18 patients (61%). It was an initial or early symptom/sign in 3 patients (17%). In three patients, parkinsonism developed four or more years after onset.

No patient exhibited features of motor neuron disease.

3.4 Neuroimaging Findings

MRI was available for review in 14 patients and brief descriptions of the MRI findings in these kindreds are presented in Table 3. Representative images highlighting the variable patterns of atrophy and the prominent subcortical white matter FLAIR hyperintensities are presented in Figure 2. Atrophy was identified in all patients, with frontotemporal predominance in 13. Parietal cortical atrophy was present in four. Subcortical white matter signal changes with a frontal predominance were identified in six individuals, and in four subjects (cases II.4 and III.2 of Kindred 2, case V.1 of Kindred 3, and case III.1 of Kindred 5), striking signal changes were present. These subcortical signal changes were most prominent in the regions of maximal cortical atrophy, and the extent of signal change increased with disease duration (Boeve et al. 2006). In two cases with focal temporal lobe atrophy (Kindred 1 case II.1 and Kindred 2 case III.3), milder subcortical signal changes were present. Longitudinal MRI scans spanning two or more years were available in six patients, which all showed progression of atrophy, ventricular, and subcortical white matter signal changes.

In Kindred 2, the topography of cortical atrophy was variable. Kindred 1 showed right cerebral hemisphere predominance in the three affected siblings, and the same can be presumed given their father's left-sided motor features. The opposite was apparent in Kindred 7, with all three siblings having left cerebral hemisphere atrophy.

3.5 Genetic Findings

As shown in Figure 3, a different *PGRN* mutation was identified in each family. (Baker et al. 2006; Gass et al. 2006). The specific mutations are listed in Table 1, indicating which individuals were genetically tested. Mutations in Kindreds 1, 2, 3, 6 and 7 cause frameshifts which create a premature stop codon, resulting in a null allele through nonsense-mediated decay. Similarly, the nonsense mutation in Kindred 3 induces nonsense-mediated decay. The mutation in family Kindred 5 destroys the 5' splice site of exon 1 thereby removing the start methionine codon, completely blocking *PGRN* protein from being generated (Boeve et al. 2006; Gass et al. 2006). Kindred 8 carried a mutation that disrupts the signal peptide sequence. RT-PCR transcript analyses for this mutation showed strongly reduced levels of mutant RNA, confirming the generation of a null allele (Gass et al. 2006).

At least one mutation carrier remains asymptomatic more than 15 years past the age of onset of their affected siblings.

3.6 Neuropathological Findings

Neuropathological data was available for 13 affected individuals; all exhibited FTLD-U with NII. Two cases had additional pathologies present; Kindred 2 case II.1 had coexistent Alzheimer pathology with moderate to frequent neuritic plaques (NIA-Reagan high likelihood) and Braak stage V neurofibrillary tangle distribution (The National Institute on Aging 1997) as well as vascular pathology, and Kindred 4 case II.1 had coexistent diffuse Lewy body pathology (McKeith et al. 2005). Prominent areas of hyperintensity on FLAIR imaging were noted in two individuals who had tissue available for review. Representative photomicrographs of these regions are shown in Figure 4, demonstrating marked microglial activation throughout the white matter as well as myelin loss and gliosis.

4. Discussion

4.1 Clinical Considerations

Our data suggest a wide spectrum of cognitive, behavioral, and motor features in FTDP-17 associated with mutations in *PGRN*. Clinical findings characteristic of FTDP-17 - cognitive impairment (executive dysfunction and/or aphasia), behavioral changes and parkinsonism – comprise the core phenotypic features, but specific clinical and radiologic features varied widely. Prominent early memory impairment –typical of amnesic MCI and Alzheimer’s disease (and an exclusionary criterion for the diagnosis of FTD) (Neary et al. 1998) – was present in several cases. Further, personality change was an early finding in only about half of the clinically evaluated patients, and was a late or absent finding in three. Levodopa-responsive parkinsonism was present in some affected individuals. A few had both visual hallucinations and parkinsonism, suggestive of Lewy body disease (McKeith et al. 2005). Some had progressive asymmetric rigidity and apraxia characteristic of the corticobasal syndrome (Boeve et al. 2003). These findings extend the range of phenotypes beyond those of previously published kindreds having *PGRN* mutations (Cruts et al. 2006; Huey et al. 2006; Masellis et al. 2006; Mukherjee et al. 2006; Snowden et al. 2006; Mesulam et al. 2007; Spina et al. 2007).

Phenotypic heterogeneity also exists within kindreds. To underscore this point, Kindred 2 had several members with primarily amnesic difficulties, one individual with PPA, another with FTDP, and one individual presented with amnesic complaints which quickly evolved into FTD. Phenotypic heterogeneity has been seen both within families and among families having identical mutations in *MAPT* as well (Bird et al. 1999).

Initial amnesic complaints occurred more frequently than would be expected in frontotemporal lobar degeneration. The proband of Kindred 5 presented with initial memory impairment suggestive of AD. Four members of Kindred 2 presented with amnesic complaints, following a course typical of AD. Another member of Kindred 2 presented with memory impairment, later following a course resembling FTD. Visuospatial deficits accompanied attention/executive impairment in three individuals clinically diagnosed with FTD. This pattern is also uncommon in FTD, but has been previously reported in a case associated with FTLD-U pathology (Meiner et al. 2005).

One sibling in Kindred 1 followed a clinical course resembling corticobasal syndrome (CBS), as did his father. His brother exhibited hemiparkinsonism late in his course, as did his sister. It has been proposed that frontotemporal lobar degeneration, CBS and progressive supranuclear palsy (PSP) represent different points of a disease spectrum on the basis of overlapping neuropathologic findings (Josephs et al. 2006b), and clinical characterization in another clinicopathologic series depended upon when in the course the subjects were evaluated (Kertesz et al. 2005). Families harboring both FTD and CBS phenotypes in association with *MAPT* mutations have been reported (Bugiani et al. 1999; Baba et al. 2007).

Others have reported families having FTD and CBS phenotypes associated with FTL-D-U pathology, (Brown et al. 1998) but this report predates the association of mutations in *PGRN* with FTL-D-U. Recently, familial CBS was associated with the c.708+1G>A splice donor site mutation in *PGRN* (Masellis et al. 2006) as well as a deletion in exon 7 (Benussi et al. 2006).

Parkinsonism was the predominant phenotype in Kindred 4, and CBS-like findings were present in many members of Kindred 1. Similar clinical features were seen in two other kindreds (Benussi et al. 2006; Masellis et al. 2006). It is unclear whether parkinsonism is more predominant in association with certain *PGRN* mutations, analogous to the parkinsonian predominant phenotype associated with the N279K mutation in *MAPT*. Presumably, this reflects more prominent extrapyramidal system pathology, but how specific mutations may direct the topography of pathology is not known. Two previous *PGRN* case-control studies reported higher frequencies of parkinsonism than this series (Josephs et al. 2007; Whitwell et al. 2007). Those series studied autopsy-confirmed cases associated with *PGRN* mutations, and the lower frequency in this series may reflect that some subjects of this study are still living and may later develop parkinsonism.

The apparent hemispheric predilection in some kindreds raises similar questions. The right cerebral hemisphere was maximally affected in four members of Kindred 1, and the left hemisphere was maximally affected in our Kindred 7, as well as the PPA3 and HDDD2 kindreds (Mukherjee et al. 2006; Mesulam et al. 2007). This hemispheric predilection has not been described in kindreds with mutations in *MAPT*. As previously suggested (Boeve et al. 2002), some genetic factors may dictate the development of neurologic disease, while others may determine the neurodegenerative topography, analogous to prion disease (Goldfarb et al. 1992).

No case had motor neuron disease (MND), and only one report has suggested the involvement of *PGRN* mutations in the development of ALS (Spina et al. 2007). In that case, a FTD patient with a strong family history of ALS was found to have a *PGRN* missense mutation (R433W). This individual had clinical manifestations suggestive of motor neuron disease as well. Neuropathologic examination identified FTL-D-U, but without NII as well as pathological changes suggestive of motor neuron disease. The relationship between this family's ALS and the *PGRN* mutation is not entirely clear. These data suggest that in contrast to *MAPT*, MND associated with mutations in *PGRN* may be quite rare (Mackenzie et al. 2006a; Josephs et al. 2007).

Sparse detail was available regarding the first generations in most families, and some later-onset cases may represent incident AD cases. However, one individual with onset at age 79 had neuropathological FTL-D-U with NII, and one mutation carrier is asymptomatic at age 73. Given the high penetrance, with 90% of subjects estimated to be symptomatic by age 70 (Gass et al. 2006), as well as the late age of onset in some individuals, it is possible that some late-onset cases had underlying FTL-D-U with NII pathology.

In aggregate, age of onset varied, with some individuals presenting in their 50's and others becoming symptomatic in their 80's. This was also true within two large kindreds (Kindreds 2 and 3), although in both of these kindreds, age of onset in the earlier generation was over 10 years older than that in subsequent generations. Generations II and III of Kindred 2 were particularly well characterized, and the mean age of onset in generation III was 15 years later than that of generation II (Table 2). In comparison to familial FTD associated with mutations in *MAPT*, the mean age of onset in our kindreds was older (64.5 years versus 45 years) and the mean disease duration somewhat shorter (6.6 years versus 8.4 years) (Baba et al. 2005).

We did not identify any gender-specificity in phenotype, but limited data regarding previous generations was available. Progranulin's role in gender-specific brain development (Suzuki and Nishihara 2002) may suggest gender-based differences in presentation and/or pattern of atrophy, warranting further study.

4.2 Neuroimaging Considerations

The MRI findings in FTD linked to charged multivesicular body protein 2B (*CHMP2B*) on chromosome 3 (Brown et al. 2004; Skibinski et al. 2005), to the valosin-containing protein (*VCP*) on chromosome 9 (Vance et al. 2006) and to *MAPT* on chromosome 17 have been frontotemporal cortical abnormalities varying from symmetric to markedly asymmetric atrophy (Basun et al. 1997; Rosso et al. 2001; Boeve et al. 2005). Most descriptions of MRI findings in sporadic and familial FTD have not reported subcortical white matter signal changes. In FTD-MND associated with chromosome 9, MRI has demonstrated "striking frontal lobe atrophy and an absence of signal change in the white matter of both hemispheres" (Vance et al. 2006). Subtle signal changes of medial temporal regions were noted in the pallidopontal nigral degeneration (PPND) kindred (Frank et al. 2007). Four of our cases with *PGRN* mutations had striking increased signal in the subcortical white matter adjacent to the cortical regions where atrophy was maximal, and in two other cases with focal temporal lobe atrophy, milder degrees of subcortical signal changes were present. Few FTDP cases with *PGRN* mutations and MRI scans have been reported, and the MR images and data presented in this report, along with our other reports (Boeve et al. 2002; Boeve et al. 2005), provide far more detail than other reports of *PGRN* mutation families. The relative sensitivity and specificity of these subcortical signal changes for cases harboring a *PGRN* mutation remains to be determined.

A recent volumetric analysis comparing individuals having FTL-D-U neuropathology suggested a more severe and widespread pattern of atrophy involving the frontal, temporal and even parietal cortex in those having *PGRN* mutations (Whitwell et al. 2007) compared to those without *PGRN* mutations. Additional analyses are warranted to further investigate the neuroimaging correlates in FTL-D-U with and without associated *PGRN* mutations and in the tauopathies with and without associated *MAPT* mutations.

4.3 Neuropathologic Considerations

Many cases of "dementia lacking distinctive histology" (Knopman et al. 1990) have subsequently been shown to exhibit positive immunostaining for ubiquitin (Josephs et al. 2004; Lipton et al. 2004; Mackenzie et al. 2006c). In our series, brain tissue was available in 13 cases. Neuropathologic examination in all 13 cases demonstrated features of FTL-D-U with NII. The presence of NII is a feature of FTL-D-U with *PGRN* mutations, but is not specific for this diagnosis as NII have been observed in FTL-D-U cases without *PGRN* mutations (Josephs et al. 2007) and in FTL-D due to mutations in *VCP* (Forman et al. 2006). None of our cases had pathological evidence of motor neuron disease. We confirm that FTL-D-U pathology can underlie the CBS, as previously reported (Grimes et al. 1999; Benussi et al. 2006; Masellis et al. 2006).

Two cases had mixed pathologies; one had FTL-D-U with Alzheimer and vascular pathology, and the other FTL-D-U plus Lewy body disease. A recent study suggested that anti-TDP-43 antibodies are specific to inclusions in FTL-D-U (Neumann et al. 2006), which we used to confirm the diagnosis of FTL-D-U in our two cases with mixed pathologies. Therefore, we suggest that TDP-43 immunohistochemistry be considered in cases with clinical features of FTD, but pathological diagnosis other than FTL-D. In two cases with striking FLAIR abnormalities, myelin loss and gliosis with associated microglial activation was demonstrated in these regions.

4.4 Disease Mechanism Considerations

All *PGRN* mutations reported in this study are expected to create functional null alleles, predominantly as a result of reading frame shifts and the creation of premature termination codons, such that no progranulin protein is expressed. This is proposed to lead to haploinsufficiency, whereby the single normal copy of the gene is incapable of producing sufficient protein to assure normal function. We suspect this lack of progranulin causes neurodegeneration, but the precise neurodegenerative mechanisms are unknown. Transgenic mouse models will soon enable investigators to test whether replacement of progranulin, or agents that increase progranulin production or secretion, can modulate neurodegeneration.

Acknowledgments

This research is supported by National Institute on Aging grants AG06786, AG16574, AG11378 and AG07216, and the Robert H. and Clarice Smith and Abigail Van Buren Alzheimer's Disease Research Program of the Mayo Foundation. RR is a postdoctoral fellow from the Fund for Scientific Research Flanders (FWO-F). We thank the Mayo Clinic Alzheimer's Disease Research Center staff for their assistance in evaluating the subjects. We particularly thank the members of these eight families for participating in neurodegenerative disease research.

References

- Baba Y, Baker MC, Le Ber I, Brice A, Maeck L, Kohlhase J, Yasuda M, Stoppe G, Bugiani O, Sperfeld AD, Tsuboi Y, Uitti RJ, Farrer MJ, Ghetti B, Hutton ML, Wszolek ZK. Clinical and genetic features of families with frontotemporal dementia and parkinsonism linked to chromosome 17 with a p301s tau mutation. *J Neural Transm.* 2007 [Epub ahead of print].
- Baba Y, Tsuboi Y, Baker MC, Uitti RJ, Hutton ML, Dickson DW, Farrer M, Putzke JD, Woodruff BK, Ghetti B, Murrell JR, Boeve BF, Petersen RC, Verpillat P, Brice A, Delisle MB, Rascol O, Arima K, Dysken MW, Yasuda M, Kobayashi T, Sunohara N, Komure O, Kuno S, Sperfeld AD, Stoppe G, Kohlhase J, Pickering-Brown S, Neary D, Bugiani O, Wszolek ZK. The effect of tau genotype on clinical features in ftdp-17. *Parkinsonism Relat Disord.* 2005; 11(4):205–208. [PubMed: 15878580]
- Baker M, Mackenzie IR, Pickering-Brown SM, Gass J, Rademakers R, Lindholm C, Snowden J, Adamson J, Sadovnick AD, Rollinson S, Cannon A, Dwosh E, Neary D, Melquist S, Richardson A, Dickson D, Berger Z, Eriksen J, Robinson T, Zehr C, Dickey CA, Crook R, McGowan E, Mann D, Boeve B, Feldman H, Hutton M. Mutations in progranulin cause tau-negative frontotemporal dementia linked to chromosome 17. *Nature.* 2006; 442(7105):916–919. [PubMed: 16862116]
- Basun H, Almkvist O, Axelman K, Brun A, Campbell TA, Collinge J, Forsell C, Froelich S, Wahlund LO, Wetterberg L, Lannfelt L. Clinical characteristics of a chromosome 17-linked rapidly progressive familial frontotemporal dementia. *Arch Neurol.* 1997; 54(5):539–544. [PubMed: 9152110]
- Benussi L, Binetti G, Sina E, Gigola L, Bettecken T, Meitinger T, Ghidoni R. A novel deletion in progranulin gene is associated with ftdp-17 and cbs. *Neurobiol Aging.* 2006 [Epub ahead of print].
- Bird TD, Nochlin D, Poorkaj P, Cherrier M, Kaye J, Payami H, Peskind E, Lampe TH, Nemens E, Boyer PJ, Schellenberg GD. A clinical pathological comparison of three families with frontotemporal dementia and identical mutations in the tau gene (p3011). *Brain.* 1999; 122(Pt 4): 741–756. [PubMed: 10219785]
- Boeve BF, Baker M, Dickson DW, Parisi JE, Giannini C, Josephs KA, Hutton M, Pickering-Brown SM, Rademakers R, Tang-Wai D, Jack CR Jr, Kantarci K, Shiung MM, Golde T, Smith GE, Geda YE, Knopman DS, Petersen RC. Frontotemporal dementia and parkinsonism associated with the IVS1+1G→A mutation in progranulin: A clinicopathologic study. *Brain.* 2006; 129(Pt 11):3103–3114. [PubMed: 17030535]
- Boeve BF, Lang AE, Litvan I. Corticobasal degeneration and its relationship to progressive supranuclear palsy and frontotemporal dementia. *Ann Neurol.* 2003; 54(Suppl 5):S15–19. [PubMed: 12833363]
- Boeve BF, Maraganore DM, Parisi JE, Ivnik RJ, Westmoreland BF, Dickson DW, Hutton M, Hardy J, Caselli RJ, Petersen RC. Corticobasal degeneration and frontotemporal dementia presentations in a

- kindred with nonspecific histopathology. *Dement Geriatr Cogn Disord*. 2002; 13(2):80–90. [PubMed: 11844889]
- Boeve BF, Tremont-Lukats IW, Waclawik AJ, Murrell JR, Hermann B, Jack CR Jr, Shiung MM, Smith GE, Nair AR, Lindor N, Koppikar V, Ghetti B. Longitudinal characterization of two siblings with frontotemporal dementia and parkinsonism linked to chromosome 17 associated with the s305n tau mutation. *Brain*. 2005; 128(Pt 4):752–772. [PubMed: 15615814]
- Brown J, Gydesen S, Johannsen P, Gade A, Skibinski G, Chakrabarti L, Brun A, Spillantini M, Yancopoulou D, Thusgaard T, Sorensen A, Fisher E, Collinge J. Frontotemporal dementia linked to chromosome 3. *Dement Geriatr Cogn Disord*. 2004; 17(4):274–276. [PubMed: 15178935]
- Brown J, Lantos PL, Rossor MN. Familial dementia lacking specific pathological features presenting with clinical features of corticobasal degeneration. *J Neurol Neurosurg Psychiatry*. 1998; 65(4):600–603. [PubMed: 9771798]
- Bugiani O, Murrell JR, Giaccone G, Hasegawa M, Ghigo G, Tabaton M, Morbin M, Primavera A, Carella F, Solaro C, Grisoli M, Savoiano M, Spillantini MG, Tagliavini F, Goedert M, Ghetti B. Frontotemporal dementia and corticobasal degeneration in a family with a p301s mutation in tau. *J Neuropathol Exp Neurol*. 1999; 58(6):667–677. [PubMed: 10374757]
- Cruts M, Gijssels I, van der Zee J, Engelborghs S, Wils H, Pirici D, Rademakers R, Vandenberghe R, Dermaut B, Martin JJ, van Duijn C, Peeters K, Sciot R, Santens P, De Pooter T, Mattheijssens M, Van den Broeck M, Cuijt I, Vennekens K, De Deyn PP, Kumar-Singh S, Van Broeckhoven C. Null mutations in progranulin cause ubiquitin-positive frontotemporal dementia linked to chromosome 17q21. *Nature*. 2006; 442(7105):920–924. [PubMed: 16862115]
- Forman MS, Mackenzie IR, Cairns NJ, Swanson E, Boyer PJ, Drachman DA, Jhaveri BS, Karlawish JH, Pestronk A, Smith TW, Tu PH, Watts GD, Markesbery WR, Smith CD, Kimonis VE. Novel ubiquitin neuropathology in frontotemporal dementia with valosin-containing protein gene mutations. *J Neuropathol Exp Neurol*. 2006; 65(6):571–581. [PubMed: 16783167]
- Foster NL, Wilhelmsen K, Sima AA, Jones MZ, D'Amato CJ, Gilman S. Frontotemporal dementia and parkinsonism linked to chromosome 17: A consensus conference. Conference participants. *Ann Neurol*. 1997; 41(6):706–715. [PubMed: 9189031]
- Frank AR, Wszolek ZK, Jack CR Jr, Boeve BF. Distinctive MRI findings in pallidopontonigral degeneration (PPND). *Neurology*. 2007; 68(8):620–621. [PubMed: 17310038]
- Gass J, Cannon A, Mackenzie IR, Boeve B, Baker M, Adamson J, Crook R, Melquist S, Kuntz K, Petersen R, Josephs K, Brown SP, Graff-Radford N, Uitti R, Dickson D, Wszolek Z, Gonzalez J, Beach TG, Bigio E, Johnson N, Weintraub S, Mesulam M, White CL 3rd, Woodruff B, Caselli R, Hsiung GY, Feldman H, Knopman D, Hutton M, Rademakers R. Mutations in progranulin are a major cause of ubiquitin-positive frontotemporal lobar degeneration. *Hum Mol Genet*. 2006; 15(20):2988–3001. [PubMed: 16950801]
- Goldfarb LG, Petersen RB, Tabaton M, Brown P, LeBlanc AC, Montagna P, Cortelli P, Julien J, Vital C, Pendelbury WW, et al. Fatal familial insomnia and familial Creutzfeldt-Jakob disease: Disease phenotype determined by a DNA polymorphism. *Science*. 1992; 258(5083):806–808. [PubMed: 1439789]
- Grimes DA, Bergeron CB, Lang AE. Motor neuron disease-inclusion dementia presenting as cortical-basal ganglionic degeneration. *Mov Disord*. 1999; 14(4):674–680. [PubMed: 10435507]
- Huey ED, Grafman J, Wassermann EM, Pietrini P, Tierney MC, Ghetti B, Spina S, Baker M, Hutton M, Elder JW, Berger SL, Heflin KA, Hardy J, Momeni P. Characteristics of frontotemporal dementia patients with a progranulin mutation. *Ann Neurol*. 2006; 60(3):374–380. [PubMed: 16983677]
- Hutton M, Lendon CL, Rizzu P, Baker M, Froelich S, Houlden H, Pickering-Brown S, Chakraverty S, Isaacs A, Grover A, Hackett J, Adamson J, Lincoln S, Dickson D, Davies P, Petersen RC, Stevens M, de Graaff E, Wauters E, van Baren J, Hillebrand M, Joosse M, Kwon JM, Nowotny P, Che LK, Norton J, Morris JC, Reed LA, Trojanowski J, Basun H, Lannfelt L, Neystat M, Fahn S, Dark F, Tannenberg T, Dodd PR, Hayward N, Kwok JB, Schofield PR, Andreadis A, Snowden J, Craufurd D, Neary D, Owen F, Oostra BA, Hardy J, Goate A, van Swieten J, Mann D, Lynch T, Heutink P. Association of missense and 5'-splice-site mutations in tau with the inherited dementia FTDP-17. *Nature*. 1998; 393(6686):702–705. [PubMed: 9641683]

- Ingram EM, Spillantini MG. Tau gene mutations: Dissecting the pathogenesis of FTDP-17. *Trends Mol Med*. 2002; 8(12):555–562. [PubMed: 12470988]
- Josephs KA, Ahmed Z, Katsuse O, Parisi JF, Boeve BF, Knopman DS, Petersen RC, Davies P, Duara R, Graff-Radford NR, Uitti RJ, Rademakers R, Adamson J, Baker M, Hutton ML, Dickson DW. Neuropathologic features of frontotemporal lobar degeneration with ubiquitin-positive inclusions with progranulin gene (PGRN) mutations. *J Neuropathol Exp Neurol*. 2007; 66(2):142–151. [PubMed: 17278999]
- Josephs KA, Holton JL, Rossor MN, Godbolt AK, Ozawa T, Strand K, Khan N, Al-Sarraj S, Revesz T. Frontotemporal lobar degeneration and ubiquitin immunohistochemistry. *Neuropathol Appl Neurobiol*. 2004; 30(4):369–373. [PubMed: 15305982]
- Josephs KA, Parisi JE, Knopman DS, Boeve BF, Petersen RC, Dickson DW. Clinically undetected motor neuron disease in pathologically proven frontotemporal lobar degeneration with motor neuron disease. *Arch Neurol*. 2006a; 63(4):506–512. [PubMed: 16606762]
- Josephs KA, Petersen RC, Knopman DS, Boeve BF, Whitwell JL, Duffy JR, Parisi JE, Dickson DW. Clinicopathologic analysis of frontotemporal and corticobasal degenerations and psp. *Neurology*. 2006b; 66(1):41–48. [PubMed: 16401843]
- Kertesz A, Kawarai T, Rogaeva E, St George-Hyslop P, Poorkaj P, Bird TD, Munoz DG. Familial frontotemporal dementia with ubiquitin-positive, tau-negative inclusions. *Neurology*. 2000; 54(4):818–827. [PubMed: 10690970]
- Kertesz A, McMonagle P, Blair M, Davidson W, Munoz DG. The evolution and pathology of frontotemporal dementia. *Brain*. 2005; 128(Pt 9):1996–2005. [PubMed: 16033782]
- Knopman DS, Mastri AR, Frey WH 2nd, Sung JH, Rustan T. Dementia lacking distinctive histologic features: A common non-alzheimer degenerative dementia. *Neurology*. 1990; 40(2):251–256. [PubMed: 2300243]
- Lipton AM, White CL 3rd, Bigio EH. Frontotemporal lobar degeneration with motor neuron disease-type inclusions predominates in 76 cases of frontotemporal degeneration. *Acta Neuropathol (Berl)*. 2004; 108(5):379–385. [PubMed: 15351890]
- Mackenzie IR, Baker M, Pickering-Brown S, Hsiung GY, Lindholm C, Dwosh E, Gass J, Cannon A, Rademakers R, Hutton M, Feldman HH. The neuropathology of frontotemporal lobar degeneration caused by mutations in the progranulin gene. *Brain*. 2006a; 129(Pt 11):3081–3090. [PubMed: 17071926]
- Mackenzie IR, Baker M, West G, Woulfe J, Qadi N, Gass J, Cannon A, Adamson J, Feldman H, Lindholm C, Melquist S, Pettman R, Sadovnick AD, Dwosh E, Whiteheart SW, Hutton M, Pickering-Brown SM. A family with tau-negative frontotemporal dementia and neuronal intranuclear inclusions linked to chromosome 17. *Brain*. 2006b; 129(Pt 4):853–867. [PubMed: 16401619]
- Mackenzie IR, Shi J, Shaw CL, Duplessis D, Neary D, Snowden JS, Mann DM. Dementia lacking distinctive histology (DLDH) revisited. *Acta Neuropathol (Berl)*. 2006c; 112(5):551–559. [PubMed: 16900341]
- Masellis M, Momeni P, Meschino W, Heffner R Jr, Elder J, Sato C, Liang Y, St George-Hyslop P, Hardy J, Bilbao J, Black S, Rogaeva E. Novel splicing mutation in the progranulin gene causing familial corticobasal syndrome. *Brain*. 2006; 129(Pt 11):3115–3123. [PubMed: 17030534]
- McKeith IG, Dickson DW, Lowe J, Emre M, O'Brien JT, Feldman H, Cummings J, Duda JE, Lippa C, Perry EK, Aarsland D, Arai H, Ballard CG, Boeve B, Burn DJ, Costa D, Del Ser T, Dubois B, Galasko D, Gauthier S, Goetz CG, Gomez-Tortosa E, Halliday G, Hansen LA, Hardy J, Iwatsubo T, Kalaria RN, Kaufer D, Kenny RA, Korczyn A, Kosaka K, Lee VM, Lees A, Litvan I, Londo E, Lopez OL, Minoshima S, Mizuno Y, Molina JA, Mukaetova-Ladinska EB, Pasquier F, Perry RH, Schulz JB, Trojanowski JQ, Yamada M. Diagnosis and management of dementia with Lewy bodies: Third report of the DLB Consortium. *Neurology*. 2005; 65(12):1863–1872. [PubMed: 16237129]
- McKhann GM, Albert MS, Grossman M, Miller B, Dickson D, Trojanowski JQ. Clinical and pathological diagnosis of frontotemporal dementia: Report of the Work Group on Frontotemporal Dementia and Pick's Disease. *Arch Neurol*. 2001; 58(11):1803–1809. [PubMed: 11708987]

- Meiner Z, Newman JP, Rosenman H, Soffer D, Steiner I. Frontotemporal dementia with ubiquitinated neuronal inclusions and visuospatial impairment. *Neurology*. 2005; 65(3):478–480. [PubMed: 16087922]
- Members of the Department of Neurology, M. C.. *Clinical examinations in neurology*. Mosby; St. Louis: 1998.
- Mesulam M, Johnson N, Krefft TA, Gass JM, Cannon AD, Adamson JL, Bigio EH, Weintraub S, Dickson DW, Hutton ML, Graff-Radford NR. Progranulin mutations in primary progressive aphasia: The ppa1 and ppa3 families. *Arch Neurol*. 2007; 64(1):43–47. [PubMed: 17210807]
- Mukherjee O, Pastor P, Cairns NJ, Chakraverty S, Kauwe JS, Shears S, Behrens MI, Budde J, Hinrichs AL, Norton J, Levitch D, Taylor-Reinwald L, Gitcho M, Tu PH, Grinberg L, Tenenholz, Liscic RM, Armendariz J, Morris JC, Goate AM. HDDD2 is a familial frontotemporal lobar degeneration with ubiquitin-positive, tau-negative inclusions caused by a missense mutation in the signal peptide of progranulin. *Ann Neurol*. 2006; 60(3):314–322. [PubMed: 16983685]
- Neary D, Snowden JS, Gustafson L, Passant U, Stuss D, Black S, Freedman M, Kertesz A, Robert PH, Albert M, Boone K, Miller BL, Cummings J, Benson DF. Frontotemporal lobar degeneration: A consensus on clinical diagnostic criteria. *Neurology*. 1998; 51(6):1546–1554. [PubMed: 9855500]
- Neumann M, Sampathu DM, Kwong LK, Truax AC, Micsenyi MC, Chou TT, Bruce J, Schuck T, Grossman M, Clark CM, McCluskey LF, Miller BL, Masliah E, Mackenzie IR, Feldman H, Feiden W, Kretschmar HA, Trojanowski JQ, Lee VM. Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science*. 2006; 314(5796):130–133. [PubMed: 17023659]
- Rademakers R, Cruts M, Dermaut B, Sleegers K, Rosso SM, Van den Broeck M, Backhovens H, van Swieten J, van Duijn CM, Van Broeckhoven C. Tau negative frontal lobe dementia at 17q21: Significant finemapping of the candidate region to a 4. *Mol Psychiatry*. 2002; 7(10):1064–1074. [PubMed: 12476321]
- Rosso SM, Kamphorst W, de Graaf B, Willemsen R, Ravid R, Niermeijer MF, Spillantini MG, Heutink P, van Swieten JC. Familial frontotemporal dementia with ubiquitin-positive inclusions is linked to chromosome 17q21-22. *Brain*. 2001; 124(Pt 10):1948–1957. [PubMed: 11571213]
- Savioz A, Riederer BM, Heutink P, Rizzu P, Tolnay M, Kovari E, Probst A, Riederer IM, Bouras C, Leuba G. Tau and neurofilaments in a family with frontotemporal dementia unlinked to chromosome 17q21-22. *Neurobiol Dis*. 2003; 12(1):46–55. [PubMed: 12609488]
- Skibinski G, Parkinson NJ, Brown JM, Chakrabarti L, Lloyd SL, Hummerich H, Nielsen JE, Hodges JR, Spillantini MG, Thusgaard T, Brandner S, Brun A, Rossor MN, Gade A, Johannsen P, Sorensen SA, Gydesen S, Fisher EM, Collinge J. Mutations in the endosomal escrtiii-complex subunit CHMP2b in frontotemporal dementia. *Nat Genet*. 2005; 37(8):806–808. [PubMed: 16041373]
- Snowden JS, Pickering-Brown SM, Mackenzie IR, Richardson AM, Varma A, Neary D, Mann DM. Progranulin gene mutations associated with frontotemporal dementia and progressive non-fluent aphasia. *Brain*. 2006; 129(Pt 11):3091–3102. [PubMed: 17003069]
- Spina S, Murrell JR, Huey ED, Wassermann EM, Pietrini P, Baraibar MA, Barbeito AG, Troncoso JC, Vidal R, Ghetti B, Grafman J. Clinicopathologic features of frontotemporal dementia with progranulin sequence variation. *Neurology*. 2007; 68(11):820–827. [PubMed: 17202431]
- Suzuki M, Nishihara M. Granulin precursor gene: A sex steroid-inducible gene involved in sexual differentiation of the rat brain. *Mol Genet Metab*. 2002; 75(1):31–37. [PubMed: 11825061]
- The National Institute on Aging; Reagan Institute Working Group on Diagnostic Criteria for the Neuropathological Assessment of Alzheimer's Disease. Consensus recommendations for the postmortem diagnosis of Alzheimer's disease. *Neurobiol Aging*. 1997; 18(4 Suppl):S1–S2. [PubMed: 9330978]
- van der Zee J, Rademakers R, Engelborghs S, Gijssels I, Bogaerts V, Vandenberghe R, Santens P, Caekebeke J, De Pooter T, Peeters K, Lubke U, Van den Broeck M, Martin JJ, Cruts M, De Deyn PP, Van Broeckhoven C, Dermaut B. A Belgian ancestral haplotype harbours a highly prevalent mutation for 17q21-linked tau-negative FTL. *Brain*. 2006; 129(Pt 4):841–852. [PubMed: 16495329]
- Vance C, Al-Chalabi A, Ruddy D, Smith BN, Hu X, Sreedharan J, Siddique T, Schelhaas HJ, Kusters B, Troost D, Baas F, de Jong V, Shaw CE. Familial amyotrophic lateral sclerosis with

frontotemporal dementia is linked to a locus on chromosome 9p13.2-21.3. *Brain*. 2006; 129(Pt 4): 868–876. [PubMed: 16495328]

Whitwell JL, Jack CR Jr, Baker M, Rademakers R, Adamson J, Boeve BF, Knopman DS, Parisi JF, Petersen RC, Dickson DW, Hutton ML, Josephs KA. Voxel-based morphometry in frontotemporal lobar degeneration with ubiquitin-positive inclusions with and without progranulin mutations. *Arch Neurol*. 2007; 64(3):371–376. [PubMed: 17353379]

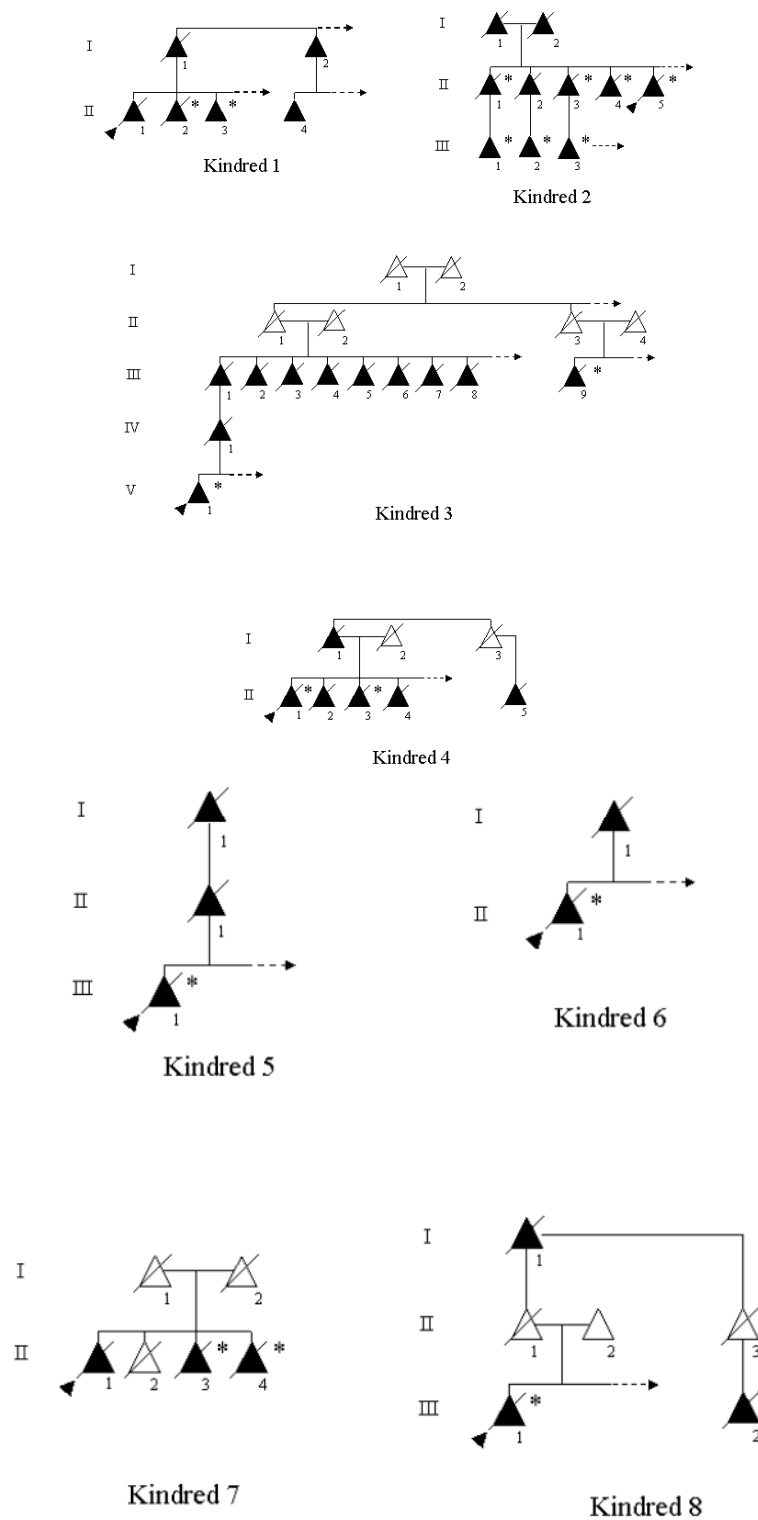


Figure 1. Pedigrees of the eight kindreds. Triangles represent individuals, shaded triangles represent affected individuals. Triangles with diagonal lines through them represent deceased individuals. The proband is indicated by a shaded arrowhead. Individuals with a confirmed

progranulin mutation are indicated by an asterisk. An arrow pointing to the right in a sibship represents additional unaffected persons, but they were purposefully excluded to maintain confidentiality.

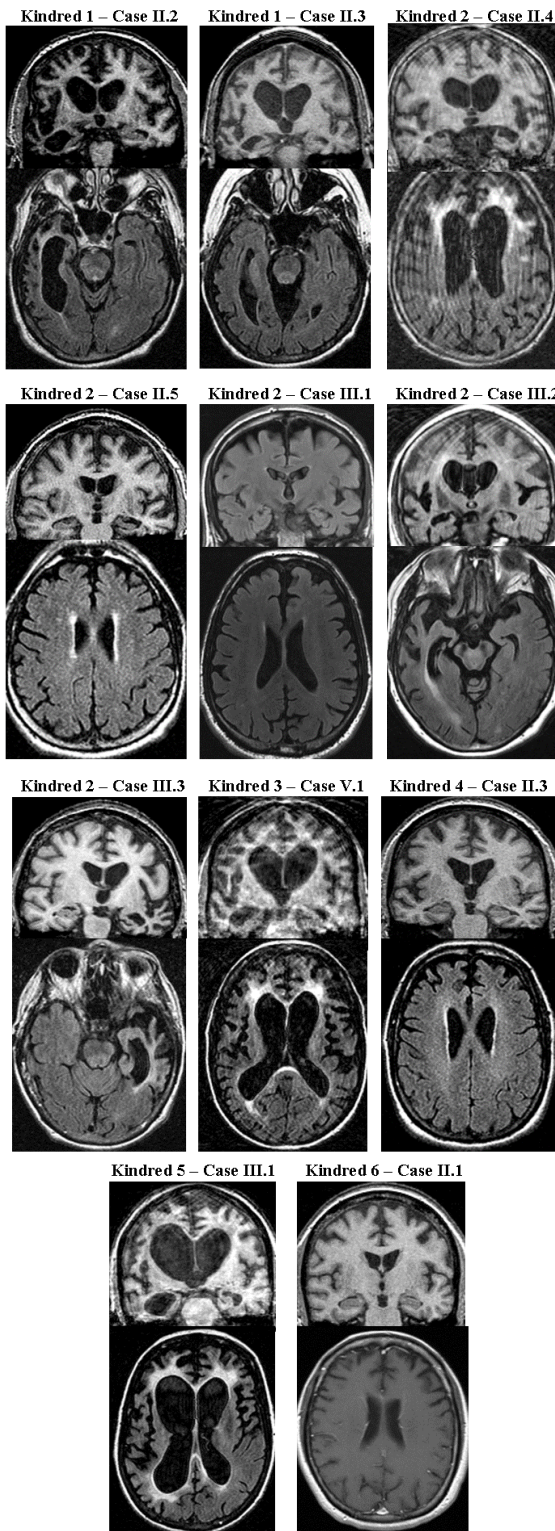


Figure 2. Representative coronal [T1-weighted or fluid attenuation inversion recovery (FLAIR)] (top), and axial T1-weighted or FLAIR (bottom) images for each affected individual for which

MRI scans were available. Note the variability in the patterns of cerebral atrophy and degree of signal changes on FLAIR images.

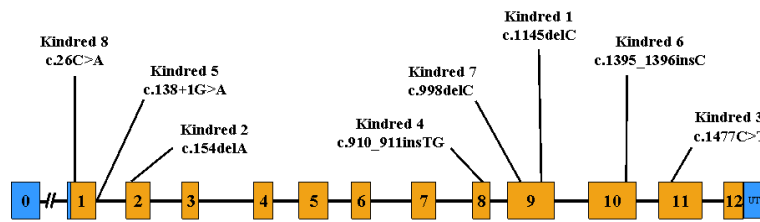


Figure 3. Schematic representation of the progranulin (*PGRN*) gene illustrating the location of the mutations in the eight kindreds. Mutations are numbered according to GenBank Accession number NM_002087.2.

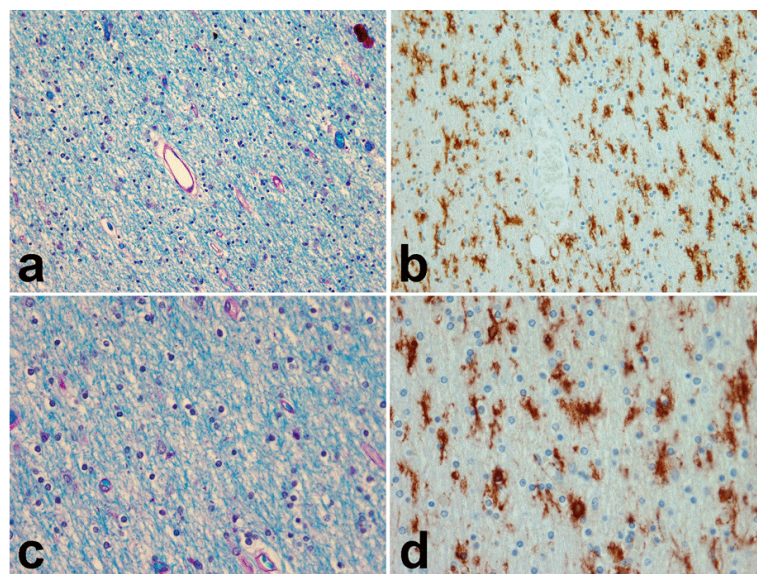


Figure 4. Photomicrographs showing myelin rarefaction and vacuolation, worse around blood vessels. Microglial stain (B and D) shows marked microglial activation throughout the white matter. Adjacent Luxol fast blue stain (A and C) shows myelin loss and gliosis.

Table 1

Demographic, clinical, neuropathologic, and genetic findings of all affected individuals

Kindred	Case	Duration (years)	Initial Diagnosis	Current/Final Diagnosis	Autopsy Findings	PGRN Mutation	Reference
Kindred 1	I.1	5	dementia NOS	encephalomalacia			
	I.2	*	dementia NOS	dementia NOS			
	II.1 (P)	6	PD	CBS	FTLD-U NII		Boeve et al, 2002
	II.2	7	FTD	FTDP	FTLD-U NII	c.1145delC	Boeve et al, 2002
Kindred 2	II.3	>4	FTD	FTD/CBS		c.1145delC	
	II.4	*	dementia NOS	dementia NOS			
	I.1	5	AD	AD			
	I.2	3	dementia NOS	dementia NOS			
Kindred 3	II.1	7	AD	AD	FTLD-U NII + AD + VD	c.154delA	
	II.2	*	dementia NOS	dementia NOS			
	II.3	2	AD	AD + Park	FTLD-U NII	c.154delA	
	II.4	7	AD	FTD	FTLD-U NII	c.154delA	
	II.5 (P)	7	amnesic MCI	AD	FTLD-U NII	c.154delA	
Kindred 3	III.1	>2	amnesic MCI + dep	amnesic MCI + dep		c.154delA	
	III.2	>7	FTD/DLB	FTDP		c.154delA	
	III.3	>7	PPA	PPA/FTD		c.154delA	
	III.1	9	dementia NOS	dementia NOS			
	III.2	7	dementia, AD	AD			
Kindred 3	III.3	1 (fire)	dementia NOS	dementia NOS			
	III.4	9	dementia NOS	dementia NOS			
	III.5	>2	dementia NOS	dementia NOS			

Kindred	Case	Duration (years)	Initial Diagnosis	Current/Final Diagnosis	Autopsy Findings	PGRN Mutation	Reference
Kindred 4	III.6	*	dementia NOS	dementia NOS			
	III.7	5	dementia NOS	dementia NOS			
	III.8	7	dementia NOS	dementia NOS			
	III.9	3	AD	AD	FTLD-U NII	c.1477C>T	
Kindred 5	IV.1	1 (MVA)	not Dx	not Dx			
	V.1 (P)	6	FTD	FTD		c.1477C>T	
	I.1	5	PD	PD			
	II.1 (P)	13	PD	PD + D	FTLD-U NII + LBD	c.910_911insTG	
Kindred 6	II.2	7	dementia NOS	dementia NOS			
	II.3	8	FTD	FTD	FTLD-U NII	c.910_911insTG	
	II.4	*	PD	PD + dementia NOS			
	II.5	*	PD	PD			
Kindred 7	I.1	13	dementia NOS	dementia NOS			
	II.1	7	dementia NOS	dementia NOS			
	III.1 (P)	7	AD	FTD	FTLD-U NII	c.138+1G>A	Boeve et al, 2006
	I.1	5	dementia NOS	dementia NOS			
Kindred 8	II.1 (P)	5	FTD	FTD	FTLD-U NII	c.1395_1396insC	Mesulam et al, 2007
	II.1	12	PPA	PPA			
	II.3	4	PPA	PPA	FTLD-U NII	c.998delG	Mesulam et al, 2007
	II.4 (P)	8	PPA	PPA	FTLD-U NII	c.998delG	Mesulam et al, 2007
Kindred 8	I.1	14	aphasia	aphasic dementia			
	III.1 (P)	>4	PPA	PPA		c.26C>A	
	III.2	*	aphasia	aphasic dementia			

Abbreviations: AD=Alzheimer's disease; CBS=corticobasal syndrome; dep=depression, DLB=dementia with Lewy bodies; FTD=frontotemporal dementia; FTDP=frontotemporal dementia and parkinsonism; FTLD-U NII=frontotemporal lobar degeneration with ubiquitin positive inclusions and neuronal intranuclear inclusions; LBD=Lewy body disease; MCI=mild cognitive impairment; MVA= motor vehicle accident; NOS=not otherwise specified; not Dx=not diagnosed; park=parkinsonism; PD=Parkinson's disease; PPA=primary progressive aphasia; VD=vascular dementia; * = unknown, (P)=proband

Table 2

Specific clinical features among affected individuals examined at Mayo Clinic

Kindred	Case	Personality Change	Hyperoral	Executive Dysfunction	Memory Dysfunction	Language Dysfunction	Parkinsonism
1	II.1	mid		mid	mid	mid	early
	II.2	early	late	mid	mid	late	mid
	II.3	early	late	early	early	absent	late
	II.3	early			early	mid	early
	II.4	early	early	early		early	late
2	II.5	mid	late	early	early	late	late
	III.1	early		early	early	absent	absent
	III.2	early	mid	early	mid	late	late
3	III.3	late		late	late	early	absent
	V.1	early	mid	later	early	early	mid
4	II.1	mid		late	mid	late	early
	II.3	early		mid	early	later	absent
5	III.1	early	mid	early	early	late	late
	II.1	early	mid	early	early	early	absent
6	II.1	absent	absent	present	present	early	absent
	II.1	present	present	late	absent	early	mid
7	II.4	present	absent	mid	absent	early	absent
	III.1	absent	absent	absent	absent	early	absent
8	III.1	absent	absent	absent	absent	early	absent

early=onset within first year of symptoms; mid=onset 1-2 years after symptom onset; late=onset 3 or more years after onset; absent=no evidence in clinical records of this feature being present. Empty fields reflect insufficient details in the clinical record to determine if that feature was present or absent.

Table 3

Descriptive MRI findings in affected individuals

Case	MRI Findings
Kindred 1 II.2	Asymmetric right greater than left frontotemporal atrophy, and lesser right parietal and occipital atrophy. Subcortical white matter changes were associated with this atrophy. Subsequent imaging confirmed progression of these findings.
Kindred 1 II.3	Mild temporal and hippocampal atrophy, and subtle subcortical white matter changes, greater on the right than the left
Kindred 2 II.4	Prominent frontotemporal atrophy and bilateral frontal white matter T2 hyperintensity with extension into the cortex
Kindred 2 II.5	Mild atrophy along the falx anteriorly, more obvious atrophy in the left mesial and inferolateral temporal lobe and mild leukoaraiosis
Kindred 2 III.1	Mild diffuse atrophy, more obvious along the anterior falx, without significant mesial temporal atrophy
Kindred 2 III.2	Moderately severe generalized cerebral atrophy, most prominent in the right temporal lobe. Mild subcortical white matter T2 hyperintensity evident in the right temporal lobe.
Kindred 2 III.3	Initial MRI showed marked atrophy of the left anterior temporal lobe and mild atrophy of the left frontal opercular region. Subsequent MRI examinations demonstrated progressive left hemispheric atrophy, most severely in the temporal lobe
Kindred 3 V.1	Moderately severe atrophy, most prominent in the frontal and temporal lobes. Increased T2 signal in the frontal subcortical white matter adjacent to the lateral ventricles
Kindred 4 II.3	Moderate anterior falcine atrophy without white matter signal changes
Kindred 5 III.1	Marked bilateral frontotemporoparietal atrophy particularly on the right, with subcortical white matter changes
Kindred 6 II.1	Mild diffuse cortical atrophy, slightly more prominent along the falx anteriorly
Kindred 7 II.3	Progressive left temporal atrophy with less severe frontal atrophy apparent on subsequent neuroimaging
Kindred 7 II.4	Progressive left temporal and parietal atrophy with enlargement of the Sylvian fissure
Kindred 8 III.1	Mild left frontotemporal atrophy