SYMPOSIUM REVIEW

Protective conditioning of the brain: expressway or roadblock?

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Abstract The brain responds to noxious stimulation with protective signalling. Over the last decades, a number of experimental strategies have been established to study endogenous brain protection. Pre-, per-, post- and remote 'conditioning' are now widely used to unravel the underlying mechanisms of endogenous neuroprotection. Some of these strategies are currently being tested in clinical trials to protect the human brain against anticipated damage or to boost protective responses during or after injury. Here we summarize the principles of 'conditioning' research and current efforts to translate this knowledge into effective treatment of patients. Conditioning to induce protected brain states provides an experimental window into endogenous brain protection and can lead to the discovery of drugs mimicking the effects of conditioning. Mechanisms of endogenous brain tolerance can be activated through a wide variety of stimuli that signal 'danger' to the brain. These danger signals lead to the induction of regulator and effector mechanisms, which suppress death and induce survival pathways, decrease metabolism, as well as increase substrate delivery. We conclude that preclinical research on endogenous brain protection has greatly benefited from conditioning strategies, but that clinical applications are challenging, and that we should not prematurely rush into ill-designed and underpowered clinical trials.

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Abbreviations Akt, Akt-kinase; CIDS, CNS injury-induced immunosupression; DAMP, damage-associated molecular patterns; EPO, erythropoeitin; ERK, extracellular signal-regulated kinase; G-CSF, granulocyte colony stimulating factor; HIF-1, hypoxia-inducible factor-1; HK, hexokinase; LDH, lactate dehydrogenase; LPS, lipopolysaccharide; MAPK, mitogen-activated protein kinase; miRNA, micro RNA; mTOR, mammalian target of rapamycin; PAMP, pathogen-associated molecular patterns; PDH, pyruvate dehydrogenase; PDK1, pyruvate dehydrogenase kinase 1; PHD, prolyl-hydroxylase; PI3K, phosphatidylinositol 3-kinases; RCT, randomized controlled trial; ROS, reactive oxygen species; TLR, toll-like receptor; VEGF, vascular endothelial growth factor.

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'That which does not kill us makes us stronger' Friedrich Nietzsche, Ecce Homo – Warum ich so weise bin 2 (1908)

The multiple flavours of 'conditioning'

Cells, tissues, organs, as well as whole organisms respond to sublethal stress by activating protective signalling cascades (Dirnagl*et al.* 2003). In aerobic organisms, strong evolutionary pressure for the development of endogenous mechanisms of protection is generated by hypoxia, as well as by infection and inflammation. Protective responses may be generated either in anticipation of a stressor, during stress against its immediate harmful consequences, or as a response to delayed secondary mechanisms after stress. Various medical disciplines, in particular cardiology, neurology/neurosurgery, anaesthesiology, as well as transplantationmedicine, are studying the pathways underlying this signalling of endogenous protective responses. It is the ultimate goal of this research to develop therapeutic organ protection based upon nature's own strategies (Dirnagl *et al.* 2009; Keep *et al.* 2010; Gidday, 2010). Earlymilestones in this quest were the discoveries that pre-exposure to hypoxia can prolong anoxic survival by preserving brain metabolism (Dahl *et al.* 1964); that brain can adapt to anoxia by hypoxic pre-exposure (Schurr *et al.* 1986); and the description of ischaemic preconditioning in ischaemic myocardium (Murry *et al.* 1986) and brain (Kitagawa *et al.* 1990). Since then, various types of organ 'conditioning' have been described (Fig. 1): *preconditioning*, in which the conditioning stimulus (e.g. ischaemia, hypoxia, metabolic inhibition or inflammation below the threshold of damage) is given several days ('delayed preconditioning') or minutes ('classic preconditioning') before a noxious stimulus presents (e.g. ischaemia); *perconditioning*, in which the conditioning stimulus is given while the noxious stimulus is still present; *postconditioning*, in which the conditioning stimulus is given shortly after the noxious stimulus (e.g. after reperfusion), and *remote conditioning*, in which not the organ which is affected by the noxious stimulus is conditioned, but another, remote organ or bodily system (e.g. limb ischaemia to induce protection of heart or brain). Robust experimental protocols have been developed for each of these types of conditioning, and a number of the underpinning signalling pathways have been established (Kirino, 2002; Gidday, 2006; Dirnagl & Meisel, 2008; Obrenovitch, 2008; Zhao, 2009; and see below). Some of these conditioning strategies are either directly applicable to patients (e.g. remote conditioning by limb ischaemia), or can be pharmacologically mimicked, such as prolyl-hydroxylase (PHD) inhibitors activating hypoxia inducible factor (HIF)-related pathways, or growth and survival factors such as erythropoeitin (EPO) or granulocyte colony stimulating factor (G-CSF). A number of clinical trials have been concluded in cardiology, and several are underway in patients with brain

disease (see below and Table 1). At least in cardiology, where clinical development of conditioning-related strategies is most advanced, their translation into effective therapies has so far been hugely disappointing (Ludman *et al.* 2010). It is therefore timely to ask what we know about the mechanisms underlying brain conditioning, and what the chances are that brain conditioning will become a clinical reality in the near future.

A window into endogenous brain protection

Pre-, per-, post-, remote as well as pharmacological ('mimics') conditioning serve as highly valuable tools to understand the mechanisms of endogenous brain protection. It appears that these mechanisms are independent of the conditioning strategy, or have at least a vast overlap. This reflects the fact that these mechanisms have evolved as unspecific responses to a number of challenges to the organism (hypoxia, injury, infection). In the following we discuss them briefly and without reference to specific conditioning strategies (Fig. 2).

Sensors of danger. Mechanisms of endogenous brain tolerance can be activated through a wide variety of stimuli that signal 'danger' to the brain. Both hypoxia and infection endanger the entire organism and are fundamental challenges for most organisms and hence for organs and their cells. Therefore, multiple redundant cascades to adapt to these conditions have evolved. Many aspects of (anti-)inflammatory and hypoxic signalling overlap, and cascades mediating endogenous tolerance are very similar in different organs. For comprehensive overviews the reader is referred to Dirnagl*et al.*(2003); Gidday (2006); Dirnagl & Meisel (2008); Obrenovitch (2008), and the references therein.

Hypoxia-inducible factor-1 (HIF-1) is the key regulator of cellular oxygen homeostasis. Under hypoxic conditions HIF-1 activates highly conserved transcriptional profiles ultimately geared to adapt cellular homeostasis to reduced oxygen availability. Among others, these adaptations include changes in cellular energy metabolism, regulation of Bcl2-family proteins, cell proliferation, cell cycle control and vasomotor control or angiogenesis (Sharp & Bernaudin, 2004; Semenza, 2009). Likewise, toll-like receptors (TLRs), which are present on many if not all mammalian cells, are responsible for inducing cellular responses to counteract infection.

HIF-1 induces adaptation to decreased oxygen on two levels. First, cellular homeostasis, including mitochondrial respiration, is adapted to hypoxic conditions rather quickly (within hours, Semenza, 2010). Second, chronic hypoxia induces angiogenesis to increase blood supply to hypoxic tissue through a variety of HIF-1-dependent factors such as vascular endothelial growth factor (VEGF;

Table 1. Examples of currently recruiting clinical trials with neurological endpoints as listed at ClinicalTrials.gov (as of 3/2011): preconditioning, remote conditioning and agents that mimic endogenous neuroprotection

Stuttering reperfusion

Pharmacological mimic

Figure 1. 'Conditioning' paradigms to protect the brain

Typically, preconditioning uses a sublethal stimulus given minutes or days before the insult against which it aims to protect. Stuttering reperfusion is the prototypical per- or postconditioning strategy, by which one aims to prevent 'reperfusion damage' by repetitively opening and blocking brain perfusion before permanent reperfusion is allowed. Remote ischaemia is another per- or postconditioning strategy which typically produces repetitive, short phases of ischaemia of a peripheral limb to induce humoral and neural mechanisms of protection of a remote organ, such as the brain. Pharmacological mimics are drugs that either boost endogenous protective signalling cascades (such as the HIF pathway), or exogenously provide the effectors of endogenous protection, such as EPO.

Semenza, 2009). In general, under normoxic conditions the HIF-1 α subunit is targeted to rapid proteasomal degradation through post-translation modification by prolyl-hydroxylation, whereas under hypoxic conditions HIF-1 α is stabilized and HIF-1-dependent transcription is initialized (Sharp & Bernaudin, 2004; Semenza, 2009). Furthermore, transcription of HIF-1 α is increased upon growth-factor signalling, in particular upon activation of the PI3K–Akt–mTOR (mTOR, mammalian target of

Figure 2. General principles of action of 'conditioning strategies' to protect the brain

A pre-, per-, post-, remote-conditioning stimulus may either: directly protect the brain via release of locally or remotely acting metabolites (e.g. adenosine); after activation of sensors (e.g. HIF-1) lead to a complex signalling cascade which may include genetic as well as epigenetic responses; or activate genetic and epigenetic responses via neuronal pathways (e.g. activating the sympathetic nervous system or the hypothalamic–pituitary axis). The signalling pathways of the various conditioning strategies may converge in similar or even identical effector mechanisms, such as suppressed death pathways, induced survival pathways, decreased metabolism ('hibernation'), and increased substrate delivery.

rapamycin) pathway (DeBerardinis *et al.* 2008). While the brain has developed some unique sensors of systemic hypoxia, such as central and arterial chemoreceptors (Sharp & Bernaudin, 2004), the molecular cascades involved in oxygen sensing are highly conserved in all cell types (Sharp & Bernaudin, 2004; Semenza, 2009, 2010).

TLRs are an integral part of the innate immune system, providing the first line of defence against pathogens at the cellular level. TLR signalling is an important mediator of ischaemic damage in the brain, but it can also mediate inflammation-induced cross tolerance such as through stimulation with lipopolysaccharide (LPS) or tumour necrosis factor-α (TNF-α) (Marsh *et al.* 2009). In general, TLRs activate transcription factors through common intracellular pathways, with distinct effects in different cell types or tissues (Marsh *et al.* 2009). TLRs are a major discriminator between 'self' and 'foreign' (Akira & Takeda, 2004). A signalling cascade resulting in activation of nuclear factor NF-κB transcription and an inflammatory response are initiated following ligation of pathogen-associated molecular patterns (PAMPs) with TLRs (Liew *et al.* 2005). Host-derived damage-associated molecular patterns (DAMPs), which are released upon ischaemic injury (Vartanian & Stenzel-Poore, 2010), can also induce inflammatory signalling through the TLR pathway (Seong & Matzinger, 2004) and contribute to ischaemic damage in the brain (Kariko *et al.* 2004; Lehnardt *et al.* 2007; Ziegler *et al.* 2007; Dirnagl *et al.* 2009). In turn, inflammatory signalling can boost HIF-1 transcription, which controls many genes involved in regulation of inflammation and host defence (Nizet & Johnson, 2009) and which has been found to be essential for the cellular innate immune response in inflammation (Cramer *et al.* 2003). Furthermore, mitochondria are very sensitive to changes in homeostasis, and are important sensors of cellular stress (see below). TLRs in the brain are constitutively expressed in astrocytes, microglia and endothelial cells. TLRs can be upregulated upon inflammatory stimulation in these cells, but also in neurons and oligodendrocytes (Marsh *et al.* 2009). Little is known about the differential role of the various TLR types and cell types which express those TLRs in mediating endogenous neuroprotection.

Regulators and effectors. Hypoxia induces substantial changes in gene expression patterns in the brain (Bernaudin *et al.* 2002). Preconditioning is thought to reprogram the brain's genomic response to a noxious stimulus (Stenzel-Poore *et al.* 2007). For example, many of the HIF-1 target genes are involved in regulating cellular metabolism, survival, proliferation and angiogenesis (Semenza, 2009). Furthermore, increasing evidence suggests that different epigenetic regulatory mechanisms are activated in the context of conditioning paradigms, and regulate the endogenous protective response. Inhibition of DNA methylation and histone deacetylation reduce ischaemic damage by altering the transcriptional profile (Endres *et al.* 2000; Meisel *et al.* 2006; Yildirim *et al.* 2008). Micro RNAs (miRNAs), small RNA molecules that function as post-transcriptional regulators of gene expression (Lagos-Quintana *et al.* 2001), are important regulators of diverse aspects of brain function, including development and maintenance of brain plasticity (Saugstad, 2010). Furthermore,miRNAs have emerged as important mediators of endogenous tolerance in the brain (Dharap & Vemuganti, 2010; Lusardi *et al.* 2010), and both HIF-1 and TLR signalling can be modulated by miRNAs (Crosby *et al.* 2009; O'Neill *et al.* 2011).

A variety of kinases involved in proliferation and survival are involved in eliciting an endogenous protective response in the brain. Among others, these include protein kinase C (PKC) (Speechly-Dick *et al.* 1994), mitogen-activated protein kinase (MAPK)/p38, extracellular signal-regulated kinase (ERK), Akt-kinase (Ruscher *et al.* 2002; Gao *et al.* 2008) and mTOR (Pagel, 2008; Swiech *et al.* 2008). The PI3K–Akt–mTOR pathway senses nutrient availability. Activation of this pathway leads to increased transcription of HIF-1 α , further supporting adaptation of metabolism to substrate deprivation (DeBerardinis *et al.* 2008; Swiech *et al.* 2008).

Ischaemia is associated with profound metabolic imbalances and much of the cellular response initiated by conditioning events is geared to alter metabolic pathways to maintain basal metabolic integrity. Therefore, under hypoxia, glycolytic flux is diverted from oxidative phosphorylation to glycolysis. HIF-1 changes the expression of genes of the entire glycolytic cascade to adapt metabolism to hypoxic conditions (Iyer *et al.* 1998; Semenza, 2009). As a consequence, the glycolytic intermediate pyruvate is shunted away from the mitochondrial tricarboxylic acid (TCA) cycle by pyruvate dehydrogenase kinase 1 (PDK1). PDK1 inhibits pyruvate dehydrogenase (PDH), thereby reducing flux through the TCA cycle and ultimately reducing excess production of reactive oxygen species (ROS) (Kim *et al.* 2006; Papandreou *et al.* 2006). In addition, lactate dehydrogenase A (LDHA) converts pyruvate to lactate, which in the brain can be rapidly taken up and shuttled away by astrocytes (Gandhi *et al.* 2009). HIF-1 control over metabolism is not only limited to states of hypoxia, but appears to be of general relevance for survival and proliferation (DeBerardinis *et al.* 2008). Furthermore, glycolytic enzymes such as mitochondrial hexokinase (HK) or glyceraldehyde-3-phosphate dehydrogenase (GAPDH) are emerging as important regulators of cell death (Majewski *et al.* 2004; Kim & Dang, 2005; Colell *et al.* 2007), with striking mechanistic similarities between neurons and cancer cells (Vaughn & Deshmukh, 2008).

Mitochondria, equipped to efficiently generate ATP through oxidative phosphorylation, also function as oxygen sensors by inhibiting PHD activity through ROS production, thereby stabilizing HIF-1 α . Mitochondria are highly susceptible to changes in oxygen concentration and abruptly react by generating ROS (Kaelin, 2005; Kim *et al.* 2006; Klimova & Chandel, 2008; Semenza, 2010). Importantly, ROS signalling not only contributes to ischaemic damage, but is also involved in endogenous protection evoked by pre-, per-, post- and remote conditioning (Gidday, 2006; Tapuria *et al.* 2008; Hausenloy, 2009; Ovize *et al.* 2010; Saxena *et al.* 2010; Semenza, 2010; Xin *et al.* 2010).

As described above, inflammation mediated by the innate immune system as well as by the adaptive immune system (Yilmaz *et al.* 2006; Hurn *et al.* 2007; Liesz *et al.* 2011) contributes to brain injury following stroke. However, inflammatory stimulation using LPS can also induce endogenous tolerance (Bastide *et al.* 2003; Kunz *et al.* 2007; Orio *et al.* 2007). Additionally, cerebral ischaemia, as well as other insults to the central nervous system, lead to immunosuppression – a phenomenon termed CNS injury-induced immunosupression (CIDS) (Meisel*et al.* 2005). CIDS might therefore serve to contain an autoaggressive immune response following stroke (Gee *et al.* 2007). In humans, ischaemic preconditioning by transient forearm ischaemia changes gene expression patterns in circulating leukocytes, thereby suppressing leukocyte activation and potentially modulating innate and adaptive immune responses (Konstantinov *et al.* 2004; Saxena *et al.* 2010).

Clinical applications: expressway or roadblock?

The discovery of ischaemic tolerance, the wealth of knowledge that has subsequently been gathered on mechanisms of endogenous organ protection, and the development of clinically applicable strategies of pre-, per- and postconditioning have precipitated a rush into clinical trials in cardiology, nephrology, anaesthesiology and neurology/neurosurgery, among other disciplines. Some of these rather small proof of concept trials, often using surrogate endpoints, have generated promising results (e.g. Chan *et al.* 2005; Bøtker *et al.* 2010; Lonborg *et al.* 2010; Schäbitz *et al.* 2010). For the brain, a putative beneficial effect of transient ischaemic attacks (as 'preconditioning equivalents') suggested the existence of endogenous neuroprotection in the human brain (Weih *et al.* 1999; Wegener *et al.* 2004), although this has been disputed (Johnston, 2004). Meanwhile, however, a number of negative or inconclusive randomized clinical trials (RCT, e.g. Hong *et al.* 2010; Rahman *et al.* 2010; Walsh *et al*. 2010) have been published on various forms of conditioning in several organ systems, including

heart and brain, and a less optimistic spirit prevails (Ludman *et al*. 2010). This is reminiscent of 'classical' neuroprotection trials, where promising preclinical and early clinical testing was not followed by evidence for efficacy in large RCTs (Green, 2008; Tymianski, 2010). Nevertheless, for conditioning strategies the jury is still out: the search term 'preconditioning OR perconditioning OR 'remote conditioning' AND brain' reveals 50 open clinical trials in the trial registry of the National Institute of Health (clinicaltrials.gov). This does not include RCTs testing pharmacological strategies of endogenous neuroprotection such as granulocyte-colony stimulating factor (G-CSF), AX200 for the treatment of ischaemic stroke (AXIS-2, NCT00927836) or erythropoietin (Safety Study of Carbamylated Erythropoietin (CEPO) to Treat Patients With Acute Ischemic Stroke, NCT00756249, publication of results pending). Table 1 lists a selection of currently recruiting RCTs with neurological endpoints in the field of 'conditioning' or endogenous neuroprotection.

It has been argued that in contrast to conventional neuroprotection trials, in which neuroprotective drugs are given after the ischaemic event, preconditioning strategies allow baseline assessment of neurological status before patients experience the index event: patients can be functionally tested before preconditioning them against an index event, such as focal neurological deficits after carotid or heart surgery, or delayed neurological deficits after subarachnoid haemorrhage. This may help cut down the variance in results and reduce the number of patients to recruit (Dirnagl *et al.* 2009), which for a Phase III neuroprotection trial in stroke may run into the thousands. However, the downside of this approach is that only a fraction of patients experience the index event (e.g. around 1% of strokes after coronary bypass surgery), potentially annihilating the advantage of an individual baseline and leading to the exposure of patients to possibly harmful treatments they do not actually need.

Outlook

Basic research on endogenous mechanisms has established a plethora of conditioning strategies and unravelled, among others, neurogenic, immunological, genetic and epigenetic mechanisms of brain protection. Nevertheless, many issues remain unsolved, including questions such as how remote preconditioning exerts its effects (humoral? neuronal?), or whether the dogma that the conditioning stimulus is subthreshold to damage is really true (it has been proposed that in many cases, damage was simply not assessed, or the tools were not sensitive enough; Dirnagl *et al.* 2003; Sommer, 2008). In the current clinical arena, many teams worldwide are testing the safety and efficacy of such diverse strategies as the prevention or amelioration of CNS damage when it can be anticipated (e.g. delayed vasospasm after subarachnoid haemorrhage), the prevention of CNS damage during potentially harmful interventions (e.g. neurosurgery), the induction of endogenous CNS protection by remote procedures (e.g. repeated limb ischaemia after acute stroke), or the pharmacological induction of endogenous CNS protection (e.g. HIF-1 induction via Desferoxamine) (see Table 1). Over the next few years some of those RCTs may provide evidence not only for the existence of endogenous neuroprotection, but also as to whether related mechanisms can be therapeutically exploited to benefit patients at risk for or with evolving CNS damage. Given the complexities and challenges of the underlying pathophysiology, as well as the design and implementation of clinical trials, and given the frustrating experiences regarding neuroprotection in the stroke field, we are well advised to learn from previous mistakes and to conduct preclinical research of the highest quality (Dirnagl, 2006) and not to prematurely rush into ill-designed and underpowered clinical trials (Weaver *et al.* 2004).

References

- Akira S & Takeda K (2004). Toll-like receptor signalling. *Nat Rev Immunol* **4**, 499–511.
- Bastide M, Gele P, Petrault O, Pu Q, Caliez A, Robin E, Deplanque D, Duriez P & Bordet R (2003). Delayed cerebrovascular protective effect of lipopolysaccharide in parallel to brain ischemic tolerance. *J Cereb Blood Flow Metab* **23**, 399–405.
- Bernaudin M, Tang Y, Reilly M, Petit E & Sharp FR (2002). Brain genomic response following hypoxia and re-oxygenation in the neonatal rat. Identification of genes that might contribute to hypoxia-induced ischemic tolerance. *J Biol Chem* **277**, 39728–39738.
- Bøtker HE, Kharbanda R, Schmidt MR, Bøttcher M, Kaltoft AK, Terkelsen CJ *et al*. (2010). Remote ischaemic conditioning before hospital admission, as a complement to angioplasty, and effect on myocardial salvage in patients with acute myocardial infarction: a randomised trial. *Lancet* **375**, 727–734.
- Chan MT, Boet R, Ng SC, Poon WS & Gin T (2005). Effect of ischemic preconditioning on brain tissue gases and pH during temporary cerebral artery occlusion. *Acta Neurochir Suppl* **95**, 93–96.
- Colell A, Ricci JE, Tait S, Milasta S, Maurer U, Bouchier-Hayes L et al. (2007). GAPDH and autophagy preserve survival after apoptotic cytochrome c release in the absence of caspase activation. *Cell* **129**, 983–997.
- Cramer T, Yamanishi Y, Clausen BE, Forster I, Pawlinski R, Mackman N *et al*. (2003). HIF-1α is essential for myeloid cell-mediated inflammation. *Cell* **112**, 645–657.
- Crosby ME, Devlin CM, Glazer PM, Calin GA & Ivan M (2009). Emerging roles of microRNAs in the molecular responses to hypoxia. *Curr Pharm Des* **15**, 3861–3866.
- Dahl NA & Balfour WM (1964). Prolonged anoxic survival due to anoxia preexposure: brain ATP, lactate, and pyruvate. *Am J Physiol* **1**, 452–456.

DeBerardinis RJ, Lum JJ, Hatzivassiliou G & Thompson CB (2008). The biology of cancer: metabolic reprogramming fuels cell growth and proliferation. *Cell Metab* **7**, 11–20.

Dharap A & Vemuganti R (2010). Ischemic pre-conditioning alters cerebral microRNAs that are upstream to neuroprotective signaling pathways. *J Neurochem* **113**, 1685–1691.

Dirnagl U (2006). Bench to bedside: the quest for quality in experimental stroke research. *J Cereb Blood Flow Metab* **26**, 1465–1478.

Dirnagl U, Becker K & Meisel A (2009). Preconditioning and tolerance against cerebral ischaemia: from experimental strategies to clinical use. *Lancet Neurol* **8**, 398–412.

Dirnagl U & Meisel A (2008). Endogenous neuroprotection: mitochondria as gateways to cerebral preconditioning? *Neuropharmacology* **55**, 334–344.

Dirnagl U, Simon RP & Hallenbeck JM (2003). Ischemic tolerance and endogenous neuroprotection. *Trends Neurosci* **26**, 248–254.

Endres M, Meisel A, Biniszkiewicz D, Namura S, Prass K, Ruscher K *et al*. (2000). DNA methyltransferase contributes to delayed ischemic brain injury. *J Neurosci* **20**, 3175–3181.

Gandhi GK, Cruz NF, Ball KK & Dienel GA (2009). Astrocytes are poised for lactate trafficking and release from activated brain and for supply of glucose to neurons. *J Neurochem* **111**, 522–536.

Gao X, Zhang H, Takahashi T, Hsieh J, Liao J, Steinberg GK & Zhao H (2008). The Akt signaling pathway contributes to postconditioning's protection against stroke; the protection is associated with the MAPK and PKC pathways. *J Neurochem* **105**, 943–955.

Gee JM, Kalil A, Shea C & Becker KJ (2007). Lymphocytes: potential mediators of postischemic injury and neuroprotection. *Stroke* **38**, 783–788.

Gidday JM (2006). Cerebral preconditioning and ischaemic tolerance. *Nat Rev Neurosci* **7**, 437–448.

Gidday JM (2010). Pharmacologic preconditioning: translating the promise. *Transl Stroke Res* **1**, 19–30.

Green AR (2008). Pharmacological approaches to acute ischaemic stroke: reperfusion certainly, neuroprotection possibly. *Br J Pharmacol* **153**, S325–S338.

Hausenloy DJ (2009). Signalling pathways in ischaemic postconditioning. *Thromb Haemost* **101**, 626–634.

Hong DM, Mint JJ, Kim JH, Sohn IS, Lim TW, Lim YJ *et al*. (2010). The effect of remote ischaemic preconditioning on myocardial injury in patients undergoing off-pump coronary artery bypass graft surgery. *Anaesth Intensive Care* **38**, 924–929.

Hurn PD, Subramanian S, Parker SM, Afentoulis ME, Kaler LJ, Vandenbark AA & Offner H (2007). T- and B-cell-deficient mice with experimental stroke have reduced lesion size and inflammation. *J Cereb Blood Flow Metab* **27**, 1798–1805.

Iyer NV, Kotch LE, Agani F, Leung SW, Laughner E, Wenger RH *et al.* (1998). Cellular and developmental control of O₂ homeostasis by hypoxia-inducible factor 1α. *Genes Dev* **12**, 149–162.

Johnston SC (2004). Ischemic preconditioning from transient ischemic attacks? Data from the Northern California TIA Study. *Stroke* **35**, 2680–2682.

Kaelin WG Jr (2005). ROS: really involved in oxygen sensing. *Cell Metab* **1**, 357–358.

Kariko K, Weissman D & Welsh FA (2004). Inhibition of toll-like receptor and cytokine signaling – a unifying theme in ischemic tolerance. *J Cereb Blood Flow Metab* **24**, 1288–1304.

Keep RF, Wang MM, Xiang J, Hua Y & Xi G (2010). Is there a place for cerebral preconditioning in the clinic? *Transl Stroke Res* **1**, 4–18.

Kim JW & Dang CV (2005). Multifaceted roles of glycolytic enzymes. *Trends Biochem Sci* **30**, 142–150.

Kim JW, Tchernyshyov I, Semenza GL & Dang CV (2006). HIF-1-mediated expression of pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to hypoxia. *Cell Metab* **3**, 177–185.

Kirino T (2002). Ischemic tolerance. *J Cereb Blood Flow Metab* **22**, 1283–1296.

Kitagawa K, Matsumoto M, Tagaya M, Hata R, Ueda H, Niinobe M *et al*. (1990). 'Ischemic tolerance' phenomenon found in the brain. *Brain Res* **528**, 21–24.

Klimova T & Chandel NS (2008). Mitochondrial complex III regulates hypoxic activation of HIF. *Cell Death Differ* **15**, 660–666.

Konstantinov IE, Arab S, Kharbanda RK, Li J, Cheung MM, Cherepanov V, Downey GP *et al*. (2004). The remote ischemic preconditioning stimulus modifies inflammatory gene expression in humans. *Physiol Genomics* **19**, 143–150.

Kunz A, Park L, Abe T, Gallo EF, Anrather J, Zhou P & Iadecola C (2007). Neurovascular protection by ischemic tolerance: role of nitric oxide and reactive oxygen species. *J Neurosci* **27**, 7083–7093.

Lagos-Quintana M, Rauhut R, Lendeckel W & Tuschl T (2001). Identification of novel genes coding for small expressed RNAs. *Science* **294**, 853–858.

Lehnardt S, Lehmann S, Kaul D, Tschimmel K, Hoffmann O, Cho S *et al*. (2007). Toll-like receptor 2 mediates CNS injury in focal cerebral ischemia. *J Neuroimmunol* **190**, 28–33.

Liesz A, Zhou W, Mracsko E, Karcher S, Bauer H, Schwarting S *et al*. (2011). Inhibition of lymphocyte trafficking shields the brain against deleterious neuroinflammation after stroke. *Brain* **134**, 704–720.

Liew FY, Xu D, Brint EK & O'Neill LA (2005). Negative regulation of toll-like receptor-mediated immune responses. *Nat Rev Immunol* **5**, 446–458.

Lønborg J, Holmvang L, Kelbæk H, Vejlstrup N, Jørgensen E, Helqvist S *et al*. (2010). ST-segment resolution and clinical outcome with ischemic postconditioning and comparison to magnetic resonance. *Am Heart J* **160**, 1085–1091.

Ludman AJ, Yellon DM & Hausenloy DJ (2010). Cardiac preconditioning for ischaemia: lost in translation. *Dis Model Mech* **3**, 35–38.

Lusardi TA, Farr CD, Faulkner CL, Pignataro G, Yang T, Lan J, Simon RP & Saugstad JA (2010). Ischemic preconditioning regulates expression of microRNAs and a predicted target, MeCP2, in mouse cortex. *J Cereb Blood Flow Metab* **30**, 744–756.

Majewski N, Nogueira V, Bhaskar P, Coy PE, Skeen JE, Gottlob K *et al*. (2004). Hexokinase-mitochondria interaction mediated by Akt is required to inhibit apoptosis in the presence or absence of Bax and Bak. *Mol Cell* **16**, 819–830.

Marsh BJ, Williams-Karnesky RL & Stenzel-Poore MP (2009). Toll-like receptor signaling in endogenous neuroprotection and stroke. *Neuroscience* **158**, 1007–1020.

Meisel A, Harms C, Yildirim F, Bosel J, Kronenberg G, Harms U, Fink KB & Endres M (2006). Inhibition of histone deacetylation protects wild-type but not gelsolin-deficient neurons from oxygen/glucose deprivation. *J Neurochem* **98**, 1019–1031.

Meisel C, Schwab JM, Prass K, Meisel A & Dirnagl U (2005). Central nervous system injury-induced immune deficiency syndrome. *Nat Rev Neurosci* **6**, 775–786.

Murry, CE, Jennings RB & Reimer KA (1986). Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation* **74**, 1124–1136.

Nizet V & Johnson RS (2009). Interdependence of hypoxic and innate immune responses. *Nat Rev Immunol* **9**, 609–617.

Obrenovitch TP (2008). Molecular physiology of preconditioning-induced brain tolerance to ischemia. *Physiol Rev* **88**, 211–247.

O'Neill LA, Sheedy FJ & McCoy CE (2011). MicroRNAs: the fine-tuners of Toll-like receptor signalling. *Nat Rev Immunol* **11**, 163–175.

Orio M, Kunz A, Kawano T, Anrather J, Zhou P & Iadecola C (2007). Lipopolysaccharide induces early tolerance to excitotoxicity via nitric oxide and cGMP. *Stroke* **38**, 2812–2817.

Ovize M, Baxter GF, Di Lisa F, Ferdinandy P, Garcia-Dorado D, Hausenloy DJ *et al*. (2010). Postconditioning and protection from reperfusion injury: where do we stand? Position paper from the Working Group of Cellular Biology of the Heart of the European Society of Cardiology. *Cardiovasc Res* **87**, 406–423.

Pagel PS (2008). Postconditioning by volatile anesthetics: salvaging ischemic myocardium at reperfusion by activation of prosurvival signaling. *J Cardiothorac Vasc Anesth* **22**, 753–765.

Papandreou I, Cairns RA, Fontana L, Lim AL & Denko NC (2006). HIF-1 mediates adaptation to hypoxia by actively downregulating mitochondrial oxygen consumption. *Cell Metab* **3**, 187–197.

Rahman IA, Mascaro JG, Steeds RP, Frenneaux MP, Nightingale P, Gosling P *et al*. (2010). Remote ischemic preconditioning in human coronary artery bypass surgery: from promise to disappointment? *Circulation* **14**, **122**, S53–S59.

Ruscher K, Freyer D, Karsch M, Isaev N, Megow D, Sawitzki B *et al*. (2002). Erythropoietin is a paracrine mediator of ischemic tolerance in the brain: evidence from an *in vitro* model. *J Neurosci* **22**, 10291–10301.

Saugstad JA (2010). MicroRNAs as effectors of brain function with roles in ischemia and injury, neuroprotection, and neurodegeneration. *J Cereb Blood Flow Metab* **30**, 1564–1576.

Saxena P, Newman MA, Shehatha JS, Redington AN & Konstantinov IE (2010). Remote ischemic conditioning: evolution of the concept, mechanisms, and clinical application. *J Card Surg* **25**, 127–134.

Schabitz WR, Laage R, Vogt G, Koch W, Kollmar R, Schwab S ¨ *et al*. (2010). AXIS: a trial of intravenous granulocyte colony-stimulating factor in acute ischemic stroke. *Stroke* **41**, 2545–2551.

Schurr A, Reid KH, Tseng MT, West C & Rigor MB (1986). Adaptation of adult brain tissue to anoxia and hypoxia in vitro. *Brain Res* **374**, 244–248.

Semenza GL (2009). Regulation of oxygen homeostasis by hypoxia-inducible factor 1. *Physiology (Bethesda)* **24**, 97–106.

Semenza GL (2010). Hypoxia-inducible factor 1: regulator of mitochondrial metabolism and mediator of ischemic preconditioning. *Biochim Biophys Acta* **1813**, 1263–1268.

Seong SY & Matzinger P (2004). Hydrophobicity: an ancient damage-associated molecular pattern that initiates innate immune responses. *Nat Rev Immunol* **4**, 469–478.

Sharp FR & Bernaudin M. (2004). HIF1 and oxygen sensing in the brain. *Nat Rev Neurosci* **5**, 437–448.

Sommer C (2008). Ischemic preconditioning: postischemic structural changes in the brain. *J Neuropathol Exp Neurol* **67**, 85–92.

Speechly-Dick ME, Mocanu MM & Yellon DM (1994). Protein kinase C. Its role in ischemic preconditioning in the rat. *Circ Res* **75**, 586–590.

Stenzel-Poore MP, Stevens SL, King JS & Simon RP (2007). Preconditioning reprograms the response to ischemic injury and primes the emergence of unique endogenous neuroprotective phenotypes: a speculative synthesis. *Stroke* **38**, 680–685.

Swiech L, Perycz M, Malik A & Jaworski J (2008). Role of mTOR in physiology and pathology of the nervous system. *Biochim Biophys Acta* **1784**, 116–132.

Tapuria N, Kumar Y, Habib MM, Abu Amara M, Seifalian AM & Davidson BR (2008). Remote ischemic preconditioning: a novel protective method from ischemia reperfusion injury – a review. *J Surg Res* **150**, 304–330.

Tymianski M (2010). Can molecular and cellular neuroprotection be translated into therapies for patients?: yes, but not the way we tried it before. *Stroke* **41**, S87–S90.

Vartanian K & Stenzel-Poore M (2010). Toll-like receptor tolerance as a mechanism for neuroprotection. *Transl Stroke Res* **1**, 252–260.

Vaughn AE & Deshmukh M (2008). Glucose metabolism inhibits apoptosis in neurons and cancer cells by redox inactivation of cytochrome c. *Nat Cell Biol* **10**, 1477–1483.

Walsh SR, Sadat U, Boyle JR, Tang TY, Lapsley M, Norden AG & Gaunt ME (2010). Remote ischemic preconditioning for renal protection during elective open infrarenal abdominal aortic aneurysm repair: randomized controlled trial. *Vasc Endovascular Surg* **44**, 334–340.

Weaver CS, Leonardi-Bee J, Bath-Hextall FJ & Bath PM (2004). Sample size calculations in acute stroke trials: a systematic review of their reporting, characteristics, and relationship with outcome. *Stroke* **35**, 1216–1224.

- Wegener S, Gottschalk B, Jovanovic V, Knab R, Fiebach JB, Schellinger PD *et al*. (2004). MRI in Acute Stroke Study Group of the German Competence Network Stroke.
- Transient ischemic attacks before ischemic stroke: preconditioning the human brain? A multicenter magnetic resonance imaging study. *Stroke* **35**, 616–621.
- Weih M, Kallenberg K, Bergk A, Dirnagl U, Harms L, Wernecke KD & Einhäupl KM (1999). Attenuated stroke severity after prodromal TIA: a role for ischemic tolerance in the brain? *Stroke* **30**, 1851–1854.
- Xin P, Zhu W, Li J, Ma S, Wang L, Liu M, Wei M & Redington AN (2010). Combined local ischemic postconditioning and remote perconditioning recapitulate cardioprotective effects of local ischemic preconditioning. *Am J Physiol Heart Circ Physiol* **298**, H1819–H1831.
- Yildirim F, Gertz K, Kronenberg G, Harms C, Fink KB, Meisel A & Endres M (2008). Inhibition of histone deacetylation protects wildtype but not gelsolin-deficient mice from ischemic brain injury. *Exp Neurol* **210**, 531–542.
- Yilmaz G, Arumugam TV, Stokes KY & Granger DN (2006). Role of T lymphocytes and interferon-gamma in ischemic stroke. *Circulation* **113**, 2105–2112.
- Zhao H (2009). Ischemic postconditioning as a novel avenue to protect against brain injury after stroke. *J Cereb Blood Flow Metab* **29**, 873–885.
- Ziegler G, Harhausen D, Schepers C, Hoffmann O, Rohr C, Prinz V *et al*. (2007). TLR2 has a detrimental role in mouse transient focal cerebral ischemia. *Biochem Biophys Res Commun* **359**, 574–579.

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