

NIH Public Access

Author Manuscript

Endocrinol Metab Clin North Am. Author manuscript; available in PMC 2012 September 1.

Published in final edited form as:

Endocrinol Metab Clin North Am. 2011 September ; 40(3): 591–614. doi:10.1016/j.ecl.2011.05.002.

Estrogens and Prostate Cancer: Etiology, Mediators, Prevention, and Management

Shuk-Mei Ho, Ph.D.,

Department of Environmental Health, Center for Environmental Genetics, and the Cancer Institute, College of Medicine, University of Cincinnati, Cincinnati, Ohio.

Ming-tsung Lee, M.Phil.,

Department of Environmental Health, College of Medicine, University of Cincinnati, Cincinnati, Ohio. Telephone 513-558-0595, Fax 513-558-0071, lee2mg@mail.uc.edu

Hung-Ming Lam, Ph.D., and

Department of Environmental Health, College of Medicine, University of Cincinnati, Cincinnati, Ohio. Telephone 513-558-0595, Fax 513-558-0071, lamhg@uc.edu

Yuet-Kin Leung, Ph.D.

Department of Environmental Health, Center for Environmental Genetics, and The Cancer Institute, College of Medicine, University of Cincinnati, Cincinnati, Ohio. Telephone 513-558-5181, Fax 513-558-0071, ricky.leung@uc.edu

Abstract

The relationship between hormones and the pathogenesis of prostate cancer (PCa) has been studied extensively. All the mainstay targets for hormonal PCa therapies are based on negating androgen action. Recent epidemiologic and experimental data have clearly pinpointed the key roles of estrogens in PCa development and progression. Racial and geographical differences, as well as age-associated changes, in estrogen synthesis and metabolism contribute significantly to the etiology by increasing the ratio of circulating estrogen to androgen, sex hormone binding globulin synthesis, and aromatase activity and reducing androgen glucuronidation and tissue bioactivation. Promotion of aberrant cell growth, evasion of apoptosis, increased oxidative stress and inflammation, and gains in adiposity and bioactivation to genotoxic carcinogens during adulthood are probable mechanisms of estrogen carcinogenicity, while "estrogen imprinting" via epigenetics in early-life also determines PCa risk. Although the effects of estrogens are known to be mediated by genomic actions of the two estrogen receptor (ER) subtypes ($E R \alpha$ and $E R \beta$), other non-canonical mediators, including the different ERβ isoforms, membrane and mitochondrial ERs, and G protein-coupled receptor 30, may have major actions diverging from classical ER actions. These new discoveries have led to renewed interest among the public and the medicinal field in estrogens and antiestrogens as singular and adjuvant PCa treatment and prevention regimens. This review summarizes current knowledge on how different estrogens/antiestrogens/estrogen mimics contribute to prostate carcinogenesis, the roles of the different mediators of estrogen in the process, and the potentials of new estrogenic/antiestrogenic compounds as targeted therapies for prevention and treatment of PCa.

Corresponding author. Address: Room 128, Kettering Complex, 3223 Eden Avenue, Department of Environmental Health, College of Medicine, University of Cincinnati, Cincinnati, OH 45267. Telephone 513-558-5701; Fax 513-558-0071; shuk-mei.ho@uc.edu. **Declaration of interest** The authors have nothing to disclose.

Keywords

Racial disparity; aging; sex steroid–binding globulin; estrogen receptor; ER spliced variants; GPR30; aromatase; antiestrogens; selective estrogen receptor modulator (SERM); nuclear receptor co-regulators; DNA methylation; developmental reprogramming; apigenin; phytoestrogen; apoptosis; oxidative stress; genomic damages; prostate cancer risk; hormonal therapy; epigenetics; genistein; resveratrol; raloxifene; toremifene; fulvestrant; estramustine; catecholestrogen

> Androgens are traditionally recognized as the major hormone promoting normal and aberrant growth of the prostate. Recent literature, however, suggests that estrogen could also be an important mediator of these processes. Estrogen alone or in synergy with androgen is responsible for the pathogenesis of prostate cancer (PCa). More important, recent experimental data suggest that estrogen or its mimics could determine the risk of PCa development as early as the prenatal stage via a process known as "estrogen imprinting." Here we a) review research findings that support a role for estrogens, estrogen mimics and estrogen metabolites in prostate carcinogenesis; b) discuss how different estrogen receptors (ER) mediate the action of estrogen in promoting the development and progression of PCa; and c) evaluate the potentials of estrogens, xenoestrogens, phytoestrogens, antiestrogens, and SERMs as therapeutics for prevention and treatment of PCa.

Epidemiologic and Animal-model Studies of the Relationship between Estrogens and Pathogenesis of PCa

Results from epidemiologic studies have suggested a role for estrogen in the pathogenesis of PCa. Racial/ethnic and geographical differences in the levels of estrogens provide a probable explanation for the disparity in the prevalence of PCa among various populations throughout the world.¹⁻³ Apropos to this view is the finding that levels of circulating estrogens in African-American men, whose incidence of PCa is the highest in United States, are higher than those in Caucasian Americans throughout their adult life.⁴⁻⁸ In contrast, Japanese men, whose incidence of PCa is low, have lower circulating levels of estrogen than do Dutch-European men.⁹ A global study on 5,003 men aged 65 years or older showed that blacks (in the US and in West Africa) had higher estrogen levels than Caucasians or Asians (in the US and in their homelands), with levels of total and free estradiol-17 β (E2) 10–16% higher and levels of estrone (E1) 27–39% higher than in the latter group.¹⁰ Moreover, the ratios of total E2 to total testosterone (T) and E1 to androstenedione were higher in blacks than in the other groups.10 A comprehensive analysis of the levels of androgens, estrogens, and their metabolites in circulation led to the conclusion that two fundamental metabolic processes, increased aromatase activity and reduced androgen glucuronidation, are major factors governing the ratio of estrogen to androgen in elderly men.¹⁰

Age is also a key risk factor for PCa.¹ The prevalence of PCa increases dramatically as men age; this is paralleled by a significant increase in the ratio of circulating estrogen to androgen levels, which may increase by up to 40% .¹¹⁻¹⁷ This age-related hormonal change, often referred to as "andropause," is caused by several endocrine events, including a decline in testicular function, and increases in adiposity, extragonadal aromatization, and the production of sex hormone–binding globulin (SHBG) as men age.^{12;18-21} The level of 5 α dihydrotestosterone (DHT) was found to decrease whereas those of estrogen (both E2 and E1) in the epithelial cells, to increase in the aging prostate (Figure 1).²² Since estrogens can be synthesized *de novo* via aromatase activity in the prostate, $\frac{3}{2}$ tissue estrogen levels may be more important than circulating estrogen levels in promoting prostate carcinogenesis and progression. In this regard, recent studies utilizing laser capture microdissection samples demonstrated that stromal rather than epithelial aromatase activity may be important in

upregulating the E2:T ratio in the tumor site via an alternative promoter activation mechanism during prostate carcinogenesis.24;25 Furthermore, aromatase-knockout mice, which cannot produce E2 locally in the prostate, have elevated levels of circulating T and DHT and, with age, are prone to the development of benign prostatic hyperplasia (BPH) but not PCa.26 Collectively, increased estrogenic influences on the prostate due to racial differences or andropause²⁷ may elevate the risk of neoplastic transformation of the prostatic epithelium in men.^{3;28;29}

Experimental models also support the suggestion that estrogens, alone or synergistically with androgens, are potent inducers of aberrant growth and neoplastic transformation in the prostate.1;2;30-33 Prenatal exposure to maternal estrogens or adult exposure to pharmacologic doses of estrogens induces a benign lesion termed *squamous metaplasia*, which is derived from the basal-cell proliferation of the prostates of various species, including humans.34 In a susceptible rat strain (Noble rats), chronic exposure to T plus E2 in adulthood promoted the evolution of a precancerous lesion similar to human prostatic intraepithelial neoplasia (PIN) and a high incidence of full-blown $PCa^{31;35-37}$ via the alteration of levels of estrogen to androgen in a manner mimicking that in aging men.35-38 Paradoxically, dietary soy that is rich in phytoestrogens can mitigate the tumor-promoting effect of T plus E2 in this rat model.³⁹ Moreover, treatment of the nude mice with T plus E2 could promote the progression of PCa and metastasis at distant organs in the tissue recombinants composed of mouse mensenchyme and a human prostatic epithelial cell line.⁴⁰ Cellular and molecular changes implicated as the mechanisms leading to prostate carcinogenesis include a) a dramatic increase in the proliferation of epithelial cells;³⁷ b) upregulation of growth factor– signaling pathways (e.g. TGF α /EGF receptor signaling;⁴¹ TGFβ signaling,⁴² IGF-1 and VEGF signaling, 43 and ER signaling); 44 c) prolactinemia or increased prolactin-receptor signaling;45 d) mitogen-activated protein kinase (MAPK) activation, possibly through Id-1;46;47 e) increased cell-survival potential through the overexpression of anti-apoptotic mediators (e.g., metallothionein⁴⁸ and TRPM-2/clusterin);^{49;50} f) elevation in oxidative stress-induced DNA damage;⁵¹⁻⁵³ g) changes in gene-expression profiles related to cell proliferation, DNA damage, activation of proto-oncogenes and transforming factors, IL-1B signaling, and TNF- α activation;^{54;55} and h) breakdown of epithelial basement membrane and stromal extracellular matrix due to the increase in gelatinolytic proteinase activity and altered expression of glycoconjugates in smooth muscles and their associated extracellular matrix.⁵⁶⁻⁵⁸ Another potential culprit associated with estrogen-induced/promoted PCa is hormone-induced chronic inflammation, $38,59-61$ although this view is not uniformly supported by all studies. 62

Influence of Estrogen Bioactivation and Detoxification on PCa Risk

The carcinogenic action of estrogen could be caused, in part, by metabolic activation of the natural estrogens E1 and E2 to genotoxic metabolites, such as 2- and 4-hydroxyl catechol estrogens, and their quinone/semiquinone intermediates that act as chemical carcinogens.⁶³ The bioactivation process is mediated principally by two cytochrome P450 enzymes, CYP1A1 and CYP1B1, whereas the enzyme catechol-O-methyltransferase (COMT) is responsible for the inactivation and removal of these genotoxic estrogen-derived intermediates.63-66 Once formed, these genotoxic metabolites can cause genomic damages through processes such as the formation of DNA adducts.⁶⁷ Supporting the suggestion that the genotoxicity of estrogen metabolites is involved in prostate carcinogenesis is the observation of increased DNA strand breakage52 and nuclear staining of 8-hydroxy-2′ $deoxy-guanosine⁶⁰$ in the prostates of rats treated with T plus E2. Conversely, increased expression or activity of COMT was shown to protect against estrogen-induced cancer by mediating the conversion of catechol estrogens into methoxyestrogens that have potent apoptotic activity against rapidly growing PCa cells,68 prompting the testing of combinatory

therapies involving methoxyestrogens and other standard therapies, such as hormone deprivation,69 docetaxel,70 and eugenol71 against PCa growth in model systems.

Furthermore, the aforementioned estrogen metabolites and their intermediates induce oxidative stress and likely promote the generation of high levels of reactive oxygen species (ROS), with damaging effects on proteins, lipids, and DNA in target tissues.29;66 Therefore, several isoforms of glutathione S-transferases (GSTs) have been reported to have protective effects against estrogen genotoxicity through the detoxification of ROS. Genetic polymorphisms that alter enzyme activity and epigenetics- or mutation-mediated silencing of the cognate genes are expected to affect the PCa risk by modulating the extent of lifelong exposure to genotoxic estrogen metabolites.29;72 Thus, polymorphisms of the *CYP1A1m1* allele, the *CYP1B1*-Leu432Val, *COMT* at codon 62 and 158, and the *GSTM1*-null genotype have been shown to modify PCa risk in certain populations.73-80 In addition, a *CYP1A1* variant (*CYP1A1v*) that resides preferentially in the nucleus and mitochondria was found to confer higher carcinogenic potential than its wild-type cytosolic counterpart.⁸¹ This finding is in agreement with the hypothesis that genotoxic estrogen metabolites produced in nuclei are potent tumor initiators.

Another metabolic pathway of relevance to estrogen carcinogenicity in the prostate is the *in situ* production of estrogen from androgen via aromatase encoded by *CYP19*. ²⁵ The enzyme and its activity have been demonstrated in the specimens of PCa and BPH.25;82 Clinical trials designed to test the efficacy of aromatase inhibitors in treating BPH⁸³⁻⁸⁵ and PCa⁸⁶⁻⁸⁹ have been reported. Polymorphisms in *CYP19* including intron 4[TTTA]n repeat⁹⁰ and *C*/T versus T/T genotype⁹¹ are associated with familial PCa. Continued investigation of the use of aromatase inhibitors as monotherapies or adjuvants for the treatment of PCa is warranted.26;92

Determination of PCa Risk during Early Life

The risk of developing PCa as an adult could be determined by early-life exposure to natural or environmental estrogens through a mechanism known as "estrogen imprinting."⁹³ Perinatal and neonatal exposure of rats $94-97$ or mice $98-100$ to estrogens or estrogen mimics induces inflammation, permanent changes in the levels of androgen and estrogen receptors, stromal hypertrophy, and elevated proliferative potentials in the prostatic epithelium of the adult aged gland.^{96;101-103} If the "estrogenized" adult glands are exposed to an elevated estrogen challenge during adult life (the "second hit"), full-blown PIN develops in the affected glands.¹⁰⁴ An environmental estrogen, bisphenol A (BPA), is equally effective in sensitizing the adult prostate to increased estrogenic influence during adulthood with regard to the induction of PIN. An unbiased screening has identified permanent alterations in the methylation status of a CpG island in the promoter of phosphodiesterase 4D4 *(Pde4e4)*, an enzyme responsible for regulating cellular cAMP.¹⁰⁴ This finding suggests estrogen imprinting may involve epigenetic reprogramming of prostatic transcription programs in early life. Whether analogous phenomena exist in humans is unclear. However, circulating E2 levels have been found to be higher in pregnant African-American women than in pregnant Caucasian-American women.105;106 These data are consistent with the hypothesis that exposure to higher levels of E2 *in utero* may explain some of the differences in PCa risk among ethnic groups. Moreover, some indicators of high levels of estrogen during pregnancy, such as high birth weight and jaundice in the newborn, are associated with increased risk of PCa, whereas indicators of low estrogen levels, such as pre-eclampsia, are related to decreased risk.^{107;108} The male offspring of women who took diethylstilbestrol (DES) during pregnancy may have a higher risk of PCa.109 These data, taken together, indicate that PCa may be considered a fetus-based disease. In addition, the window of susceptibility for early-life reprogramming may extend to perinatal and peripubertal periods

and exposures to environmental estrogens may have the same impact as natural estrogens. Of relevance to these hypotheses are studies reporting significant exposure of human fetuses to BPA, likely caused by maternal use of BPA-containing products.¹¹⁰⁻¹¹⁶ Perinatal exposure to BPA has also been documented.117-119 Paradoxically, concerns about developmental reprogramming of PCa risk by soy infant formulas containing high concentrations of phytoestrogens have seldom been raised.120-122 Yet, the results of numerous human studies examining the risks and benefits of adult consumption of soy and phytoestrogens for prevention of PCa have been inconclusive.123-127 In summary, if PCa risk can be reprogrammed in early life, cancer prevention strategies should be directed at the aforementioned early developmental stages.

Estrogen Receptors (ERα and ERβ) as Functionally Divergent Mediators of Estrogen Action in the Prostate

The actions of estrogens are now believed to be mediated primarily by two ER subtypes, ER α and ER β and their variant forms.^{28;32;128;129} Both of the ERs have six common domains (A-F). They share a highly homologous DNA-binding domain (97% amino acid homology) but have very dissimilar N- and C -termini.¹³⁰ In addition, their ligand-binding domains are 56% homologous. These structural differences are the basis of the reported significant functional differences between the two receptor subtypes. Furthermore, it has been reported that the binding of the two receptors to the same ligand can initiate recruitments of different co-regulators and trigger the utilization of different *cis*-regulatory elements, thus further increasing the functional diversity of these two receptors. $130-135$ Finally, the recent discovery of variant forms of $ER\alpha$ and $ER\beta$ (isoforms) caused by alternative splicing or mutations has added complexity to estrogen action because these variants clearly show distinct functional disparity and patterns of tissue-/cell type-specific distribution.129;132;136;137

Many reports have shown differential expression patterns of the two receptors in the epithelial and stromal compartments of the normal and malignant human prostate (Figure 1).^{33;137-144} In the normal human prostate, the wild-type ERβ (also known as ERβ1) is localized mainly in the basal epithelial compartment, where $ER\alpha$ is almost never found, while ER α , along with ER β , is expressed in the stroma.¹⁴¹ Both ER subtypes are absent in the luminal epithelial compartment.¹⁴¹ A gradual loss of ERβ1 expression is apparent during the progression to high-grade PCa in the primary site $115;139;140;145;146$ The progressive silencing of ERβ is accompanied by hypermethylation of a CpG island in the proximal promoter of the gene.¹⁴⁷ Paradoxically, ERβ, but not ER α , is expressed at high levels in PCa metastasized to the bone and regional lymph nodes,¹⁴¹ with a concomitant loss of methylation in the CpG island of $ER\beta$.¹⁴⁷ Fine mapping of the CpG island revealed that this reversible silencing and reactivation of ERβ expression is attributable to cytosine methylation of an AP2 site within the CpG island of the ERβ promoter.¹⁴⁸ Moreover, antiestrogen-based therapies involving DES or PC-SPES lowered the level of ERβ1 expression in bone metastases of the treated patients.¹⁴⁹

It is now established that ERβ1 exerts an antiproliferative effect on the prostate epithelia^{131;133} and inhibits epithelial-mesenchymal transition.¹⁵⁰ However, less is known about the biological roles of its spliced variants. We recently demonstrated that ERβ2 and ERβ5 promote metastasis and that their expression is associated with shorter metastasis-free survival in patients with PCa.¹³⁷ In support of a divergent functional role of various ER β isoforms, when a pan-ERβ antibody was used, the retention of ERβ expression in primary PCa was associated with increased mortality.¹³⁹ The co-expression of ERβ with endothelial nitric oxide synthase (eNOS) and hypoxia-inducible factor 2α (HIF-2 α) suggests that an estrogen-mediated NO-enriched environment may influence the aggressive phenotype of

PCa significantly.¹⁵¹ Thus, ER β appears to play differential roles in human prostate carcinogenesis through differential expression of the various spliced variants and possibly alternative promoter utilization.¹⁵² Differential binding of the receptor to different ligands and crosstalk with other transcriptional factor-signaling cascades may also introduce additional divergence in ERβ action.131;153;154

Studies in animal models have provided strong evidence of the antiproliferative role of ERβ in the prostate gland. ERβ knockout mice have been found to develop prostatic hyperplasia in old age, a phenomenon not seen in ER α knockout mice.¹⁵⁵ Jiang et al.¹⁵⁶ observed an age-dependent decline in ER β expression in the canine prostate. Chang and Prins¹⁵⁷ reported that neonatal exposure of rats to estrogen causes the downregulation of ERβ; the upregulation of ERα; and the development of hyperplastic, dysplastic, and neoplastic lesions in the adult ventral prostates. Consistent with the hypothesis of the antiproliferative role ERβ, Risbriger et al. demonstrated that the administration of ERβ agonists, but not of an ERα agonist, to aromatase knockout mice suppressed the prostate epithelial cell growth and promoted apoptosis¹⁵⁸ and that the action of the receptor is mediated via TNF- α signaling.¹³³

Apart from ERβ expression, ERα, which is expressed primarily in the stromal compartment of the prostate, may contribute to the pathogenesis of PCa. Higher levels of $ER\alpha$ were observed in the prostatic stroma of Hispanic and Asian men than in that of Caucasian and African-American men, who are at a higher risk for PCa.159 In a genotyping and allelic frequency analysis of six different polymorphic loci of ERα in a Japanese population, polymorphism in codon 10 was found to be associated with a higher PCa risk.¹⁶⁰ These findings are in stark contrast to results from studies of genetic polymorphisms of $ER\beta^{161-164}$ that reported no strong association of various polymorphic loci with PCa risk, with the exception of one promoter single-nucleotide polymorphism (SNP).^{165;166}

Finally, during fetal development of the human prostate, $ER\beta$ is the first to appear in the prostate (by the seventh week of gestation) and is the only ER subtype expressed in the epithelial and stromal cells during the early ductal morphogenesis.^{$34,167$} ERB expression begins by week 15 of gestation and is strongly associated with the squamous metaplasia in the distal periurethral ducts and utricle.¹⁶⁷ These findings suggested that ERβ, perhaps in concert with the AR, mediates the very early stage of fetal prostate development, followed by the action of ERα. Thus, selective ERβ-activating compounds may play an important role in prenatal estrogen imprinting of PCa risk.

Non-canonical Actions of Estrogen and a New Therapeutic Target (G Protein-Coupled Receptor 30)

Apart from acting as transcription factors, ERα and ERβ can trigger rapid non-genomic signal transduction at the cell membrane level through the activation of specific kinase activity or induction of a calcium influx.¹⁶⁸⁻¹⁷⁰ These membrane ER-mediated events can activate gene-transcription activities with or without synergy with the classical genomic actions of nuclear ERs ¹⁷¹ In this regard, membrane $ER\alpha$ has been shown to tether onto the epidermal growth factor receptor (EGFR) signaling pathway for the activation of the MAPK¹⁷² and phosphoinositide 3-kinase (PI3K) signaling.¹⁷³ In addition, ERβ found localized in the mitochondria has been demonstrated to act as a mitochondrial transcription factor that regulates mitochondrial gene expression.¹⁷⁴ ER β has been reported to move from the mitochondria to the nucleus during neoplastic transformation.¹⁷⁵ However, the functions of cell membrane or mitochondrial ERα and ERβ in the prostate and PCa remain unknown and need further elucidation.

Estrogen has recently been found to be able to exert its action via the G protein-coupled receptor 30 (GPR30).176 GPR30 has been localized in the cell membrane and endoplasmic recticulum of various estrogen-sensitive tissues and cell lines¹⁷⁶⁻¹⁷⁸ and to mediate the nongenomic action of E2 in breast, endometrial, and ovarian cancer cells through the activation of Erk1/2 and cAMP pathways.179;180 GPR30 is expressed in the immortalized normal prostate stroma cell line WPWY-1 but does not appear to mediate the growth response induced by high-dose E2.181 A recent study demonstrated positive staining of GPR30 in both plasma membrane and cytoplasm in the normal and cancerous prostate, and its predominant expression in the luminal epithelium.182 Activation of GPR30 by a nonestrogenic, synthetic, GPR30-specific ligand G-1 inhibited PCa cell growth *in vitro* by activating Erk1/2 and upregulating p21-mediated G2-M arrest and attenuated the growth of human PCa xenografts in nude mice.¹⁸³ These new findings suggest that GPR30 is a novel estrogen mediator in the prostate and that a specific GPR30 ligand such as G-1 may have therapeutic potential for PCa.

Estrogen-related receptors (ERRs) are orphan nuclear receptors that are highly homologous to ERs, especially in the DNA- and ligand-binding domains.184 However, they do not bind natural estrogens and may have yet-to-be discovered endogenous ligands. There are at least three EERs, namely ERR-α, -β,¹⁸⁵ and -γ.^{186;187} They regulate gene expression through transactivation via *cis*-elements such as the ERE, steroidogenic factor-1 response element (SFRE), and estrogen-related receptor response element (ERRE).¹⁸⁸⁻¹⁹² Similar to ERs, different ERRs have distinct differential expression patterns in normal and cancerous prostate tissues/cells.¹⁹³ Finally, although they do not bind natural estrogens, ERR β and ERRγ do bind DES, tamoxifen, 4-hydroxytamoxifen, flavones, and isoflavone phytoestrogens194-196 and therefore should be considered in devising estrogen- or antiestrogen-based therapies for PCa.

Prevention of and Therapy for PCa with Selective ER Modulators (SERMs), Aromatase Inhibitors, Phytoestrogens, and Other Estrogen-/Antiestrogenbased Treatments

The first effective drug therapy for PCa was found about 70 years ago. DES, a xenoestrogen, was applied for the treatment of metastatic PCa¹⁹⁷ but fell out of favor because of its cardiovascular toxicity and other serious adverse effects.33;198;199 Other ER-based treatments for PCa include antiestrogens, selective ER modulators (SERMs), and aromatase inhibitors. SERMs are estrogenic compounds that act as either ER agonists or antagonists according to the presence of different co-regulators in a cell-/tissue-specific manner. Consequently, the ERs can be either stimulated or inhibited in the cell/tissue.200 However, clinical trials have shown that different generations of SERMs, such as tamoxifen, toremifene, raloxifene, and fulvestrant, have limited efficacy as alternatives to DES.^{33;201-207} Their lack of efficacy could be due to their original design for targeting the transactivation of $ER\alpha$ on an ERE and/or blocking the traditional genomic action of estrogen. With our growing knowledge of the wide range of genomic and non-genomic actions of estrogen and the variable expression of $ER\beta$ and its isoforms in the prostate and PCa, as well as of the significance of the non-canonical ER, GPR30, the future design of estrogen/antiestrogen therapies and chemopreventive agents will have to take into account the complex nature of estrogen action in the development and progression of PCa.

In this regard, it is important to recognize that many specificities or off-target effects of estrogenic/antiestrogenic drugs are due to their broad spectrum of activities not restricted to their estrogenicity. Traditionally, estrogen-induced PCa regression is believed to be mediated by its action on the hypothalamic-pituitary axis, thereby inhibiting testosterone

synthesis.²⁰⁸ However, it is now known that many estrogens/antiestrogens/phytoestrogens/ SERMs including DES, 2-methoxy-E2, genistein, resveratrol, licochalcone, raloxifene, toremifene, fulvestrant, and estramustine, have antitumor effects independent of this pathway.209-225 The ability of these compounds to suppress PCa cell growth has been attributed to a broad range of actions, including direct cytotoxicity, $22\overline{3}$ interruption of cellcycle progression, $^{210;221;226}$ induction of apoptosis, $^{211;221;224;225}$ depolymerization of microtubules,²⁰⁹ inhibition of DNA synthesis,²¹³ inhibition of topoisomerase II,²¹⁷ blockade of tyrosine kinase, 2^{17} ;218 disruption of apoptotic regulators, 2^{22} and activation of deathdomain receptors.214;219 Some of these estrogenic/antiestrogenic compounds are also potent inhibitors of angiogenesis and metastasis, through their actions in upregulating expression of genes related to anti-angiogenesis or anti-metastasis, 2^{15} activating the cell adhesion– signaling molecule focal adhesion kinase, 216 and reduction in metastatic spreading via the lymphatic system.²²⁰

As a recent development in the treatment of PCa, the application of estrogen patches has grown in popularity.227-229 The advantages include reduction in cardiovascular toxicity and the maintenance of adequate hormonal levels for a convenient time period.²³⁰ as well as the additional advantage of allevating hot flashes and improving bone density after endocrine treatment of PCa. 231;232 Estrogen patches have recently been tested in a phase II clinical trial as a first-line hormonal therapy in patients with locally advanced or metastatic PCa. Preliminary results produced the levels of testosterone and a PSA response similar to those of castration.²²⁸

Aside from treating PCa, the potential of SERMs for preventing PCa has turned out to be a promising proposition. The rationale stems from several lines of investigation. SERMS were shown to decrease testosterone levels by suppressing the hypothalamic-pituitary axis.²³³ Furthermore, age-related increases in PCa prevalence parallel increases in serum estrogen levels, and the incidence of PCa is low in countries with diets rich in phytoestrogens.²³⁴ A multicentered, double-blind randomized study involving 514 men with high-grade PIN and no cancer reported a significant reduction in PCa incidence in patients treated daily with 20 mg of toremifene for 6 -12 months as compared with placebo.²³⁵ These promising results have prompted the initiation of a large phase III study to examine the potential for toremifene as a chemopreventive agent.

Phytoestrogens are another important class of estrogen-based chemopreventive agents. The most common phytoestrogens, soy isoflavones such as genistein, equol, and daidzein, which are abundant in soy beans and its products, have been found to have estrogenic or antiestrogenic activity.236-239 They have been some of the most popular agents in studies of the potential of PCa chemoprevention and therapy because of the low incidence of PCa, along with the high levels of phytochemicals, in the diets of Asian populations.²⁴⁰ Although there is an abundance of *in vitro* and *in vivo* experimental data about the anti-tumor properties of phytoestrogens, the value of their protective or therapeutic effects in cohort studies still remains controversial.²⁴¹⁻²⁴⁸ Nevertheless, a number of phase I clinical trials evaluating the use of phytoestrogen supplementation in patients with PCa have generally seen beneficial effects without any toxicity.212;249-252

Summary

Traditionally, androgens have been considered to be the major sex hormones regulating the normal and malignant growth of the prostate. However, recent epidemiologic findings and experimental data suggest that estrogens and their mimics can be responsible for the pathogenesis of PCa. The carcinogenicity of estrogens in the prostate during adulthood is believed to be mediated by the combined effects of the hormone-induced unscheduled cell

proliferation and epigenetic silencing of anti-tumor genes, along with the bioactivation of estrogens to genotoxic carcinogens. Thus, individuals or ethnic groups with polymorphisms in genes encoding ERs and/or estrogen-metabolizing enzymes can modify the risk for PCa caused by altered responsiveness to the hormone and exposure to its carcinogenic metabolites over a lifetime. The age-dependent hormonal shift from androgen to estrogen could also be an important contributing factor to increased estrogen bioavailability. Although PCa has a long latency and starts to develop in men around middle age, recent data strongly suggest that PCa risk could be determined even as early as during pre- and perinatal life stages by a process known as the "estrogen imprinting." Thus, primary PCa prevention should probably begin in early life. Among the various cellular mediators, ERβ appears to be a key determinant in the pathogenesis, progression, and metastasis of PCa. Therapeutic approaches targeting its activation/inactivation may have important ramifications in the prevention and treatment of PCa. Epigenetic mechanisms such as DNA methylation play important roles in regulating the expression of the two ER subtypes. A change in the methylation status of proximal promoters of these genes constitutes an "on/off" switch for reversible gene regulation. Moreover, the differential expression of different ER-spliced variants (isoforms) could explain some conflicting observations related to estrogen action in the initiation and progression of PCa. Apart from the canonical genomic action of $ER\alpha$ and ERβ, the therapeutic potential of ER and its variants that function in multiple non-genomic pathways, such as membrane $ER\alpha$, mitochondrial ER , ERRs, and GPR30, may further contribute to the pathogenesis of PCa. Finally, various estrogenic/antiestrogenic/SERM-like compounds have demonstrable efficacies in causing PCa regression through the utilization of pathways independent of the hypothalamic-pituitary axis. As the treatment of advanced PCa with transdermal estrogen has gained in popularity, several SERMS, including toremifene, have shown promise as chemopreventive or therapeutic agents in clinical trials. Although data from clinical trials are not conclusive, phytoestrogen supplements, including dietary soy, continue to be used by patients as complementary alternative medicine for PCa. With a greater understanding of the molecular mechanism underlying estrogen carcinogenicity in the prostate, the applicability of estrogen/antiestrogen-based prevention and treatment therapies, as first-line or adjuvant therapies, will gain ground in the clinic. Thus, it is timely in devising a new generation of estrogenic/antiestrogenic therapies with higher specificity against PCa and fewer off-target effects.

Acknowledgments

We thank Nancy K. Voynow for her professional editorial assistance and Dr. Xiang Zhang for helpful critiques of the manuscript. Research supported in part by NIH grants CA112532; ES006096; ES015584; ES018758; and ES018789.

Funding support: NIH grants: DK061084, CA112532, CA015776, ES006096, and ES015584 to Shuk-mei Ho; Department of Defense Prostate Cancer Program grant: PC030595 to Shuk-mei Ho and Department of Defense Breast Cancer Program grant: BC094017 to Ming-tsung Lee

Reference List

- 1. Bostwick DG, Burke HB, Djakiew D, Euling S, Ho SM, Landolph J, et al. Human prostate cancer risk factors. Cancer. 2004; 101(10 Suppl):2371–2490. [PubMed: 15495199]
- 2. Ho SM. Estrogens and anti-estrogens: key mediators of prostate carcinogenesis and new therapeutic candidates. J Cell Biochem. 2004; 91(3):491–503. [PubMed: 14755680]
- 3. Ho SM, Leung YK, Chung I. Estrogens and antiestrogens as etiological factors and therapeutics for prostate cancer. Ann N Y Acad Sci. 2006:1089177–193.
- 4. Abdelrahaman E, Raghavan S, Baker L, Weinrich M, Winters SJ. Racial difference in circulating sex hormone-binding globulin levels in prepubertal boys. Metabolism. 2005; 54(1):91–96. [PubMed: 15562386]

- 5. Hui SL, DiMeglio LA, Longcope C, Peacock M, McClintock R, Perkins AJ, et al. Difference in bone mass between black and white American children: attributable to body build, sex hormone levels, or bone turnover? J Clin Endocrinol Metab. 2003; 88(2):642–649. [PubMed: 12574194]
- 6. Richards RJ, Svec F, Bao W, Srinivasan SR, Berenson GS. Steroid hormones during puberty: racial (black-white) differences in androstenedione and estradiol--the Bogalusa Heart Study. J Clin Endocrinol Metab. 1992; 75(2):624–631. [PubMed: 1639961]
- 7. Rohrmann S, Nelson WG, Rifai N, Brown TR, Dobs A, Kanarek N, et al. Serum estrogen, but not testosterone, levels differ between black and white men in a nationally representative sample of Americans. J Clin Endocrinol Metab. 2007; 92(7):2519–2525. [PubMed: 17456570]
- 8. Ross R, Bernstein L, Judd H, Hanisch R, Pike M, Henderson B. Serum testosterone levels in healthy young black and white men. J Natl Cancer Inst. 1986; 76(1):45–48. [PubMed: 3455741]
- 9. de Jong FH, Oishi K, Hayes RB, Bogdanowicz JF, Raatgever JW, van der Maas PJ, et al. Peripheral hormone levels in controls and patients with prostatic cancer or benign prostatic hyperplasia: results from the Dutch-Japanese case-control study. Cancer Res. 1991; 51(13):3445–3450. [PubMed: 1711411]
- 10. Orwoll ES, Nielson CM, Labrie F, Barrett-Connor E, Cauley JA, Cummings SR, et al. Evidence for Geographical and Racial Variation in Serum Sex Steroid Levels in Older Men. J Clin Endocrinol Metab. 2010
- 11. Ellem SJ, Risbridger GP. Aromatase and regulating the estrogen:androgen ratio in the prostate gland. J Steroid Biochem Mol Biol. 2010; 118(4-5):246–251. [PubMed: 19896534]
- 12. Feldman HA, Longcope C, Derby CA, Johannes CB, Araujo AB, Coviello AD, et al. Age trends in the level of serum testosterone and other hormones in middle-aged men: longitudinal results from the Massachusetts male aging study. J Clin Endocrinol Metab. 2002; 87(2):589–598. [PubMed: 11836290]
- 13. Gray A, Berlin JA, McKinlay JB, Longcope C. An examination of research design effects on the association of testosterone and male aging: results of a meta-analysis. J Clin Epidemiol. 1991; 44(7):671–684. [PubMed: 1829756]
- 14. Gray A, Feldman HA, McKinlay JB, Longcope C. Age, disease, and changing sex hormone levels in middle-aged men: results of the Massachusetts Male Aging Study. J Clin Endocrinol Metab. 1991; 73(5):1016–1025. [PubMed: 1719016]
- 15. Griffiths K. Estrogens and prostatic disease. International Prostate Health Council Study Group. Prostate. 2000; 45(2):87–100. [PubMed: 11027407]
- 16. Roberts RO, Jacobson DJ, Rhodes T, Klee GG, Leiber MM, Jacobsen SJ. Serum sex hormones and measures of benign prostatic hyperplasia. Prostate. 2004; 61(2):124–131. [PubMed: 15305335]
- 17. Vermeulen A, Rubens R, Verdonck L. Testosterone secretion and metabolism in male senescence. J Clin Endocrinol Metab. 1972; 34(4):730–735. [PubMed: 5012774]
- 18. Gapstur SM, Gann PH, Kopp P, Colangelo L, Longcope C, Liu K. Serum androgen concentrations in young men: a longitudinal analysis of associations with age, obesity, and race. The CARDIA male hormone study. Cancer Epidemiol Biomarkers Prev. 2002; 11(10 Pt 1):1041–1047. [PubMed: 12376505]
- 19. Krieg JM, Vermeulen A. The decline of androgen levels in elderly men and its clinical and therapeutic implications. Endocr Rev. 2005; 26(6):833–876. [PubMed: 15901667]
- 20. Moretti C, Frajese GV, Guccione L, Wannenes F, De Martino MU, Fabbri A, et al. Androgens and body composition in the aging male. J Endocrinol Invest. 2005; 28(3 Suppl):56–64. [PubMed: 16042362]
- 21. Starka L, Pospisilova H, Hill M. Free testosterone and free dihydrotestosterone throughout the life span of men. J Steroid Biochem Mol Biol. 2009; 116(1-2):118–120. [PubMed: 19465126]
- 22. Krieg M, Nass R, Tunn S. Effect of aging on endogenous level of 5 alpha-dihydrotestosterone, testosterone, estradiol, and estrone in epithelium and stroma of normal and hyperplastic human prostate. J Clin Endocrinol Metab. 1993; 77(2):375–381. [PubMed: 7688377]
- 23. Farnsworth WE. Roles of estrogen and SHBG in prostate physiology. Prostate. 1996; 28(1):17–23. [PubMed: 8545277]

- 24. Ellem SJ, Schmitt JF, Pedersen JS, Frydenberg M, Risbridger GP. Local aromatase expression in human prostate is altered in malignancy. J Clin Endocrinol Metab. 2004; 89(5):2434–2441. [PubMed: 15126575]
- 25. Ellem SJ, Risbridger GP. Aromatase and prostate cancer. Minerva Endocrinol. 2006; 31(1):1–12. [PubMed: 16498360]
- 26. McPherson SJ, Wang H, Jones ME, Pedersen J, Iismaa TP, Wreford N, et al. Elevated androgens and prolactin in aromatase-deficient mice cause enlargement, but not malignancy, of the prostate gland. Endocrinology. 2001; 142(6):2458–2467. [PubMed: 11356695]
- 27. Bain J. Testosterone and the aging male: to treat or not to treat? Maturitas. 2010; 66(1):16–22. [PubMed: 20153946]
- 28. Prins GS, Korach KS. The role of estrogens and estrogen receptors in normal prostate growth and disease. Steroids. 2008; 73(3):233–244. [PubMed: 18093629]
- 29. Singh PB, Matanhelia SS, Martin FL. A potential paradox in prostate adenocarcinoma progression: oestrogen as the initiating driver. Eur J Cancer. 2008; 44(7):928–936. [PubMed: 18381236]
- 30. Bosland MC. Use of animal models in defining efficacy of chemoprevention agents against prostate cancer. Eur Urol. 1999; 35(5-6):459–463. [PubMed: 10325505]
- 31. Ho, SM.; Lane, K.; Lee, K. Neoplastic transformation of the prostate. In: Naz, Rajesh K., editor. Prostate: Basic and Clinical Aspects. CRC Press; New York: 1997. p. 74-114.
- 32. Ricke WA, Wang Y, Cunha GR. Steroid hormones and carcinogenesis of the prostate: the role of estrogens. Differentiation. 2007; 75(9):871–882. [PubMed: 17924963]
- 33. Taplin ME, Ho SM. Clinical review 134: The endocrinology of prostate cancer. J Clin Endocrinol Metab. 2001; 86(8):3467–3477. [PubMed: 11502765]
- 34. Adams JY, Leav I, Lau KM, Ho SM, Pflueger SM. Expression of estrogen receptor beta in the fetal, neonatal, and prepubertal human prostate. Prostate. 2002; 52(1):69–81. [PubMed: 11992621]
- 35. Ho SM, Leav I, Merk FB, Yu M, Kwan PW, Ziar J. Induction of atypical hyperplasia, apoptosis, and type II estrogen-binding sites in the ventral prostates of Noble rats treated with testosterone and pharmacologic doses of estradiol-17 beta. Lab Invest. 1995; 73(3):356–365. [PubMed: 7564268]
- 36. Leav I, Ho SM, Ofner P, Merk FB, Kwan PW, Damassa D. Biochemical alterations in sex hormone-induced hyperplasia and dysplasia of the dorsolateral prostates of Noble rats. J Natl Cancer Inst. 1988; 80(13):1045–1053. [PubMed: 2457709]
- 37. Leav I, Merk FB, Kwan PW, Ho SM. Androgen-supported estrogen-enhanced epithelial proliferation in the prostates of intact Noble rats. Prostate. 1989; 15(1):23–40. [PubMed: 2477830]
- 38. Yatkin E, Bernoulli J, Talvitie EM, Santti R. Inflammation and epithelial alterations in rat prostate: impact of the androgen to oestrogen ratio. Int J Androl. 2009; 32(4):399–410. [PubMed: 19515173]
- 39. Hsu A, Bruno RS, Lohr CV, Taylor AW, Dashwood RH, Bray TM, et al. Dietary soy and tea mitigate chronic inflammation and prostate cancer via NFkappaB pathway in the Noble rat model. J Nutr Biochem. 2010
- 40. Ricke WA, Ishii K, Ricke EA, Simko J, Wang Y, Hayward SW, et al. Steroid hormones stimulate human prostate cancer progression and metastasis. Int J Cancer. 2006; 118(9):2123–2131. [PubMed: 16331600]
- 41. Kaplan PJ, Leav I, Greenwood J, Kwan PW, Ho SM. Involvement of transforming growth factor alpha (TGFalpha) and epidermal growth factor receptor (EGFR) in sex hormone-induced prostatic dysplasia and the growth of an androgen-independent transplantable carcinoma of the prostate. Carcinogenesis. 1996; 17(12):2571–2579. [PubMed: 9006091]
- 42. Wong YC, Xie W, Tsao SW. Structural changes and alteration in expression of TGF-beta1 and its receptors in prostatic intraepithelial neoplasia (PIN) in the ventral prostate of noble rats. Prostate. 2000; 45(4):289–298. [PubMed: 11102953]
- 43. Wang YZ, Wong YC. Sex hormone-induced prostatic carcinogenesis in the noble rat: the role of insulin-like growth factor-I (IGF-I) and vascular endothelial growth factor (VEGF) in the development of prostate cancer. Prostate. 1998; 35(3):165–177. [PubMed: 9582085]

- 44. Lau KM, Leav I, Ho SM. Rat estrogen receptor-alpha and -beta, and progesterone receptor mRNA expression in various prostatic lobes and microdissected normal and dysplastic epithelial tissues of the Noble rats. Endocrinology. 1998; 139(1):424–427. [PubMed: 9421443]
- 45. Leav I, Merk FB, Lee KF, Loda M, Mandoki M, McNeal JE, et al. Prolactin receptor expression in the developing human prostate and in hyperplastic, dysplastic, and neoplastic lesions. Am J Pathol. 1999; 154(3):863–870. [PubMed: 10079264]
- 46. Leav I, Galluzzi CM, Ziar J, Stork PJ, Ho SM, Loda M. Mitogen-activated protein kinase and mitogen-activated kinase phosphatase-1 expression in the Noble rat model of sex hormoneinduced prostatic dysplasia and carcinoma. Lab Invest. 1996; 75(3):361–370. [PubMed: 8804359]
- 47. Ling MT, Wang X, Ouyang XS, Lee TK, Fan TY, Xu K, et al. Activation of MAPK signaling pathway is essential for Id-1 induced serum independent prostate cancer cell growth. Oncogene. 2002; 21(55):8498–8505. [PubMed: 12466969]
- 48. Ghatak S, Oliveria P, Kaplan P, Ho SM. Expression and regulation of metallothionein mRNA levels in the prostates of noble rats: lack of expression in the ventral prostate and regulation by sex hormones in the dorsolateral prostate. Prostate. 1996; 29(2):91-100. [PubMed: 8700805]
- 49. Ho SM, Leav I, Ghatak S, Merk F, Jagannathan VS, Mallery K. Lack of association between enhanced TRPM-2/clusterin expression and increased apoptotic activity in sex-hormone-induced prostatic dysplasia of the Noble rat. Am J Pathol. 1998; 153(1):131–139. [PubMed: 9665473]
- 50. Ouyang XS, Wang X, Lee DT, Tsao SW, Wong YC. Up-regulation of TRPM-2, MMP-7 and ID-1 during sex hormone-induced prostate carcinogenesis in the Noble rat. Carcinogenesis. 2001; 22(6): 965–973. [PubMed: 11375906]
- 51. Ghatak S, Ho SM. Age-related changes in the activities of antioxidant enzymes and lipid peroxidation status in ventral and dorsolateral prostate lobes of noble rats. Biochem Biophys Res Commun. 1996; 222(2):362–367. [PubMed: 8670210]
- 52. Ho SM, Roy D. Sex hormone-induced nuclear DNA damage and lipid peroxidation in the dorsolateral prostates of Noble rats. Cancer Lett. 1994; 84(2):155–162. [PubMed: 8076372]
- 53. Tam NN, Ghatak S, Ho SM. Sex hormone-induced alterations in the activities of antioxidant enzymes and lipid peroxidation status in the prostate of Noble rats. Prostate. 2003; 55(1):1–8. [PubMed: 12640655]
- 54. Tam NN, Szeto CY, Sartor MA, Medvedovic M, Ho SM. Gene expression profiling identifies lobe-specific and common disruptions of multiple gene networks in testosterone-supported, 17beta-estradiol- or diethylstilbestrol-induced prostate dysplasia in Noble rats. Neoplasia. 2008; 10(1):20–40. [PubMed: 18231636]
- 55. Thompson CJ, Tam NN, Joyce JM, Leav I, Ho SM. Gene expression profiling of testosterone and estradiol-17 beta-induced prostatic dysplasia in Noble rats and response to the antiestrogen ICI 182,780. Endocrinology. 2002; 143(6):2093–2105. [PubMed: 12021174]
- 56. Chan FL, Ho SM. Comparative study of glycoconjugates of the rat prostatic lobes by lectin histochemistry. Prostate. 1999; 38(1):1–16. [PubMed: 9973104]
- 57. Chan FL, Choi HL, Ho SM. Analysis of glycoconjugate patterns of normal and hormone-induced dysplastic Noble rat prostates, and an androgen-independent Noble rat prostate tumor, by lectin histochemistry and protein blotting. Prostate. 2001; 46(1):21–32. [PubMed: 11170128]
- 58. Li SC, Chen GF, Chan PS, Choi HL, Ho SM, Chan FL. Altered expression of extracellular matrix and proteinases in Noble rat prostate gland after long-term treatment with sex steroids. Prostate. 2001; 49(1):58–71. [PubMed: 11550211]
- 59. Bernoulli J, Yatkin E, Konkol Y, Talvitie EM, Santti R, Streng T. Prostatic inflammation and obstructive voiding in the adult Noble rat: impact of the testosterone to estradiol ratio in serum. Prostate. 2008; 68(12):1296–1306. [PubMed: 18500685]
- 60. Tam NN, Leav I, Ho SM. Sex hormones induce direct epithelial and inflammation-mediated oxidative/nitrosative stress that favors prostatic carcinogenesis in the noble rat. Am J Pathol. 2007; 171(4):1334–1341. [PubMed: 17717140]
- 61. Yatkin E, Bernoulli J, Lammintausta R, Santti R. Fispemifene [Z-2-{2-[4-(4-chloro-1,2 diphenylbut-1-enyl)-phenoxy]ethoxy}-ethanol], a novel selective estrogen receptor modulator, attenuates glandular inflammation in an animal model of chronic nonbacterial prostatitis. J Pharmacol Exp Ther. 2008; 327(1):58–67. [PubMed: 18583549]

- 62. Bernoulli J, Yatkin E, Laakso A, Anttinen M, Bosland M, Vega K, et al. Histopathological evidence for an association of inflammation with ductal pin-like lesions but not with ductal adenocarcinoma in the prostate of the noble rat. Prostate. 2008; 68(7):728–739. [PubMed: 18302197]
- 63. Cavalieri EL, Rogan EG. A unified mechanism in the initiation of cancer. Ann N Y Acad Sci. 2002:959341–354.
- 64. Cavalieri E, Frenkel K, Liehr JG, Rogan E, Roy D. Estrogens as endogenous genotoxic agents-- DNA adducts and mutations. J Natl Cancer Inst Monogr. 2000; (27):75–93. [PubMed: 10963621]
- 65. Yager JD, Liehr JG. Molecular mechanisms of estrogen carcinogenesis. Annu Rev Pharmacol Toxicol. 1996:36203–232.
- 66. Yager JD. Endogenous estrogens as carcinogens through metabolic activation. J Natl Cancer Inst Monogr. 2000; (27):67–73. [PubMed: 10963620]
- 67. Jefcoate CR, Liehr JG, Santen RJ, Sutter TR, Yager JD, Yue W, et al. Tissue-specific synthesis and oxidative metabolism of estrogens. J Natl Cancer Inst Monogr. 2000; (27):95–112. [PubMed: 10963622]
- 68. Lakhani NJ, Sarkar MA, Venitz J, Figg WD. 2-Methoxyestradiol, a promising anticancer agent. Pharmacotherapy. 2003; 23(2):165–172. [PubMed: 12587805]
- 69. Sato F, Fukuhara H, Basilion JP. Effects of hormone deprivation and 2-methoxyestradiol combination therapy on hormone-dependent prostate cancer in vivo. Neoplasia. 2005; 7(9):838– 846. [PubMed: 16229806]
- 70. Reiner T, de Las PA, Gomez LA, Perez-Stable C. Low dose combinations of 2-methoxyestradiol and docetaxel block prostate cancer cells in mitosis and increase apoptosis. Cancer Lett. 2009; 276(1):21–31. [PubMed: 19046802]
- 71. Ghosh R, Ganapathy M, Alworth WL, Chan DC, Kumar AP. Combination of 2-methoxyestradiol (2-ME2) and eugenol for apoptosis induction synergistically in androgen independent prostate cancer cells. J Steroid Biochem Mol Biol. 2009; 113(1-2):25–35. [PubMed: 19084597]
- 72. Nock NL, Cicek MS, Li L, Liu X, Rybicki BA, Moreira A, et al. Polymorphisms in estrogen bioactivation, detoxification and oxidative DNA base excision repair genes and prostate cancer risk. Carcinogenesis. 2006; 27(9):1842–1848. [PubMed: 16569655]
- 73. Acevedo C, Opazo JL, Huidobro C, Cabezas J, Iturrieta J, Quinones SL. Positive correlation between single or combined genotypes of CYP1A1 and GSTM1 in relation to prostate cancer in Chilean people. Prostate. 2003; 57(2):111–117. [PubMed: 12949934]
- 74. Caceres DD, Iturrieta J, Acevedo C, Huidobro C, Varela N, Quinones L. Relationship among metabolizing genes, smoking and alcohol used as modifier factors on prostate cancer risk: exploring some gene-gene and gene-environment interactions. Eur J Epidemiol. 2005; 20(1):79– 88. [PubMed: 15756908]
- 75. Quinones LA, Irarrazabal CE, Rojas CR, Orellana CE, Acevedo C, Huidobro C, et al. Joint effect among p53, CYP1A1, GSTM1 polymorphism combinations and smoking on prostate cancer risk: an exploratory genotype-environment interaction study. Asian J Androl. 2006; 8(3):349–355. [PubMed: 16625286]
- 76. Silig Y, Pinarbasi H, Gunes S, Ayan S, Bagci H, Cetinkaya O. Polymorphisms of CYP1A1, GSTM1, GSTT1, and prostate cancer risk in Turkish population. Cancer Invest. 2006; 24(1):41– 45. [PubMed: 16466991]
- 77. Sobti RC, Onsory K, Al-Badran AI, Kaur P, Watanabe M, Krishan A, et al. CYP17, SRD5A2, CYP1B1, and CYP2D6 gene polymorphisms with prostate cancer risk in North Indian population. DNA Cell Biol. 2006; 25(5):287–294. [PubMed: 16716118]
- 78. Suzuki M, Mamun MR, Hara K, Ozeki T, Yamada Y, Kadowaki T, et al. The Val158Met polymorphism of the catechol-O-methyltransferase gene is associated with the PSA-progressionfree survival in prostate cancer patients treated with estramustine phosphate. Eur Urol. 2005; 48(5):752–759. [PubMed: 16126332]
- 79. Tanaka Y, Sasaki M, Shiina H, Tokizane T, Deguchi M, Hirata H, et al. Catechol-Omethyltransferase gene polymorphisms in benign prostatic hyperplasia and sporadic prostate cancer. Cancer Epidemiol Biomarkers Prev. 2006; 15(2):238–244. [PubMed: 16492910]

Ho et al. Page 14

- 80. Tang YM, Green BL, Chen GF, Thompson PA, Lang NP, Shinde A, et al. Human CYP1B1 Leu432Val gene polymorphism: ethnic distribution in African-Americans, Caucasians and Chinese; oestradiol hydroxylase activity; and distribution in prostate cancer cases and controls. Pharmacogenetics. 2000; 10(9):761–766. [PubMed: 11221602]
- 81. Leung YK, Lau KM, Mobley J, Jiang Z, Ho SM. Overexpression of cytochrome P450 1A1 and its novel spliced variant in ovarian cancer cells: alternative subcellular enzyme compartmentation may contribute to carcinogenesis. Cancer Res. 2005; 65(9):3726–3734. [PubMed: 15867368]
- 82. Hiramatsu M, Maehara I, Ozaki M, Harada N, Orikasa S, Sasano H. Aromatase in hyperplasia and carcinoma of the human prostate. Prostate. 1997; 31(2):118–124. [PubMed: 9140125]
- 83. Ito K, Fukabori Y, Shibata Y, Suzuki K, Mieda M, Gotanda K, et al. Effects of a new steroidal aromatase inhibitor, TZA-2237, and/or chlormadinone acetate on hormone-induced and spontaneous canine benign prostatic hyperplasia. Eur J Endocrinol. 2000; 143(4):543–554. [PubMed: 11022202]
- 84. Radlmaier A, Eickenberg HU, Fletcher MS, Fourcade RO, Santos JM Reis, van Aubel OG, et al. Estrogen reduction by aromatase inhibition for benign prostatic hyperplasia: results of a doubleblind, placebo-controlled, randomized clinical trial using two doses of the aromatase-inhibitor atamestane. Atamestane Study Group. Prostate. 1996; 29(4):199–208. [PubMed: 8876703]
- 85. Suzuki K, Okazaki H, Ono Y, Kurokawa K, Suzuki T, Onuma E, et al. Effect of dual inhibition of 5-alpha-reductase and aromatase on spontaneously developed canine prostatic hypertrophy. Prostate. 1998; 37(2):70–76. [PubMed: 9759700]
- 86. Brodie A, Njar V, Macedo LF, Vasaitis TS, Sabnis G. The Coffey Lecture: steroidogenic enzyme inhibitors and hormone dependent cancer. Urol Oncol. 2009; 27(1):53–63. [PubMed: 19111799]
- 87. Djurendic E, Daljev J, Sakac M, Canadi J, Santa SJ, Andric S, et al. Synthesis of some epoxy and/ or N-oxy 17-picolyl and 17-picolinylidene-androst-5-ene derivatives and evaluation of their biological activity. Steroids. 2008; 73(1):129–138. [PubMed: 17963806]
- 88. Kruit WH, Stoter G, Klijn JG. Effect of combination therapy with aminoglutethimide and hydrocortisone on prostate-specific antigen response in metastatic prostate cancer refractory to standard endocrine therapy. Anticancer Drugs. 2004; 15(9):843–847. [PubMed: 15457124]
- 89. Sanderson T, Renaud M, Scholten D, Nijmeijer S, van den Berg M, Cowell S, et al. Effects of lactone derivatives on aromatase (CYP19) activity in H295R human adrenocortical and (anti)androgenicity in transfected LNCaP human prostate cancer cells. Eur J Pharmacol. 2008; 593(1-3):92–98. [PubMed: 18639541]
- 90. Suzuki K, Nakazato H, Matsui H, Koike H, Okugi H, Ohtake N, et al. Association of the genetic polymorphism of the CYP19 intron 4[TTTA]n repeat with familial prostate cancer risk in a Japanese population. Anticancer Res. 2003; 23(6D):4941–4946. [PubMed: 14981949]
- 91. Suzuki K, Nakazato H, Matsui H, Koike H, Okugi H, Kashiwagi B, et al. Genetic polymorphisms of estrogen receptor alpha, CYP19, catechol-O-methyltransferase are associated with familial prostate carcinoma risk in a Japanese population. Cancer. 2003; 98(7):1411–1416. [PubMed: 14508827]
- 92. Li X, Nokkala E, Yan W, Streng T, Saarinen N, Warri A, et al. Altered structure and function of reproductive organs in transgenic male mice overexpressing human aromatase. Endocrinology. 2001; 142(6):2435–2442. [PubMed: 11356692]
- 93. Rajfer J, Coffey DS. Sex steroid imprinting of the immature prostate. Long-term effects. Invest Urol. 1978; 16(3):186–190. [PubMed: 711410]
- 94. Arai Y, Suzuki Y, Nishizuka Y. Hyperplastic and metaplastic lesions in the reproductive tract of male rats induced by neonatal treatment with diethylstilbestrol. Virchows Arch A Pathol Anat Histol. 1977; 376(1):21–28. [PubMed: 145081]
- 95. Arai Y, Mori T, Suzuki Y, Bern HA. Long-term effects of perinatal exposure to sex steroids and diethylstilbestrol on the reproductive system of male mammals. Int Rev Cytol. 1983:84235–268.
- 96. Prins GS. Neonatal estrogen exposure induces lobe-specific alterations in adult rat prostate androgen receptor expression. Endocrinology. 1992; 130(6):3703–3714. [PubMed: 1597166]
- 97. Vorherr H, Messer RH, Vorherr UF, Jordan SW, Kornfeld M. Teratogenesis and carcinogenesis in rat offspring after transplacental and transmammary exposure to diethylstilbestrol. Biochem Pharmacol. 1979; 28(12):1865–1877. [PubMed: 454458]

- 98. McLachlan JA, Newbold RR, Bullock B. Reproductive tract lesions in male mice exposed prenatally to diethylstilbestrol. Science. 1975; 190(4218):991–992. [PubMed: 242076]
- 99. McLachlan JA. Prenatal exposure to diethylstilbestrol in mice: toxicological studies. J Toxicol Environ Health. 1977; 2(3):527–537. [PubMed: 846001]
- 100. Pylkkanen L, Santti R, Newbold R, McLachlan JA. Regional differences in the prostate of the neonatally estrogenized mouse. Prostate. 1991; 18(2):117–129. [PubMed: 2006118]
- 101. Prins GS, Sklarew RJ, Pertschuk LP. Image analysis of androgen receptor immunostaining in prostate cancer accurately predicts response to hormonal therapy. J Urol. 1998; 159(3):641–649. [PubMed: 9474117]
- 102. Prins GS, Birch L, Couse JF, Choi I, Katzenellenbogen B, Korach KS. Estrogen imprinting of the developing prostate gland is mediated through stromal estrogen receptor alpha: studies with alphaERKO and betaERKO mice. Cancer Res. 2001; 61(16):6089–6097. [PubMed: 11507058]
- 103. vom Saal FS, Timms BG, Montano MM, Palanza P, Thayer KA, Nagel SC, et al. Prostate enlargement in mice due to fetal exposure to low doses of estradiol or diethylstilbestrol and opposite effects at high doses. Proc Natl Acad Sci U S A. 1997; 94(5):2056–2061. [PubMed: 9050904]
- 104. Ho SM, Tang WY, Belmonte de FJ, Prins GS. Developmental exposure to estradiol and bisphenol A increases susceptibility to prostate carcinogenesis and epigenetically regulates phosphodiesterase type 4 variant 4. Cancer Res. 2006; 66(11):5624–5632. [PubMed: 16740699]
- 105. Henderson BE, Feigelson HS. Hormonal carcinogenesis. Carcinogenesis. 2000; 21(3):427–433. [PubMed: 10688862]
- 106. Potischman N, Troisi R, Thadhani R, Hoover RN, Dodd K, Davis WW, et al. Pregnancy hormone concentrations across ethnic groups: implications for later cancer risk. Cancer Epidemiol Biomarkers Prev. 2005; 14(6):1514–1520. [PubMed: 15941965]
- 107. Ekbom A, Hsieh CC, Lipworth L, Wolk A, Ponten J, Adami HO, et al. Perinatal characteristics in relation to incidence of and mortality from prostate cancer. BMJ. 1996; 313(7053):337–341. [PubMed: 8760741]
- 108. Ekbom A, Wuu J, Adami HO, Lu CM, Lagiou P, Trichopoulos D, et al. Duration of gestation and prostate cancer risk in offspring. Cancer Epidemiol Biomarkers Prev. 2000; 9(2):221–223. [PubMed: 10698486]
- 109. Schrager S, Potter BE. Diethylstilbestrol exposure. Am Fam Physician. 2004; 69(10):2395–2400. [PubMed: 15168959]
- 110. Chapin RE, Adams J, Boekelheide K, Gray LE Jr. Hayward SW, Lees PS, et al. NTP-CERHR expert panel report on the reproductive and developmental toxicity of bisphenol A. Birth Defects Res B Dev Reprod Toxicol. 2008; 83(3):157–395. [PubMed: 18613034]
- 111. Ikezuki Y, Tsutsumi O, Takai Y, Kamei Y, Taketani Y. Determination of bisphenol A concentrations in human biological fluids reveals significant early prenatal exposure. Hum Reprod. 2002; 17(11):2839–2841. [PubMed: 12407035]
- 112. Kuroda N, Kinoshita Y, Sun Y, Wada M, Kishikawa N, Nakashima K, et al. Measurement of bisphenol A levels in human blood serum and ascitic fluid by HPLC using a fluorescent labeling reagent. J Pharm Biomed Anal. 2003; 30(6):1743–1749. [PubMed: 12485715]
- 113. Ranjit N, Siefert K, Padmanabhan V. Bisphenol-A and disparities in birth outcomes: a review and directions for future research. J Perinatol. 2010; 30(1):2–9. [PubMed: 19587689]
- 114. Schonfelder G, Wittfoht W, Hopp H, Talsness CE, Paul M, Chahoud I. Parent bisphenol A accumulation in the human maternal-fetal-placental unit. Environ Health Perspect. 2002; 110(11):A703–A707. [PubMed: 12417499]
- 115. Tsurusaki T, Aoki D, Kanetake H, Inoue S, Muramatsu M, Hishikawa Y, et al. Zone-dependent expression of estrogen receptors alpha and beta in human benign prostatic hyperplasia. J Clin Endocrinol Metab. 2003; 88(3):1333–1340. [PubMed: 12629127]
- 116. Yamada H, Furuta I, Kato EH, Kataoka S, Usuki Y, Kobashi G, et al. Maternal serum and amniotic fluid bisphenol A concentrations in the early second trimester. Reprod Toxicol. 2002; 16(6):735–739. [PubMed: 12401500]

Ho et al. Page 16

- 117. Braniste V, Jouault A, Gaultier E, Polizzi A, Buisson-Brenac C, Leveque M, et al. Impact of oral bisphenol A at reference doses on intestinal barrier function and sex differences after perinatal exposure in rats. Proc Natl Acad Sci U S A. 2010; 107(1):448–453. [PubMed: 20018722]
- 118. Prins GS, Tang WY, Belmonte J, Ho SM. Developmental exposure to bisphenol A increases prostate cancer susceptibility in adult rats: epigenetic mode of action is implicated. Fertil Steril. 2008; 89(2 Suppl):e41. [PubMed: 18308059]
- 119. Rubin BS, Soto AM. Bisphenol A: Perinatal exposure and body weight. Mol Cell Endocrinol. 2009; 304(1-2):55–62. [PubMed: 19433248]
- 120. Badger TM, Gilchrist JM, Pivik RT, Andres A, Shankar K, Chen JR, et al. The health implications of soy infant formula. Am J Clin Nutr. 2009; 89(5):1668S–1672S. [PubMed: 19357221]
- 121. Joeckel RJ, Phillips SK. Overview of infant and pediatric formulas. Nutr Clin Pract. 2009; 24(3): 356–362. [PubMed: 19483065]
- 122. Tuohy PG. Soy infant formula and phytoestrogens. J Paediatr Child Health. 2003; 39(6):401–405. [PubMed: 12919490]
- 123. Hamilton-Reeves JM, Rebello SA, Thomas W, Kurzer MS, Slaton JW. Effects of soy protein isolate consumption on prostate cancer biomarkers in men with HGPIN, ASAP, and low-grade prostate cancer. Nutr Cancer. 2008; 60(1):7–13. [PubMed: 18444130]
- 124. Ide H, Tokiwa S, Sakamaki K, Nishio K, Isotani S, Muto S, et al. Combined inhibitory effects of soy isoflavones and curcumin on the production of prostate-specific antigen. Prostate. 2010; 70(10):1127–1133. [PubMed: 20503397]
- 125. Kwan W, Duncan G, Van PC, Liu M, Lim J. A phase II trial of a soy beverage for subjects without clinical disease with rising prostate-specific antigen after radical radiation for prostate cancer. Nutr Cancer. 2010; 62(2):198–207. [PubMed: 20099194]
- 126. Pendleton JM, Tan WW, Anai S, Chang M, Hou W, Shiverick KT, et al. Phase II trial of isoflavone in prostate-specific antigen recurrent prostate cancer after previous local therapy. BMC Cancer. 2008; 8132
- 127. Vaishampayan U, Hussain M, Banerjee M, Seren S, Sarkar FH, Fontana J, et al. Lycopene and soy isoflavones in the treatment of prostate cancer. Nutr Cancer. 2007; 59(1):1–7. [PubMed: 17927495]
- 128. Risbridger GP, Ellem SJ, McPherson SJ. Estrogen action on the prostate gland: a critical mix of endocrine and paracrine signaling. J Mol Endocrinol. 2007; 39(3):183–188. [PubMed: 17766643]
- 129. Warner M, Gustafsson JA. The role of estrogen receptor beta (ERbeta) in malignant diseases--a new potential target for antiproliferative drugs in prevention and treatment of cancer. Biochem Biophys Res Commun. 2010; 396(1):63–66. [PubMed: 20494112]
- 130. Katzenellenbogen BS, Sun J, Harrington WR, Kraichely DM, Ganessunker D, Katzenellenbogen JA. Structure-function relationships in estrogen receptors and the characterization of novel selective estrogen receptor modulators with unique pharmacological profiles. Ann N Y Acad Sci. 2001:9496–15.
- 131. Dondi D, Piccolella M, Biserni A, Della TS, Ramachandran B, Locatelli A, et al. Estrogen receptor beta and the progression of prostate cancer: role of 5alpha-androstane-3beta,17beta-diol. Endocr Relat Cancer. 2010; 17(3):731–742. [PubMed: 20562232]
- 132. Leung YK, Mak P, Hassan S, Ho SM. Estrogen receptor (ER)-beta isoforms: a key to understanding ER-beta signaling. Proc Natl Acad Sci U S A. 2006; 103(35):13162–13167. [PubMed: 16938840]
- 133. McPherson SJ, Hussain S, Balanathan P, Hedwards SL, Niranjan B, Grant M, et al. Estrogen receptor-beta activated apoptosis in benign hyperplasia and cancer of the prostate is androgen independent and TNFalpha mediated. Proc Natl Acad Sci U S A. 2010; 107(7):3123–3128. [PubMed: 20133657]
- 134. Osborne CK, Schiff R, Fuqua SA, Shou J. Estrogen receptor: current understanding of its activation and modulation. Clin Cancer Res. 2001; 7(12 Suppl):4338s–4342s. [PubMed: 11916222]
- 135. Tremblay GB, Giguere V. Coregulators of estrogen receptor action. Crit Rev Eukaryot Gene Expr. 2002; 12(1):1–22. [PubMed: 12433063]

- 136. Barone I, Brusco L, Fuqua SA. Estrogen receptor mutations and changes in downstream gene expression and signaling. Clin Cancer Res. 2010; 16(10):2702–2708. [PubMed: 20427689]
- 137. Leung YK, Lam HM, Wu S, Song D, Levin L, Cheng L, et al. Estrogen receptor beta2 and beta5 are associated with poor prognosis in prostate cancer, and promote cancer cell migration and invasion. Endocr Relat Cancer. 2010; 17(3):675–689. [PubMed: 20501637]
- 138. Fixemer T, Remberger K, Bonkhoff H. Differential expression of the estrogen receptor beta (ERbeta) in human prostate tissue, premalignant changes, and in primary, metastatic, and recurrent prostatic adenocarcinoma. Prostate. 2003; 54(2):79–87. [PubMed: 12497580]
- 139. Horvath LG, Henshall SM, Lee CS, Head DR, Quinn DI, Makela S, et al. Frequent loss of estrogen receptor-beta expression in prostate cancer. Cancer Res. 2001; 61(14):5331–5335. [PubMed: 11454669]
- 140. Latil A, Bieche I, Vidaud D, Lidereau R, Berthon P, Cussenot O, et al. Evaluation of androgen, estrogen (ER alpha and ER beta), and progesterone receptor expression in human prostate cancer by real-time quantitative reverse transcription-polymerase chain reaction assays. Cancer Res. 2001; 61(5):1919–1926. [PubMed: 11280747]
- 141. Leav I, Lau KM, Adams JY, McNeal JE, Taplin ME, Wang J, et al. Comparative studies of the estrogen receptors beta and alpha and the androgen receptor in normal human prostate glands, dysplasia, and in primary and metastatic carcinoma. Am J Pathol. 2001; 159(1):79–92. [PubMed: 11438457]
- 142. Royuela M, de Miguel MP, Bethencourt FR, Sanchez-Chapado M, Fraile B, Arenas MI, et al. Estrogen receptors alpha and beta in the normal, hyperplastic and carcinomatous human prostate. J Endocrinol. 2001; 168(3):447–454. [PubMed: 11241176]
- 143. Torlakovic E, Lilleby W, Torlakovic G, Fossa SD, Chibbar R. Prostate carcinoma expression of estrogen receptor-beta as detected by PPG5/10 antibody has positive association with primary Gleason grade and Gleason score. Hum Pathol. 2002; 33(6):646–651. [PubMed: 12152165]
- 144. Weihua Z, Warner M, Gustafsson JA. Estrogen receptor beta in the prostate. Mol Cell Endocrinol. 2002; 193(1-2):1–5. [PubMed: 12160995]
- 145. Pasquali D, Staibano S, Prezioso D, Franco R, Esposito D, Notaro A, et al. Estrogen receptor beta expression in human prostate tissue. Mol Cell Endocrinol. 2001; 178(1-2):47–50. [PubMed: 11403893]
- 146. Pasquali D, Rossi V, Esposito D, Abbondanza C, Puca GA, Bellastella A, et al. Loss of estrogen receptor beta expression in malignant human prostate cells in primary cultures and in prostate cancer tissues. J Clin Endocrinol Metab. 2001; 86(5):2051–2055. [PubMed: 11344205]
- 147. Zhu X, Leav I, Leung YK, Wu M, Liu Q, Gao Y, et al. Dynamic regulation of estrogen receptorbeta expression by DNA methylation during prostate cancer development and metastasis. Am J Pathol. 2004; 164(6):2003–2012. [PubMed: 15161636]
- 148. Zhang X, Leung YK, Ho SM. AP-2 regulates the transcription of estrogen receptor (ER)-beta by acting through a methylation hotspot of the 0N promoter in prostate cancer cells. Oncogene. 2007; 26(52):7346–7354. [PubMed: 17525739]
- 149. Lai JS, Brown LG, True LD, Hawley SJ, Etzioni RB, Higano CS, et al. Metastases of prostate cancer express estrogen receptor-beta. Urology. 2004; 64(4):814–820. [PubMed: 15491740]
- 150. Mak P, Leav I, Pursell B, Bae D, Yang X, Taglienti CA, et al. ERbeta impedes prostate cancer EMT by destabilizing HIF-1alpha and inhibiting VEGF-mediated snail nuclear localization: implications for Gleason grading. Cancer Cell. 2010; 17(4):319–332. [PubMed: 20385358]
- 151. Nanni S, Benvenuti V, Grasselli A, Priolo C, Aiello A, Mattiussi S, et al. Endothelial NOS, estrogen receptor beta, and HIFs cooperate in the activation of a prognostic transcriptional pattern in aggressive human prostate cancer. J Clin Invest. 2009; 119(5):1093–1108. [PubMed: 19363294]
- 152. Leung, YK.; Lee, MT.; Wang, J.; Ho, SM. Post-transcriptional regulation of estrogen receptor beta isoforms in prostate cancer. ENDO2010 92th Annual Meeting; 2010.
- 153. Lee, MT.; Leung, YK.; Chung, I.; Ho, SM. Regulation of ERbeta1 transcriptional activity by acetylation. ENDO 2010 92th Annual Meeting; 2010.
- 154. Mak P, Leung YK, Tang WY, Harwood C, Ho SM. Apigenin suppresses cancer cell growth through ERbeta. Neoplasia. 2006; 8(11):896–904. [PubMed: 17132221]

- 155. Krege JH, Hodgin JB, Couse JF, Enmark E, Warner M, Mahler JF, et al. Generation and reproductive phenotypes of mice lacking estrogen receptor beta. Proc Natl Acad Sci U S A. 1998; 95(26):15677–15682. [PubMed: 9861029]
- 156. Jiang J, Chang HL, Sugimoto Y, Lin YC. Effects of age on growth and ERbeta mRNA expression of canine prostatic cells. Anticancer Res. 2005; 25(6B):4081–4090. [PubMed: 16312047]
- 157. Chang WY, Prins GS. Estrogen receptor-beta: implications for the prostate gland. Prostate. 1999; 40(2):115–124. [PubMed: 10386472]
- 158. Risbridger GP, Bianco JJ, Ellem SJ, McPherson SJ. Oestrogens and prostate cancer. Endocr Relat Cancer. 2003; 10(2):187–191. [PubMed: 12790781]
- 159. Haqq C, Li R, Khodabakhsh D, Frolov A, Ginzinger D, Thompson T, et al. Ethnic and racial differences in prostate stromal estrogen receptor alpha. Prostate. 2005; 65(2):101–109. [PubMed: 15880569]
- 160. Tanaka Y, Sasaki M, Kaneuchi M, Shiina H, Igawa M, Dahiya R. Polymorphisms of estrogen receptor alpha in prostate cancer. Mol Carcinog. 2003; 37(4):202–208. [PubMed: 12891629]
- 161. Chae YK, Huang HY, Strickland P, Hoffman SC, Helzlsouer K. Genetic polymorphisms of estrogen receptors alpha and beta and the risk of developing prostate cancer. PLoS One. 2009; 4(8):e6523. [PubMed: 19654868]
- 162. Nicolaiew N, Cancel-Tassin G, Azzouzi AR, Grand BL, Mangin P, Cormier L, et al. Association between estrogen and androgen receptor genes and prostate cancer risk. Eur J Endocrinol. 2009; 160(1):101–106. [PubMed: 18952763]
- 163. McIntyre MH, Kantoff PW, Stampfer MJ, Mucci LA, Parslow D, Li H, et al. Prostate cancer risk and ESR1 TA, ESR2 CA repeat polymorphisms. Cancer Epidemiol Biomarkers Prev. 2007; 16(11):2233–2236. [PubMed: 18006911]
- 164. Chen YC, Kraft P, Bretsky P, Ketkar S, Hunter DJ, Albanes D, et al. Sequence variants of estrogen receptor beta and risk of prostate cancer in the National Cancer Institute Breast and Prostate Cancer Cohort Consortium. Cancer Epidemiol Biomarkers Prev. 2007; 16(10):1973– 1981. [PubMed: 17932344]
- 165. Hedelin M, Balter KA, Chang ET, Bellocco R, Klint A, Johansson JE, et al. Dietary intake of phytoestrogens, estrogen receptor-beta polymorphisms and the risk of prostate cancer. Prostate. 2006; 66(14):1512–1520. [PubMed: 16921512]
- 166. Thellenberg-Karlsson C, Lindstrom S, Malmer B, Wiklund F, Augustsson-Balter K, Adami HO, et al. Estrogen receptor beta polymorphism is associated with prostate cancer risk. Clin Cancer Res. 2006; 12(6):1936–1941. [PubMed: 16551880]
- 167. Shapiro E, Huang H, Masch RJ, McFadden DE, Wilson EL, Wu XR. Immunolocalization of estrogen receptor alpha and beta in human fetal prostate. J Urol. 2005; 174(5):2051–2053. [PubMed: 16217392]
- 168. Kampa M, Pelekanou V, Castanas E. Membrane-initiated steroid action in breast and prostate cancer. Steroids. 2008; 73(9-10):953–960. [PubMed: 18249430]
- 169. Losel R, Wehling M. Nongenomic actions of steroid hormones. Nat Rev Mol Cell Biol. 2003; 4(1):46–56. [PubMed: 12511868]
- 170. Zhalilo LI, Novominskaia IM. Tissue respiratory activity with a varying potassium ion content in the incubation medium. Fiziol Zh. 1976; 22(3):410–412. [PubMed: 1278483]
- 171. Hammes SR, Levin ER. Extranuclear steroid receptors: nature and actions. Endocr Rev. 2007; 28(7):726–741. [PubMed: 17916740]
- 172. Zhang Z, Maier B, Santen RJ, Song RX. Membrane association of estrogen receptor alpha mediates estrogen effect on MAPK activation. Biochem Biophys Res Commun. 2002; 294(5): 926–933. [PubMed: 12074565]
- 173. Song RX, Zhang Z, Santen RJ. Estrogen rapid action via protein complex formation involving ERalpha and Src. Trends Endocrinol Metab. 2005; 16(8):347–353. [PubMed: 16126407]
- 174. O'Lone R, Knorr K, Jaffe IZ, Schaffer ME, Martini PG, Karas RH, et al. Estrogen receptors alpha and beta mediate distinct pathways of vascular gene expression, including genes involved in mitochondrial electron transport and generation of reactive oxygen species. Mol Endocrinol. 2007; 21(6):1281–1296. [PubMed: 17374850]

- 175. Chen JQ, Russo PA, Cooke C, Russo IH, Russo J. ERbeta shifts from mitochondria to nucleus during estrogen-induced neoplastic transformation of human breast epithelial cells and is involved in estrogen-induced synthesis of mitochondrial respiratory chain proteins. Biochim Biophys Acta. 2007; 1773(12):1732–1746. [PubMed: 17604135]
- 176. Revankar CM, Cimino DF, Sklar LA, Arterburn JB, Prossnitz ER. A transmembrane intracellular estrogen receptor mediates rapid cell signaling. Science. 2005; 307(5715):1625–1630. [PubMed: 15705806]
- 177. Langer G, Bader B, Meoli L, Isensee J, Delbeck M, Noppinger PR, et al. A critical review of fundamental controversies in the field of GPR30 research. Steroids. 2010; 75(8-9):603–610. [PubMed: 20034504]
- 178. Thomas P, Dressing G, Pang Y, Berg H, Tubbs C, Benninghoff A, et al. Progestin, estrogen and androgen G-protein coupled receptors in fish gonads. Steroids. 2006; 71(4):310–316. [PubMed: 16289637]
- 179. Maggiolini M, Picard D. The unfolding stories of GPR30, a new membrane-bound estrogen receptor. J Endocrinol. 2010; 204(2):105–114. [PubMed: 19767412]
- 180. Park II, Zhang Q, Liu V, Kozlowski JM, Zhang J, Lee C. 17Beta-estradiol at low concentrations acts through distinct pathways in normal versus benign prostatic hyperplasia-derived prostate stromal cells. Endocrinology. 2009; 150(10):4594–4605. [PubMed: 19608654]
- 181. Zhang Z, Duan L, Du X, Ma H, Park I, Lee C, et al. The proliferative effect of estradiol on human prostate stromal cells is mediated through activation of ERK. Prostate. 2008; 68(5):508–516. [PubMed: 18213633]
- 182. Lam, HM.; Ouyang, B.; Ho, SM. Activation of the G-coupled Protein Receptor 30 (GPR30) by the non-estrogenic ligand G-1 inhibits prostate cancer cell growth and regulation of its expression via an epigenetic mechanism. ENDO 2009 91st Annual Meeting; 2010.
- 183. Chan QK, Lam HM, Ng CF, Lee AY, Chan ES, Ng HK, et al. Activation of GPR30 inhibits the growth of prostate cancer cells through sustained activation of Erk1/2, c-jun/c-fos-dependent upregulation of p21, and induction of G(2) cell-cycle arrest. Cell Death Differ. 2010; 17(9): 1511–1523. [PubMed: 20203690]
- 184. Cheung CP, Yu S, Wong KB, Chan LW, Lai FM, Wang X, et al. Expression and functional study of estrogen receptor-related receptors in human prostatic cells and tissues. J Clin Endocrinol Metab. 2005; 90(3):1830–1844. [PubMed: 15598686]
- 185. Giguere V, Yang N, Segui P, Evans RM. Identification of a new class of steroid hormone receptors. Nature. 1988; 331(6151):91–94. [PubMed: 3267207]
- 186. Chen F, Zhang Q, McDonald T, Davidoff MJ, Bailey W, Bai C, et al. Identification of two hERR2-related novel nuclear receptors utilizing bioinformatics and inverse PCR. Gene. 1999; 228(1-2):101–109. [PubMed: 10072763]
- 187. Heard DJ, Norby PL, Holloway J, Vissing H. Human ERRgamma, a third member of the estrogen receptor-related receptor (ERR) subfamily of orphan nuclear receptors: tissue-specific isoforms are expressed during development and in the adult. Mol Endocrinol. 2000; 14(3):382–392. [PubMed: 10707956]
- 188. Heard DJ, Norby PL, Holloway J, Vissing H. Human ERRgamma, a third member of the estrogen receptor-related receptor (ERR) subfamily of orphan nuclear receptors: tissue-specific isoforms are expressed during development and in the adult. Mol Endocrinol. 2000; 14(3):382–392. [PubMed: 10707956]
- 189. Hong H, Yang L, Stallcup MR. Hormone-independent transcriptional activation and coactivator binding by novel orphan nuclear receptor ERR3. J Biol Chem. 1999; 274(32):22618–22626. [PubMed: 10428842]
- 190. Vanacker JM, Pettersson K, Gustafsson JA, Laudet V. Transcriptional targets shared by estrogen receptor-related receptors (ERRs) and estrogen receptor (ER) alpha, but not by ERbeta. EMBO J. 1999; 18(15):4270–4279. [PubMed: 10428965]
- 191. Xie W, Hong H, Yang NN, Lin RJ, Simon CM, Stallcup MR, et al. Constitutive activation of transcription and binding of coactivator by estrogen-related receptors 1 and 2. Mol Endocrinol. 1999; 13(12):2151–2162. [PubMed: 10598588]

- 192. Zhang Z, Teng CT. Estrogen receptor-related receptor alpha 1 interacts with coactivator and constitutively activates the estrogen response elements of the human lactoferrin gene. J Biol Chem. 2000; 275(27):20837–20846. [PubMed: 10779508]
- 193. Cheung CP, Yu S, Wong KB, Chan LW, Lai FM, Wang X, et al. Expression and functional study of estrogen receptor-related receptors in human prostatic cells and tissues. J Clin Endocrinol Metab. 2005; 90(3):1830–1844. [PubMed: 15598686]
- 194. Suetsugi M, Su L, Karlsberg K, Yuan YC, Chen S. Flavone and isoflavone phytoestrogens are agonists of estrogen-related receptors. Mol Cancer Res. 2003; 1(13):981–991. [PubMed: 14638870]
- 195. Coward P, Lee D, Hull MV, Lehmann JM. 4-Hydroxytamoxifen binds to and deactivates the estrogen-related receptor gamma. Proc Natl Acad Sci U S A. 2001; 98(15):8880–8884. [PubMed: 11447273]
- 196. Tremblay GB, Bergeron D, Giguere V. 4-Hydroxytamoxifen is an isoform-specific inhibitor of orphan estrogen-receptor-related (ERR) nuclear receptors beta and gamma. Endocrinology. 2001; 142(10):4572–4575. [PubMed: 11564725]
- 197. Huggins C, Hodges CV. Studies on prostatic cancer. I. The effect of castration, of estrogen and androgen injection on serum phosphatases in metastatic carcinoma of the prostate. CA Cancer J Clin. 1972; 22(4):232–240. [PubMed: 4625049]
- 198. Cox RL, Crawford ED. Estrogens in the treatment of prostate cancer. J Urol. 1995; 154(6):1991– 1998. [PubMed: 7500443]
- 199. Denis LJ, Griffiths K. Endocrine treatment in prostate cancer. Semin Surg Oncol. 2000; 18(1):52– 74. [PubMed: 10617897]
- 200. Shiau AK, Barstad D, Loria PM, Cheng L, Kushner PJ, Agard DA, et al. The structural basis of estrogen receptor/coactivator recognition and the antagonism of this interaction by tamoxifen. Cell. 1998; 95(7):927–937. [PubMed: 9875847]
- 201. Bergan RC, Reed E, Myers CE, Headlee D, Brawley O, Cho HK, et al. A Phase II study of highdose tamoxifen in patients with hormone-refractory prostate cancer. Clin Cancer Res. 1999; 5(9): 2366–2373. [PubMed: 10499606]
- 202. Chadha MK, Ashraf U, Lawrence D, Tian L, Levine E, Silliman C, et al. Phase II study of fulvestrant (Faslodex) in castration resistant prostate cancer. Prostate. 2008; 68(13):1461–1466. [PubMed: 18618738]
- 203. Hamilton M, Dahut W, Brawley O, Davis P, Wells-Jones T, Kohler D, et al. A phase I/II study of high-dose tamoxifen in combination with vinblastine in patients with androgen-independent prostate cancer. Acta Oncol. 2003; 42(3):195–201. [PubMed: 12852695]
- 204. Lissoni P, Vigano P, Vaghi M, Frontini L, Giuberti C, Manganini V, et al. A phase II study of tamoxifen in hormone-resistant metastatic prostate cancer: possible relation with prolactin secretion. Anticancer Res. 2005; 25(5):3597–3599. [PubMed: 16101186]
- 205. Shazer RL, Jain A, Galkin AV, Cinman N, Nguyen KN, Natale RB, et al. Raloxifene, an oestrogen-receptor-beta-targeted therapy, inhibits androgen-independent prostate cancer growth: results from preclinical studies and a pilot phase II clinical trial. BJU Int. 2006; 97(4):691–697. [PubMed: 16536755]
- 206. Smith MR, Morton RA, Barnette KG, Sieber PR, Malkowicz SB, Rodriguez D, et al. Toremifene to reduce fracture risk in men receiving androgen deprivation therapy for prostate cancer. J Urol. 2010; 184(4):1316–1321. [PubMed: 20723926]
- 207. Stein S, Zoltick B, Peacock T, Holroyde C, Haller D, Armstead B, et al. Phase II trial of toremifene in androgen-independent prostate cancer: a Penn cancer clinical trials group trial. Am J Clin Oncol. 2001; 24(3):283–285. [PubMed: 11404501]
- 208. el-Rayes BF, Hussain MH. Hormonal therapy for prostate cancer: past, present and future. Expert Rev Anticancer Ther. 2002; 2(1):37–47. [PubMed: 12113064]
- 209. Dahllof B, Billstrom A, Cabral F, Hartley-Asp B. Estramustine depolymerizes microtubules by binding to tubulin. Cancer Res. 1993; 53(19):4573–4581. [PubMed: 8402630]
- 210. Fu Y, Hsieh TC, Guo J, Kunicki J, Lee MY, Darzynkiewicz Z, et al. Licochalcone-A, a novel flavonoid isolated from licorice root (Glycyrrhiza glabra), causes G2 and late-G1 arrests in

androgen-independent PC-3 prostate cancer cells. Biochem Biophys Res Commun. 2004; 322(1): 263–270. [PubMed: 15313200]

- 211. Kim HT, Kim BC, Kim IY, Mamura M, Seong DH, Jang JJ, et al. Raloxifene, a mixed estrogen agonist/antagonist, induces apoptosis through cleavage of BAD in TSU-PR1 human cancer cells. J Biol Chem. 2002; 277(36):32510–32515. [PubMed: 12084714]
- 212. Kumar NB, Cantor A, Allen K, Riccardi D, Besterman-Dahan K, Seigne J, et al. The specific role of isoflavones in reducing prostate cancer risk. Prostate. 2004; 59(2):141–147. [PubMed: 15042614]
- 213. Kuwajerwala N, Cifuentes E, Gautam S, Menon M, Barrack ER, Reddy GP. Resveratrol induces prostate cancer cell entry into s phase and inhibits DNA synthesis. Cancer Res. 2002; 62(9): 2488–2492. [PubMed: 11980638]
- 214. LaVallee TM, Zhan XH, Johnson MS, Herbstritt CJ, Swartz G, Williams MS, et al. 2 methoxyestradiol up-regulates death receptor 5 and induces apoptosis through activation of the extrinsic pathway. Cancer Res. 2003; 63(2):468–475. [PubMed: 12543804]
- 215. Li Y, Sarkar FH. Gene expression profiles of genistein-treated PC3 prostate cancer cells. J Nutr. 2002; 132(12):3623–3631. [PubMed: 12468598]
- 216. Liu Y, Kyle E, Lieberman R, Crowell J, Kellof G, Bergan RC. Focal adhesion kinase (FAK) phosphorylation is not required for genistein-induced FAK-beta-1-integrin complex formation. Clin Exp Metastasis. 2000; 18(3):203–212. [PubMed: 11315093]
- 217. Matsukawa Y, Marui N, Sakai T, Satomi Y, Yoshida M, Matsumoto K, et al. Genistein arrests cell cycle progression at G2-M. Cancer Res. 1993; 53(6):1328–1331. [PubMed: 8443813]
- 218. Misra RR, Hursting SD, Perkins SN, Sathyamoorthy N, Mirsalis JC, Riccio ES, et al. Genotoxicity and carcinogenicity studies of soy isoflavones. Int J Toxicol. 2002; 21(4):277–285. [PubMed: 12171629]
- 219. Mor G, Kohen F, Garcia-Velasco J, Nilsen J, Brown W, Song J, et al. Regulation of fas ligand expression in breast cancer cells by estrogen: functional differences between estradiol and tamoxifen. J Steroid Biochem Mol Biol. 2000; 73(5):185–194. [PubMed: 11070347]
- 220. Neubauer BL, Best KL, Counts DF, Goode RL, Hoover DM, Jones CD, et al. Raloxifene (LY156758) produces antimetastatic responses and extends survival in the PAIII rat prostatic adenocarcinoma model. Prostate. 1995; 27(4):220–229. [PubMed: 7479389]
- 221. Qadan LR, Perez-Stable CM, Anderson C, D'Ippolito G, Herron A, Howard GA, et al. 2- Methoxyestradiol induces G2/M arrest and apoptosis in prostate cancer. Biochem Biophys Res Commun. 2001; 285(5):1259–1266. [PubMed: 11478793]
- 222. Rafi MM, Rosen RT, Vassil A, Ho CT, Zhang H, Ghai G, et al. Modulation of bcl-2 and cytotoxicity by licochalcone-A, a novel estrogenic flavonoid. Anticancer Res. 2000; 20(4):2653– 2658. [PubMed: 10953339]
- 223. Schulz P, Bauer HW, Brade WP, Keller A, Fittler F. Evaluation of the cytotoxic activity of diethylstilbestrol and its mono- and diphosphate towards prostatic carcinoma cells. Cancer Res. 1988; 48(10):2867–2870. [PubMed: 2834049]
- 224. Shimada K, Nakamura M, Ishida E, Kishi M, Konishi N. Requirement of c-jun for testosteroneinduced sensitization to N-(4-hydroxyphenyl)retinamide-induced apoptosis. Mol Carcinog. 2003; 36(3):115–122. [PubMed: 12619033]
- 225. Yo YT, Shieh GS, Hsu KF, Wu CL, Shiau AL. Licorice and licochalcone-A induce autophagy in LNCaP prostate cancer cells by suppression of Bcl-2 expression and the mTOR pathway. J Agric Food Chem. 2009; 57(18):8266–8273. [PubMed: 19711916]
- 226. Kumar AP, Garcia GE, Slaga TJ. 2-methoxyestradiol blocks cell-cycle progression at G(2)/M phase and inhibits growth of human prostate cancer cells. Mol Carcinog. 2001; 31(3):111–124. [PubMed: 11479920]
- 227. Bland LB, Garzotto M, DeLoughery TG, Ryan CW, Schuff KG, Wersinger EM, et al. Phase II study of transdermal estradiol in androgen-independent prostate carcinoma. Cancer. 2005; 103(4):717–723. [PubMed: 15641029]
- 228. Langley RE, Godsland IF, Kynaston H, Clarke NW, Rosen SD, Morgan RC, et al. Early hormonal data from a multicentre phase II trial using transdermal oestrogen patches as first-line

hormonal therapy in patients with locally advanced or metastatic prostate cancer. BJU Int. 2008; 102(4):442–445. [PubMed: 18422771]

- 229. Ockrim JL, Lalani EN, Laniado ME, Carter SS, Abel PD. Transdermal estradiol therapy for advanced prostate cancer--forward to the past? J Urol. 2003; 169(5):1735–1737. [PubMed: 12686820]
- 230. Ockrim JL, Lalani e, Kakkar AK, Abel PD. Transdermal estradiol therapy for prostate cancer reduces thrombophilic activation and protects against thromboembolism. J Urol. 2005; 174(2): 527–533. [PubMed: 16006886]
- 231. Gerber GS, Zagaja GP, Ray PS, Rukstalis DB. Transdermal estrogen in the treatment of hot flushes in men with prostate cancer. Urology. 2000; 55(1):97–101. [PubMed: 10654902]
- 232. Ockrim JL, Lalani EN, Banks LM, Svensson WE, Blomley MJ, Patel S, et al. Transdermal estradiol improves bone density when used as single agent therapy for prostate cancer. J Urol. 2004; 172(6 Pt 1):2203–2207. [PubMed: 15538232]
- 233. Taneja SS, Smith MR, Dalton JT, Raghow S, Barnette G, Steiner M, et al. Toremifene--a promising therapy for the prevention of prostate cancer and complications of androgen deprivation therapy. Expert Opin Investig Drugs. 2006; 15(3):293–305.
- 234. Denis L, Morton MS, Griffiths K. Diet and its preventive role in prostatic disease. Eur Urol. 1999; 35(5-6):377–387. [PubMed: 10325492]
- 235. Price D, Stein B, Sieber P, Tutrone R, Bailen J, Goluboff E, et al. Toremifene for the prevention of prostate cancer in men with high grade prostatic intraepithelial neoplasia: results of a doubleblind, placebo controlled, phase IIB clinical trial. J Urol. 2006; 176(3):965–970. [PubMed: 16890670]
- 236. Jian L. Soy, isoflavones, and prostate cancer. Mol Nutr Food Res. 2009; 53(2):217–226. [PubMed: 18985655]
- 237. Lampe JW. Emerging research on equol and cancer. J Nutr. 2010; 140(7):1369S–1372S. [PubMed: 20505018]
- 238. Markiewicz L, Garey J, Adlercreutz H, Gurpide E. In vitro bioassays of non-steroidal phytoestrogens. J Steroid Biochem Mol Biol. 1993; 45(5):399–405. [PubMed: 8499347]
- 239. Milligan SR, Balasubramanian AV, Kalita JC. Relative potency of xenobiotic estrogens in an acute in vivo mammalian assay. Environ Health Perspect. 1998; 106(1):23–26. [PubMed: 9417770]
- 240. Sonn GA, Aronson W, Litwin MS. Impact of diet on prostate cancer: a review. Prostate Cancer Prostatic Dis. 2005; 8(4):304–310. [PubMed: 16130015]
- 241. Ganry O. Phytoestrogens and prostate cancer risk. Prev Med. 2005; 41(1):1–6. [PubMed: 15916986]
- 242. Hebert JR, Hurley TG, Olendzki BC, Teas J, Ma Y, Hampl JS. Nutritional and socioeconomic factors in relation to prostate cancer mortality: a cross-national study. J Natl Cancer Inst. 1998; 90(21):1637–1647. [PubMed: 9811313]
- 243. Hussain M, Banerjee M, Sarkar FH, Djuric Z, Pollak MN, Doerge D, et al. Soy isoflavones in the treatment of prostate cancer. Nutr Cancer. 2003; 47(2):111–117. [PubMed: 15087261]
- 244. Lee MM, Gomez SL, Chang JS, Wey M, Wang RT, Hsing AW. Soy and isoflavone consumption in relation to prostate cancer risk in China. Cancer Epidemiol Biomarkers Prev. 2003; 12(7):665– 668. [PubMed: 12869409]
- 245. Ozasa K, Nakao M, Watanabe Y, Hayashi K, Miki T, Mikami K, et al. Serum phytoestrogens and prostate cancer risk in a nested case-control study among Japanese men. Cancer Sci. 2004; 95(1): 65–71. [PubMed: 14720329]
- 246. Stattin P, Adlercreutz H, Tenkanen L, Jellum E, Lumme S, Hallmans G, et al. Circulating enterolactone and prostate cancer risk: a Nordic nested case-control study. Int J Cancer. 2002; 99(1):124–129. [PubMed: 11948503]
- 247. Strom SS, Yamamura Y, Duphorne CM, Spitz MR, Babaian RJ, Pillow PC, et al. Phytoestrogen intake and prostate cancer: a case-control study using a new database. Nutr Cancer. 1999; 33(1): 20–25. [PubMed: 10227039]

Ho et al. Page 23

- 248. Travis RC, Spencer EA, Allen NE, Appleby PN, Roddam AW, Overvad K, et al. Plasma phytooestrogens and prostate cancer in the European Prospective Investigation into Cancer and Nutrition. Br J Cancer. 2009; 100(11):1817–1823. [PubMed: 19436304]
- 249. Dalais FS, Meliala A, Wattanapenpaiboon N, Frydenberg M, Suter DA, Thomson WK, et al. Effects of a diet rich in phytoestrogens on prostate-specific antigen and sex hormones in men diagnosed with prostate cancer. Urology. 2004; 64(3):510–515. [PubMed: 15351581]
- 250. Jarred RA, Keikha M, Dowling C, McPherson SJ, Clare AM, Husband AJ, et al. Induction of apoptosis in low to moderate-grade human prostate carcinoma by red clover-derived dietary isoflavones. Cancer Epidemiol Biomarkers Prev. 2002; 11(12):1689–1696. [PubMed: 12496063]
- 251. Miltyk W, Craciunescu CN, Fischer L, Jeffcoat RA, Koch MA, Lopaczynski W, et al. Lack of significant genotoxicity of purified soy isoflavones (genistein, daidzein, and glycitein) in 20 patients with prostate cancer. Am J Clin Nutr. 2003; 77(4):875–882. [PubMed: 12663286]
- 252. Takimoto CH, Glover K, Huang X, Hayes SA, Gallot L, Quinn M, et al. Phase I pharmacokinetic and pharmacodynamic analysis of unconjugated soy isoflavones administered to individuals with cancer. Cancer Epidemiol Biomarkers Prev. 2003; 12(11 Pt 1):1213–1221. [PubMed: 14652284]

Figure. 1.

Expression of various hormone receptors and levels of hormones during the development and progression of prostate cancer (PCa). In the normal human prostate (far left panel), androgen receptor (AR), estrogen receptor alpha (ERα), different estrogen receptor beta (ERβ) isoforms, and G protein–coupled receptor 30 (GPR30) are expressed differentially in the stroma, and luminal and basal epithelium of the prostate. The epithelial cell compartment expresses only $ER\beta$ and AR, whereas the stromal compartment expresses both $ER\alpha$ and AR. GPR30 is expressed in both compartments. During tumor progression from high grade PIN (second to the far left panel) to prostate cancer (second to the far right panel), expression of AR and ER α remains unchanged, while expression of the antiproliferative ER β 1 is lost in the basal epithelial cells along with the disruption of the basement membrane (pink line). Two ERβ isoforms, ERβ2 and ERβ5, which have been shown to have pro-metastatic potential, are expressed in the normal epithelium, but their co-expression in PCa specimens is associated with a poor prognosis and shorter metastasis-free survival. In prostate metastases (far right panel), ERβ1 is re-expressed at high levels. Other non-canonical ERs such as GPR30 and ERRs are differentially expressed in normal and malignant prostate tissues but their roles in prostate carcinogenesis remains to be elucidated. Throughout the development and progression of PCa, tissue estrogen levels [estradiol-17β (E2) and estrone (E1)] increase with concomitant decreases in levels of 5α-dihydrotestosterone (DHT). These changes are due in part to an increase in CYP19 (aromatase) activity and a loss of SRD5A (5α-reductase) expression in the primary site (insert). SRD5A expression is completely lost in metastases. Additionally, an age-related reduction in testicular testosterone (T) synthesis, an increase in levels of sex hormone–binding globulin, and increases in adiposity and peripheral aromatase activity, collectively contribute to an increase in the estrogen to androgen ratio in circulation that further raise prostatic estrogen levels. In summary, a lifetime over-exposure to estrogens and/or catecholestrogens could be an etiological factor in prostate carcinogenesis. The localizations of these enzymes as well as mediators of estrogen/ androgen actions are indicated.