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Risk factors for neoplastic progression in Barrett's esophagus

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Abstract

Barrett's esophagus (BE) confers a significant increased risk for development of esophageal adenocarcinoma (EAC), with the pathogenesis appearing to progress through a "metaplasia-dysplasia-carcinoma" (MDC) sequence. Many of the genetic insults driving this MDC sequence have recently been characterized, providing targets for candidate biomarkers with potential clinical utility to stratify risk in individual patients. Many clinical risk factors have been investigated, and associations with a variety of genetic, specific gastrointestinal and other modifiable factors have been proposed in the literature. This review summarizes the current understanding of the mechanisms involved in neoplastic progression of BE to EAC and critically appraises the relative roles and contributions of these putative risk factors from the published evidence currently available.

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Key words: Barrett's esophagus; Esophageal adeno-

carcinoma; Metaplasia-dysplasia-carcinoma; Neoplastic progression; Risk factors

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INTRODUCTION

Barrett's esophagus (BE) describes a condition where native esophageal stratified squamous epithelium is replaced by metaplastic columnar epithelium, with cephalad displacement of the squamocolumnar junction. BE represents the only identified precursor lesion and most important risk factor for esophageal adenocarcinoma (EAC)^[1]. Patients with BE have an estimated 30- to 125-fold greater risk of developing EAC than the general population^[2]. A systematic review of 27 studies suggested annual progression rates of 0.5%^[3], whereas a review of 8 UK studies by Jankowski *et al*^[4] showed cancer risk of 1.0% per year.

BE PATHOGENESIS AND MECHANISMS OF NEOPLASTIC PROGRESSION

BE is an acquired condition where healing from esophageal mucosal injury [typically triggered by gastro-esophageal reflux disease (GERD)] is metaplastic, with replacement of damaged squamous cells by columnar epithelium. Ordinarily, esophageal healing involves regeneration of squamous cells; it remains unclear why the response is metaplastic in some individuals, since only a minority of patients with GERD develop BE. Progression of BE to EAC occurs by a metaplasia-dysplasia-carcinoma (MDC) sequence. Metaplastic columnar epithelial cells are predisposed to genetic damage with potential for developing

dysplasia^[5]. Dysplasia represents a histological spectrum from low- to high-grade, defined by degree of cytological and architectural disruption present, with genetic instability resulting in progressive acquisition of genetic abnormalities towards a frankly neoplastic phenotype. These can be considered within the framework of Hanahan and Weinberg's^[6] model of "cancer hallmarks" necessary for carcinogenesis, whereby cancer cells must acquire growth self-sufficiency, insensitivity to anti-growth signals, avoidance of apoptosis, limitless replicative potential, sustained angiogenesis, and invasive and metastatic potential^[7].

Many genetic insults conferring these advantages in the BE MDC sequence have been characterized. Initiating events probably involve genes regulating cell cycle progression, notably *p16*. Mutations, loss of heterozygosity (LOH) or promoter hypermethylation (i.e. silencing) of *p16* have been identified in 80% of BE, whilst *p16* hypermethylation correlated with the degree of dysplasia in some studies^[8]. Additional changes identified include upregulation of cyclins D1 and E, transforming growth factor- α and epidermal growth factor (EGF), each contributing towards growth autonomy^[9,10]. These mutations should trigger apoptosis *via* *p53*-dependent pathways. However, subsequent accrual of *p53* lesions confers resistance to apoptosis, and has been identified in 52%-93% of EACs (compared with 1%-5% non-malignant BE cell lines)^[11]. Inactivation of *p53* increases clonal genomic instability, predisposing to widespread DNA changes and evolution of ploidy lesions, late events in cancer progression. Many other genetic and molecular alterations have been described^[8,9,12-64] (Table 1).

The concept of a linear, stepwise evolution of tumor suppressor gene mutations in which clonal expansion of a solitary mutated clone expands to fill the entire Barrett's segment has been termed the "selective sweep to fixation" model. However, an alternative model has been proposed by Leedham *et al*^[65], who performed genetic analysis of individual crypts rather than a flow purified whole biopsy specimen. This technique permitted identification of certain mutations masked by whole biopsy segment analysis (attributed to dilution effect of the normal stroma on whole biopsy analysis), whilst also revealing a greater degree of genotypical and phenotypical heterogeneity within the same biopsy sample than previously appreciated. The demonstrated lack of a single founder mutation present in every crypt suggested that the clonal expansion arose from multiple independent clones rather than a single common founder mutation^[65,66] (Figure 1).

This enhanced understanding prompted research into > 200 candidate novel biomarkers of disease progression in BE/EAC. Several, including 17p LOH, cyclin D1, tetraploidy and aneuploidy, have undergone phase 3/4 validation and in future might have clinical/prognostic utility as intermediate markers of progression^[67]. However, Leedham's recent findings call into question the reliability of "surveillance" biomarker identification *via* genetic analysis of whole biopsy specimens, since minority clones within the sample (harboring neoplastic potential) might not be detected^[65].

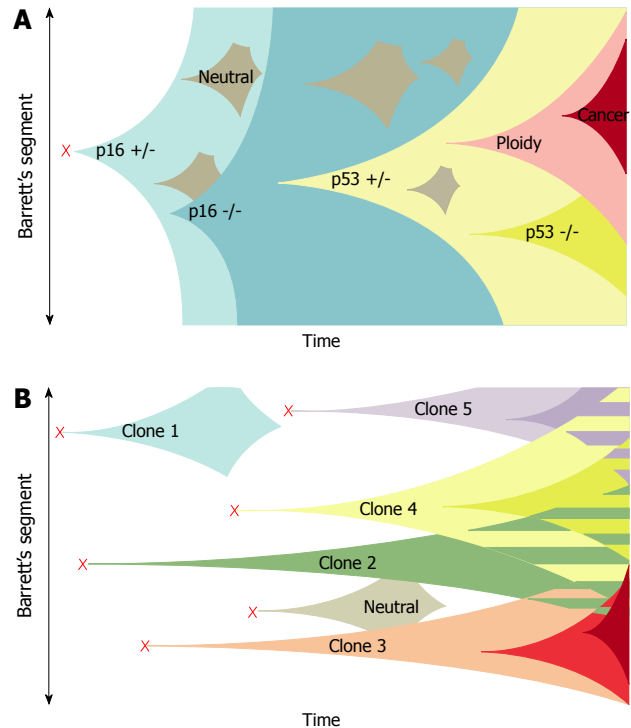


Figure 1 Clonal evolution models in Barrett's esophagus. A: The current model of clonal evolution adapted from Maley *et al*^[66]. Founder mutation (red cross) occurs in a single progenitor and provides a growth advantage that predisposes to a selective sweep. Successive selective sweeps result in progression along the metaplasia dysplasia pathway. Clone bifurcation is responsible for the genetic heterogeneity in this model; B: The newly proposed model of evolution based on the mutation of multiple progenitor cells situated in esophageal gland squamous ducts located throughout the length of the esophagus (red crosses). Multiple independent clones then arise and evolve separately. The presence of multiple different clones gives rise to a mosaic interdigitating clonal pattern of the Barrett's segment represented as the striped areas^[65].

Currently, dysplasia remains the only validated marker for identifying BE patients at risk, and forms the basis of EAC surveillance. However, this is imperfect. The tempo of progression towards EAC is highly variable and it remains unclear whether relentless progression through the MDC sequence is inevitable; some evidence suggests that high-grade dysplasia may remain stable for years or even regress^[68]. Patients with BE may develop EAC during surveillance without detection of earlier MDC stages. This might relate to pace of progression, sampling error or lesions skipping directly from non-dysplastic disease to cancer. Other limitations of dysplasia as a prognostic marker include inter-observer variability in histological interpretation, and that inflammation may mimic dysplastic changes^[69].

RISK FACTORS FOR NEOPLASTIC PROGRESSION

Until molecular biomarkers enter clinical practice it remains important to identify other clinical risk factors for malignant progression to effectively allocate resources and individualize surveillance programs, targeting those at highest risk. Identifying modifiable risk factors will also

Table 1 Published evidence from selected studies investigating genetic and epigenetic changes implicated in the metaplasia-dysplasia-carcinoma sequence of Barrett's esophagus

Factor	Summary of major findings/conclusions	Ref.
Growth self-sufficiency		
Cyclin D1	<p>↑ nuclear cyclin D1 immunostaining in 46% BE specimens: -?cyclin D1 overexpression early event in MDC sequence</p> <p>↑ nuclear cyclin D1 immunostaining in 64% EAC specimens</p> <p>Cyclin D1 expression correlates with degree of dysplasia in BE</p> <p>Cyclin D1 expression 43% BE mucosa (<i>vs</i> 0% normal mucosa)</p> <p>Polyphenon E inhibits growth of BE and EAC cells <i>via</i> downregulation of cyclin D1 expression</p>	<p>Arber <i>et al</i>^[9]</p> <p>Arber <i>et al</i>^[13]</p> <p>Coppola <i>et al</i>^[14]</p> <p>Umansky <i>et al</i>^[15]</p> <p>Song <i>et al</i>^[16]</p>
Cyclin E	<p>↑ cyclin E expression in neoplastic cells in BE</p> <p>Cyclin E expression 37% BE mucosa (<i>vs</i> 0% normal mucosa)</p>	<p>Coppola <i>et al</i>^[14]</p> <p>Umansky <i>et al</i>^[15]</p>
p27 ^{Kip-1}	<p>83% EAC specimens displayed low p27 protein levels (despite high p27 mRNA): -p27 inactivated in most BE-associated EAC (post-transcriptional modification) → loss of cell cycle inhibition</p> <p>Experimentally-induced BE and EAC development in mouse model significantly enhanced by p27 gene knockout</p>	<p>Singh <i>et al</i>^[17]</p> <p>Ellis <i>et al</i>^[18]</p>
EGF (and EGF-R)	<p>↑ EGF in cytoplasm of BE epithelial cells (<i>vs</i> gastric mucosa)</p> <p>EGF-R expression area in inflamed mucosa (43.1%) significantly > normal mucosa (29.5%); all BE showed positive EGF-R staining</p> <p>EGF/EGF-R expression significantly ↑ in BE and EAC mucosa (<i>vs</i> normal mucosa) by flow cytometry (<i>P</i> < 0.01)</p> <p>EGF-R expression positive in 64% of BE-associated EAC; ↑ staining associated with poorer survival (<i>P</i> = 0.004)</p>	<p>Jankowski <i>et al</i>^[19]</p> <p>Jankowski <i>et al</i>^[20]</p> <p>Jankowski <i>et al</i>^[21]</p> <p>Yacoub <i>et al</i>^[22]</p>
TGF-α	<p>EGF A61G G/G genotype associated with >double EAC risk in BE pts (<i>vs</i> A/A or A/G) (OR 2.2)</p> <p>↑ TGF-α expression in cells from BE and EAC mucosa (<i>vs</i> normal gastric mucosa) by flow cytometry (<i>P</i> < 0.01)</p>	<p>Lanuti <i>et al</i>^[23]</p> <p>Jankowski <i>et al</i>^[21]</p>
HGF (and HGF-R)	<p>TGF-α expression positive in 100% of BE-associated EAC</p> <p>HGF expression significantly ↑ in BE specimens (<i>vs</i> normal esophageal mucosa)</p> <p>Intense HGF-R immunostaining in 100% EAC and dysplastic BE specimens (<i>vs</i> minimal staining in non-dysplastic BE or normal mucosa); HGF-R mRNA and protein levels ↑ in EAC cell lines</p>	<p>Yacoub <i>et al</i>^[22]</p> <p>Konturek <i>et al</i>^[24]</p> <p>Herrera <i>et al</i>^[25]</p>
Erb family tyrosine kinases	<p>Membranous c-erbB2 overexpressed in 26% EAC (<i>vs</i> 0% BE with dysplasia): -?later event in MDC sequence</p> <p>c-erbB-2 gene amplification in 14% EAC <i>vs</i> 11% HG-dysplasia <i>vs</i> 0% metaplasia/LG-dysplasia specimens</p>	<p>Hardwick <i>et al</i>^[26]</p> <p>Geddert <i>et al</i>^[27]</p>
FGF	<p>Immunostaining intensity for FGF sequentially ↑ from metaplasia/LG-dysplasia (negligible) → HG-dysplasia (weak/moderate) → EAC (moderate/strong)</p> <p>FGF-1 mRNA and protein expression sequentially ↑ in HG-dysplasia/EAC (<i>vs</i> metaplasia/LG-dysplasia/controls)</p>	<p>Soslow <i>et al</i>^[28]</p> <p>Soslow <i>et al</i>^[29]</p>
Src family tyrosine kinases	<p>Src-specific activity 3-4-fold ↑ in BE and 6-fold ↑ in EAC (<i>vs</i> controls): -?Src activation early event in MDC sequence</p> <p>Strong Src expression in 85% EAC <i>vs</i> 93% BE HG-dysplasia <i>vs</i> 72% BE LG-dysplasia <i>vs</i> 27% BE specimens</p>	<p>Kumble <i>et al</i>^[30]</p> <p>Iravani <i>et al</i>^[31]</p>
Insensitivity to anti-growth signals		
p16	<p>9p21 (p16) LOH observed in 89% EAC specimens (<i>vs</i> 0% non-dysplastic BE); homozygous p16 deletion in only 25%</p> <p>p16 promoter hypermethylation (inactivation) in 75% BE with HG-dysplasia <i>vs</i> 56% LG-dysplasia (<i>vs</i> 3% non-dysplastic BE)</p>	<p>González <i>et al</i>^[32]</p> <p>Klump <i>et al</i>^[8]</p>
APC	<p>5q (APC) LOH seen in 80% EAC specimens (and surrounding mucosa)</p> <p>APC gene LOH observed in 60% EAC specimens (<i>vs</i> 0% non-dysplastic BE)</p> <p>APC promoter hypermethylation in 92% EAC <i>vs</i> 40% BE (<i>vs</i> 0% normal esophageal tissues)</p>	<p>Barrett <i>et al</i>^[33]</p> <p>González <i>et al</i>^[32]</p> <p>Kawakami <i>et al</i>^[34]</p>
Avoidance of apoptosis		
p53	<p>Positive p53 immunostaining in 87% EAC <i>vs</i> 55% BE with HG-dysplasia <i>vs</i> 9% LG-dysplasia <i>vs</i> 0% non-dysplastic BE</p> <p>17p (p53) LOH found in 91% BE pts who developed aneuploid cell populations: -17p allelic losses precede aneuploidy</p> <p>p53 overexpression in 64% EAC <i>vs</i> 31% dysplastic BE <i>vs</i> 0% non-dysplastic BE; trend of ↑ p53 expression with ↑ tumour grade: -?p53 mutation early event in malignant progression</p> <p>p53 immunoreactivity only in EAC/BE with HG-dysplasia (not in BE with LG-/no dysplasia); mutated p53 in 69%: -?late event in MDC sequence (during transition to HG-dysplasia)</p> <p>p53 protein expression in 85% EAC specimens <i>vs</i> 60% BE with HG-dysplasia <i>vs</i> 7% LG-dysplasia (<i>P</i> < 0.001)</p> <p>p53 mutations identified in 75% EAC specimens; p53 overexpression in 58% EAC <i>vs</i> 60% BE with HG-dysplasia <i>vs</i> 12% LG-dysplasia <i>vs</i> 0% non-dysplastic BE</p>	<p>Younes <i>et al</i>^[35]</p> <p>Blount <i>et al</i>^[36]</p> <p>Symmans <i>et al</i>^[37]</p> <p>Rice <i>et al</i>^[38]</p> <p>Rioux-Leclercq <i>et al</i>^[39]</p> <p>Chung <i>et al</i>^[40]</p>
Fas (CD95)	<p>↓ surface expression of Fas observed in EAC specimens; impaired translocation of Fas to membrane wild-type Fas protein retained in cytoplasm in EAC cell line: -?potential mechanism by which EAC cells evade Fas-mediated apoptosis</p>	<p>Hughes <i>et al</i>^[41]</p>
Bcl-xl/Bax/Bcl-2	<p>↓ surface expression of Fas and resistance to Fas-mediated apoptosis observed in EAC cell lines</p> <p>Bcl-xl positive in all dysplasia and EAC cells, but negative in 47% non-dysplastic BE: -?switch to anti-apoptotic phenotype in transformation from metaplasia to EAC</p>	<p>Mahidhara <i>et al</i>^[42]</p> <p>van der Woude <i>et al</i>^[43]</p>

COX-2	Bel-2 expression in 84% LG-dysplasia vs 0% HG-dysplasia or EAC	Rioux-Leclercq <i>et al</i> ^[39]
	Cytoplasmic Bcl-xl immunostaining in 59% EAC vs 71% BE/HG-dysplasia vs 60% LG-dysplasia vs 27% non-dysplastic	Iravani <i>et al</i> ^[31]
	↑ COX-2 mRNA levels in 80% BE and 100% EAC specimens (<i>vs</i> normal gastric controls) ($P < 0.001$);	Wilson <i>et al</i> ^[44]
	COX-2 immunostaining strongly positive in 100% BE samples (> gastric controls)	Lagorce <i>et al</i> ^[45]
	COX-2 immunopositivity in 91% non-dysplastic BE vs 94% dysplastic vs 97% EAC	Cheong <i>et al</i> ^[46]
Limitless replicative potential	Natural/synthetic COX-2 inhibitors suppressed proliferation, induced apoptosis and blocked cell cycle in EAC cell lines	Majka <i>et al</i> ^[47]
	Cox-2 mRNA strongly upregulated in experimentally-induced BE epithelium in rat model (<i>vs</i> absent in control animals); COX-2 overexpression observed in human BE patients with dysplasia	
Telomerase	Telomerase RNA positive in 100% EAC/BE with HG-dysplasia vs 90% LG-dysplasia vs 70% non-dysplastic BE: marked ↑ telomerase RNA accompanies transition along MDC sequence	Morales <i>et al</i> ^[48]
	human telomerase reverse transcriptase (catalytic subunit of telomerase) expression ↑ at all stages of BE <i>vs</i> normal controls, and in EAC ($P = 0.003$) and dysplastic BE ($P = 0.056$) <i>vs</i> non-dysplastic BE	Lord <i>et al</i> ^[49]
	Telomerase activity (by telomeric repeat amplification protocol assay) ↑ in EAC samples <i>vs</i> adjacent mucosa ($P = 0.0002$) and in EAC <i>vs</i> BE ($P = 0.001$); no difference BE <i>vs</i> adjacent mucosa	Barclay <i>et al</i> ^[50]
	Telomerase inhibition (by small interference RNAs) induced senescence in 40% and apoptosis in 86% in BE cell lines	Shammas <i>et al</i> ^[51]
Sustained angiogenesis	VEGF expression correlated with higher vascularisation in BE and EAC specimens	Couvelard <i>et al</i> ^[52]
	VEGF-A expressed in BE epithelium; VEGFR-2 strongly expressed in immature endothelial cells feeding BE epithelium; ↑ VEGF-C expression in BE (<i>vs</i> absent in normal epithelium); ↑ VEGFR-3 in EAC: ?aberrant neovasculature early in MDC sequence	Auvinen <i>et al</i> ^[53]
	VEGF expressed in 64% EAC specimens; significantly correlated with angiolymphatic invasion/survival	Saad <i>et al</i> ^[54]
Invasive/metastatic potential	VEGF expression significantly ↑ in EAC (> dysplastic BE > BE > normal epithelium)	Griffiths <i>et al</i> ^[55]
CAMs	↓ expression in EAC specimens of E-cadherin (in 74%), α-catenin (60%) and β-catenin (72%)	Krishnadath <i>et al</i> ^[56]
	Abnormal expression of β-catenin ($P = 0.022$), α-catenin ($P < 0.01$) and E-cadherin ($P = 0.049$) significantly associated with higher degrees of BE-related dysplasia	Washington <i>et al</i> ^[57]
	↓ expression of E-cadherin with progression along MDC sequence ($P < 0.01$); in contrast P-cadherin absent from BE (± dysplasia) but expressed in 67% EAC specimens	Bailey <i>et al</i> ^[58]
Cathepsins	Slug (E-cadherin repressor) immunostaining and mRNA levels overexpressed in EAC <i>vs</i> BE metaplasia specimens: -?Slug upregulation represents mechanism of E-cadherin silencing	Jethwa <i>et al</i> ^[59]
	Detected amplicon at chromosome 8p22-23 resulting in cathepsin B overexpression (observed in 73% EAC samples)	Hughes <i>et al</i> ^[60]
CD44	↑ cathepsin C expression in EAC (<i>vs</i> BE <i>vs</i> normal) in rat model	Cheng <i>et al</i> ^[61]
	Stepwise ↑ cathepsin D mRNA levels in GERD→BE→EAC tissue	Breton <i>et al</i> ^[62]
	CD44-H and -V6 variant frequently expressed in BE; differing expression patterns along spectrum normal→dysplastic BE→EAC: -?CD44H and V6 involved in carcinogenesis of BE mucosa	Lagorce-Pages <i>et al</i> ^[63]
	↓ CD44 expression in EAC/HG-dysplasia (<i>vs</i> BE/LG-dysplasia)	Darlavoix <i>et al</i> ^[64]

BE: Barrett's esophagus; MDC: Metaplasia-dysplasia-carcinoma; EAC: Esophageal adenocarcinoma; EGF: Epidermal growth factor; EGF-R: EGF receptor; pts: Patients; OR: Odds ratio; TGF: Transforming growth factor; HGF: Hepatocyte growth factor; HGF-R: HGF receptor; mRNA: Messenger RNA; FGF: Fibroblast growth factor; HG: High grade; LG: Low grade; LOH: Loss of heterozygosity; APC: Adenomatous polyposis coli; COX-2: Cyclooxygenase-2; VEGF: Vascular endothelial growth factor; VEGF-R: VEGF receptor; CAM: Cell adhesion molecule; GERD: Gastro-esophageal reflux disease.

Table 2 Clinical and demographic risk factors for neoplastic progression of Barrett's esophagus

Innate factors	Gastrointestinal factors	Other modifiable factors
Age	Bile and acid reflux	Obesity
Gender	Anti-reflux surgery	Diet
Ethnicity	Proton pump inhibition	Alcohol
	Pharmacological lower esophageal sphincter relaxation	Smoking
	Salivary nitrates	Socioeconomic status
	Barrett's segment length	Pharmacological COX-2 inhibition

COX-2: Cyclooxygenase-2.

inform disease prevention strategies. Epidemiological studies of EAC have described a "birth cohort effect", with higher incidence rates observed in recent cohorts

suggesting exposure to an exogenous risk factor in early life contributing increased risk in all ages of the cohort^[70] (Figure 2). Multiple risk factors for neoplastic progression of BE have been investigated (Table 2).

INNATE HOST FACTORS

Age is a well-recognized risk for both BE and EAC. Corley *et al*^[71] reported an incidence of BE of 2/100 000 for 21-30-year-old and 31/100 000 for 61-70-year-old, whilst El-Serag *et al*^[70] calculated the risk of EAC to increase by 6.6% for each 5-year age increase. Evidence specifically linking age to risk of neoplastic progression within BE is lacking, but it seems intuitive to propose advancing age as an independent risk factor.

BE displays a male preponderance of approximately 2:1, rising to 4:1 for BE-associated EAC, suggesting an independent influence of gender on risk of neoplastic pro-

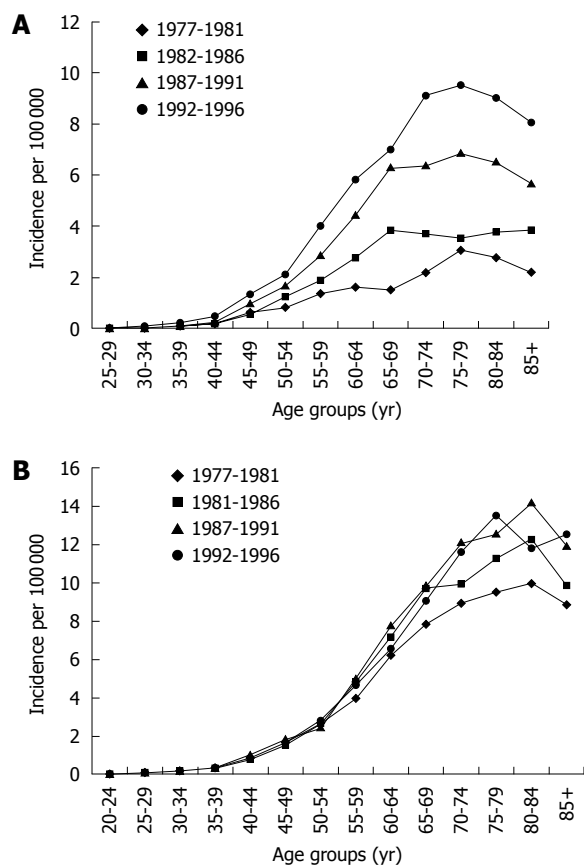


Figure 2 Age distribution of cases diagnosed with oesophageal adenocarcinoma (A) and gastric cardia adenocarcinoma (B) in the USA between 1977-1996, displaying the "birth cohort effect". Each individual curve represents the age-specific incidence rates in a five year period (from El-Serag *et al*^[70]).

gression^[71,72]. Why male gender should confer additional risk is unknown; some have speculated that male propensity toward visceral pattern of obesity might be relevant^[73].

A higher prevalence of BE in Caucasians has long been recognized^[74]; again, this association strengthens with development of BE-associated EAC^[75]. Analysis of the US Surveillance, Epidemiology and End Results registry found that the annual incidence of EAC for Caucasian males was double that for Hispanic males and four times higher than Black, Asian, Pacific Island and Native American males^[76]. Although selection bias and differing endoscopy uptakes between ethnic groups might partially explain this, other factors seem to be involved. Whilst environmental influences are probably important, hitherto-unknown genetic variations influencing protection against reflux-induced mucosal damage seem likely. A US study found similar GERD prevalence in Caucasian and Black Americans from the same geographical population, yet the latter displayed significantly less esophagitis and almost no BE^[77].

GASTROINTESTINAL FACTORS

Bile/acid reflux

The relationship between GERD and BE is well established, and whilst reflux of gastric acid is known to

induce chronic mucosal esophageal injury the contribution of bile salts and acids (from duodenal refluxate) is increasingly recognized. Vaezi and Richter demonstrated patients with complicated BE (dysplasia/stricture/ulceration) reflux significantly greater amounts of both gastric and bile acids than those with uncomplicated BE, and postulated that complications might result from synergism between the two^[78]. Bile salts induce esophageal injury over a wide pH range, and patients with BE display significantly more bile salts in aspiration studies than patients with mild reflux only^[79]. Menges *et al*^[80] observed a strong correlation between duration of esophageal exposure to acid and bile with severity of pathological change in BE. Furthermore, proton pump inhibitor (PPI) therapy predisposes to upper gastrointestinal bacterial colonization and consequent bile salt-deconjugation, which, in this high pH environment, has been linked to chronic inflammation^[81].

Refluxate-mediated inflammation might promote carcinogenesis *via* both the arachidonic acid (AA) pathway and induction of oxidative stress. Low pH and bile salts promote expression of cyclooxygenase-2 (COX-2), catalyzing conversion of AA into various prostaglandins, including PGE₂. PGE₂ increases proliferation of BE epithelial cells and inhibits tumor surveillance through suppressing natural killer cell function. Consequently, abnormal cells displaying genomic instability may accumulate. COX-2 expression has been shown to increase with neoplastic progression of BE, supporting a role for the AA pathway in EAC carcinogenesis^[44]. Chronic mucosal injury also induces production of reactive oxygen species (ROS), depletes antioxidants and increases expression of oxidative stress-related genes. High levels of oxygen radicals and lipid peroxidation products have been demonstrated in BE epithelial cells, with reduced levels of vitamin C and glutathione, indicating compromised oxidant defences^[82]. ROS have well-established mutagenic capacity, whilst subsequent apoptosis of mutated cells is additionally suppressed by capacity of bile salts to induce proteasomal degradation of p53^[83].

The Factors Influencing the Barrett's Adenocarcinoma Relationship (FINBAR) study suggested GERD symptom chronicity and frequency appeared better predictors for neoplastic progression than severity^[84]. However, a significant proportion of EAC patients (40%-50%) do not recall ever having prior reflux^[85]. Furthermore, reflux of gastroduodenal contents correlates poorly with heartburn symptoms, BE is frequently asymptomatic and development of less sensitive Barrett's epithelium may ameliorate symptoms. Thus, symptom-based risk scores for assessing progression risk have so far not proved useful in clinical practice.

PPIs

PPIs increase pH of gastric refluxate, attenuating acid-induced damage. Ouatu-Lascar *et al*^[86] showed "normalization" of intraesophageal pH with acid suppression favors differentiation and reduces cellular proliferation in BE biopsy specimens. However, PPIs have not prevented

recent increases in EAC, and the observation of EAC with PPI administration in animal models raises concern they might actually favor progression of BE^[87]. This might be mediated *via* interaction of gastrin with its cholecystokinin receptor, CCK₂R. PPIs elevate serum gastrin levels, which on binding to CCK₂R, stimulate expression of EGF and trefoil peptide, inducing COX-2 expression. Gastrin exposure increases proliferation in esophageal cell culture, and BE mucosa expresses more CCK₂R than normal squamous mucosa. CCK₂R stimulation also inactivates pro-apoptotic factors^[88].

Despite this, the clinical relevance in humans remains unproven. Three large studies have examined PPI usage and EAC risk in BE patients, each reporting a strong inverse correlation. Two observed a decreased risk with longer duration of PPI, and one showed an increased risk with delayed PPI use^[89]. Obszynska *et al*^[90] investigated effects of hypergastrinemia induced by different PPI doses in cell models and BE patients. Despite increased cell proliferation *in vitro*, COX2 induction and enhanced epithelial restitution, they found no evidence of longer-term harm using surrogate biomarkers of proliferation or apoptosis *in vivo*. The Aspirin Esomeprazole Chemoprevention Trial (AsPECT) is currently investigating effects of different PPI doses in combination with aspirin on EAC risk.

Anti-reflux surgery

Theoretically, anti-reflux surgery should prevent reflux of duodenal contents, against which PPIs have no effect, potentially mitigating against progression of BE. Unfortunately this is not supported by the available evidence. Two large cohort studies failed to show cancer protection in GERD patients^[91,92], whilst a meta-analysis by Corey *et al*^[93] concluded no reduction in progression risk for BE. However, different surgical procedures were employed and effectiveness of reflux control was not always assessed.

Lower esophageal sphincter-relaxing drugs

Pharmacological lower esophageal sphincter (LES) relaxation might promote development/progression of BE by increasing reflux, suggested by the observation that drugs with these effects (e.g. tricyclic antidepressants) have increased in use alongside the rise in EAC. A Swedish population-based study by Lagergren *et al*^[94] reported a positive association between EAC and long-term use of LES-relaxing drugs, with the strongest association for anticholinergics; this association disappeared after adjustment for reflux symptoms.

Helicobacter pylori infection

An increase in BE-associated EAC alongside falling rates of *Helicobacter pylori* (*H. pylori*) infection has led some to propose a protective effect of *H. pylori*, mediated by its influence in reducing gastric acidity. The virulent *cagA* strain is particularly associated with high-grade gastric inflammation and atrophy^[95]. A meta-analysis by Rokkas *et al*^[96] reported statistically significant inverse relationships between *H. pylori* infection and both EAC and BE [odds ratio (OR),

0.52% and 0.64%, respectively]. Furthermore, a large prospective study of BE patients and GERD controls found less *H. pylori* infection with increasing "severity" of disease: 44% in GERD; 35% in uncomplicated BE; 14%-15% in BE with high-grade dysplasia/EAC^[97].

However, another study, controlling for demographic and lifestyle factors, failed to demonstrate reduced EAC with *cagA*+ infection^[98]. A confounding factor might be the degree of bile acid reflux, since excessive bile reflux may prevent *H. pylori* colonization and contribute to chronic mucosal injury^[88]. The protective role for *H. pylori* is debatable and since *H. pylori* is a World Health Organisation class 1 mutagen for gastric adenocarcinoma it is difficult to argue against its eradication whenever it is detected.

Salivary nitrates

Dietary nitrate, concentrated in saliva and reduced to nitrites by oral flora, produces intraesophageal nitric oxide (NO) during reflux. Achlorhydria induced by PPI or atrophic gastritis may cause overgrowth of nitrate-reducing bacteria in the upper gut, providing another source of nitrite^[88]. Clemons demonstrated the capacity of NO to induce double-strand DNA breaks in esophageal BE cells *in vitro*, which could promote neoplastic progression^[99]. Increasing agricultural nitrate use in the latter 20th century caused significant increases in nitrate content of leafy vegetables and drinking water^[100] and could have partially contributed to the increase in EAC incidence.

Barrett's segment length

Although EAC can develop in BE segments of any length, several observational studies support the intuitive notion that longer segments confer greater risk^[101]. However, a meta-analysis by Thomas *et al*^[102] showed only a non-significant trend towards reduced progression with shorter BE segments, and evidence remains insufficient to advocate surveillance strategies based on segment length alone.

OTHER MODIFIABLE RISK FACTORS

Obesity

Increasing obesity has also paralleled increased rates of BE and EAC. Strong links between obesity and both GERD and erosive esophagitis have been established^[103]. It is logical that this might predispose to BE, but a meta-analysis specifically comparing body mass index (BMI) in BE cases with population controls showed only a modest risk increase^[104]. However, elevated BMI is a strong risk factor for EAC (OR, 1.8 and 2.4 for BMI > 25 and BMI > 30, respectively)^[105]. Increased risk may relate more to distribution of body fat than BMI alone, with visceral (abdominal) obesity conferring greater risk^[106]. Other studies noted an association between obesity in early life and EAC risk, suggesting adiposity may act early in the disease process^[84,107].

Although a small prospective study by Oberg and colleagues failed to identify any association between BMI

Table 3 Selected published evidence linking adipokines (and ghrelin) with Barrett's esophagus and progression to esophageal adenocarcinoma

Adipokine	Evidence in BE and EAC	
	Relevant study findings	Ref.
Adiponectin (↓ in obesity)	↓ adiponectin receptors in Barrett's mucosa compared with normal mucosa from controls	Konturek <i>et al</i> ^[110]
	↑ Bax (pro-apoptotic), ↓ Bcl-2 (anti-apoptotic) and ↑ apoptosis of EAC cell lines on incubation with adiponectin Plasma adiponectin levels inversely associated with BE risk in 50 matched cases (OR 4.7 for each 10 µg/mL ↓ in level) (independent of BMI) No difference in adiponectin levels between 51 BE patients and 67 controls	Konturek <i>et al</i> ^[110] Rubenstein <i>et al</i> ^[111]
Leptin (↑ in obesity)	Leptin receptors expressed in esophagus	Kendall <i>et al</i> ^[112]
	↑ proliferation and ↓ apoptosis (<i>via</i> various signalling pathways) in EAC cell lines Leptin levels strongly associated with ↑ risk of BE in males (no association in females) Gastric (fundic) leptin levels positively associated with BE (no association with serum leptin)	Francois <i>et al</i> ^[113] Ogunwobi <i>et al</i> ^[114] Kendall <i>et al</i> ^[112] Francois <i>et al</i> ^[113]
Ghrelin (↓ in obesity)	↑ gastric emptying (so may ↓ gastric reflux)	Dornonville <i>et al</i> ^[115]
	↓ TNF- α -induced COX-2 and interleukin-1- β expression in BE cell line Ghrelin expression negligible in archived EAC cell specimens (<i>vs</i> rich expression in normal mucosa) ↑ serum ghrelin associated with ↓ EAC risk (in overweight subjects)	Konturek <i>et al</i> ^[110] Mottershead <i>et al</i> ^[116] de Martel <i>et al</i> ^[117]

BE: Barrett's esophagus; EAC: Esophageal adenocarcinoma; OR: Odds ratio; BMI: Body mass index; COX-2: Cyclooxygenase-2; TNF: Tumor necrosis factor.

and progression from BE to low- or high-grade dysplasia^[108], it had limited power, and a larger study from the Seattle Barrett's Esophagus Program revealed strong correlations between waist-to-hip ratio and intermediate biomarkers of progression^[109]; again, associations were less apparent for elevated BMI *per se*.

Obesity causes GERD through several mechanical and physiological mechanisms. However, part of the association between obesity and EAC is independent of GERD, suggesting a role for reflux-independent mechanisms, probably linked to important endocrine actions of adipose tissue. Many recent studies have linked several adipokines (metabolically active factors) to plausible actions in the MDC process^[110-117] (Table 3).

Kristal *et al*^[118] investigated whether weight loss (alongside other dietary measures) impacted upon measured biomarkers of cellular proliferation in BE. Despite weight loss (mean 3.6 kg) at 18 mo no differences in biomarkers were observed. This study was relatively small, and the lack of response might relate to the relatively modest weight loss, and/or choice of proliferation markers employed.

Diet

Several studies have shown an association between a diet high in fruit and vegetables and reduced EAC. A large population-based Swedish study found individuals in the highest exposure quartile of fruit and vegetable intake to have approximately 50% less EAC compared to the lowest quartile^[119]. However, Kristal *et al*'s study observed no effect on biomarkers of BE cell proliferation despite a net increase in fruit and vegetable consumption^[118], whilst the FINBAR study observed a reduction in EAC with increased fruit, but not vegetable, consumption^[84]. A protective effect for the natural anti-oxidants in fruit was proposed. A well-controlled, prospective study by Dong *et al*^[120] showed patients who took multivitamin pills had significantly decreased risk of tetraploidy [hazard ratio (HR), 0.19] and frank EAC (HR, 0.38). Significant inverse associations with EAC were also observed for supple-

mental vitamins C (HR, 0.25) and E (HR, 0.25), both well-recognized antioxidants.

Chen *et al*^[121] observed a significant inverse association between zinc intake and EAC risk compared with controls (OR, 0.5); inverse associations were also noted for vitamin A, β -cryptoxanthin, riboflavin, folate, fiber, protein and carbohydrate, whilst saturated fat intake was positively associated with EAC. Rudolph *et al*^[122] investigated selenium levels in 396 BE patients: those with levels in the upper three quartiles were less likely to display high-grade dysplasia (OR, 0.5), aneuploidy (OR, 0.4) or 17p LOH (OR, 0.5) than the lowest quartile. No association was observed with *p16* LOH (an early event in the MDC sequence), indicating selenium's protective effects might occur late in progression to EAC.

Alcohol

Data supporting links between alcohol and BE/EAC are sparse. The UK BE registry found no association between alcohol consumption in patients with BE compared with reflux esophagitis^[123]. Although at least eleven studies have investigated the relationship between alcohol and EAC only six have shown a positive association, and in most it was weak^[124-134]. One study even seemed to suggest wine to be protective^[133].

Smoking

Studies of smoking and BE/EAC are contradictory. An Australian population-based case-control study found smoking was associated with 2- to 3-fold increased risk of BE and BE with dysplasia^[135]. However, there was no dose-response effect. Other small studies found no clear association^[131]. Whilst smoking is a strong risk factor for esophageal squamous cell carcinoma, studies of EAC have been inconsistent, yielding conclusions ranging from complete absence of association^[132-134] to a significant OR of 3.4 for current smokers^[128]. Problems with study methodology occur and certainly smoking has rarely been a primary endpoint for studies of BE/EAC.

Socioeconomic status

There are no clear associations between socioeconomic status and neoplastic progression of BE. Some studies suggest increased EAC risk in higher socioeconomic groups, others the reverse^[72].

COX-2 inhibition

Given the role of the AA pathway in neoplastic progression, pharmacological inhibition of COX-2 might modify the natural history of BE. Various studies have investigated whether aspirin and non-steroidal anti-inflammatory drugs (NSAIDs) might confer protection against EAC. A meta-analysis by Corley *et al.*^[136] including 1813 EAC patients suggested a protective association (OR, 0.67). Both intermittent and frequent use appeared advantageous, with evidence of a dose-effect, whilst aspirin conferred greater protection than NSAIDs.

However the Chemoprevention for Barrett's Oesophagus Trial randomized 100 BE patients with dysplasia to either celecoxib 200 mg twice daily or placebo, with negative results^[137]. A retrospective analysis of the UK BE registry with a total follow-up of 3683 patient-years also failed to demonstrate a protective effect of aspirin^[138]. AspECT should provide further useful information.

CONCLUSION

The etiology of progression of BE is probably multi-factorial, with contributions from environmental risk factors interacting with genetically-determined characteristics. Obesity and ongoing bile and acid reflux are emerging as potentially modifiable risk factors, though designing practical interventions has so far proved difficult. Developments in understanding the MDC process in BE may provide future testable therapeutic targets.

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REFERENCES

- 1 Reid BJ. Barrett's esophagus and esophageal adenocarcinoma. *Gastroenterol Clin North Am* 1991; **20**: 817-834
- 2 Hage M, Siersema PD, van Dekken H, Steyerberg EW, Dees J, Kuipers EJ. Oesophageal cancer incidence and mortality in patients with long-segment Barrett's oesophagus after a mean follow-up of 12.7 years. *Scand J Gastroenterol* 2004; **39**: 1175-1179
- 3 Shaheen NJ, Crosby MA, Bozyski EM, Sandler RS. Is there publication bias in the reporting of cancer risk in Barrett's esophagus? *Gastroenterology* 2000; **119**: 333-338
- 4 Jankowski JA, Provenzale D, Moayyedi P. Esophageal adenocarcinoma arising from Barrett's metaplasia has regional

- 5 variations in the west. *Gastroenterology* 2002; **122**: 588-590
- 6 Boulton RA, Usselman B, Mohammed I, Jankowski J. Barrett's esophagus: environmental influences in the progression of dysplasia. *World J Surg* 2003; **27**: 1014-1017
- 7 Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000; **100**: 57-70
- 8 Morales CP, Souza RF, Spechler SJ. Hallmarks of cancer progression in Barrett's oesophagus. *Lancet* 2002; **360**: 1587-1589
- 9 Klump B, Hsieh CJ, Holzmann K, Gregor M, Porschen R. Hypermethylation of the CDKN2/p16 promoter during neoplastic progression in Barrett's esophagus. *Gastroenterology* 1998; **115**: 1381-1386
- 10 Arber N, Lightdale C, Rotterdam H, Han KH, Sgambato A, Yap E, Ahsan H, Finegold J, Stevens PD, Green PH, Hibshoosh H, Neugut AI, Holt PR, Weinstein IB. Increased expression of the cyclin D1 gene in Barrett's esophagus. *Cancer Epidemiol Biomarkers Prev* 1996; **5**: 457-459
- 11 Jankowski J, Hopwood D, Wormsley KG. Expression of epidermal growth factor, transforming growth factor alpha and their receptor in gastro-oesophageal diseases. *Dig Dis* 1993; **11**: 1-11
- 12 Ireland AP, Clark GW, DeMeester TR. Barrett's esophagus. The significance of p53 in clinical practice. *Ann Surg* 1997; **225**: 17-30
- 13 Wijnhoven BP, Tilanus HW, Dinjens WN. Molecular biology of Barrett's adenocarcinoma. *Ann Surg* 2001; **233**: 322-337
- 14 Arber N, Gammon MD, Hibshoosh H, Britton JA, Zhang Y, Schonberg JB, Rotterdam H, Fabian I, Holt PR, Weinstein IB. Overexpression of cyclin D1 occurs in both squamous carcinomas and adenocarcinomas of the esophagus and in adenocarcinomas of the stomach. *Hum Pathol* 1999; **30**: 1087-1092
- 15 Coppola D, Falcone R, Livingston S, Karl R, Nicosia S, Cacho CM. Cyclin D1 expression correlates with degrees of dysplasia in Barrett's esophagus. *Lab Invest* 1997; **76**: 298-302
- 16 Umansky M, Yasui W, Hallak A, Brill S, Shapira I, Halpern Z, Hibshoosh H, Rattan J, Meltzer S, Tahara E, Arber N. Proton pump inhibitors reduce cell cycle abnormalities in Barrett's esophagus. *Oncogene* 2001; **20**: 7987-7991
- 17 Song S, Krishnan K, Liu K, Bresalier RS. Polyphenon E inhibits the growth of human Barrett's and aerodigestive adenocarcinoma cells by suppressing cyclin D1 expression. *Clin Cancer Res* 2009; **15**: 622-631
- 18 Singh SP, Lipman J, Goldman H, Ellis FH, Aizenman L, Cangi MG, Signoretti S, Chiaur DS, Pagano M, Loda M. Loss or altered subcellular localization of p27 in Barrett's associated adenocarcinoma. *Cancer Res* 1998; **58**: 1730-1735
- 19 Ellis FH, Xu X, Kulke MH, LoCicero J, Loda M. Malignant transformation of the esophageal mucosa is enhanced in p27 knockout mice. *J Thorac Cardiovasc Surg* 2001; **122**: 809-814
- 20 Jankowski J, Coghill G, Tregaskis B, Hopwood D, Wormsley KG. Epidermal growth factor in the oesophagus. *Gut* 1992; **33**: 1448-1453
- 21 Jankowski J, Murphy S, Coghill G, Grant A, Wormsley KG, Sanders DS, Kerr M, Hopwood D. Epidermal growth factor receptors in the oesophagus. *Gut* 1992; **33**: 439-443
- 22 Jankowski J, Hopwood D, Wormsley KG. Flow-cytometric analysis of growth-regulatory peptides and their receptors in Barrett's oesophagus and oesophageal adenocarcinoma. *Scand J Gastroenterol* 1992; **27**: 147-154
- 23 Yacoub L, Goldman H, Odze RD. Transforming growth factor-alpha, epidermal growth factor receptor, and MiB-1 expression in Barrett's-associated neoplasia: correlation with prognosis. *Mod Pathol* 1997; **10**: 105-112
- 24 Lanuti M, Liu G, Goodwin JM, Zhai R, Fuchs BC, Asomaning K, Su L, Nishioka NS, Tanabe KK, Christiani DC. A functional epidermal growth factor (EGF) polymorphism, EGF serum levels, and esophageal adenocarcinoma risk and outcome. *Clin Cancer Res* 2008; **14**: 3216-3222
- 25 Konturek PC, Nikiforuk A, Kania J, Raithel M, Hahn EG, Mühlendorfer S. Activation of NFkappaB represents the central

- event in the neoplastic progression associated with Barrett's esophagus: a possible link to the inflammation and overexpression of COX-2, PPARgamma and growth factors. *Dig Dis Sci* 2004; **49**: 1075-1083
- 25 **Herrera LJ**, El-Hefnawy T, Queiroz de Oliveira PE, Raja S, Finkelstein S, Gooding W, Luketich JD, Godfrey TE, Hughes SJ. The HGF receptor c-Met is overexpressed in esophageal adenocarcinoma. *Neoplasia* 2005; **7**: 75-84
 - 26 **Hardwick RH**, Shepherd NA, Moorghen M, Newcomb PV, Alderson D. c-erbB-2 overexpression in the dysplasia/carcinoma sequence of Barrett's oesophagus. *J Clin Pathol* 1995; **48**: 129-132
 - 27 **Geddert H**, Zerriouh M, Wolter M, Heise JW, Gabbert HE, Sarbia M. Gene amplification and protein overexpression of c-erb-b2 in Barrett carcinoma and its precursor lesions. *Am J Clin Pathol* 2002; **118**: 60-66
 - 28 **Soslow RA**, Ying L, Altorki NK. Expression of acidic fibroblast growth factor in Barrett's esophagus and associated esophageal adenocarcinoma. *J Thorac Cardiovasc Surg* 1997; **114**: 838-843
 - 29 **Soslow RA**, Nabeya Y, Ying L, Blundell M, Altorki NK. Acidic fibroblast growth factor is progressively increased in the development of oesophageal glandular dysplasia and adenocarcinoma. *Histopathology* 1999; **35**: 31-37
 - 30 **Kumble S**, Omary MB, Cartwright CA, Triadafilopoulos G. Src activation in malignant and premalignant epithelia of Barrett's esophagus. *Gastroenterology* 1997; **112**: 348-356
 - 31 **Iravani S**, Zhang HQ, Yuan ZQ, Cheng JQ, Karl RC, Jove R, Coppola D. Modification of insulin-like growth factor 1 receptor, c-Src, and Bcl-XL protein expression during the progression of Barrett's neoplasia. *Hum Pathol* 2003; **34**: 975-982
 - 32 **González MV**, Artímez ML, Rodrigo L, López-Larrea C, Menéndez MJ, Alvarez V, Pérez R, Fresno MF, Pérez MJ, Sampedro A, Coto E. Mutation analysis of the p53, APC, and p16 genes in the Barrett's oesophagus, dysplasia, and adenocarcinoma. *J Clin Pathol* 1997; **50**: 212-217
 - 33 **Barrett MT**, Galipeau PC, Sanchez CA, Emond MJ, Reid BJ. Determination of the frequency of loss of heterozygosity in esophageal adenocarcinoma by cell sorting, whole genome amplification and microsatellite polymorphisms. *Oncogene* 1996; **12**: 1873-1878
 - 34 **Kawakami K**, Brabender J, Lord RV, Groshen S, Greenwald BD, Krasna MJ, Yin J, Fleisher AS, Abraham JM, Beer DG, Sidransky D, Huss HT, Demeester TR, Eads C, Laird PW, Ilson DH, Kelsen DP, Harpole D, Moore MB, Danenberg KD, Danenberg PV, Meltzer SJ. Hypermethylated APC DNA in plasma and prognosis of patients with esophageal adenocarcinoma. *J Natl Cancer Inst* 2000; **92**: 1805-1811
 - 35 **Younes M**, Lebovitz RM, Lechago LV, Lechago J. p53 protein accumulation in Barrett's metaplasia, dysplasia, and carcinoma: a follow-up study. *Gastroenterology* 1993; **105**: 1637-1642
 - 36 **Blount PL**, Galipeau PC, Sanchez CA, Neshat K, Levine DS, Yin J, Suzuki H, Abraham JM, Meltzer SJ, Reid BJ. 17p allelic losses in diploid cells of patients with Barrett's esophagus who develop aneuploidy. *Cancer Res* 1994; **54**: 2292-2295
 - 37 **Symmans PJ**, Linehan JM, Brito MJ, Filipe MI. p53 expression in Barrett's oesophagus, dysplasia, and adenocarcinoma using antibody DO-7. *J Pathol* 1994; **173**: 221-226
 - 38 **Rice TW**, Goldblum JR, Falk GW, Tubbs RR, Kirby TJ, Casey G. p53 immunoreactivity in Barrett's metaplasia, dysplasia, and carcinoma. *J Thorac Cardiovasc Surg* 1994; **108**: 1132-1137
 - 39 **Rioux-Leclercq N**, Turlin B, Sutherland F, Heresbach N, Launois B, Campion JP, Ramee MP. Analysis of Ki-67, p53 and Bcl-2 expression in the dysplasia-carcinoma sequence of Barrett's esophagus. *Oncol Rep* 1999; **6**: 877-882
 - 40 **Chung SM**, Kao J, Hyjek E, Chen YT. p53 in esophageal adenocarcinoma: a critical reassessment of mutation frequency and identification of 72Arg as the dominant allele. *Int J Oncol* 2007; **31**: 1351-1355
 - 41 **Hughes SJ**, Nambu Y, Soldes OS, Hamstra D, Rehemtulla A, Iannettoni MD, Orringer MB, Beer DG. Fas/APO-1 (CD95) is not translocated to the cell membrane in esophageal adenocarcinoma. *Cancer Res* 1997; **57**: 5571-5578
 - 42 **Mahidhara RS**, Queiroz De Oliveira PE, Kohout J, Beer DG, Lin J, Watkins SC, Robbins PD, Hughes SJ. Altered trafficking of Fas and subsequent resistance to Fas-mediated apoptosis occurs by a wild-type p53 independent mechanism in esophageal adenocarcinoma. *J Surg Res* 2005; **123**: 302-311
 - 43 **van der Woude CJ**, Jansen PL, Tiebosch AT, Beuving A, Homan M, Kleibeuker JH, Moshage H. Expression of apoptosis-related proteins in Barrett's metaplasia-dysplasia-carcinoma sequence: a switch to a more resistant phenotype. *Hum Pathol* 2002; **33**: 686-692
 - 44 **Wilson KT**, Fu S, Ramanujam KS, Meltzer SJ. Increased expression of inducible nitric oxide synthase and cyclooxygenase-2 in Barrett's esophagus and associated adenocarcinomas. *Cancer Res* 1998; **58**: 2929-2934
 - 45 **Lagorce C**, Paraf F, Vidaud D, Couvelard A, Wendum D, Martin A, Fléjou JF. Cyclooxygenase-2 is expressed frequently and early in Barrett's oesophagus and associated adenocarcinoma. *Histopathology* 2003; **42**: 457-465
 - 46 **Cheong E**, Ivory K, Doleman J, Parker ML, Rhodes M, Johnson IT. Synthetic and naturally occurring COX-2 inhibitors suppress proliferation in a human oesophageal adenocarcinoma cell line (OE33) by inducing apoptosis and cell cycle arrest. *Carcinogenesis* 2004; **25**: 1945-1952
 - 47 **Majka J**, Rembiasz K, Migaczewski M, Budzynski A, Ptak-Belowska A, Pabianczyk R, Urbanczyk K, Zub-Pokrowiecka A, Matlok M, Brzozowski T. Cyclooxygenase-2 (COX-2) is the key event in pathophysiology of Barrett's esophagus. Lesson from experimental animal model and human subjects. *J Physiol Pharmacol* 2010; **61**: 409-418
 - 48 **Morales CP**, Lee EL, Shay JW. In situ hybridization for the detection of telomerase RNA in the progression from Barrett's esophagus to esophageal adenocarcinoma. *Cancer* 1998; **83**: 652-659
 - 49 **Lord RV**, Salonga D, Danenberg KD, Peters JH, DeMeester TR, Park JM, Johansson J, Skinner KA, Chandrasoma P, DeMeester SR, Bremner CG, Tsai PI, Danenberg PV. Telomerase reverse transcriptase expression is increased early in the Barrett's metaplasia, dysplasia, adenocarcinoma sequence. *J Gastrointest Surg* 2000; **4**: 135-142
 - 50 **Barclay JY**, Morris A, Nwokolo CU. Telomerase, hTERT and splice variants in Barrett's oesophagus and oesophageal adenocarcinoma. *Eur J Gastroenterol Hepatol* 2005; **17**: 221-227
 - 51 **Shammas MA**, Koley H, Batchu RB, Bertheau RC, Protopopov A, Munshi NC, Goyal RK. Telomerase inhibition by siRNA causes senescence and apoptosis in Barrett's adenocarcinoma cells: mechanism and therapeutic potential. *Mol Cancer* 2005; **4**: 24
 - 52 **Couvelard A**, Paraf F, Gratio V, Scoazec JY, Hénin D, Degott C, Fléjou JF. Angiogenesis in the neoplastic sequence of Barrett's oesophagus. Correlation with VEGF expression. *J Pathol* 2000; **192**: 14-18
 - 53 **Auvinen MI**, Sihvo EI, Ruohtula T, Salminen JT, Koivistoinen A, Siivola P, Rönholm R, Rämö JO, Bergman M, Salo JA. Incipient angiogenesis in Barrett's epithelium and lymphangiogenesis in Barrett's adenocarcinoma. *J Clin Oncol* 2002; **20**: 2971-2979
 - 54 **Saad RS**, El-Gohary Y, Memari E, Liu YL, Silverman JF. Endoglin (CD105) and vascular endothelial growth factor as prognostic markers in esophageal adenocarcinoma. *Hum Pathol* 2005; **36**: 955-961
 - 55 **Griffiths EA**, Pritchard SA, McGrath SM, Valentine HR, Price PM, Welch IM, West CM. Increasing expression of hypoxia-inducible proteins in the Barrett's metaplasia-dysplasia-adenocarcinoma sequence. *Br J Cancer* 2007; **96**: 1377-1383
 - 56 **Krishnadath KK**, Tilanus HW, van Blankenstein M, Hop WC, Kremers ED, Dinjens WN, Bosman FT. Reduced expres-

- sion of the cadherin-catenin complex in oesophageal adenocarcinoma correlates with poor prognosis. *J Pathol* 1997; **182**: 331-338
- 57 **Washington K**, Chiappori A, Hamilton K, Shyr Y, Blanke C, Johnson D, Sawyers J, Beauchamp D. Expression of beta-catenin, alpha-catenin, and E-cadherin in Barrett's esophagus and esophageal adenocarcinomas. *Mod Pathol* 1998; **11**: 805-813
- 58 **Bailey T**, Biddlestone L, Shepherd N, Barr H, Warner P, Jankowski J. Altered cadherin and catenin complexes in the Barrett's esophagus-dysplasia-adenocarcinoma sequence: correlation with disease progression and dedifferentiation. *Am J Pathol* 1998; **152**: 135-144
- 59 **Jethwa P**, Naqvi M, Hardy RG, Hotchin NA, Roberts S, Spychal R, Tselepis C. Overexpression of Slug is associated with malignant progression of esophageal adenocarcinoma. *World J Gastroenterol* 2008; **14**: 1044-1052
- 60 **Hughes SJ**, Glover TW, Zhu XX, Kuick R, Thoraval D, Orlinger MB, Beer DG, Hanash S. A novel amplicon at 8p22-23 results in overexpression of cathepsin B in esophageal adenocarcinoma. *Proc Natl Acad Sci USA* 1998; **95**: 12410-12415
- 61 **Cheng P**, Gong J, Wang T, Chen J, Liu GS, Zhang R. Gene expression in rats with Barrett's esophagus and esophageal adenocarcinoma induced by gastroduodenoesophageal reflux. *World J Gastroenterol* 2005; **11**: 5117-5122
- 62 **Breton J**, Gage MC, Hay AW, Keen JN, Wild CP, Donnellan C, Findlay JB, Hardie LJ. Proteomic screening of a cell line model of esophageal carcinogenesis identifies cathepsin D and aldo-keto reductase 1C2 and 1B10 dysregulation in Barrett's esophagus and esophageal adenocarcinoma. *J Proteome Res* 2008; **7**: 1953-1962
- 63 **Lagorce-Pages C**, Paraf F, Dubois S, Belghiti J, Fléjou JF. Expression of CD44 in premalignant and malignant Barrett's oesophagus. *Histopathology* 1998; **32**: 7-14
- 64 **Darlavoix T**, Seelentag W, Yan P, Bachmann A, Bosman FT. Altered expression of CD44 and DKK1 in the progression of Barrett's esophagus to esophageal adenocarcinoma. *Virchows Arch* 2009; **454**: 629-637
- 65 **Leedham SJ**, Preston SL, McDonald SA, Elia G, Bhandari P, Poller D, Harrison R, Novelli MR, Jankowski JA, Wright NA. Individual crypt genetic heterogeneity and the origin of metaplastic glandular epithelium in human Barrett's oesophagus. *Gut* 2008; **57**: 1041-1048
- 66 **Maley CC**, Galipeau PC, Li X, Sanchez CA, Paulson TG, Reid BJ. Selectively advantageous mutations and hitchhikers in neoplasms: p16 lesions are selected in Barrett's esophagus. *Cancer Res* 2004; **64**: 3414-3427
- 67 **Paulson TG**, Reid BJ. Focus on Barrett's esophagus and esophageal adenocarcinoma. *Cancer Cell* 2004; **6**: 11-16
- 68 **Ramel S**. Barrett's esophagus: model of neoplastic progression. *World J Surg* 2003; **27**: 1009-1013
- 69 **Falk GW**, Goldblum JR. Extent of low-grade dysplasia in Barrett's esophagus: is it useful for risk stratification? *Am J Gastroenterol* 2007; **102**: 494-496
- 70 **El-Serag HB**, Mason AC, Petersen N, Key CR. Epidemiological differences between adenocarcinoma of the oesophagus and adenocarcinoma of the gastric cardia in the USA. *Gut* 2002; **50**: 368-372
- 71 **Corley DA**, Kubo A, Levin TR, Block G, Habel L, Rumore G, Quesenberry C, Buffler P. Race, ethnicity, sex and temporal differences in Barrett's oesophagus diagnosis: a large community-based study, 1994-2006. *Gut* 2009; **58**: 182-188
- 72 **Wong A**, Fitzgerald RC. Epidemiologic risk factors for Barrett's esophagus and associated adenocarcinoma. *Clin Gastroenterol Hepatol* 2005; **3**: 1-10
- 73 **von Rahden BH**, Stein HJ, Siewert JR. Barrett's esophagus and Barrett's carcinoma. *Curr Oncol Rep* 2003; **5**: 203-209
- 74 **Rogers EL**, Goldkind SF, Iseri OA, Bustin M, Goldkind L, Hamilton SR, Smith RL. Adenocarcinoma of the lower esophagus. A disease primarily of white men with Barrett's esophagus. *J Clin Gastroenterol* 1986; **8**: 613-618
- 75 **Pondugula K**, Wani S, Sharma P. Barrett's esophagus and esophageal adenocarcinoma in adults: long-term GERD or something else? *Curr Gastroenterol Rep* 2007; **9**: 468-474
- 76 **Kubo A**, Corley DA. Marked multi-ethnic variation of esophageal and gastric cardia carcinomas within the United States. *Am J Gastroenterol* 2004; **99**: 582-588
- 77 **El-Serag HB**, Petersen NJ, Carter J, Graham DY, Richardson P, Genta RM, Rabeneck L. Gastroesophageal reflux among different racial groups in the United States. *Gastroenterology* 2004; **126**: 1692-1699
- 78 **Vaezi MF**, Richter JE. Synergism of acid and duodenogastroesophageal reflux in complicated Barrett's esophagus. *Surgery* 1995; **117**: 699-704
- 79 **Nehra D**, Howell P, Williams CP, Pye JK, Beynon J. Toxic bile acids in gastro-oesophageal reflux disease: influence of gastric acidity. *Gut* 1999; **44**: 598-602
- 80 **Menges M**, Müller M, Zeitz M. Increased acid and bile reflux in Barrett's esophagus compared to reflux esophagitis, and effect of proton pump inhibitor therapy. *Am J Gastroenterol* 2001; **96**: 331-337
- 81 **Theisen J**, Nehra D, Citron D, Johansson J, Hagen JA, Crookes PF, DeMeester SR, Bremner CG, DeMeester TR, Peters JH. Suppression of gastric acid secretion in patients with gastroesophageal reflux disease results in gastric bacterial overgrowth and deconjugation of bile acids. *J Gastrointest Surg* 2000; **4**: 50-54
- 82 **Wild CP**, Hardie LJ. Reflux, Barrett's oesophagus and adenocarcinoma: burning questions. *Nat Rev Cancer* 2003; **3**: 676-684
- 83 **Qiao D**, Gaitonde SV, Qi W, Martinez JD. Deoxycholic acid suppresses p53 by stimulating proteasome-mediated p53 protein degradation. *Carcinogenesis* 2001; **22**: 957-964
- 84 **Anderson LA**, Watson RG, Murphy SJ, Johnston BT, Comber H, Mc Guigan J, Reynolds JV, Murray LJ. Risk factors for Barrett's oesophagus and oesophageal adenocarcinoma: results from the FINBAR study. *World J Gastroenterol* 2007; **13**: 1585-1594
- 85 **Chak A**, Faulx A, Eng C, Grady W, Kinnard M, Ochs-Balcom H, Falk G. Gastroesophageal reflux symptoms in patients with adenocarcinoma of the esophagus or cardia. *Cancer* 2006; **107**: 2160-2166
- 86 **Ouatu-Lascar R**, Fitzgerald RC, Triadafilopoulos G. Differentiation and proliferation in Barrett's esophagus and the effects of acid suppression. *Gastroenterology* 1999; **117**: 327-335
- 87 **Attwood SE**, Harrison LA, Preston SL, Jankowski JA. Esophageal adenocarcinoma in " mice and men " : back to basics! *Am J Gastroenterol* 2008; **103**: 2367-2372
- 88 **Buttar NS**, Wang KK. Mechanisms of disease: Carcinogenesis in Barrett's esophagus. *Nat Clin Pract Gastroenterol Hepatol* 2004; **1**: 106-112
- 89 **Islami F**, Kamangar F, Boffetta P. Use of proton pump inhibitors and risk of progression of Barrett's esophagus to neoplastic lesions. *Am J Gastroenterol* 2009; **104**: 2646-2648
- 90 **Obszynska JA**, Atherfold PA, Nanji M, Glancy D, Santander S, Graham TA, Otto WR, West K, Harrison RF, Jankowski JA. Long-term proton pump induced hypergastrinaemia does induce lineage-specific restitution but not clonal expansion in benign Barrett's oesophagus in vivo. *Gut* 2010; **59**: 156-163
- 91 **Ye W**, Chow WH, Lagergren J, Yin L, Nyrén O. Risk of adenocarcinomas of the esophagus and gastric cardia in patients with gastroesophageal reflux diseases and after antireflux surgery. *Gastroenterology* 2001; **121**: 1286-1293
- 92 **Tran T**, Spechler SJ, Richardson P, El-Serag HB. Fundoplication and the risk of esophageal cancer in gastroesophageal reflux disease: a Veterans Affairs cohort study. *Am J Gastroenterol* 2005; **100**: 1002-1008
- 93 **Corey KE**, Schmitz SM, Shaheen NJ. Does a surgical antireflux procedure decrease the incidence of esophageal adeno-

- carcinoma in Barrett's esophagus? A meta-analysis. *Am J Gastroenterol* 2003; **98**: 2390-2394
- 94 **Lagergren J**, Bergström R, Adami HO, Nyrén O. Association between medications that relax the lower esophageal sphincter and risk for esophageal adenocarcinoma. *Ann Intern Med* 2000; **133**: 165-175
- 95 **Maaroos HI**, Vorobjova T, Sipponen P, Tammur R, Uibo R, Wadström T, Keevallik R, Villako K. An 18-year follow-up study of chronic gastritis and Helicobacter pylori association of CagA positivity with development of atrophy and activity of gastritis. *Scand J Gastroenterol* 1999; **34**: 864-869
- 96 **Rokkas T**, Pistiolas D, Sechopoulos P, Robotis I, Margantinis G. Relationship between Helicobacter pylori infection and esophageal neoplasia: a meta-analysis. *Clin Gastroenterol Hepatol* 2007; **5**: 1413-1417, 1417.e1-1417.e2
- 97 **Weston AP**, Badr AS, Topalovski M, Cherian R, Dixon A, Hassanein RS. Prospective evaluation of the prevalence of gastric Helicobacter pylori infection in patients with GERD, Barrett's esophagus, Barrett's dysplasia, and Barrett's adenocarcinoma. *Am J Gastroenterol* 2000; **95**: 387-394
- 98 **Wu AH**, Crabtree JE, Bernstein L, Hawtin P, Cockburn M, Tseng CC, Forman D. Role of Helicobacter pylori CagA+ strains and risk of adenocarcinoma of the stomach and esophagus. *Int J Cancer* 2003; **103**: 815-821
- 99 **Clemons NJ**, McColl KE, Fitzgerald RC. Nitric oxide and acid induce double-strand DNA breaks in Barrett's esophagus carcinogenesis via distinct mechanisms. *Gastroenterology* 2007; **133**: 1198-1209
- 100 **Forman D**, Al-Dabbagh S, Doll R. Nitrates, nitrites and gastric cancer in Great Britain. *Nature* 1985; **313**: 620-625
- 101 **Falk GW**. Risk factors for esophageal cancer development. *Surg Oncol Clin N Am* 2009; **18**: 469-485
- 102 **Thomas T**, Abrams KR, De Caestecker JS, Robinson RJ. Meta analysis: Cancer risk in Barrett's oesophagus. *Aliment Pharmacol Ther* 2007; **26**: 1465-1477
- 103 **Hampel H**, Abraham NS, El-Serag HB. Meta-analysis: obesity and the risk for gastroesophageal reflux disease and its complications. *Ann Intern Med* 2005; **143**: 199-211
- 104 **Cook MB**, Greenwood DC, Hardie LJ, Wild CP, Forman D. A systematic review and meta-analysis of the risk of increasing adiposity on Barrett's esophagus. *Am J Gastroenterol* 2008; **103**: 292-300
- 105 **Kubo A**, Corley DA. Body mass index and adenocarcinomas of the esophagus or gastric cardia: a systematic review and meta-analysis. *Cancer Epidemiol Biomarkers Prev* 2006; **15**: 872-878
- 106 **Murray L**, Romero Y. Role of obesity in Barrett's esophagus and cancer. *Surg Oncol Clin N Am* 2009; **18**: 439-452
- 107 **de Jonge PJ**, Steyerberg EW, Kuipers EJ, Honkoop P, Wolters LM, Kerkhof M, van Dekken H, Siersema PD. Risk factors for the development of esophageal adenocarcinoma in Barrett's esophagus. *Am J Gastroenterol* 2006; **101**: 1421-1429
- 108 **Oberg S**, Wenner J, Johansson J, Walther B, Willén R. Barrett esophagus: risk factors for progression to dysplasia and adenocarcinoma. *Ann Surg* 2005; **242**: 49-54
- 109 **Vaughan TL**, Kristal AR, Blount PL, Levine DS, Galipeau PC, Prevo LJ, Sanchez CA, Rabinovitch PS, Reid BJ. Nonsteroidal anti-inflammatory drug use, body mass index, and anthropometry in relation to genetic and flow cytometric abnormalities in Barrett's esophagus. *Cancer Epidemiol Biomarkers Prev* 2002; **11**: 745-752
- 110 **Konturek PC**, Burnat G, Rau T, Hahn EG, Konturek S. Effect of adiponectin and ghrelin on apoptosis of Barrett adenocarcinoma cell line. *Dig Dis Sci* 2008; **53**: 597-605
- 111 **Rubenstein JH**, Dahlkemper A, Kao JY, Zhang M, Morgenstern H, McMahon L, Inadomi JM. A pilot study of the association of low plasma adiponectin and Barrett's esophagus. *Am J Gastroenterol* 2008; **103**: 1358-1364
- 112 **Kendall BJ**, Macdonald GA, Hayward NK, Prins JB, Brown I, Walker N, Pandeya N, Green AC, Webb PM, Whiteman DC. Leptin and the risk of Barrett's oesophagus. *Gut* 2008; **57**: 448-454
- 113 **Francois F**, Roper J, Goodman AJ, Pei Z, Ghumman M, Mourad M, de Perez AZ, Perez-Perez GI, Tseng CH, Blaser MJ. The association of gastric leptin with oesophageal inflammation and metaplasia. *Gut* 2008; **57**: 16-24
- 114 **Ogunwobi O**, Mutungi G, Beales IL. Leptin stimulates proliferation and inhibits apoptosis in Barrett's esophageal adenocarcinoma cells by cyclooxygenase-2-dependent, prostaglandin-E2-mediated transactivation of the epidermal growth factor receptor and c-Jun NH2-terminal kinase activation. *Endocrinology* 2006; **147**: 4505-4516
- 115 **Dornonville de la Cour C**, Lindström E, Norlén P, Håkanson R. Ghrelin stimulates gastric emptying but is without effect on acid secretion and gastric endocrine cells. *Regul Pept* 2004; **120**: 23-32
- 116 **Mottershead M**, Kareris E, Barclay JY, Suortamo S, Newbold M, Randeve H, Nwokolo CU. Immunohistochemical and quantitative mRNA assessment of ghrelin expression in gastric and oesophageal adenocarcinoma. *J Clin Pathol* 2007; **60**: 405-409
- 117 **de Martel C**, Haggerty TD, Corley DA, Vogelstein JH, Orentreich N, Parsonnet J. Serum ghrelin levels and risk of subsequent adenocarcinoma of the esophagus. *Am J Gastroenterol* 2007; **102**: 1166-1172
- 118 **Kristal AR**, Blount PL, Schenk JM, Sanchez CA, Rabinovitch PS, Odze RD, Standley J, Vaughan TL, Reid BJ. Low-fat, high fruit and vegetable diets and weight loss do not affect biomarkers of cellular proliferation in Barrett esophagus. *Cancer Epidemiol Biomarkers Prev* 2005; **14**: 2377-2383
- 119 **Terry P**, Lagergren J, Hansen H, Wolk A, Nyrén O. Fruit and vegetable consumption in the prevention of oesophageal and cardia cancers. *Eur J Cancer Prev* 2001; **10**: 365-369
- 120 **Dong LM**, Kristal AR, Peters U, Schenk JM, Sanchez CA, Rabinovitch PS, Blount PL, Odze RD, Ayub K, Reid BJ, Vaughan TL. Dietary supplement use and risk of neoplastic progression in esophageal adenocarcinoma: a prospective study. *Nutr Cancer* 2008; **60**: 39-48
- 121 **Chen H**, Tucker KL, Graubard BI, Heineman EF, Markin RS, Potischman NA, Russell RM, Weisenburger DD, Ward MH. Nutrient intakes and adenocarcinoma of the esophagus and distal stomach. *Nutr Cancer* 2002; **42**: 33-40
- 122 **Rudolph RE**, Vaughan TL, Kristal AR, Blount PL, Levine DS, Galipeau PC, Prevo LJ, Sanchez CA, Rabinovitch PS, Reid BJ. Serum selenium levels in relation to markers of neoplastic progression among persons with Barrett's esophagus. *J Natl Cancer Inst* 2003; **95**: 750-757
- 123 **Caygill CP**, Johnston DA, Lopez M, Johnston BJ, Watson A, Reed PI, Hill MJ. Lifestyle factors and Barrett's esophagus. *Am J Gastroenterol* 2002; **97**: 1328-1331
- 124 **Gammon MD**, Schoenberg JB, Ahsan H, Risch HA, Vaughan TL, Chow WH, Rotterdam H, West AB, Dubrow R, Stanford JL, Mayne ST, Farrow DC, Niwa S, Blot WJ, Fraumeni JF. Tobacco, alcohol, and socioeconomic status and adenocarcinomas of the esophagus and gastric cardia. *J Natl Cancer Inst* 1997; **89**: 1277-1284
- 125 **Kabat GC**, Ng SK, Wynder EL. Tobacco, alcohol intake, and diet in relation to adenocarcinoma of the esophagus and gastric cardia. *Cancer Causes Control* 1993; **4**: 123-132
- 126 **Zhang ZF**, Kurtz RC, Sun M, Karpeh M, Yu GP, Gargon N, Fein JS, Georgopoulos SK, Harlap S. Adenocarcinomas of the esophagus and gastric cardia: medical conditions, tobacco, alcohol, and socioeconomic factors. *Cancer Epidemiol Biomarkers Prev* 1996; **5**: 761-768
- 127 **Brown LM**, Silverman DT, Pottner LM, Schoenberg JB, Greenberg RS, Swanson GM, Liff JM, Schwartz AG, Hayes RB, Blot WJ. Adenocarcinoma of the esophagus and esophagogastric junction in white men in the United States: alcohol, tobacco, and socioeconomic factors. *Cancer Causes Control* 1994; **5**: 333-340

- 128 **Vaughan TL**, Davis S, Kristal A, Thomas DB. Obesity, alcohol, and tobacco as risk factors for cancers of the esophagus and gastric cardia: adenocarcinoma versus squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev* 1995; **4**: 85-92
- 129 **Menke-Pluymers MB**, Hop WC, Dees J, van Blankenstein M, Tilanus HW. Risk factors for the development of an adenocarcinoma in columnar-lined (Barrett) esophagus. The Rotterdam Esophageal Tumor Study Group. *Cancer* 1993; **72**: 1155-1158
- 130 **Achkar JP**, Post AB, Achkar E, Carey WD. Risk of extraesophageal malignancy in patients with adenocarcinoma arising in Barrett's esophagus. *Am J Gastroenterol* 1995; **90**: 39-43
- 131 **Gray MR**, Donnelly RJ, Kingsnorth AN. The role of smoking and alcohol in metaplasia and cancer risk in Barrett's columnar lined oesophagus. *Gut* 1993; **34**: 727-731
- 132 **Levi F**, Ollyo JB, La Vecchia C, Boyle P, Monnier P, Savary M. The consumption of tobacco, alcohol and the risk of adenocarcinoma in Barrett's oesophagus. *Int J Cancer* 1990; **45**: 852-854
- 133 **Lagergren J**, Bergström R, Lindgren A, Nyrén O. The role of tobacco, snuff and alcohol use in the aetiology of cancer of the oesophagus and gastric cardia. *Int J Cancer* 2000; **85**: 340-346
- 134 **Avidan B**, Sonnenberg A, Schnell TG, Chejfec G, Metz A, Sontag SJ. Hiatal hernia size, Barrett's length, and severity of acid reflux are all risk factors for esophageal adenocarcinoma. *Am J Gastroenterol* 2002; **97**: 1930-1936
- 135 **Smith KJ**, O'Brien SM, Smithers BM, Gotley DC, Webb PM, Green AC, Whiteman DC. Interactions among smoking, obesity, and symptoms of acid reflux in Barrett's esophagus. *Cancer Epidemiol Biomarkers Prev* 2005; **14**: 2481-2486
- 136 **Corley DA**, Kerlikowske K, Verma R, Buffler P. Protective association of aspirin/NSAIDs and esophageal cancer: a systematic review and meta-analysis. *Gastroenterology* 2003; **124**: 47-56
- 137 **Heath EI**, Canto MI, Piantadosi S, Montgomery E, Weinstein WM, Herman JG, Dannenberg AJ, Yang VW, Shar AO, Hawk E, Forastiere AA. Secondary chemoprevention of Barrett's esophagus with celecoxib: results of a randomized trial. *J Natl Cancer Inst* 2007; **99**: 545-557
- 138 **Gatenby PA**, Ramus JR, Caygill CP, Winslet MC, Watson A. Aspirin is not chemoprotective for Barrett's adenocarcinoma of the oesophagus in multicentre cohort. *Eur J Cancer Prev* 2009; **18**: 381-384

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