

Nucleotide sequence of the *Euglena gracilis* chloroplast genes for isoleucine, phenylalanine and cysteine transfer RNAs and ribosomal protein S14

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The nucleotide sequence of 780 bp of *Euglena gracilis* Z chloroplast DNA encoding three tRNA genes, *trnI-CAU*, *trnF-GAA*, *trnC-GCA*, and the gene for ribosomal protein S14 (*rps14*) has been determined. The genes are located in the chloroplast DNA Eco-A fragment within the overlap of fragments HaeII-E and PvuII-E and C (1). The genes are all of the same polarity, organized as *trnI*-39 bp spacer-*rps14*-52 spacer-*trnF*-1 bp spacer-*trnC*. The *trnI* is unusual in having a mismatched base pair (A-A) in the DHU stem and a long variable loop. Although *trnI* has a methionyl-tRNA anticodon (CAU), it shares the most sequence identity (67%) with the spinach isoleucyl-tRNA (also-CAU), which is charged with isoleucine (2). The *Euglena rps14* locus (101 codons) is unique in having a single 106 bp intron (positions 220-325), similar to previously described introns (5). The derived amino acid sequence is 48% and 37% identical to the corresponding sequences from liverwort (3) and *E. coli* (4), respectively.

<i>trnI</i> <u>GCGTCTATAATTTGTAAGGCCATTATGGCAGAGTGACGATAGCACGGGACTCATATACTCGCTCCGGAAAGGACGTCGCTGGTTGAATCCAGCTGAAT</u>	100
<i>rps14</i> . exon 1 <u>GCATCTTATTCGAGCATTATTAACCTAAAAATTTTTATTTGTCAAAAAAAAGCTTATAGCAAGACAGAGAAAACGAAATAATTAGTATAATAC</u>	200
<u>H S K K S L I A R Q R K R I I L V L I</u>	
<u>ATCCCCATAACCGTTATGTATACAGGACAAATGGAAAAGACGAAAAATCTTTGAGAAAAATTGCGGATTACTCTTTTGCAAAAATTACCTAGAA</u>	300
<u>H S H N R Y V Y R T N G K D E K S F E K K L R I Y S F L Q K L P R N</u>	
<u>TAGTTTGCCTTGCCGATTAATTGATTTCTATTGTCATTCAAATTAGCTGTTAATAATTGAAAATTTATAAAATTTTATATTATTTTT</u>	400
<u>S L R C R L</u>	
<i>rps14</i> .exon 2 <u>GAGTATTTTATTTGTTAATCTTCAATCGTTGTTACGTAACAGGGAGATCACGTGGATATTAGGACTTTGGATTATCACGACATATTCTCGA</u>	500
<u>R N R C Y V T G R S R G Y F R T F G L S R H I L R</u>	
<u>GATATGGCTCATTATGGTTACTTCCAGGTAAACCAAGCGAGTTGGAAACTTAAATTACTTATAATAATAAGTATAAAATTATAATAATA</u>	600
<u>D M A H Y G L L P G V T K A S W *</u>	
<i>trnF</i> <u>ATAGCTGGGATAGCTCAGTTGGTAGAGCGGAGGACTGAAATCCTTGTGTCACCAAGTCATCTGGTTCTAGCATGGCGCATGGCCAAAGCGGTAGG</u>	700
<u>CAGAAAGATTGCAAATCTTTATTCGGCAGTTCTGGGTGCGCTTTAAGAGAAATTGAGAAAACAATAAAACAAA</u>	780

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References

1. Cushman, J.C., Hallick, R.B., and Price, C.A. (1988) *Curr. Gen.* 13, 159-179.
2. Kashdan, M.A. and Dudock, B.S. (1982) *J. Biol. Chem.* 257, 11191-11194.
3. Umesono, K., Inokuchi, H., Ohyama, K., and Ozeki, H. (1984) *Nucleic Acids Res.* 12, 9551-9565.
4. Ceretti, D.P., Dean, D., Davis, G.R., Bedwell, D.M., and Nomura, M. (1983) *Nucleic Acids Res.* 11, 2599-2616.
5. Christopher, D.A., Cushman, J.C., Price, C.A., and Hallick, R.B. (1988) *Curr. Genet.* 14, 275-286.