

Neuronal patterning by BMPs: a requirement for GDF7 in the generation of a discrete class of commissural interneurons in the mouse spinal cord

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Inductive factors are known to direct the regional differentiation of the vertebrate central nervous system (CNS) but their role in the specification of individual neuronal cell types is less clear. We have examined the function of GDF7, a BMP family member expressed selectively by roof plate cells, in the generation of neuronal cell types in the dorsal spinal cord. We find that GDF7 can promote the differentiation in vitro of two dorsal sensory interneuron classes, D1A and D1B neurons. In *Gdf7*-null mutant embryos, the generation of D1A neurons is eliminated but D1B neurons and other identified dorsal interneurons are unaffected. These findings show that GDF7 is an inductive signal from the roof plate required for the specification of neuronal identity in the dorsal spinal cord and that GDF7 and other BMP family members expressed by the roof plate have non-redundant functions in vivo. More generally, these results suggest that BMP signaling may have a prominent role in the assignment of neuronal identity within the mammalian CNS.

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The patterning of the vertebrate central nervous system (CNS) depends on the subdivision of the neural epithelium into distinct regional domains and the generation of a diverse array of neuronal cell types within each domain. The early regionalization of the neural epithelium appears to be regulated by inductive factors secreted by signaling centers in and adjacent to the neural tube (Lumsden and Krumlauf 1996; Tanabe and Jessell 1996). Genetic studies and in vitro assays have provided evidence that signaling factors of the Wnt, FGF, and Hedgehog families have prominent roles in the specification of these regional domains (McMahon and Bradley 1990; Thomas and Capecchi 1990; Chiang et al. 1996; Crossley et al. 1996; Ikeya et al. 1997; Lee et al. 1997; Meyers et al. 1998; Ye et al. 1998).

The factors responsible for the specification of individual neuronal subtypes within local regions of the developing CNS, however, remain less well defined. Sonic hedgehog (Shh) has been implicated in neuronal cell-type specification on the basis of in vitro studies in which the generation of distinct classes of ventral neuronal cell

types is elicited by exposure of neural progenitor cells to different Shh concentrations (Roelink et al. 1995; Marti et al. 1995; Ericson et al. 1996, 1997). Nevertheless, the elimination of Shh function in mice results in the loss of the entire ventral half of the embryonic CNS (Chiang et al. 1996), precluding a genetic assessment of the requirement for Shh signaling in the generation of individual neuronal cell types.

The TGF β superfamily (Kingsley 1994) constitutes the largest known class of inductive signals and their capacity for heterodimer formation extends further the potential diversity of TGF β -related signals. Evidence for a role of TGF β -related proteins in the control of neuronal differentiation in the CNS has emerged in part from studies of cell patterning in the dorsal spinal cord (Basler et al. 1993; Liem et al. 1995, 1997). Members of the BMP subclass of TGF β -related proteins are expressed by epidermal ectoderm cells that flank the neural plate (Liem et al. 1995; Lyons et al. 1995; Arkell and Beddington 1997; Dudley and Robertson 1997), and the induction of neural crest and roof plate differentiation by the epidermal ectoderm (Moury and Jacobson 1989; Dickinson et al. 1995; Liem et al. 1995) appears to be mediated by BMPs (Liem et al. 1995, 1997). Roof plate cells at the dorsal midline of the neural tube subsequently express several

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BMPs (Jones et al. 1991; Liem et al. 1995, 1997; Lyons et al. 1995; Arkell and Beddington 1997; Dudley and Robertson 1997) and in vitro assays have shown that BMPs expressed by roof plate cells can promote the differentiation of dorsal spinal cord interneurons (Liem et al. 1997). These findings have led to the idea that neuronal patterning in the dorsal spinal cord depends on a cascade of TGF β -related inductive signaling that is initiated by the epidermal ectoderm and propagated by the roof plate.

Gene targeting studies in mice have demonstrated a number of essential functions for BMPs in mammalian development, notably in the differentiation of mesodermal tissues (King et al. 1994; Storm et al. 1994; Dudley et al. 1995; Luo et al. 1995; Winnier et al. 1995; Zhang and Bradley 1996). These studies, however, have not been informative in defining the contribution of BMPs to neural tube patterning. Two major issues concerning the role of BMPs as dorsalizing signals in the CNS remain unresolved. Are the BMPs expressed by the roof plate required in vivo for the generation of dorsal neuronal cell types? And, if so, do roof plate-derived BMPs have distinct or overlapping functions in the generation of dorsal neuronal cell types?

We have taken a genetic approach to define the requirement for individual BMPs in the control of neuronal identity in the dorsal spinal cord. This study focuses on GDF7, a BMP family member that we have found to be expressed selectively by roof plate cells in the embryonic CNS. GDF7 can mimic the ability of the roof plate

to induce the differentiation in vitro of two classes of dorsal commissural interneurons, D1A and D1B neurons. To define the requirement for GDF7 signaling we generated a targeted mutation in the mouse *Gdf7* gene. *Gdf7* mutant embryos lack D1A neurons, but D1B neurons and other types of dorsal interneurons differentiate normally. Our results establish that GDF7 activity is required for the specification of neuronal identity in the spinal cord. They also show that roof plate signaling has an essential role in dorsal neural patterning, distinct from that provided by epidermal ectoderm signals.

Results

Selective expression of *Gdf7* by roof plate cells

To identify inductive factors that control the differentiation of dorsal interneurons in the mouse spinal cord, we analyzed the neural expression of BMP family members between E8.5 and E12.5, the period of generation of most dorsal interneurons (Nornes and Das 1974; Altman and Bayer 1984). Of nine BMP genes analyzed, only one, *Gdf7* (Storm et al. 1994), was found to be expressed selectively by roof plate cells at the dorsal midline of the neural tube. *Gdf7* expression in roof plate cells was first evident at E9 at hindbrain and rostral spinal cord levels (Fig. 1A) and was subsequently detected at all axial levels (Fig. 1B; data not shown) from the diencephalon to the sacral spinal cord. *Gdf7* expression persisted throughout the neu-

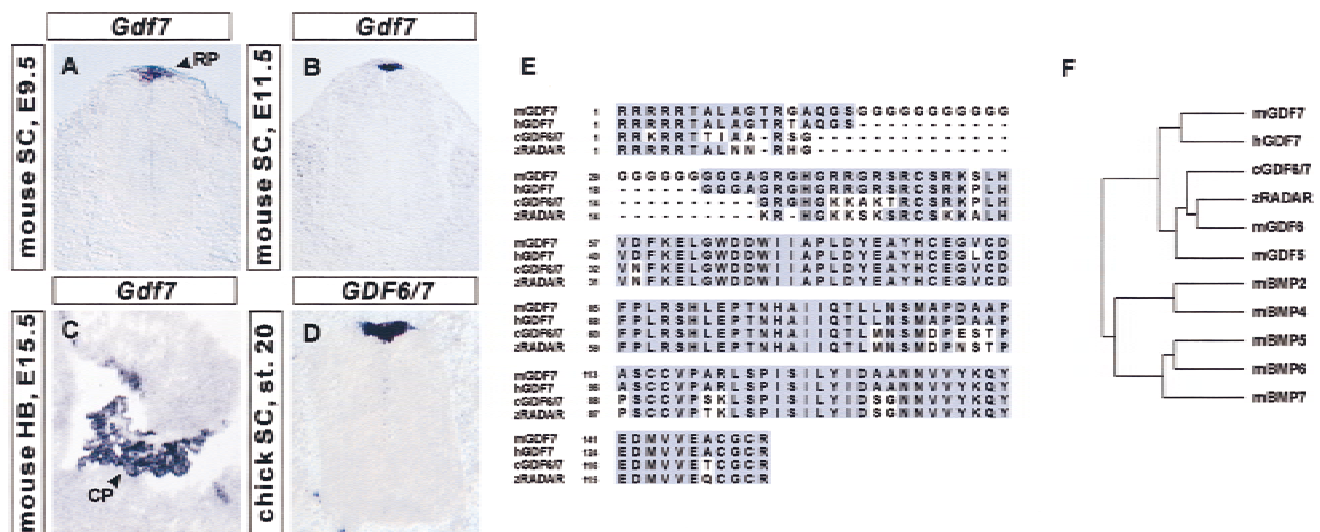


Figure 1. Expression of *Gdf7* in the embryonic CNS. (A–C) Distribution of *Gdf7* in the embryonic mouse CNS; (D) distribution of the related chick *GDF6/7* gene in the developing chick spinal cord (SC). (A,B) Transverse sections through the mouse caudal neural tube showing *Gdf7* expression in the roof plate (RP) at E9.5 (A) and at E11.5 (B). (C) Parasagittal section through the E15.5 mouse hindbrain (HB) showing *Gdf7* expression in the choroid plexus (CP) of the fourth ventricle. (D) Transverse section through stage 20 chick spinal cord showing selective expression of *GDF6/7* in the roof plate. Although chick *GDF6/7* is slightly more similar in sequence to mouse *Gdf6* than mouse *Gdf7* (see F), its expression pattern resembles that of *Gdf7*. (E) Sequence alignment of the predicted amino acid sequences of the mature carboxy-terminal domains of mouse (m) GDF7 (Storm et al. 1994), human (h) GDF7, chick (c) GDF6/7 and zebrafish (z) RADAR (Rissi et al. 1995). (F) Sequence relationship between mouse GDF7, human GDF7, chick GDF6/7, zebrafish RADAR, and other mouse BMP and GDF family members (Hogan 1996). Sequence similarity was determined by comparison of predicted amino acid sequences following the first cysteine residue of the mature carboxy-terminal domains. This dendrogram was generated by use of the Genetics Computer Group program PileUp.

ral axis until E15.5, the latest stage examined (data not shown). The related genes *Gdf5* and *Gdf6* (Storm et al. 1994) are not expressed in the embryonic spinal cord (data not shown).

Two other BMP genes, *Bmp6* and *Bmp7*, are also expressed in roof plate cells (Fig. 4C,D, below; Jones et al. 1991; Lyons et al. 1995; Arkell and Beddington 1997; Dudley and Robertson 1997). Within the roof plate, the expression of *Gdf7*, *Bmp6*, and *Bmp7* overlapped temporally and spatially, but *Bmp6* and *Bmp7* were both expressed more widely than *Gdf7* in other regions of the CNS (data not shown). In addition, expression of *Bmp7* was detected in epidermal ectoderm cells flanking the neural folds (data not shown; Dudley et al. 1995; Lyons et al. 1995; Arkell and Beddington 1997). Beginning at E13, expression of *Gdf7*, as well as *Bmp6* and *Bmp7* (Furuta et al. 1997), was detected in choroid plexus cells. *Gdf7* was expressed solely in the choroid plexus of the fourth ventricle (Fig. 1C), whereas expression of *Bmp6* and *Bmp7* was also detected in lateral ventricle choroid plexus (data not shown; Furuta et al. 1997).

The patterns of neural expression of many members of the BMP family vary among different vertebrate species (Jones et al. 1991; Basler et al. 1993; Dudley and Robertson 1997; Liem et al. 1997). To determine whether the pattern of *Gdf7* expression is conserved, we isolated a chick gene that encodes a BMP family member closely related to mouse GDF6 and GDF7 (Fig. 1E,F). This gene is also expressed selectively by roof plate cells (Fig. 1D). A closely related zebrafish gene, *Radar*, is expressed in the dorsal midline of the hindbrain and spinal cord (Rissi et al. 1995). Thus, the roof-plate-specific expression of a member of the *Gdf7/Gdf6/Radar* subfamily of *Bmp* genes appears conserved in vertebrate evolution.

Expression of mATH1 and LH2 transcription factors defines two classes of dorsal commissural interneurons

The time of onset and selectivity of expression of *Gdf7* by roof plate cells suggested a role for this gene in the differentiation of interneurons generated close to the dorsal midline of the spinal cord. To begin to address this issue, we used the expression of transcription factors to define successive steps in the differentiation of distinct classes of dorsal interneurons in the spinal cord of mouse and chick embryos. The basic helix-loop-helix (bHLH) gene *mATH1*, and its chick homolog *cATH1*, are expressed by neural progenitor cells that are located adjacent to the roof plate of the spinal cord (Fig. 2A,E,I,M-P; Akazawa et al. 1995; Ben-Arie et al. 1996). Double labeling showed that laterally positioned *mATH1*⁺/*cATH1*⁺ progenitors express the LIM homeodomain proteins LH2A (*Lhx2a*) and LH2B (*Lhx2b*) (Fig. 2M-P; data not shown), markers of the D1 class of commissural neurons (Liem et al. 1997). During the early stages of neurogenesis in the dorsal spinal cord, neuronal precursors migrate in a strict mediolateral plate (Leber and Sanes 1995). Thus, these data support the idea that *mATH1*⁺/*cATH1*⁺ progenitors give rise to D1 commissural neu-

rons. The analysis of *mATH1/lacZ* transgenic mice has provided independent evidence for the generation of D1 commissural neurons from *mATH1*⁺ dorsal progenitors (Helms and Johnson 1998).

In the E10.5 mouse spinal cord, the domain of *LH2A* and *LH2B* expression overlapped completely, and expression of both genes was detected in all cells in this region (Fig. 2B,C; data not shown). Thus individual dorsal neurons appear initially to express both *LH2A* and *LH2B*. Between E10.5 and E12, a population of neurons that expresses *LH2B* but not *LH2A*, defined as D1B neurons, appears more ventrally and settles in the deep dorsal horn (Fig. 2F-H). Cell lineage tracing studies have revealed that neurons that are generated at dorsal positions in the spinal cord later migrate circumferentially, in a dorsal-to-ventral direction (Leber and Sanes 1995). The time of appearance and position of D1B neurons, together with the ventral migration of dorsally generated neurons, provide strong evidence that D1B neurons arise from *LH2A*⁺/*LH2B*⁺ neurons generated at the lateral margins of the *mATH1*⁺ progenitor domain.

After E12, the expression of *LH2A* and *LH2B* segregates into distinct neuronal populations in the dorsal spinal cord. At this stage, neurons that emerge from the *mATH1*⁺ progenitor domain express *LH2A* but not *LH2B* (Fig. 2J-L). These neurons, defined as D1A neurons, also appear to migrate ventrally to populate the deep dorsal horn, but settle in a position distinct from that of D1B neurons (Fig. 2J-L; data not shown). The expression of *mATH1* overlaps that of the LH2 proteins throughout the period of D1 neuron generation (Fig. 2M-P; data not shown). Thus, both D1A and D1B neurons appear to derive from *mATH1*⁺ progenitors but acquire distinct profiles of transcription factor expression and occupy different positions in the dorsal spinal cord. A third class of dorsal interneurons (termed D2 neurons) expresses the LIM homeobox gene *Isl1* and appears to arise from a more ventral position (Liem et al. 1997).

GDF7 induces the differentiation of dorsal commissural neurons in vitro

To examine whether GDF7 can promote the generation of D1A and D1B interneurons, stage 10 chick intermediate neural plate (ii) explants (Yamada et al. 1993) were cultured alone or in the presence of GDF7 for 36–48 hr. Explants grown alone did not give rise to *cATH1*⁺ or *LH2A*⁺/*LH2B*⁺ cells (Fig. 2Q,S). Treatment with GDF7 induced *cATH1*⁺ cells (Fig. 2R) as well as neurons that expressed *LH2A* and/or *LH2B* (Fig. 2T). Because the available anti-LH2 antibody detects both *LH2A* and *LH2B* proteins, we used an RT-PCR assay to distinguish the induction of *LH2A* and *LH2B* gene expression. Treatment of [i] explants with GDF7 induced both *LH2A* and *LH2B* (Fig. 2U). Thus, GDF7 induces the expression of transcription factors that define the D1A and D1B classes of interneurons.

Gdf7 mutant mice develop hydrocephalus

To address the requirement for GDF7 function in cell

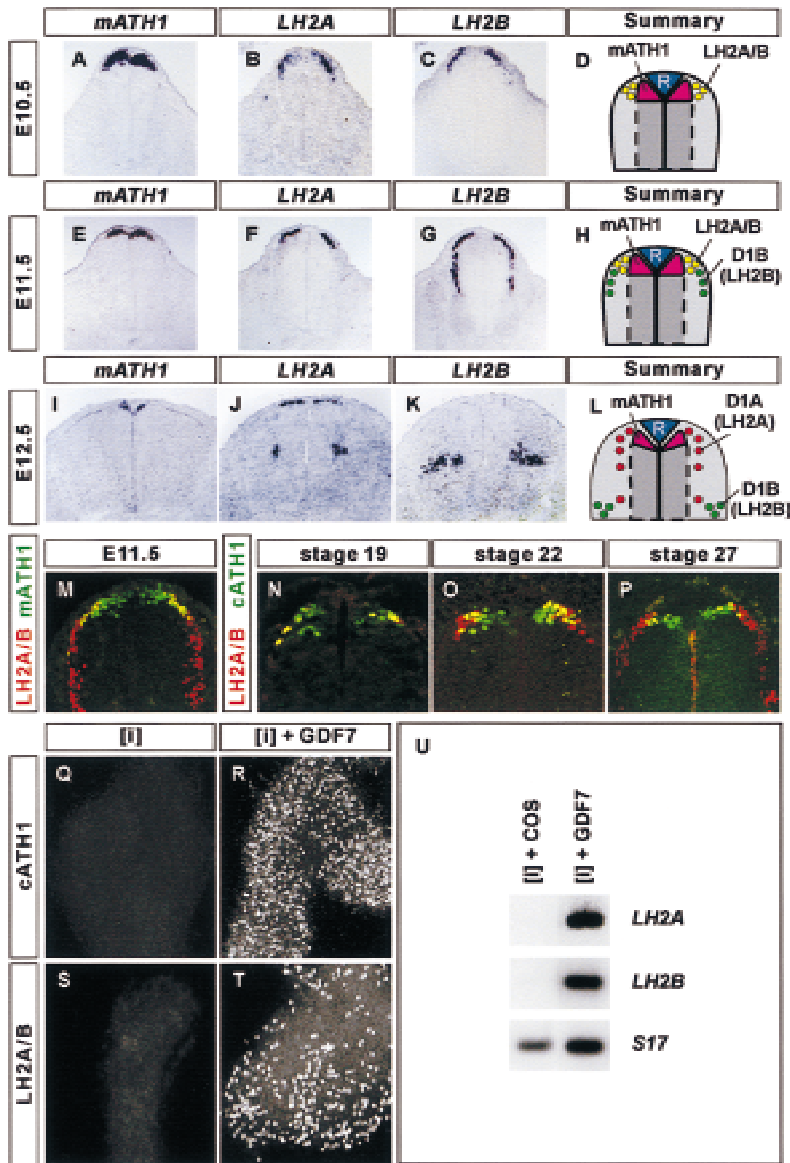


Figure 2. GDF7 promotes the generation of D1A and D1B sensory interneurons. (A–L) In situ hybridization of mouse cervical spinal cord showing the expression of *mATH1* (A,E,I), *LH2A* (B,F,J), and *LH2B* (C,G,K) during the differentiation of dorsal sensory interneurons. At E10.5 (summarized in D), *mATH1* (A) is expressed by neural progenitors adjacent to the roof plate. *LH2A* (B) and *LH2B* (C) are co-expressed by neurons at the lateral margins of the *mATH1*⁺ progenitor domain. At E11.5 (summarized in H), co-expression of *LH2A* (F) and *LH2B* (G) persists in neurons lateral to the *mATH1*⁺ progenitors (E), and a group of more ventrally located D1B neurons express *LH2B* but not *LH2A*. At E12.5 (summarized in L), expression of *LH2A* (J) and *LH2B* (K) has segregated into distinct neuronal populations. Most D1A neurons, which express *LH2A* alone, are adjacent to the *mATH1*⁺ progenitors (J), whereas D1B neurons, which express *LH2B* alone, are located in the deep dorsal horn. Expression of *LH2A* and *LH2B* persists in D1A and D1B neurons until roughly E14.5. (M–P) Confocal images showing *mATH1/cATH1* (green) and *LH2A/LH2B* (red) immunoreactivity in the mouse (M) and in the chick (N–P) spinal cord. Double-labeled cells appear yellow. (M) In the E11.5 mouse spinal cord, *LH2A/LH2B* immunoreactivity is detected in the most lateral *mATH1*⁺ progenitors. In the chick spinal cord, lateral *cATH1*⁺ progenitors co-express *LH2A* and/or *LH2B* at stage 19 (N), stage 22 (O), and stage 27 (P). Thus, both D1A and D1B neurons appear to derive from *mATH1*⁺/*cATH1*⁺ progenitors. (Q,R) Stage 10 chick [i] explants cultured for 48 hr with GDF7 generate *cATH1*⁺ neural cells ([i] + control supernatant, 0 cells; [i] + GDF7, 526 ± 105 cells; mean ± S.D., n = 4–6 explants). (S,T) chick [i] explants cultured for 48 hr with GDF7 generate *LH2A/LH2B*-immunoreactive neurons ([i] + control supernatant, 0 cells; [i] + GDF7, 425 ± 88 cells; mean ± S.D., n = 4–6 explants). (U) RT-PCR analysis of *LH2A*, *LH2B*, and ribosomal protein *S17* gene (control) expression in stage 10 chick [i] explants cultured for 36 hr with COS cell (control) supernatant or supernatant containing GDF7. GDF7 treatment induces expression of both *LH2A* and *LH2B*.

patterning in the dorsal spinal cord, we generated a null mutation in the mouse *Gdf7* gene (Fig. 3A–C). Mice heterozygous for the targeted *Gdf7* allele (termed *Gdf7*^{mi}) were overtly normal and fertile. These mice were interbred to generate homozygous *Gdf7*^{mi}/*Gdf7*^{mi} mutant offspring. In two different genetic backgrounds, *Gdf7*^{mi} homozygous mutant mice were born in normal numbers and survived the immediate postnatal period (Table 1).

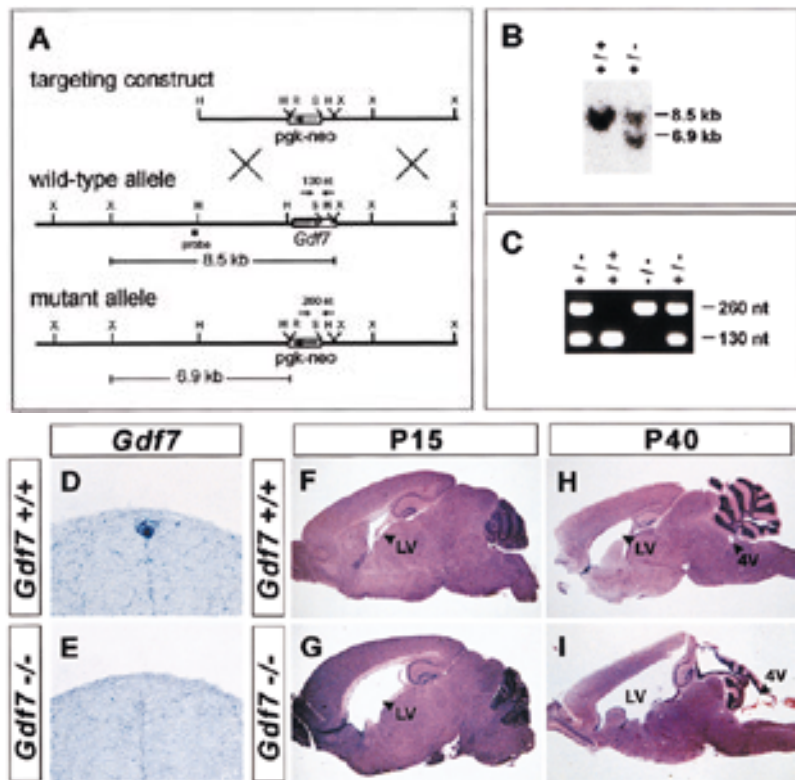
From 14 to 21 days after birth, however, a significant fraction of the homozygous mutant animals developed severe hydrocephalus (Fig. 3F–I). The penetrance of this phenotype varied between 20% and 37%, depending on the genetic background (Table 1). Similar hydrocephalic abnormalities were not detected in heterozygous (*Gdf7*^{mi}/+) or homozygous wild-type siblings in either

genetic background. Further details of the hydrocephalic phenotype are provided in the legend to Figure 3.

A selective defect in neurogenesis in the dorsal spinal cord of Gdf7 mutant mice

To analyze the role of GDF7-mediated signaling in embryonic neuronal patterning, we examined neural differentiation in the dorsal spinal cord of *Gdf7*^{mi} mutant embryos. We first determined whether the development of the roof plate itself is perturbed in *Gdf7*^{mi} mutants. The roof plate expression of the homeobox genes *Msx1* and *Msx2* and the secreted growth factor gene *Wnt1* was similar in wild-type and *Gdf7*^{mi} homozygous mutant embryos analyzed from E11.5 to E13.5 (Fig. 4A,B; data

Figure 3. Generation of a *Gdf7*-null allele and hydrocephalic abnormalities in *Gdf7* mutant mice. (A) Strategy for targeted disruption of the mouse *Gdf7* locus by homologous recombination. The diagram indicates the position of the *Gdf7* exon encoding half of the pro-domain and the entire mature carboxy-terminal region (coding region, solid rectangle; 3' UTR, open rectangle). Restriction sites: (H) *Hind*III; (R) *Eco*RV; (S) *Sph*I; (X) *Xba*I. (B) Southern blot analysis of DNA from ES clones digested with *Xba*I and *Eco*RV and probed with the fragment indicated in A. The presence of the wild-type allele is indicated by an 8.5-kb band and the disrupted allele by a 6.9-kb band. (C) PCR analysis of DNA from progeny of heterozygote matings with primers indicated in A. The wild-type allele is indicated by a 130-bp band and the mutant allele by a 260-bp band.



(D, E) In situ hybridization of wild-type (D) and *Gdf7*^{mut1} homozygous mutant (E) embryos with a *Gdf7* coding region probe. Hybridization was not detected in the roof plate in *Gdf7*^{mut1} homozygous mutant embryos. (F–I) Parasagittal sections of brains from wild-type (F, H) and *Gdf7*^{mut1} homozygous mutant (G, I) mice stained with hematoxylin/eosin. Twenty-five percent of *Gdf7* mutant brains ($n = 8$) analyzed at postnatal (P) day 15 showed a marked dilation of the lateral ventricles (LV). No obvious defects in morphology of the cerebellum and cerebral cortex were detected at this age. By P40, a severe hydrocephalus that is associated with dilation of the fourth (4V) and lateral ventricles (cf. H and I) was observed in 25%–37% of *Gdf7* mutants (see Table 1). Hydrocephalic mutant animals showed considerable variation in the extent to which the different ventricles were enlarged. In some mutants (I) the fourth ventricle was dilated, and the organization and foliation of the cerebellum was disrupted. In all affected brains examined, however, the cerebral cortex was thinned and the hippocampus was displaced dorsally. Despite these abnormalities, histological analysis showed that the cellular architecture of the cerebellum and cerebral cortex was grossly normal in *Gdf7*-null mutants (data not shown). The late onset of brain defects and the relatively normal cellular organization in *Gdf7* mutants suggest that the disruption of brain morphology is a secondary consequence of hydrocephalus. The expression of *Gdf7* in the fourth ventricle choroid plexus (Fig. 1C) raises the possibility that defects in choroid plexus development may underlie the hydrocephalus observed in *Gdf7*^{mut1} mutant mice. Histological analysis indicated that the fourth ventricle choroid plexus epithelium was still present in affected *Gdf7*^{mut1} mutant mice and expression of *Msx1*, *Bmp6*, and *Bmp7* in the choroid plexus appeared normal (data not shown). Thus, the cellular basis of the hydrocephalus is unclear, but it is likely to reflect a requirement for *Gdf7* function in choroid plexus cells.

not shown). Moreover, the expression of *Bmp6* and *Bmp7* by roof plate cells was unchanged in *Gdf7*^{mut1} mutant embryos examined from E10.5 to E12.5 (Fig. 4C, D;

Table 1. Analysis of progeny from heterozygote matings (*Gdf7*^{mut1/+} × *Gdf7*^{mut1/+})

			Hydrocephalic -/-
+/+	+/-	-/-	(% of total -/-)
No. of progeny: mixed background (129Sv × C57BL/6)			
66 (26%)	127 (50%)	60 (24%)	22 (37%)
No. of progeny: inbred background (129Sv)			
15 (24%)	32 (52%)	15 (24%)	3 (20%)

Animals were genotyped ~10 days after birth. No differences were noted in the results of crosses of animals that derive from two independent ES cell targeting events, and both sets of results are combined here. Hydrocephalus was not observed in any wild-type or heterozygous animals in these crosses.

data not shown). Thus, the differentiation of the roof plate and its expression of other signaling molecules appear not to be affected by the loss of *Gdf7* function.

Next, we examined the development of the *mATH1*⁺ progenitor population in *Gdf7*^{mut1} mutants and in wild-type and heterozygous siblings. The early phase (E10–E10.5) of generation of *mATH1*⁺ cells in the dorsal spinal cord appeared normal in *Gdf7*^{mut1} mutants (Fig. 4E, H). However, by E11.5, the number of *mATH1*⁺ progenitors in *Gdf7*^{mut1} mutant embryos was reduced to ~40% (Fig. 4F, H) and by E12.5, to ~10% (Fig. 4G, H) of the number found in wild-type embryos. The loss of the later phase of *mATH1* expression was a fully penetrant phenotype.

This analysis left unresolved whether the elimination of late *mATH1* expression in *Gdf7*^{mut1} mutants reflects the deletion of this progenitor population or simply the loss of an individual molecular marker. To examine this issue, we made use of the observation that *mATH1*⁺ cells are normally interposed between *Msx1*⁺ roof plate cells and a more ventral group of progenitor cells defined by

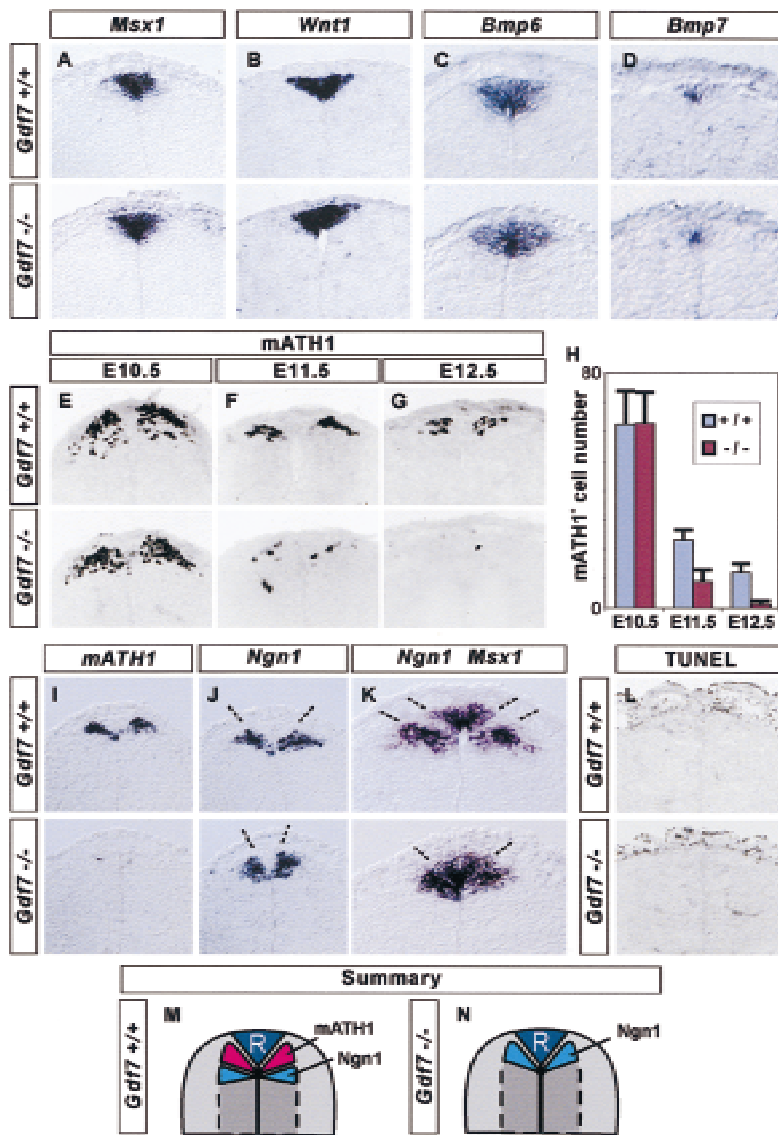


Figure 4. Loss of late *mATH1*⁺ progenitors in *Gdf7* mutant mice. The development of roof plate cells and dorsal neural progenitors in the cervical (forelimb level) spinal cord in wild-type (top panels) and in *Gdf7*^{mut} homozygous mutant embryos (bottom panels). (A–D) In situ hybridization showing that roof plate expression of *Msx1* (A), *Wnt1* (B), *Bmp6* (C), and *Bmp7* (D) is normal in *Gdf7* mutant embryos examined at E12.5 (A,B) and at E11.5 (C,D). (E–H) Generation of *mATH1*⁺ progenitors adjacent to the roof plate. At E10.5 (E,H), *mATH1*⁺ cell number is normal in *Gdf7* mutants. By E11.5 (F,H), the number of *mATH1*⁺ cells in *Gdf7* mutant embryos is significantly reduced (to ~40%), and at E12.5 (G,H), *Gdf7* mutants have only 10% as many *mATH1*⁺ cells as wild-type siblings. The depletion of *mATH1*⁺ cells in the cervical spinal cord was detected in all E12.5 *Gdf7* homozygous mutants examined ($n = 8\text{--}20$ sections from 18 embryos). (H) Quantitative analysis of the *mATH1*⁺ progenitor population in wild-type and homozygous mutant embryos (mean \pm S.D., $n = 10\text{--}25$ sections from two to six embryos of each genotype). In addition, we detected no difference in the number of *mATH1*⁺ progenitors in *Gdf7*^{mut/+} heterozygotes as compared with wild-type siblings. (I–K) In situ hybridization showing the location of *mATH1*⁺ and *Ngn1*⁺ cells in wild-type and *Gdf7*^{mut} mutant embryos at E12.5. In wild-type embryos, *mATH1*⁺ progenitors (I, top) are interposed between *Ngn1*⁺ progenitors (J) and *Msx1*⁺ roof plate cells (summarized in M). Thus, the *mATH1* progenitor domain appears as a region of unlabeled cells (between broken lines) in the wild-type spinal cord hybridized with *Ngn1* and *Msx1* probes (K, top). In *Gdf7*^{mut} mutant embryos, the loss of *mATH1*⁺ cells (I, bottom) is accompanied by a dorsal shift in the position of *Ngn1*⁺ progenitors (J), and the unlabeled region between *Ngn1*⁺ neural cells and *Msx1*⁺ roof plate cells is eliminated (K, top, summarized in N). The distance between the domains of *Ngn1* progenitors (indicated by broken lines in J) is $89 \pm 14 \mu\text{m}$ in wild-type and $43 \pm 5 \mu\text{m}$ in

Gdf7^{mut} homozygote embryos (mean \pm S.D.; $n = 10$ sections from four embryos of each genotype). There is no difference in the mean width of the *Msx1*⁺ roof plate in wild-type and mutant embryos (see A). (L) Detection of apoptotic cell death by end labeling of fragmented DNA (TUNEL assay) in the dorsal spinal cord in E12.5 wild-type and *Gdf7*^{mut} mutant embryos. Very few dying cells are detected in or near the *mATH1* progenitor domain in wild-type embryos or in mutant littermates that lack late *mATH1*⁺ cells. In contrast, abundant cell death was detected in other tissues, for example, in the tail bud and in the distal limb, in both wild-type and mutant embryos (data not shown). (M,N) Summary showing positions of neural progenitors in the dorsal spinal cord in wild-type and *Gdf7*^{mut} mutant embryos at E12.5.

expression of the bHLH gene *Neurogenin1* (*Ngn1*) (Fig. 4M). If the cells that normally express *mATH1* are still present in *Gdf7*^{mut} mutant embryos, we would expect that the *Msx1*⁺ roof plate cells and the *Ngn1*⁺ progenitors are separated by a domain of cells that express neither gene. Instead we observed that the loss of *mATH1* expression (Fig. 4I) was accompanied by a dorsal shift in the *Ngn1*⁺ cell population to a position immediately adjacent to the roof plate (Fig. 4J–K). The size of the *Ngn1*⁺ precursor domain in *Gdf7*^{mut} mutants was, however, unchanged (Fig. 4J). These observations show that the late population of *mATH1*⁺ progenitor cells is deleted in

Gdf7^{mut} mutants.

To address whether the absence of late *mATH1*⁺ progenitors in *Gdf7*^{mut} mutants is a consequence of apoptotic cell death, we compared patterns of cell death in the spinal cord of E12.5 wild-type and mutant embryos. Very few apoptotic cells were detected in the dorsal spinal cord in E12.5 wild-type embryos, and there was no increase in the number of dying cells in *Gdf7*^{mut} mutant embryos (Fig. 4L). Patterns of cell death were indistinguishable in the cervical or thoracic spinal cord, where the loss of *mATH1*⁺ cells in *Gdf7*^{mut} mutants is pronounced at E12.5, and at the more developmentally de-

layed lumbar level, where the loss of *mATH1*⁺ cells was first observed at E13.5. Thus the loss of *Gdf7* function does not appear to result in increased apoptosis in the region of D1 neuron generation in the dorsal spinal cord.

To address whether the loss of late *mATH1* progenitor cells in *Gdf7*^{mut} mutants is accompanied by defects in the generation of dorsal interneurons, we examined the differentiation of D1A and D1B neurons, the two cell populations defined by the differential expression of the LH2 transcription factors. In E11.5 wild-type embryos, *LH2A* and *LH2B* are co-expressed by neurons lateral to the roof plate whereas more ventrally, D1B neurons express *LH2B* but not *LH2A* (Fig. 5A,B). In *Gdf7*^{mut} homozygous mutant embryos examined at E11.5, we found no change in expression of *LH2A* or *LH2B* (Fig. 5A,B), in the number of *LH2A*⁺/*LH2B*⁺ neurons close to the roof plate (Fig. 5C,D) or in the number of ventrally located D1B (*LH2B*⁺/*LH2A*⁻) neurons (data not shown).

Older *Gdf7*^{mut} mutant embryos, however, exhibit a striking defect in dorsal interneuron generation. In E12.5 wild-type embryos, *LH2A* and *LH2B* expression has segregated into two distinct populations of neurons. Most D1A (*LH2A*⁺/*LH2B*⁻) neurons are located lateral to the roof plate, whereas D1B neurons are found in a more ventral position deep in the dorsal horn (Fig. 5F,I). The number of dorsal D1A neurons in E12.5 *Gdf7*^{mut} homo-

zygotes was reduced to ~8% of that in wild-type littermates (Fig. 5F-I). In contrast, the number of *LH2A*⁺/*LH2B*⁺ neurons in the deep dorsal horn, which at E12.5 consist primarily of D1B neurons, was not significantly altered in *Gdf7*^{mut} mutants (Fig. 5I-L). A similar depletion of D1A neurons was seen in *Gdf7*^{mut} mutant embryos examined at E13.5 and at E14.5, again with no apparent change in the number of D1B neurons (data not shown). Several other classes of dorsal cells were also unaffected in *Gdf7*^{mut} mutants, including *Isl1*⁺ (D2) interneurons (wild type, 123 ± 17 D2 neurons/section; *Gdf7*^{mut} homozygotes, 106 ± 13 D2 neurons/section; mean ± s.d. for *n* = 12 sections of four embryos) and dorsal spinal cord interneurons expressing the LIM homeobox genes *Lmx1b* or *Lim1/2* (data not shown).

Taken together, these data show that *Gdf7* function is required selectively in the pathway of D1A interneuron differentiation.

Other TGFβ-related signals are required for the induction of D1B interneurons

Our analysis of neurogenesis in the dorsal spinal cord of *Gdf7*^{mut} mutant embryos indicates that the generation of D1B neurons does not require GDF7 signaling. Previous studies have demonstrated that the ability of chick roof

Figure 5. A selective loss of D1A neurons in *Gdf7* mutant mice. Analysis of dorsal neuron populations in the cervical (forelimb level) spinal cord in wild-type embryos (top panels) and in *Gdf7*^{mut} homozygous mutant embryos (bottom panels). (A–E) D1 neuron populations in the E11.5 spinal cord (summarized in E). In situ hybridization shows that at this stage, expression of *LH2A* (A) and *LH2B* (B) is normal in *Gdf7* mutant embryos. The number of dorsal *LH2A*/*LH2B*-immunoreactive neurons (red box in C) is not changed in *Gdf7* mutants. (D) Quantitative analysis of the dorsal *LH2A*⁺/*LH2B*⁺ cell population (mean ± s.d., *n* = 10–25 sections from two to six embryos of each genotype). We also found no alteration in the number of more ventrally located D1B neurons in *Gdf7*^{mut} mutants. (F–M) D1 neuron populations at E12.5 (summarized in M). At this stage, expression of *LH2A* (F) is greatly reduced, whereas *LH2B* (J) is unaffected in *Gdf7* mutant embryos. Similarly, the number of dorsal *LH2A*/*LH2B*-immunoreactive cells (D1A neurons; boxed in I, shown enlarged in G) is reduced to ~8% the number in wild-type and the number of ventral *LH2A*/*LH2B*-immunoreactive cells (mostly D1B neurons; boxed in I, enlarged in K) is not significantly altered in *Gdf7* mutants. (H,L) Quantitative analysis of the D1A and D1B cell populations (mean ± s.d., *n* = 10–25 sections from two to six embryos of each genotype).

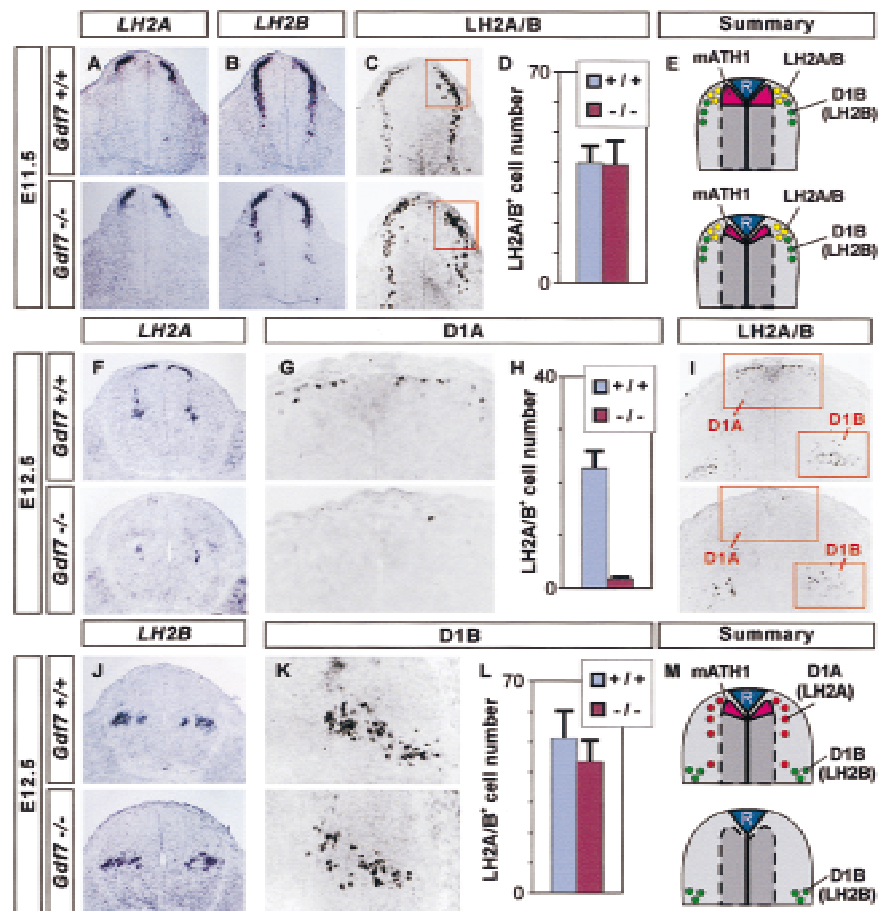


plate to induce $LH2A^+/LH2B^+$ neurons (D1 neurons) is blocked only partially by treatment with the BMP binding proteins Noggin and Follistatin (Liem et al. 1997). This partial block could reflect the expression by the chick roof plate of TGF β family members such as Dsl-1 that are insensitive to Noggin and Follistatin (Liem et al. 1997) or that some D1 neurons can be induced by non-TGF β signals from the roof plate.

To begin to address whether the generation of D1B neurons depends on inductive signals mediated by other TGF β family members, we examined whether the inductive activities of the three known BMPs expressed by mouse roof plate cells can be inhibited by BMP-binding proteins. Noggin binds and inactivates BMP4 (Zimmerman et al. 1996) and Follistatin blocks signaling by activin and BMP7 (Nakamura et al. 1990; Yamashita et al. 1995). We found that the GDF7-mediated induction of $LH2A/LH2B^+$ neurons in chick [i] explants was blocked by Noggin but not by Follistatin (Fig. 6A). Exposure of [i] explants to either BMP6 (Fig. 6B) or BMP7 (Liem et al. 1997) also induced $LH2A/LH2B^+$ neurons. The activity of BMP6 and BMP7 in these assays, in contrast to that of GDF7, was blocked by Follistatin but not by Noggin (Fig. 6B; Liem et al. 1997). Thus, the three known BMP family members expressed by the mouse roof plate are sensitive to the inhibitory actions of either Noggin or Follistatin.

Next, we examined whether inductive signals from mouse roof plate that promote $LH2A/LH2B^+$ neuron generation are inhibited by Noggin and Follistatin. Many $LH2A/LH2B^+$ neurons were induced in [i] explants grown in contact with mouse roof plate (Fig. 6D,F). The induction of $LH2A/LH2B^+$ neurons by the roof plate was markedly suppressed by treatment with either Noggin or Follistatin and was inhibited by ~97% with combined application of both antagonists (Fig. 6E,F). Thus, the generation of both the D1A and D1B classes of neurons in response to roof plate signals appears to depend on inductive signaling mediated by TGF β family members. The persistence of D1B neuronal differentiation in $Gdf7^{m1}$ mutant embryos is therefore likely to result from residual TGF β -related signaling by the roof plate.

The generation of D1A neurons in $Gdf7$ mutants is restored by exogenous GDF7 or BMP7

Our results indicate that GDF7 has an essential signaling function in the generation of D1A neurons. However, both $Bmp6$ and $Bmp7$ are expressed normally by roof plate cells in $Gdf7^{m1}$ mutants (Fig. 4C,D), raising the issue of why BMP6 and BMP7 fail to compensate for the loss of GDF7 activity in vivo. One possible explanation is that in mouse neural tissue, GDF7 alone is capable of promoting D1A neuron formation. To test this idea, we compared the ability of GDF7 and BMP7 to induce D1A neurons in wild-type neural tissue and to rescue D1A neuron generation in neural tissue isolated from $Gdf7^{m1}$ mutant embryos.

First, we asked whether exogenous GDF7 can restore late $mATH1$ and $LH2A$ expression, the two markers of D1A neuron differentiation, in $Gdf7^{m1}$ mutant neural

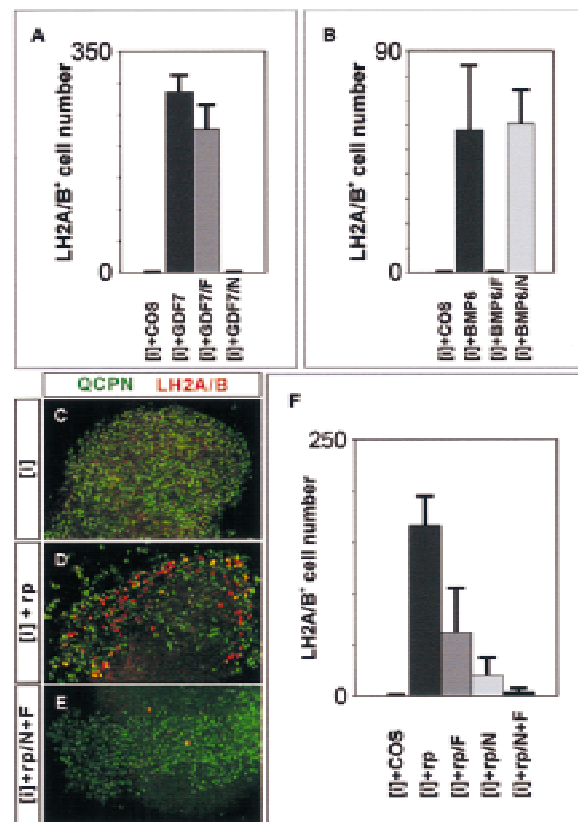
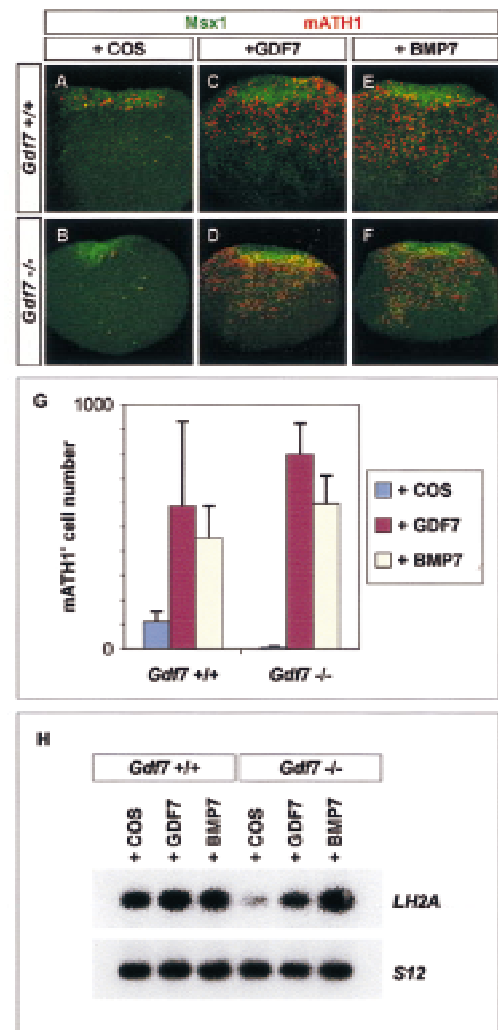


Figure 6. Roof plate induction of D1A and D1B neurons requires TGF β -related signaling. (A) GDF7 induction of $LH2A/LH2B^+$ neurons in chick [i] explants is blocked by addition of Noggin (N; 20 nM) but not by Follistatin (F; 80 nM). (B) Follistatin, but not Noggin, blocks the BMP6-mediated induction of $LH2A/LH2B^+$ neurons. (C–E) Quail neural plate [i] explants [stained with anti-quail cell-specific perinuclear marker (QCPN), green] cultured alone (C) for 48 hr do not generate $LH2A/LH2B^+$ neurons (red), whereas explants cultured with mouse roof plate (D) generate many $LH2A/LH2B^+$ neurons. Addition of Noggin alone (20 nM) or Follistatin alone (80 nM) partially blocks the induction of $LH2A/LH2B^+$ neurons by mouse roof plate (F). Addition of both Noggin and Follistatin reduces mouse roof plate-mediated induction of $LH2A/LH2B^+$ neurons by ~95% (E,F). Histograms show mean \pm S.D. for 6–12 explants.

tissue. Explants of mouse dorsal neural tube containing roof plate cells ([d+rp] explants) were isolated from $Gdf7^{m1}$ homozygous mutants and wild-type siblings at E11.5, a time when a reduction in $mATH1$ expression is first detected in mutant embryos. These explants were cultured for 60 hr, either alone or in the presence of GDF7. When cultured alone, $Gdf7^{m1}$ mutant [d+rp] explants generated only ~10% as many $mATH1^+$ cells as wild-type [d+rp] explants (Fig. 7A,B,G), consistent with the loss of late $mATH1^+$ cells in $Gdf7^{m1}$ mutants in vivo. Moreover, there was a threefold reduction in $LH2A$ expression in [d+rp] explants from $Gdf7^{m1}$ mutants, relative to the level detected in wild-type explants (Fig. 7H). Addition of GDF7 to wild-type [d+rp] explants resulted in a marked increase in the number of $mATH1^+$ cells

Figure 7. Treatment with *Gdf7* or BMP7 restores *mATH1* and *LH2A* expression in *Gdf7* mutant neural tissue. (A–F) Expression of *mATH1* (red) in explants of mouse forelimb level dorsal neural tube ([d+rp] explants) isolated from wild-type (A, C, E) and *Gdf7^{mut}* homozygous mutant (B, D, F) embryos. *Msx1⁺* roof plate cells are green. *Gdf7* mutant [d+rp] explants isolated at E11.5 (44 somites) (B) and cultured in COS (control) supernatant for 60 hr generate <10% as many *mATH1⁺* cells as explants from age-matched wild-type siblings (A, G). Wild-type explants respond to added GDF7 with the generation of ~5-fold more *mATH1⁺* cells (C, G). Culture with GDF7 increases the number of *mATH1⁺* cells in mutant explants to a level similar to that in treated wild-type explants (D, G). Treatment with BMP7 similarly increases *mATH1* cell number in wild-type and mutant explants (E, F, G). In these experiments, explants were treated with equivalent doses of GDF7 and BMP7, as judged by their activity in inducing *LH2A⁺/LH2B⁺* neurons in chick neural plate assays (data not shown). In untreated wild-type and mutant [d+rp] explants, *mATH1⁺* cells are restricted to the region adjacent to the roof plate (A, B). After treatment with GDF7 or BMP7, *mATH1⁺* cells are found at all dorsoventral positions within the explants, independent of genotype (C–F). Thus, at E11.5, cells more distant from the roof plate respond to GDF7 or BMP7 with the expression of *mATH1*. GDF7 treatment of [d+rp] explants isolated from E12.5 wild-type or mutant embryos did not increase *mATH1⁺* cell number (data not shown), suggesting that GDF7 signaling normally acts prior to E12.5 to induce or maintain *mATH1⁺* progenitors. (G) Histograms show mean \pm s.d. of results from four to six explants of each genotype. (H) RT-PCR analysis of *LH2A* and ribosomal protein *S12* gene (control) expression in E11.5 wild-type and *Gdf7* mutant [d+rp] explants cultured for 60 hr with COS cell (control) supernatant or supernatant containing GDF7 or BMP7. Treatment of mutant explants with GDF7 or BMP7 restores *LH2A* expression to wild-type levels.



(Fig. 7C,G). GDF7 treatment of *Gdf7^{mut}* mutant [d+rp] explants increased the number of *mATH1⁺* cells to a level similar to that in treated wild-type explants (Fig. 7D,G) and restored *LH2A* expression to wildtype levels (Fig. 7H). Thus, GDF7 treatment of [d+rp] explants isolated from E11.5 *Gdf7^{mut}* mutant embryos rescues the late phases of both *mATH1* and *LH2A* expression.

To examine whether other BMPs expressed by the roof plate mimic the ability of GDF7 to restore late *mATH1* and *LH2A* expression, wild-type or *Gdf7^{mut}* mutant explants were cultured in the presence of BMP7. This treatment increased *mATH1⁺* cell number to a level similar to that obtained with GDF7 (Fig. 7E,F,G). In addition, exposure of *Gdf7^{mut}* mutant explants to BMP7 restored *LH2A* expression to wild-type levels (Fig. 7H). Thus, there is no apparent difference in the ability of GDF7 and BMP7, or by inference BMP6, to rescue late *mATH1* and *LH2A* expression in *Gdf7^{mut}* mutant explants. These results argue against the idea that the selective requirement for GDF7 function in the generation of D1A neurons results from the inability of neural progenitor cells to respond to BMP6 and BMP7.

Discussion

BMPs have been implicated in the induction and patterning of neuronal cell types in the dorsal spinal cord but their precise functions in this process have not been established. We have examined whether BMPs are required in vivo to promote the differentiation of dorsal interneurons and whether different BMPs have redundant or distinct functions in the generation of specific dorsal cell types. Our results show that *Gdf7*-null mutant mice lack a single identified class of dorsal commissural neurons, D1A neurons, and thus establish that BMP-mediated signals from the roof plate are required for the differentiation of specific neuronal subtypes in the dorsal spinal cord. Furthermore, our studies provide evidence that the multiple BMPs expressed by the roof plate do indeed have nonredundant functions in controlling the generation of selective subsets of spinal neurons. The diversity of TGF β -related molecules expressed in discrete regions within the embryonic CNS thus raises the possibility that this class of signals is employed more widely in the specification of neuronal subtype identity.

GDF7 is required for the generation of D1A interneurons

Genetic studies on the function of other vertebrate BMP family members have established certain essential roles for this class of secreted proteins, but have not resolved their contribution, if any, to the control of neuronal identity. Targeted mutations in mouse *Bmp2* and *Bmp4* result in early embryonic lethality, which has so far precluded an analysis of their later function in neuronal development (Winnier et al. 1995; Zhang and Bradley 1996). Eye growth and development is perturbed in *Bmp7* mutants (Dudley et al. 1995; Luo et al. 1995) but neuronal patterning defects in the brain and spinal cord have not been reported. Mutations of other BMP family members appear not to be associated with neural abnormalities (King et al. 1994; Storm et al. 1994). One interpretation of these results is that there is functional redundancy between co-expressed BMP family members (Dudley and Robertson 1997). Alternatively, the BMPs that have been analyzed to date may not play essential roles in the particular neural structures that have been studied. Our results establish that GDF7 has a critical function in the development of a specific subset of dorsal spinal cord interneurons and thus provide genetic evidence for the involvement of BMP signaling in the control of neuronal identity and pattern.

The expression of *Bmp6* and *Bmp7* by roof plate cells is not changed in *Gdf7* mutants. Therefore, the loss of D1A neurons in *Gdf7* mutants could reflect a specific requirement for GDF7 signaling. Alternatively, the inductive activity of GDF7 and other roof-plate-derived BMPs could be equivalent, with the diffusion of multiple BMPs from the roof plate establishing a BMP activity gradient in the dorsal neural tube. If the specification of D1A neurons were to require the highest level of BMP activity, the loss of GDF7 function might simply reduce net BMP activity to a level below the threshold sufficient to generate D1A neurons. An examination of dorsal neuronal patterning in *Bmp6* and *Bmp7* mutants has, however, revealed no apparent defect in the differentiation of D1A or D1B neurons (K.J. Lee, T.M. Jessell, A. Dudley, M. Solloway, and E. Robertson, unpubl.). Thus, the loss of D1A interneurons in *Gdf7* mutants is most easily explained by a selective dependence on GDF7 signaling, in turn indicating that roof-plate-derived BMPs have non-equivalent functions in vivo.

It remains unclear why *Bmp6* and *Bmp7* fail to substitute for *Gdf7* in vivo. The specific requirement for GDF7 could result either from selectivity in the ability of these BMPs to promote D1A neuron generation or from a difference in the presentation of these signaling molecules in vivo. We find that D1A neuron generation can be restored in *Gdf7^{mut1}* mutant explants by treatment with either GDF7 or BMP7. Similarly, GDF7, BMP6, and BMP7 have indistinguishable inductive activities on chick neural tissue. Thus, the exposure of neural tissue to recombinant BMPs in vitro appears to obscure the distinction in the functions of GDF7 and other roof-plate-derived BMPs in vivo.

GDF7, BMP6, and BMP7 have apparently similar activities on cultured neural tissue, yet serve different functions in vivo. One possible explanation for this observation is that although *Bmp6* and *Bmp7* are expressed by roof-plate cells, the level of BMP6 and BMP7 produced is below the threshold for D1A neuron generation. A second possibility is that the activities of BMPs in vivo require accessory factors that are available in the vicinity of the roof plate only for GDF7. One potential class of accessory factor may be a BMP heterodimer partner that enhances inductive activity (Aono et al. 1995; Suzuki et al. 1997), and the roof plate might express such a factor that dimerizes with GDF7, but not with other BMPs. A second class of accessory factor might be one required to facilitate the secretion, diffusion, or presentation of BMPs. The *Drosophila* BMP-binding protein Sog has been suggested to function in vivo not simply as a BMP antagonist, but rather as part of a transport mechanism for the BMP family member Dpp (Holley et al. 1996). Indeed, Noggin, a protein that we have found to bind GDF7 but not BMP6 and BMP7, is expressed by roof-plate cells (Shimamura et al. 1995; McMahon et al. 1998). It is possible, therefore, that Noggin facilitates the diffusion or presentation of GDF7, and the failure of BMP6 and BMP7 to substitute for GDF7 reflects the absence of corresponding accessory factors for these BMPs. Mice lacking *Noggin* function have defects in ventral neural tube patterning (McMahon et al. 1998), but the differentiation of dorsal interneurons has not been analyzed in these mutants.

The generation of other classes of dorsal interneurons depends on TGF β -related signals independent of GDF7

The generation of D1B neurons is unaffected in *Gdf7^{mut1}* mutants, indicating that other signals are sufficient for D1B neuron generation. One likely source of signals that direct D1B neuron generation is the roof plate. In addition, because D1B neurons are generated prior to D1A neurons, earlier BMP signals produced by epidermal ectoderm cells (Liem et al. 1995; Lyons et al. 1995; Dudley and Robertson 1997) may also contribute to the generation of D1B neurons. The induction of *LH2A/LH2B⁺* neurons by roof-plate cells (Fig. 6) and by epidermal ectoderm (Liem et al. 1997) is blocked by treatment with Follistatin and Noggin. Thus, inductive signals for D1B neuron differentiation, whether provided by the roof plate or by the epidermal ectoderm, appear to be mediated by TGF β family members.

Other classes of dorsal interneurons, notably D2 interneurons, also differentiate normally in the absence of GDF7 function. A different TGF β -related protein, activinB, has been implicated in the induction of D2 interneurons (Liem et al. 1997). Members of different subclasses of the TGF β superfamily can also act as qualitatively distinct signals in the promotion of diverse neural crest derivatives (Shah et al. 1996). Collectively, these observations support the idea that the activities of distinct TGF β family members contribute to the diversifi-

cation of cell types in both the central and peripheral nervous systems.

GDF7 signaling and the pathway of D1 interneuron specification

Our results indicate that the loss of *mATH1* expression reflects the deletion of the neuronal progenitors normally marked by *mATH1*. It follows that the D1A neurons that normally derive from these progenitors are also lacking. The elimination of the late *mATH1*⁺ cells does not appear to result from apoptosis of these dorsal neural progenitors as no increase in cell death was observed in the dorsal spinal cord in *Gdf7*^{mut} mutants. In addition, we found no increase in the *Ngn1*⁺ population or in the number of D1B or D2 interneurons in *Gdf7* mutants. Thus, the loss of *mATH1*⁺ cells and D1A neurons is likely to result from a failure in the generation of this cell population rather than from cell death or from a switch in cell fate.

There are two possible steps at which GDF7 may influence the differentiation of D1 interneurons. Neither the initial generation of *mATH1*⁺ progenitors in the dorsal neural tube nor the differentiation of D1B interneurons from these progenitors is affected by loss of GDF7 function. In contrast, the maintenance of the *mATH1*⁺ progenitor population and the differentiation of D1A neurons from this late population of *mATH1*⁺ progenitors depends on GDF7 signaling. Thus, GDF7 activity could be required to induce late *mATH1*⁺ progenitors from the pool of dorsal *Pax3*⁺, *Pax7*⁺ precursors (Fig. 8, Model 1). Alternatively, GDF7 could be required for the conversion of early D1B neuron-producing *mATH1*⁺

cells to a second *mATH1*⁺ progenitor cell type that gives rise instead to D1A neurons (Fig. 8, Model 2).

An essential role for the roof plate as a secondary signaling center

The patterning of cell types in both the dorsal and ventral halves of the spinal cord appears to involve a cascade of inductive signaling initiated in adjacent non-neural tissue and propagated by midline glial cells (Tanabe and Jessell 1996). Ventrally, Shh secreted by the notochord induces floor plate cells, thereby establishing a secondary, neural source of Shh. Shh function is required for ventral cell patterning in the spinal cord (Marti et al. 1995; Chiang et al. 1996; Ericson et al. 1996). However, it remains unclear whether early Shh signaling by the notochord and late Shh signaling by the floor plate have separate roles in the patterning of the ventral spinal cord.

In the dorsal spinal cord, BMPs produced by cells of the epidermal ectoderm induce roof-plate cells, which in turn express additional BMPs (Liem et al. 1997). Inductive signaling in the dorsal neural tube, however, appears to differ from that in the ventral neural tube, in that certain BMPs such as *Gdf7* are expressed solely by roof-plate cells and not by the epidermal ectoderm. Our genetic analysis of GDF7 function has established a requirement for roof-plate-derived signals in the differentiation of one subclass of dorsal interneurons. Thus, primary and secondary dorsal signaling centers, the epidermal ectoderm and roof plate, have distinct roles in the regulation of neuronal identity and pattern in the mammalian CNS.

Materials and methods

Cloning of chick and human Gdf7-related genes

A 75-nucleotide fragment of chick *GDF6/7* was cloned by degenerate PCR with the primers: 5'-CCGCTGGATTACGAGGCATAC-3' and 5'-GTTGGTGGGCTCCAGGTGGG-3'. This fragment was used to screen a chicken genomic DNA library (Stratagene), and two overlapping clones were isolated and mapped. A fragment of human *GDF7* (gift of Se-Jin Lee) was used to isolate overlapping clones from a human genomic DNA library. Human *GDF7* was mapped by FISH (BIOS Labs) to 2p23-2p24, a position that is syntenic to that reported previously for mouse *Gdf7* on the proximal end of chromosome 12 (Storm et al. 1994).

Generation of a targeted mutation in Gdf7

A mouse 129/Sv genomic library (Stratagene) was screened with a 400-nucleotide *Gdf7* genomic fragment (Storm et al. 1994), and overlapping phage clones were isolated and mapped (Fig. 3). The *Gdf7* targeting vector was constructed by flanking *pgk-neo* with a 3.5-kb *HindIII* fragment and a 5.5-kb *SphI-NotI* fragment (where the *NotI* site is provided by the phage λ polylinker). This targeting vector is designed to delete the *Gdf7* exon that encodes half of the propeptide and the mature carboxy-terminal domain. Linearized plasmid (30 μ g) was used to electroporate 3×10^7 E14 ES cells (Hooper et al. 1987) at 800 V and 3 μ F with a BioRad Gene Pulser. One hundred and sixty G418-resistant

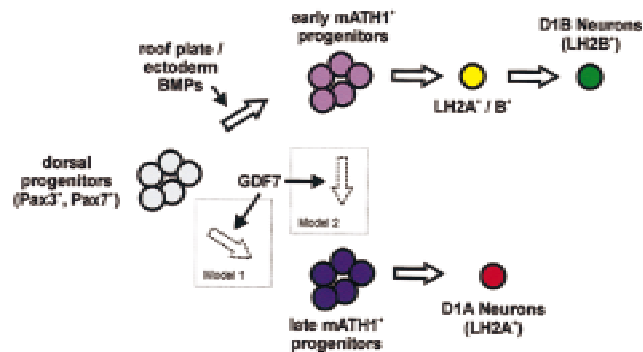


Figure 8. Potential roles for GDF7 in the regulation of interneuron fate in the dorsal spinal cord. The generation of late *mATH1*⁺ progenitors and D1A neurons depends on GDF7 activity. In contrast, other BMP signals from the epidermal ectoderm and/or roof plate appear to be sufficient for the induction of early *mATH1*⁺ progenitors and the differentiation of D1B neurons. GDF7 could function in the generation of D1A neurons in one of two ways: GDF7 may act on dorsal *Pax3*⁺, *Pax7*⁺ neural plate cells to induce the late population of *mATH1*⁺ progenitors that differentiate into D1A neurons (Model 1). Alternatively, GDF7 may be required to convert the early D1B-fated *mATH1*⁺ progenitors into the late class of *mATH1*⁺ progenitors that generates D1A neurons (Model 2).

colonies were isolated and expanded. Genomic DNA was prepared and digested with *Xba*I and *Eco*RV and probed with a 200-nucleotide *Sma*I fragment. Six recombinant clones were identified and confirmed by further Southern blot analysis. Two of these clones were injected into C57BL/6J blastocysts to produce a total of three chimeric founders that transmitted the mutated allele.

Indistinguishable phenotypes were observed in animals derived from the independently targeted clones, both in a mixed (129Sv × C57B/6) and an inbred (129Sv) background. Genotyping of embryos and offspring was performed by PCR with the following primers: Neo220, 5'-ACTTCCTGACTAGGGGAGGAG-3'; GDF-C3, 5'-ATTCTCTACATCGATGCCGCC-3'; KO-B1, 5'-ATGACACCTGCTCTTGAGCTAGGG-3'. Thirty-three cycles of PCR were performed with annealing at 65°C. The mutant allele generated a 260-nucleotide product (between Neo220 and KO-B1) and the wild-type allele a 130-nucleotide product (between GDF-C3 and KO-B1).

In situ hybridization

In situ hybridization was performed essentially as described (Schaeren-Wiemers and Gerfin-Moser 1993). For detection of mouse *Gdf7*, a 400-nucleotide genomic fragment (Storm et al. 1994) or a 1.2-kb *Hind*III fragment was used as a probe. A 5-kb *Xba*I genomic fragment was used to detect chick *GDF6/7*. Other probes for in situ hybridization were *Bmp2* and *Bmp6* (Lyons et al. 1989), *Bmp4* (Jones et al. 1991), *Bmp5* (King et al. 1994), *Bmp7* (Lyons et al. 1995), *Gdf5* and *Gdf6* (Storm et al. 1994), mouse *Dsl1* (Basler et al. 1993), *mATH1* (Helms and Johnson 1998), rat *LH2A* (Xu et al. 1993), mouse *LH2B* [gift of L. Carlsson (Umeå University, Sweden) and H. Okamoto (Keio University School of Medicine, Japan)], chick *LH2A* (gift of J.C. Izpisua-Belmonte, Salk Institute, La Jolla, CA), chick *LH2B* (Liem et al. 1997), *Wnt1* (Parr et al. 1993), *Msx1* (MacKenzie et al. 1991), *Msx2* (Monaghan et al. 1991), *Ngn1* (Ma et al. 1996), and *Lmx1b* (Chen et al. 1998).

Immunofluorescence and cell death (TUNEL) assays

Immunocytochemistry was performed as described (Yamada et al. 1993; Pfaff et al. 1996). Antibodies used were: L1, rabbit anti-LH2A/LH2B (Liem et al. 1997); mouse anti-LH2A/LH2B (G. Tremml and T.M. Jessell, unpubl.), guinea pig anti-LH2B (K.J. Lee and T.M. Jessell, unpubl.); anti-mATH1/cATH1 (Helms and Johnson 1998); K5, anti-Isl1 (Tsuchida et al. 1994); T4, anti-Lim1/2 (Tsuchida et al. 1994); 4G1, anti-Msx1/2 (Riddle et al. 1995). Apoptotic cell death was detected in cryostat-sectioned embryonic tissue by fluorescein-dUTP labeling of DNA strand breaks with the In Situ Cell Death Detection kit (Boehringer Mannheim).

RT-PCR

RT-PCR was performed as described (Tanabe et al. 1995). The primers for amplification of chick *LH2A* were: CLH2A3, 5'-TGCAACAAGATGCTGACCACC-3' and CLH2A4, 5'-TTGACGTAGGAGGTTCCGTCTG-3'; for amplification of chick *LH2B*: GT52, 5'-GAGTCCCTTTTGCAAGGAGAATAC-3' and GT53, 5'-CGCTTTATCGACACCCCATTC-3'; for amplification of mouse *LH2A*: LH2A12, 5'-TCAACCTG-GAGTCGGAACACTCAC-3' and LH2A4, 5'-TTGTCTTTTGGC-TGCTGGGG-3'.

Neural explant culture and induction assays

Explants of chick neural plate ([i] explants) were isolated and cultured as described previously (Yamada et al. 1993; Liem et al. 1995). Explants of mouse neural tube ([d+rp] explants) from the cervical (forelimb) region were dissected in dispase, and surrounding tissues were removed. The neural tube was opened at the ventral midline, and the roof plate and dorsal one-quarter of the neural tube was isolated. Mouse explants were cultured in collagen matrices in Iscove's Modified Dulbecco's Medium (GIBCO) supplemented with Mito serum extender (Collaborative Research).

Mouse GDF7, human BMP7, and mouse BMP6 were obtained by transfection of COS-7 cells (Basler et al. 1993). To prepare a Myc-tagged GDF7 expression construct, the GDF7 carboxy-terminal mature peptide coding region was first amplified by PCR with the primers: 5'-CGGAATTCGATATCCGAGGAGGACCTGAGCCGCTGCAGTCGCAAGTC-3' and 5'-AGCACTAGTATCCTACCTGCAGCCGAGGCCT-3'. This fragment was digested with *Eco*RV and *Spe*I (underlined sites) and ligated to a *Asp*718-*Eco*RV fragment encoding the amino-terminal propeptide region of DSL1 (Basler et al. 1993) to recreate the myc epitope at the junction between DSL1 and GDF7. A similar construct that expresses a fusion of the GDF7 mature peptide with the propeptide region of BMP4 was also prepared. Both fusions were cloned into the expression vector pMT21 and used to transfect COS-7 cells. The resulting supernatants containing DSL1-GDF7 or BMP4-GDF7 had qualitatively identical activities in all assays. Noggin was provided by Richard Harland (University of California, Berkeley) and Follistatin by the National Hormone and Pituitary Program.

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