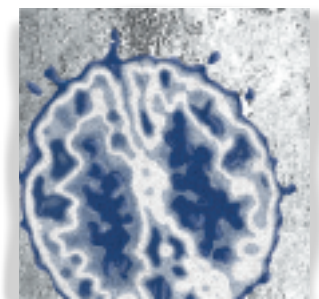


Basic research

Fleshing out the amyloid cascade hypothesis: the molecular biology of Alzheimer's disease

Simon Lovestone, PhD



Alzheimer's disease (AD) is a disorder of two pathologies—plaques and tangles. The former have as a key constituent amyloid protein and the latter the microtubule-associated protein tau. Genetics has demonstrated that changes in either protein are sufficient to cause dementia. The amyloid cascade hypothesis proposes that plaque-related changes precede tangle-related changes and positions amyloid as central to the degeneration of AD. All the evidence suggests this is correct, including evidence that presenilins alter the processing of the amyloid precursor protein and evidence that disrupting the normal properties of tau underlies the related frontotemporal dementias. The amyloid cascade hypothesis has provided the basis for nearly a decade of intensive basic science—the skeleton of that hypothesis can now be fleshed out, and confidence is growing that this will result in useful disease-modifying therapies in the future.

Keywords: amyloid; tau; Alzheimer's disease; tauopathy; GSK-3; APP; presenilin; Notch

Author affiliations: Institute of Psychiatry, De Crespigny Park, London, UK

Address for correspondence: Prof Simon Lovestone, Institute of Psychiatry, De Crespigny Park, London SE5 8AF, UK
(e-mail: s.lovestone@iop.kcl.ac.uk)

Basic research into Alzheimer's disease (AD) more than two decades ago demonstrated early and profound loss of cholinergic neurons, a finding that led to the first therapeutic advance with the development and licensing of the first specific treatments: the acetylcholinesterase inhibitors. Whatever the therapeutic efficiency of these compounds, their impact in the field of dementia care cannot be overestimated. However, today's basic research has the power to go beyond the cholinergic hypothesis, and there is every hope that the current process of fleshing out the bones of the amyloid cascade hypothesis will yield effective disease-modifying treatments.

The amyloid cascade hypothesis

In 1992, soon after the discovery of mutations in the amyloid precursor protein gene, John Hardy proposed the amyloid cascade hypothesis, which in its most basic form states that amyloid is at the center of the pathophysiology, that amyloid deposits in AD result from a multitude of genetic or environmental insults and are at the origin of the neurodegeneration that leads to dementia.¹ Although many new questions have arisen—for instance, is the pathogenic amyloid intracellular and soluble or extracellular and fibrillar?—the hypothesis not only stands, but has been confirmed with each new advance of recent years. Furthermore, important aspects of basic research are omitted from the cascade, or at least cannot at present be easily fitted into the cascade, including the role of inflammation and the putative pathogenic events resulting from risk factors such as prior affective disorder or hypertension. Nevertheless, most of the molecular and cellular biology of AD can be discussed in the context of this important framework.

Basic research

Selected abbreviations and acronyms

AD	<i>Alzheimer's disease</i>
apo E	<i>apolipoprotein E</i>
APP	<i>amyloid precursor protein</i>
BACE	<i>beta-site APP-cleaving enzyme</i>
DRAP	<i>Down's region aspartic protease</i>
FTDP-17	<i>frontotemporal dementia and parkinsonism linked to chromosome 17</i>
GSK-3	<i>glycogen synthase kinase-3</i>
NFT	<i>neurofibrillary tangle</i>
PHF	<i>paired helical filaments</i>
PKC	<i>protein kinase C</i>
PP2A	<i>type 2A protein phosphatase</i>
PS-1, -2	<i>presenilin-1 and -2</i>
PSP	<i>progressive supranuclear palsy</i>
TPK1	<i>tau protein kinase 1</i>

APP and the formation of plaques

The core component of plaques is a 4-kd peptide known as A β .^{2,3} In plaques, the peptide forms fibrils in a beta-pleated sheet configuration, thus assuming the properties of amyloid characterized by its unique birefringence with Congo red staining. A β is derived from amyloid precursor protein (APP), the gene for which is on chromosome 21. The discovery that mutations in the APP gene cause a rare form of autosomal dominant AD confirmed the process of A β formation from APP as central to the etiopathogenesis of AD.^{4,8} APP is a ubiquitous and large single-pass membrane-spanning protein, the function of which is not clear, although there are suggestions that it may have a role in cell-to-cell contact signaling or neurite outgrowth.^{9,10} When derived from APP, A β is a peptide of between 40 and 43 amino acids that has a tendency to aggregate *in vitro*. This tendency is enhanced in the longer forms of the peptide, suggesting that these slightly larger peptides are more pathogenic (and that inhibiting fibril formation may therefore be therapeutic).¹¹⁻¹³ Although the process *in vivo* is not understood, it is assumed that A β peptide is formed intracellularly and then aggregates either within the cell or after release into the extracellular space. However, some early work did find intracellular fibrils in cells expressing the c-terminal fragment of APP, and increasing attention is being paid to the possibility of intracellular A β toxicity.¹⁴⁻¹⁶

These deposits of A β form diffuse plaques visible on immunohistochemistry in affected regions of the brain. Technically, as these diffuse plaques consist only of fibrillized extracellular peptide that is not in a beta-pleated sheet configuration and hence not birefringent, they cannot properly be said to be amyloid. Careful studies of Down's syndrome brains suggest a sequential series of steps whereby diffuse plaques form the neuritic or classic plaque containing true amyloid, which in time evolves to form the burnt-out plaque where only the amyloid deposit remains.¹⁷

Understanding the process whereby A β is generated from APP is of the utmost importance and is the most obvious target for therapy. APP is metabolized through two opposing pathways involving three proteases.¹⁸ The first, often called the nonamyloidogenic pathway, results in cleavage of APP within the A β sequence moiety by the putative α -secretase. It is thought that α -secretase cleavage occurs at the extracellular membrane, but it is clear that it results in the secretion of the large extracellular portion of APP known as sAPP α . The function of this secreted peptide is not fully understood, but α -secretase cleavage certainly prevents the formation of A β as the cleavage site is within this part of the protein. Although the enzyme itself has not yet been identified, the regulation of the activity of α -secretase has been extensively examined. Phorbol ester activation of protein kinase C (PKC) increases sAPP α secretion into the medium of transfected cells, and in neurons very considerably so.¹⁹⁻²³ Interestingly, the same observation was made when acetylcholine receptors linked through second messengers to PKC were stimulated. Stimulation of other PKC-linked receptors also stimulates sAPP α release, whereas stimulation of muscarinic receptors linked to cyclic adenosine monophosphate does not.²⁴ These findings are intriguing and may have therapeutic significance, especially as a similarly beneficial effect of muscarinic stimulation is seen in a process thought to underlie the formation of tangles.

In contrast to nonamyloidogenic processing of APP, the production of A β necessitates two protease activities. The previously named enzyme β -secretase was recently identified and renamed BACE (for beta-site APP-cleaving enzyme).²⁵ Interestingly, a very similar protease was found near the region on chromosome 21 critical for Down's syndrome (Down's region aspartic protease [DRAP], or BACE2). These proteases cleave APP

within the extracellular domain, probably in the endosomal-lysosomal pathway following reinternalization of extracellular membrane-bound APP that escapes α -secretase cleavage.²⁶ Action of the putative protease γ -secretase at a second site releases free A β of between 40 and 42 amino acids, depending on the exact site of cleavage. The γ -secretase site is unusual in that it is buried within the lipid bilayer.

Mutations in APP and the formation of A β

Activity of all three secretases can be found in normal brain. A β and APPs can be detected from normal cells, and, in humans, A β is detectable by enzyme-linked immunosorbent assay (ELISA) in cerebrospinal fluid (CSF) as well as in serum. These, then, are not pathological processes per se, but rather they suggest that disease results from a tendency towards the amyloidogenic combination of secretases resulting, over a lifetime, in increased A β formation and increased plaque formation. What then are the known influences on these, essentially normal, processes? The first influence on APP metabolism to be discovered was the mutations in APP. Autosomal dominant AD in a few rare families results from mutations that cluster adjacent to the regions of α -, β -, or γ -secretase cleavage. The first set of mutations to be discovered were those clustering at, or adjacent to, the γ -secretase cleavage site (APP717). Expression of these mutated APP cDNAs in cells confirmed that the mutation does indeed alter APP metabolism, as relatively more of the longer forms of A β were generated in mutation-carrying cells.^{27,28} Mutations at the c-terminal end of the A β sequence within APP also alter APP metabolism, presumably by interfering with BACE. These mutations, the double Swedish mutation (APP670/671), also alter APP metabolism in cultured cells, and the amount of A β in serum or CSF of patients carrying either the mutations near the γ - or the β -secretase site is increased.^{29,30} Two very interesting mutations occur within the A β region close to the α -secretase site. One, at APP693, is associated with a rare disorder, hereditary cerebral hemorrhage with amyloidosis, Dutch type, and the other, at APP692, with presenile dementia and cerebral hemorrhage due to cerebral amyloid angiopathy—a clearly related, but not identical, disorder. In the APP692 disorder, but not in APP693 disease, there was not only angiopathy but large plaques and neurofibrillary tangles.³¹ In cells, the effect of the APP692

mutation is to increase both A β 40 and A β 42 secretion, whereas APP693 does not. Thus there is, in the APP mutations, convincing evidence in favor of the amyloid cascade hypothesis—mutations associated with AD increase either all A β or the longer and more fibrillogenic forms of A β , whereas mutations associated with other disease do not.

Presenilins and APP metabolism

Mutations in two very closely homologous genes—presenilin-1 and -2 (PS-1 and -2)—also cause early-onset autosomal dominant AD.^{32,33} The proteins encoded by these genes are multipass membrane-associated proteins that are certainly present in endoplasmic reticulum and possibly in nuclear envelope and plasma membrane as well.³⁴⁻³⁷ The normal biology of the presenilins is under extensive examination, and transgenic animals have already provided some insight into this. Overexpression of APP with the disease causing mutations results in plaque-like deposits of amyloid in mice, and this process is accelerated in mice overexpressing mutated PS-1.^{38,39} Knockouts of PS-1, however, are embryonically lethal. Studies from neurons from these animals, among other data, strongly suggest that the presenilins function as γ -secretase or as regulators of γ -secretase, as these neurons produce low levels of A β , resulting from low levels of γ -secretase activity.^{40,41} Whether the Alzheimer-related mutations increase γ -secretase activity or have some other gain-of-function activity is not entirely clear, but from the transgenic animals, studies in transfected cells, and studies in fibroblasts from families carrying these mutations, it is clear that the PS-1 mutations somehow increase the production, especially of the longer forms of the amyloid peptides, offering more evidence that the amyloid cascade hypothesis is correct to position APP processing as a central event in pathogenesis.⁴²

Neurofibrillary tangles and the tau question

If there has been any real controversy associated with the amyloid cascade hypothesis, this has been with the question of tau and neurofibrillary tangles (NFTs). These neuronal inclusion bodies are a defining feature of AD and are also found in other degenerative disorders such as dementia pugilistica and certain frontotemporal

Basic research

dementias. The number of NFTs correlates extremely well with dementia severity, in contrast to plaques where some analyses of total amyloid load correlate with dementia, but other neuropathological studies show no such correlation.⁴³⁻⁴⁵ Furthermore, NFTs show an anatomical localization in those regions where function is lost, occurring first in the transentorhinal region and spreading to hippocampal regions and then to cortex, but never occurring in cerebellum.^{46,47} Plaques, on the other hand, show no such consistent progression, and while they do occur in some quantity in the hippocampus where function is lost, they also occur in cerebellum, where no such loss is noted in dementia.⁴⁸ Finally, NFTs are intraneuronal lesions, the neurons containing NFTs show loss of vital intracellular organization with the loss of normal neuronal cytoskeleton, and there is convincing neuropathological evidence that the presence of NFTs heralds the death of that neuron. All this circumstantial evidence points very firmly in the direction of NFTs being essential pathological components of the cascade resulting in dementia. Nonetheless, there was some dissension from this view—perhaps NFTs were a nonessential by-product of neurodegeneration, an epiphenomenon.

Under the electron microscope, NFTs can be seen to consist principally of paired helical filaments together with a smaller proportion of straight filaments. These filaments are composed of the microtubule-associated protein tau, present in a highly phosphorylated state, and are abnormal, being found only in dementia. In the normal state, tau is expressed to a significant extent only in neurons where it is present in axons. Here it acts to stabilize microtubules, which are an essential component of the cellular cytoskeleton and in neurons assume a straight track parallel to axons. Microtubules are essential for fast axonal transport, the process whereby vesicles and other organelles such as mitochondria are transported from the cell body to distal parts of the neuron including synapses. The consequences of loss of fast axonal transport from the neuron or destruction of microtubules are not fully understood, but would be expected to result in loss of function of the neuron if not loss of viability. Tau, therefore, has an important role in regulating the stability and function of neurons. *In vitro*, tau binds to tubulin (the building block of the microtubule itself) and promotes the formation of tubulin polymers and the extension of these polymers into microtubules. Six different isoforms of tau are generated from

a single gene in the central nervous system, and there is some evidence that these isoforms have different abilities to promote microtubule assembly *in vitro*. There is developmental regulation of the expression of these isoforms, as in the fetal forms, which bind microtubules that are in excess relatively weakly, with a change to stronger binding isoforms on maturation. However, such regulation is a relatively slow process and real-time regulation of the properties of tau is almost certainly altered by the phosphorylation state of tau.

Tau phosphorylation—regulation of microtubule stability and role in Alzheimer's disease

Tau is a highly phosphorylated protein, and its ability to bind microtubules is regulated by this phosphorylation—the more phosphates, the less tau promotes microtubule assembly.⁴⁹ There is some controversy as to whether it is the amount of phosphorylation that is important or whether there are specific sites in tau that are critical in tau-tubulin interactions.⁵⁰ In the fetus, tau is very highly phosphorylated, and even in normal adult human brain examined in biopsy samples the amount of phosphorylation is relatively high.^{51,52} It is likely that acute regulation by a combination of kinases and phosphatases of tau phosphorylation controls the properties of neurons, which in turn alters the rate of transport within the neuron and, perhaps, other, structural, properties of tau. Even though tau is phosphorylated in normal adult neurons, and more so in normal fetal neurons, in the PHF-tau aggregates of AD, tau is even more phosphorylated. The amount of phosphorylation is higher in total terms, and there may be specific sites of tau that are phosphorylated only in AD. Functional studies of tau from human brain reflect this phosphorylation, with tau from fetal brain being less able to promote microtubule association *in vitro* than normal brain, and tau from AD brain being even less able to stabilize microtubule formation than fetal tau.⁴⁹ It is not yet clear whether tau phosphorylation and the functional deficiencies seen in tau from AD brain precedes or follows aggregation. However, careful pathological studies suggest that phosphorylated epitopes of tau appear in neurons together with the appearance of tau in the cell bodies of affected neurons (tau normally being seen only in axons) before the presence of aggregates of tau in NFTs.^{46,53} It is at least a viable hypothesis that an alteration in the phosphorylation state of tau results in a failure to bind microtubules, a consequent

accumulation in cell bodies, and eventual loss of microtubules and aggregation of tau into NFTs.

This hypothesis led to an intensive search for the kinases and phosphatases that might regulate tau. Of the phosphatases, type 2A protein phosphatase (PP2A) would appear to be the most viable candidate. In vitro, PP2A readily phosphorylates tau, it is found associated with microtubules, and, in cells, inhibition of PP2A results in an increase in the phosphorylation state of tau.⁵⁴⁻⁵⁶ A parallel investigation of the kinases responsible for tau phosphorylation has proved more controversial. Many kinases act on the common serine and threonine sites phosphorylated in paired helical filaments (PHF)-tau. However, in cells, we demonstrated that it is only glycogen synthase kinase-3 (GSK-3) that is able to phosphorylate tau readily at epitopes also phosphorylated in AD.^{57,58} Simultaneously, Ishiguro and colleagues purified a kinase from brain that readily phosphorylated tau, which they named tau protein kinase 1 (TPK1).⁵⁹ On purification, TPK1 was found to be GSK-3, and, although other kinases certainly do phosphorylate tau and may even be necessary to prime tau for subsequent phosphorylation, it does appear now that GSK-3 is the predominant kinase at these sites in brain.⁶⁰ Functional studies have added weight to the growing evidence for a role of GSK-3 in the phosphorylation of tau in vivo as GSK-3 activity alters the properties of tau, reducing its ability to bind and promote microtubule assembly in vitro and, in cells, reduces the ability of tau to alter the morphology and stability of microtubules.⁶¹

Regulation of the phosphorylation of tau

Interesting findings have emerged from studies of GSK-3 regulation, which might begin to tie together the two strands of AD basic science—the amyloid strand and the tau strand. Most enticingly, A β is neurotoxic to neurons in culture and matured and fibrillized A β peptides increase tau phosphorylation.^{62,63} Inhibiting GSK-3 activity protects neurons, suggesting that GSK-3 might be an intermediary step between amyloid and tau phosphorylation.^{64,65} One approach to inhibition of GSK-3 that has been used in these studies is lithium. Lithium results in developmental abnormalities in experimental models that mimic a signal transduction cascade known as Wingless (wnt in mammals). Wingless or wnt signaling results in GSK-3 inhibition, and this led Klein and Melton to hypothesize and then demonstrate that

lithium mimics Wingless signal by inhibiting GSK-3.⁶⁶ In nonneuronal cells, in neurons, and in animals, lithium has now been shown to reduce tau phosphorylation as would be expected if GSK-3 is a predominant tau-kinase.⁶⁷⁻⁷² This inhibition of GSK-3 alters the properties of tau in neurons and in living nonneuronal cells, and does so within the therapeutic range of lithium. This body of work does raise the interesting question as to whether GSK-3 is the target of lithium in the therapy of affective disorders, especially as another agent used in bipolar disorder, sodium valproate, also inhibits GSK-3.⁷³ Attention has recently turned to a pathway that interacts with Wingless signaling—the Notch pathway. Notch is a transmembrane protein essential for neurogenesis, but also present, and presumably therefore active, in adult brain.⁷⁴⁻⁷⁶ Activation of Notch involves cleavage within the membrane domain, very reminiscent of the γ -secretase cleavage of APP.⁷⁷ A role for presenilins in Notch activity was first suggested by homology as the equivalent of presenilins in *Caenorhabditis elegans*, SEL12, is associated with LIN12, the *C. elegans* equivalent of Notch. Human presenilins are able to compensate for loss of SEL12, but mutated human presenilins lose this ability.^{78,79} In a number of different mammalian model experiments, the presenilin protein has now been shown to activate Notch.⁷⁹⁻⁸⁴ The evidence that presenilins are involved in Notch signaling is now compelling, and this is intriguing, as Notch signaling and Wingless signaling interact.⁸⁵⁻⁸⁷ In the Wingless signal cascade, inhibition of GSK-3 results in accumulation of a protein called β -catenin, and, to add to the complexity of this area, presenilins bind to catenins and affect β -catenin signaling.⁸⁸⁻⁹² Much needs to be done to untangle this complicated set of observations, not all of which are consistent. However, it does appear to be the case that Wingless and Notch signaling interact, and that, in doing so, GSK-3 activity is regulated, and that the presenilins are involved—certainly with Notch signaling, and possibly with Wingless signaling.

In addition to Wingless/wnt signaling, GSK-3 is inhibited by insulin signaling through protein kinase B (PKB) and PI3-kinase. As predicted, insulin not only reduces tau phosphorylation in neurons, but also increases tau-microtubule interactions.⁹³ Just as GSK-3 might be the missing link between amyloid and tau, so too might GSK-3 be the missing link between an important finding from epidemiology and etiopathogenesis. Diabetes has now been shown to be a significant risk factor for AD.⁹⁴

Basic research

This finding is not explained simply by the confounding factor of increasing vascular risk in people with diabetes, and the finding that insulin resistance also increases risk of AD suggests that the pathogenic factor might be a failing of insulin signaling.^{95,96} If insulin signaling is deficient in some way, then might GSK-3 escape normal regulation? If this were so, then the predicted result would be increased tau phosphorylation and increased neuronal vulnerability.

Tau and the tauopathies

All doubt about the role of tau in dementia was finally laid to rest, however, when mutations in tau were shown to be the cause of some familial dementias.⁹⁷⁻⁹⁹ Mutations in tau, both missense coding mutations and intronic, were found in some families with frontotemporal dementia and parkinsonism linked to chromosome 17 (FTDP-17). These families have a clinical appearance of a frontal lobe dementia, very similar in presentation to Pick's disease, but with some parkinsonism. On neuropathology, many have tau inclusion bodies in either glia or neurons or both.¹⁰⁰ A new classification of certain dementia disorders has now arisen, including Pick's disorder, progressive supranuclear palsy (PSP), and the frontotemporal dementias, which have variable amounts of tau pathology, in some cases caused by tau mutations—these disorders being now known as the tauopathies.^{101,102} Ironically, it took the tauopathies to confirm the amyloid cascade hypothesis—mutations in APP give rise to both plaques and tangles, while mutations in tau give rise to tangles only. This is exactly the design of a genetic experiment to investigate sequential biochemical steps in a model organism. It follows, without any doubt at all, that the direction of effect is from amyloid through tau to dementia, and that tau is an essential part of the cascade. It does not follow that there are not other mechanisms whereby dementia can occur, and it might be that in some instances a remote event might give rise independently to both plaque and tangle pathology, although Occam's razor argues against this. The effect of the intronic tau mutations appears to be to alter the proportion of isoforms with 3- and 4- microtubule binding domains expressed in brain. The mutations cluster at the splice site for these alternative isoforms and disrupt splicing.¹⁰³⁻¹⁰⁵ This is very much in line with the biochemistry from pathological samples in these cases, which suggests that in frontotemporal

dementia there is a disruption in the normal equal expression of 3- and 4- repeat isoforms. Both in vitro and in vivo studies of the exonic missense mutations suggest that these disrupt microtubule binding.¹⁰⁶⁻¹⁰⁸ In our own studies, we showed that the mutations reduce the ability of tau to promote microtubule extension in cells in exactly the same manner as phosphorylation.¹⁰⁹ Other in vitro studies have suggested that the mutations in tau increase its propensity to self-aggregation.¹¹⁰

A molecular model of Alzheimer's disease

The amyloid cascade hypothesis can now be elaborated in some detail. Normally, APP is processed via both amyloidogenic and nonamyloidogenic routes. A number of events perturb the balance to a greater or lesser extent. Mutations in APP profoundly bias metabolism toward the amyloidogenic route, and head injury increases amyloidogenesis perhaps by simply increasing the total levels of APP expression. Somehow, amyloid production increases tau phosphorylation. Perhaps the most likely hypothesis at the present time is that amyloid peptide increases GSK-3 activity, although whether this is through intra- or extracellular amyloid is uncertain. GSK-3 activity increases tau phosphorylation, which then fails to bind microtubules, resulting in loss of microtubule stability and accumulation of tau in the cell body, which predisposes to tau aggregation. Mutations in tau also cause increased aggregation and reduced binding to microtubules in a manner analogous to phosphorylation. Mutations in presenilins certainly cause increased amyloid production from APP and might also have other effects including through Notch and/or Wingless signaling that might impact upon tau phosphorylation.

What else is known about AD that impacts upon the cascade? Most obviously omitted from this scheme is apolipoprotein E (apo E), the only confirmed genetic association with late-onset AD.^{111,112} Studies of the biology of apo E have proved very difficult to conduct, with disparate results partly accounted for by technical differences in the preparation of apo E protein. Apo E has been shown to interact with amyloid peptide, but some studies show greater interaction with apo E2 and others with apo E4.¹¹³⁻¹¹⁵ Depending upon the true result in vivo, apo E binding might enhance amyloid fibrillization and hence plaque formation, or enhance amyloid clearance and hence plaque destruction. Alternatively, apo E might affect tau phosphorylation. Tau binds apo E in an iso-

form-dependent manner, and it was hypothesized that such binding would alter the phosphorylation state of tau.¹¹⁶⁻¹¹⁸ We have confirmed this is in fact the case (unpublished observations), although whether this occurs in vivo is uncertain. Indeed it is not even known if tau and apo E would meet in vivo. Some studies suggest extracellular apo E is internalized into the cytoplasm compartment.^{119,120} At least one study suggests it is not.¹²¹ In neurons, apo E appears to be in the cytoplasm, but this might result from expression of apo E in a form that is not immediately secreted.¹²²⁻¹²⁴ Other cellular approaches do suggest tau alters microtubules and affects neuronal growth, both compatible with, but not proving, an effect of apo E on tau.^{120,125-127} It might be that apo E has no effect on either tau or amyloid, affecting instead local cholesterol transport, neuronal viability, and resilience to damage. At present, apo E can be slotted into the cascade in too many places to be sure which is the most likely.

Epidemiology has identified a few nonaging, nongenetic factors that do fit in with the hypothesis. Head injury, for example, might influence AD by increasing amyloid production. Diabetes or insulin-resistance syndrome might affect AD by reducing inhibition of GSK-3 and increasing tau pathology. It will be interesting over the forthcoming years to see how other factors, and the genetic factors in particular, which will be identified following the systematic genome scans, enhance our understanding of the cascade. For now, however, it is clear that substantial parts of the cascade of events leading to neuronal death and dementia are understood, and, most importantly, the race is now on to convert these targets for therapies into compounds that might delay, prevent, or possibly even reverse this devastating disease. □

Reforzando la hipótesis de la cascada del amiloide: la biología molecular de la Enfermedad de Alzheimer

La Enfermedad de Alzheimer (EA) es un trastorno de dos patologías: placas y ovillos. Las primeras están constituidas por proteínas de amiloide y los segundos por microtúbulos asociados a la proteína tau. La genética ha demostrado que los cambios en cualquiera de las proteínas son suficientes para causar demencia. La hipótesis de la cascada de amiloide propone que los cambios relacionados con las placas preceden a los cambios asociados con los ovillos y las posiciones del amiloide como elementos centrales en la degeneración de la EA. Todas las evidencias actuales sugieren que esta hipótesis es correcta; hay evidencias que señalan que las presenilinas alteran el procesamiento de la proteína precursora de amiloide y otras que demuestran que los trastornos en las propiedades normales de la proteína tau subyacen a las demencias frontotemporales. La hipótesis de la cascada de amiloide ha proporcionado las bases, por casi una década, de numerosos trabajos en ciencias básicas (el esqueleto de esta hipótesis ahora puede ser reforzado y existe gran confianza en que esto se traducirá en terapias útiles que modificarán la enfermedad en el futuro).

Développement de l'hypothèse de la cascade amyloïde : biologie moléculaire de la maladie d'Alzheimer

La maladie d'Alzheimer (MA) est un trouble résultant de deux pathologies : les plaques séniles et la dégénérescence neurofibrillaire. La première a comme constituant clé la protéine amyloïde et la seconde la protéine tau liée aux microtubules. La recherche en génétique a démontré que des modifications d'une des deux protéines sont suffisantes pour provoquer une démence. Selon l'hypothèse de la cascade amyloïde, les modifications liées à la plaque précèderaient celles liées à la dégénérescence neurofibrillaire, conférant ainsi au peptide amyloïde une place centrale dans le processus dégénératif de la MA. Cette hypothèse semble actuellement étayée par l'ensemble des études, tant celles montrant que les présénilines altèrent le processus du peptide précurseur de l'amyloïde que celles indiquant que l'altération des propriétés normales de la protéine tau est à l'origine du développement des démences frontotemporales. L'hypothèse de la cascade amyloïde a servi de moteur à presque une décennie de recherche intensive dans le domaine des sciences fondamentales. Il reste désormais à concrétiser cette hypothèse avec l'espoir grandissant de voir se développer dans l'avenir des traitements de fond pour cette maladie.

Basic research

REFERENCES

- Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. *Science*. 1992;256:184-185.
- Storey E, Cappai R. The amyloid precursor protein of Alzheimer's disease and the A β peptide. *Neuropathol Appl Neurobiol*. 1999;25:81-97.
- Wilson CA, Doms RW, Lee VMY. Intracellular APP processing and A β production in Alzheimer disease. *J Neuropathol Exp Neurol*. 1999;58:787-794.
- Rubinsztein DC. The genetics of Alzheimer's disease. *Prog Neurobiol*. 1997;52:447-454.
- Goate A, Chartier-Harlin MC, Mullan M, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature*. 1991;349:704-706.
- Kamino K, Orr HT, Payami H, et al. Linkage and mutational analysis of familial Alzheimer disease kindreds for the APP gene region. *Am J Hum Genet*. 1992;51:998-1014.
- Clark RF, Goate AM. Molecular genetics of Alzheimer's disease. *Arch Neurol*. 1993;50:1164-1172.
- Lannfelt L, Viitanen M, Johansson K, et al. Low frequency of the APP 670/671 mutation in familial Alzheimer's disease in Sweden. *Neurosci Lett*. 1993;153:85-87.
- Milward EA, Papadopoulos R, Fuller SJ, et al. The amyloid protein precursor of Alzheimer's disease is a mediator of the effects of nerve growth factor on neurite outgrowth. *Neuron*. 1992;9:129-137.
- Shea TB, Beermann ML, Honda T, Nixon RA. Secretion of amyloid precursor protein and laminin by cultured astrocytes is influenced by culture conditions. *J Neurosci Res*. 1994;37:197-207.
- Walsh DM, Hartley DM, Kusumoto Y, et al. Amyloid β -protein fibrillogenesis—structure and biological activity of protofibrillar intermediates. *J Biol Chem*. 1999;274:25945-25952.
- Lazo ND, Downing DT. Amyloid fibrils may be assembled from β -helical protofibrils. *Biochemistry*. 1998;37:1731-1735.
- Bandiera T, Lansen J, Post C, Varasi M. Inhibitors of A β peptide aggregation as potential anti-Alzheimer agents. *Curr Med Chem*. 1997;4:159-170.
- Maruyama K, Terakado K, Usami M, Yoshikawa K. Formation of amyloid-like fibrils in COS cells overexpressing part of the Alzheimer amyloid protein precursor. *Nature*. 1990;347:566-569.
- Yamaguchi H, Yamazaki T, Ishiguro K, et al. Ultrastructural localization of Alzheimer amyloid β A4 protein precursor in the cytoplasm of neurons and senile plaque-associated astrocytes. *Acta Neuropathol (Berl)*. 1992;85:15-22.
- Ferreira A, Caceres A, Kosik KS. Intraneuronal compartments of the amyloid precursor protein. *J Neurosci*. 1993;13:3112-3123.
- Mann DM, Brown A, Prinza D, et al. An analysis of the morphology of senile plaques in Down's syndrome patients of different ages using immunocytochemical and lectin histochemical techniques. *Neuropathol Appl Neurobiol*. 1989;15:317-329.
- Gandy S, Greengard P. Regulated cleavage of the Alzheimer amyloid precursor protein: molecular and cellular basis. *Biochimie*. 1994;76:300-303.
- Gabuzda D, Busciglio J, Yankner BA. Inhibition of β -amyloid production by activation of protein kinase C. *J Neurochem*. 1993;61:2326-2329.
- Nitsch RM, Slack BE, Farber SA, et al. Receptor-coupled amyloid precursor protein processing. *Ann NY Acad Sci*. 1993;695:122-127.
- Govoni S, Racchi M, Bergamaschi S, et al. Defective protein kinase C α leads to impaired secretion of soluble β -amyloid precursor protein from Alzheimer's disease fibroblasts. *Ann NY Acad Sci*. 1996;777:332-337.
- Benussi L, Govoni S, Gasparini L, et al. Specific role for protein kinase C α in the constitutive and regulated secretion of amyloid precursor protein in human skin fibroblast. *Neurosci Lett*. 1998;240:97-101.
- LeBlanc AC, Koutroumanis M, Goodyer CG. Protein kinase C activation increases release of secreted amyloid precursor protein without decreasing A β production in human primary neuron cultures. *J Neurosci*. 1998;18:2907-2913.
- Nitsch RM, Deng MH, Growdon JH, Wurtman RJ. Serotonin 5-HT $_2$ a and 5-HT $_2$ e receptors stimulate amyloid precursor protein ectodomain secretion. *J Biol Chem*. 1996;271:4188-4194.
- Vassar R, Bennett BD, Babu-Khan S, et al. Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. *Science*. 1999;286:735-741.
- Selkoe DJ, Yamazaki T, Citron M, et al. The role of APP processing and trafficking pathways in the formation of amyloid β -protein. *Ann NY Acad Sci*. 1996;777:57-64.
- Suzuki N, Cheung TT, Cai XD, et al. An increased percentage of long amyloid β protein secreted by familial amyloid β protein precursor (β APP $_{717}$) mutants. *Science*. 1994;264:1336-1340.
- Tamaoka A, Odaka A, Ishibashi Y, et al. APP717 missense mutation affects the ratio of amyloid β protein species (A β 1-42/43 and A β 1-40) in familial Alzheimer's disease brain. *J Biol Chem*. 1994;269:32721-32724.
- Citron M, Vigo-Pelfrey C, Teplow DB, et al. Excessive production of amyloid β -protein by peripheral cells of symptomatic and presymptomatic patients carrying the Swedish familial Alzheimer disease mutation. *Proc Natl Acad Sci USA*. 1994;91:11993-11997.
- Kosaka T, Imagawa M, Seki K, et al. The β APP717 Alzheimer mutation increases the percentage of plasma amyloid- β protein ending at A β 42(43). *Neurology*. 1997;48:741-745.
- Cras P, van Harskamp F, Hendriks L, et al. Presenile Alzheimer dementia characterized by amyloid angiopathy and large amyloid core type senile plaques in the APP 692Ala- \rightarrow Gly mutation. *Acta Neuropathol (Berl)*. 1998;96:253-260.
- Cruts M, Hendriks L, Van Broeckhoven C. The presenilin genes: a new gene family involved in Alzheimer disease pathology. *Hum Mol Genet*. 1996;5(special issue):1449-1455.
- Da Silva HAR, Patel AJ. Presenilins and early-onset familial Alzheimer's disease. *Neuroreport*. 1997;8:1-12.
- Cook DG, Sung JC, Golde TE, et al. Expression and analysis of presenilin 1 in a human neuronal system: localization in cell bodies and dendrites. *Proc Natl Acad Sci USA*. 1996;93:9223-9228.
- Kovacs DM, Fausett HJ, Page KJ, et al. Alzheimer-associated presenilins 1 and 2: Neuronal expression in brain and localization to intracellular membranes in mammalian cells. *Nat Med*. 1996;2:224-229.
- Moussaoui S, Czech C, Pradier L, et al. Immunohistochemical analysis of presenilin-1 expression in the mouse brain. *FEBS Lett*. 1996;383:219-222.
- De Strooper B, Beullens M, Contreras B, et al. Phosphorylation, subcellular localization, and membrane orientation of the Alzheimer's disease-associated presenilins. *J Biol Chem*. 1997;272:3590-3598.
- Duff K, Eckman C, Zehr C, et al. Increased amyloid- β 42(43) in brains of mice expressing mutant presenilin 1. *Nature*. 1996;383:710-713.
- Borchelt DR, Ratovitski T, Van Lare J, et al. Accelerated amyloid deposition in the brains of transgenic mice coexpressing mutant presenilin 1 and amyloid precursor proteins. *Neuron*. 1997;19:939-945.
- Annaert WG, Levesque L, Craessaerts K, et al. Presenilin 1 controls gamma-secretase processing of amyloid precursor protein in pre-Golgi compartments of hippocampal neurons. *J Cell Biol*. 1999;147:277-294.
- De Strooper B, Saftig P, Craessaerts K, et al. Deficiency of presenilin-1 inhibits the normal cleavage of amyloid precursor protein. *Nature*. 1998;391:387-390.
- Haass C, De Strooper B. Review: Neurobiology - The presenilins in Alzheimer's disease—proteolysis holds the key. *Science*. 1999;286:916-919.
- Haroutunian V, Purohit DP, Perl DP, et al. Neurofibrillary tangles in nondemented elderly subjects and mild Alzheimer disease. *Arch Neurol*. 1999;56:713-718.
- Iraizoz I, Guizarro JL, Gonzalo LM, de Lacalle S. Neuropathological changes in the nucleus basalis correlate with clinical measures of dementia. *Acta Neuropathol (Berl)*. 1999;98:186-196.
- Nagy Z, Esiri MM, Jobst KA, et al. Relative roles of plaques and tangles in the dementia of Alzheimer's disease: correlations using three sets of neuropathological criteria. *Dementia*. 1995;6:21-31.
- Braak E, Braak H, Mandelkow EM. A sequence of cytoskeleton changes related to the formation of neurofibrillary tangles and neuropil threads. *Acta Neuropathol (Berl)*. 1994;87:554-567.
- Braak H, Braak E. Evolution of neuronal changes in the course of Alzheimer's disease. *J Neural Transm*. 1998;105(suppl 53):127-140.
- Braak H, Braak E, Bohl J, Lang W. Alzheimer's disease: amyloid plaques in the cerebellum. *J Neurol Sci*. 1989;93:277-287.
- Lovestone S, Reynolds CH. The phosphorylation of tau: a critical stage in neurodevelopmental and neurodegenerative processes. *Neuroscience*. 1997;78:309-324.
- Biernat J, Gustke N, Drewes G, Mandelkow EM, Mandelkow E. Phosphorylation of Ser262 strongly reduces binding of tau to microtubules: distinction between PHF-like immunoreactivity and microtubule binding. *Neuron*. 1993;11:153-163.
- Matsuo ES, Shin R-W, Billingsley ML, et al. Biopsy-derived adult human brain tau is phosphorylated at many of the same sites as Alzheimer's disease paired helical filament tau. *Neuron*. 1994;13:989-1002.
- Garver TD, Lehman RAW, Billingsley ML. Microtubule assembly competence analysis of freshly-biopsied human tau, dephosphorylated tau, and Alzheimer tau. *J Neurosci Res*. 1996;44:12-20.

53. Braak H, Braak E, Strothjohann M. Abnormally phosphorylated tau protein related to the formation of neurofibrillary tangles and neuropil threads in the cerebral cortex of sheep and goat. *Neurosci Lett*. 1994;171:1-4.
54. Drewes G, Mandelkow EM, Baumann K, et al. Dephosphorylation of tau protein and Alzheimer paired helical filaments by calcineurin and phosphatase-2A. *FEBS Lett*. 1993;336:425-432.
55. Merrick SE, Trojanowski JQ, Lee VMY. Selective destruction of stable microtubules and axons by inhibitors of protein serine/threonine phosphatases in cultured human neurons (NT2N cells). *J Neurosci*. 1997;17:5726-5737.
56. Trojanowski JQ, Lee VMY. Phosphorylation of paired helical filament tau in Alzheimer's disease neurofibrillary lesions: focusing on phosphatases. *FASEB J*. 1995;9:1570-1576.
57. Lovestone S, Reynolds CH, Latimer D, et al. Alzheimer's disease-like phosphorylation of the microtubule-associated protein tau by glycogen synthase kinase-3 in transfected mammalian cells. *Curr Biol*. 1994;4:1077-1086.
58. Latimer DA, Lovestone S, Reynolds CH, et al. Phosphorylation of tau in transiently transfected control and raf-transformed 3T3 cells. *Neurobiol Aging*. 1994;15. Abstract 103.
59. Ishiguro K, Omori A, Takamatsu M, et al. Phosphorylation sites on tau by tau protein kinase I, a bovine derived kinase generating an epitope of paired helical filaments. *Neurosci Lett*. 1992;148:202-206.
60. Ishiguro K, Shiratsuchi A, Sato S, et al. Glycogen synthase kinase 3 β is identical to tau protein kinase I generating several epitopes of paired helical filaments. *FEBS Lett*. 1993;325:167-172.
61. Lovestone S, Hartley CL, Pearce J, Anderton BH. Phosphorylation of tau by glycogen synthase kinase-3 β in intact mammalian cells: the effects on organisation and stability of microtubules. *Neuroscience*. 1996;73:1145-1157.
62. Busciglio J, Lorenzo A, Yeh J, Yankner BA. β -Amyloid fibrils induce tau phosphorylation and loss of microtubule binding. *Neuron*. 1995;14:879-888.
63. Le WD, Xie WJ, Kong R, Appel SH. β -Amyloid-induced neurotoxicity of a hybrid septal cell line associated with increased tau phosphorylation and expression of β -amyloid precursor protein. *J Neurochem*. 1997;69:978-985.
64. Takashima A, Honda T, Yasutake K, et al. Activation of tau protein kinase I glycogen synthase kinase-3 β by amyloid β peptide (25-35) enhances phosphorylation of tau in hippocampal neurons. *Neurosci Res*. 1998;31:317-323.
65. Alvarez G, Muñoz-Montaño JR, Satrustegui J, et al. Lithium protects cultured neurons against β -amyloid-induced neurodegeneration. *FEBS Lett*. 1999;453:260-264.
66. Klein PS, Melton DA. A molecular mechanism for the effect of lithium on development. *Proc Natl Acad Sci USA*. 1996;93:8455-8459.
67. Leroy K, Menu R, Conreur JL, et al. The function of the microtubule-associated protein tau is variably modulated by graded changes in glycogen synthase kinase-3 β activity (In Process Citation). *FEBS Lett*. 2000;465:34-38.
68. Lovestone S, Davis DR, Webster MT, et al. Lithium reduces tau phosphorylation—effects in living cells and in neurons at therapeutic concentrations. *Biol Psychiatry*. 1999;45:995-1003.
69. Takahashi M, Yasutake K, Tomizawa K. Lithium inhibits neurite growth and tau protein kinase I/glycogen synthase kinase-3 β -dependent phosphorylation of juvenile tau in cultured hippocampal neurons. *J Neurochem*. 1999;73:2073-2083.
70. Hong M, Chen DC, Klein PS, Lee VM. Lithium reduces tau phosphorylation by inhibition of glycogen synthase kinase-3. *J Biol Chem*. 1997;272:25326-25332.
71. Muñoz-Montaño JR, Moreno FJ, Avila J, Díaz-Nido J. Lithium inhibits Alzheimer's disease-like tau protein phosphorylation in neurons. *FEBS Lett*. 1997;411:183-188.
72. Stambolic V, Ruel L, Woodgett JR. Lithium inhibits glycogen synthase kinase-3 activity and mimics wingless signalling in intact cells. *Curr Biol*. 1996;6:1664-1668.
73. Chen G, Huang LD, Jiang YM, Manji HK. The mood-stabilizing agent valproate inhibits the activity of glycogen synthase kinase-3. *J Neurochem*. 1999;72:1327-1330.
74. Berezovska O, Xia MQ, Hyman BT. Notch is expressed in adult brain, is coexpressed with presenilin-1, and is altered in Alzheimer disease. *J Neuropathol Exp Neurol*. 1998;57:738-745.
75. Beatus P, Lendahl U. Notch and neurogenesis. *J Neurosci Res*. 1998;54:125-136.
76. Fortini ME, Artavanis-Tsakonas S. Notch: neurogenesis is only part of the picture. *Cell*. 1993;75:1245-1247.
77. Weinmaster G. The ins and outs of Notch signaling. *Mol Cell Neurosci*. 1997;9:91-102.
78. Levitan D, Doyle TG, Brousseau D, et al. Assessment of normal and mutant human presenilin function in *Caenorhabditis elegans*. *Proc Natl Acad Sci USA*. 1996;93:14940-14944.
79. Baumeister R, Leimer U, Zweckbronner I, et al. Human presenilin-1, but not familial Alzheimer's disease (FAD) mutants, facilitate *Caenorhabditis elegans* notch signaling independently of proteolytic processing. *Genes Funct*. 1997;1:149-159.
80. Berezovska O, Frosch M, McLean P, et al. The Alzheimer-related gene presenilin 1 facilitates notch 1 in primary mammalian neurons. *Mol Brain Res*. 1999;69:273-280.
81. De Strooper B, Annaert W, Cupers P, et al. A presenilin-1-dependent gamma-secretase-like protease mediates release of Notch intracellular domain. *Nature*. 1999;398:518-522.
82. Ray WJ, Yao M, Nowotny P, et al. Evidence for a physical interaction between presenilin and Notch. *Proc Natl Acad Sci USA*. 1999;96:3263-3268.
83. Song WH, Nadeau P, Yuan ML, et al. Proteolytic release and nuclear translocation of Notch-1 are induced by presenilin-1 and impaired by pathogenic presenilin-1 mutations. *Proc Natl Acad Sci USA*. 1999;96:6959-6963.
84. Wong PC, Zheng H, Chen H, et al. Presenilin 1 is required for Notch1 Dll1 expression in the paraxial mesoderm. *Nature*. 1997;387:288-292.
85. Axelrod JD, Matsuno K, Artavanis-Tsakonas S, Perrimon N. Interaction between wingless and notch signaling pathways mediated by Dishevelled. *Science*. 1996;271:1826-1832.
86. Blair SS. Notch and wingless signals collide. *Science*. 1996;271:1822-1823.
87. Couso JP, Martinez Arias A. Notch is required for wingless signaling in the epidermis of *Drosophila*. *Cell*. 1994;79:259-272.
88. Kang DE, Soriano S, Frosch MP, et al. Presenilin 1 facilitates the constitutive turnover of β -catenin: differential activity of Alzheimer's disease-linked PS1 mutants in the β -catenin-signaling pathway. *J Neurosci*. 1999;19:4229-4237.
89. Nishimura M, Yu G, Levesque G, et al. Presenilin mutations associated with Alzheimer disease cause defective intracellular trafficking of β -catenin, a component of the presenilin protein complex. *Nat Med*. 1999;5:164-169.
90. Wehl CC, Miller RJ, Roos RP. The role of β -catenin stability in mutant PS1-associated apoptosis. *Neuroreport*. 1999;10:2527-2532.
91. Zhang ZH, Hartmann H, Do VM, et al. Destabilization of β -catenin by mutations in presenilin-1 potentiates neuronal apoptosis. *Nature*. 1998;395:698-702.
92. Zhou JH, Liyanage U, Medina M, et al. Presenilin 1 interaction in the brain with a novel member of the Armadillo family. *Neuroreport*. 1997;8:1489-1494.
93. Hong Mand Lee VMY. Insulin and insulin-like growth factor-1 regulate tau phosphorylation in cultured human neurons. *J Biol Chem*. 1997;272:19547-19553.
94. Stewart R, Liolitsa D. Type 2 diabetes mellitus, cognitive impairment and dementia. *Diabet Med*. 1999;16:93-112.
95. Kuusisto J, Koivisto K, Mykkanen L, et al. Association between features of the insulin resistance syndrome and Alzheimer's disease independently of apolipoprotein E4 phenotype: cross-sectional population based study. *BMJ*. 1997;315:1045-1049.
96. Lovestone S. Diabetes and dementia: is the brain another site of end-organ damage? *Neurology*. 1999;53:1907-1909.
97. Dumanchin C, Camuzat A, Campion D, et al. Segregation of a missense mutation in the microtubule-associated protein tau gene with familial frontotemporal dementia and parkinsonism. *Hum Mol Genet*. 1998;7:1825-1829.
98. Hutton M, Lendon CL, Rizzu P, et al. Association of missense and 5'-splice-site mutations in tau with the inherited dementia FTDP-17. *Nature*. 1998;393:702-705.
99. Spillantini MG, Murrell JR, Goedert M, et al. Mutation in the tau gene in familial multiple system tauopathy with presenile dementia. *Proc Natl Acad Sci USA*. 1998;95:7737-7741.
100. Hulette CM, Pericak-Vance MA, Roses AD, et al. Neuropathological features of frontotemporal dementia and parkinsonism linked to chromosome 17q21-22 (FTDP-17): Duke family 1684. *J Neuropathol Exp Neurol*. 1999;58:859-866.
101. Spillantini MG, Bird TD, Ghetti B. Frontotemporal dementia and Parkinsonism linked to chromosome 17: a new group of tauopathies. *Brain Pathol*. 1998;8:387-402.
102. Trojanowski JQ, Lee VMY. Transgenic models of tauopathies and synucleinopathies. *Brain Pathol*. 1999;9:733-739.

Basic research

103. D'Souza I, Poorkaj P, Hong M, et al. Missense and silent tau gene mutations cause frontotemporal dementia with parkinsonism-chromosome 17 type, by affecting multiple alternative RNA splicing regulatory elements. *Proc Natl Acad Sci USA*. 1999;96:5598-5603.
104. Hasegawa M, Smith MJ, Iijima M, Tabira T, Goedert M. FTDP-17 mutations N279K and S305N in tau produce increased splicing of exon 10. *FEBS Lett*. 1999;443:93-96.
105. Varani L, Hasegawa M, Spillantini MG, et al. Structure of tau exon 10 splicing regulatory element RNA and destabilization by mutations of frontotemporal dementia and parkinsonism linked to chromosome 17. *Proc Natl Acad Sci USA*. 1999;96:8229-8234.
106. Hasegawa M, Smith MJ, Goedert M. Tau proteins with FTDP-17 mutations have a reduced ability to promote microtubule assembly. *FEBS Lett*. 1998;437:207-210.
107. Hong M, Zhukareva V, Vogelsberg-Ragaglia V, et al. Mutation-specific functional impairments in distinct tau isoforms of hereditary FTDP-17. *Science*. 1998;282:1914-1917.
108. Matsumura N, Yamazaki T, Ihara Y. Stable expression in Chinese hamster ovary cells of mutated tau genes causing frontotemporal dementia and parkinsonism linked to chromosome 17 (FTDP-17). *Am J Pathol*. 1999;154:1649-1656.
109. Dayanandan R, Van Slegtenhorst M, Mack TG, et al. Mutations in tau reduce its microtubule binding properties in intact cells and affect its phosphorylation. *FEBS Lett*. 1999;446:228-232.
110. Yen SH, Hutton M, DeTure M, Ko LW, Nacharaju P. Fibrillogenesis of tau: insights from tau missense mutations in FTDP-17. *Brain Pathol*. 1999;9:695-705.
111. Roses AD. The Alzheimer diseases. *Curr Opin Neurobiol*. 1996;6:644-650.
112. Roses AD. Apolipoprotein E alleles as risk factors in Alzheimer's disease. *Annu Rev Med*. 1996;47:387-400.
113. Strittmatter WJ, Weisgraber KH, Huang DY, et al. Binding of human apolipoprotein E to synthetic amyloid β peptide: isoform-specific effects and implications for late-onset Alzheimer disease. *Proc Natl Acad Sci USA*. 1993;90:8098-8102.
114. Castaño EM, Prelli F, Wisniewski T, et al. Fibrillogenesis in Alzheimer's disease of amyloid? Peptides and apolipoprotein E. *Biochem J*. 1995;306:599-604.
115. LaDu MJ, Falduto MT, Manelli AM, et al. Isoform-specific binding of apolipoprotein E to β -amyloid. *J Biol Chem*. 1994;269:23403-23406.
116. Huang DY, Goedert M, Jakes R, et al. Isoform-specific interactions of apolipoprotein E with the microtubule-associated protein MAP2c: implications for Alzheimer's disease. *Neurosci Lett*. 1994;182:55-58.
117. Huang DY, Weisgraber KH, Goedert M, et al. ApoE3 binding to tau tandem repeat I is abolished by tau serine₂₆₂ phosphorylation. *Neurosci Lett*. 1995;192:209-212.
118. Scott BL, Welch K, DeSerrano V, et al. Human apolipoprotein E accelerates microtubule polymerization in vitro. *Neurosci Lett*. 1998;245:105-108.
119. Lovestone S, Anderton BH, Hartley C, Jensen TG, Jorgensen AL. The intracellular fate of tau is apolipoprotein E isoform-specific and tau-dependent. *Neuroreport*. 1996;7:1005-1008.
120. Bellosta S, Nathan BP, Orth M, et al. Stable expression and secretion of apolipoproteins E3 and E4 in mouse neuroblastoma cells produces differential effects on neurite outgrowth. *J Biol Chem*. 1995;270:27063-27071.
121. DeMattos RB, Thorngate FE, Williams DL. A test of the cytosolic apolipoprotein E hypothesis fails to detect the escape of apolipoprotein E from the endocytic pathway into the cytosol and shows that direct expression of apolipoprotein E in the cytosol is cytotoxic. *J Neurosci*. 1999;19:2464-2473.
122. Han SH, Einstein G, Weisgraber KH, et al. Apolipoprotein E is localized to the cytoplasm of human cortical neurons: a light and electron microscopic study. *J Neuropathol Exp Neurol*. 1994;53:535-544.
123. LaFerla FM, Troncoso JC, Strickland DK, Kawas CH, Jay G. Neuronal cell death in Alzheimer's disease correlates with apoE uptake and intracellular Abeta stabilization. *J Clin Invest*. 1997;100:310-320.
124. Roses AD, Gilbert J, Xu PT, et al. Cis-acting human ApoE tissue expression element is associated with human pattern of intraneuronal ApoE in transgenic mice. *Neurobiol Aging*. 1998;19(suppl):S53-S58.
125. DeMattos RB, Curtiss LK, Williams DL. A minimally lipidated form of cell-derived apolipoprotein E exhibits isoform-specific stimulation of neurite outgrowth in the absence of exogenous lipids or lipoproteins. *J Biol Chem*. 1998;273:4206-4212.
126. Nathan BP, Bellosta S, Sanan DA, et al. Differential effects of apolipoproteins E3 and E4 on neuronal growth in vitro. *Science*. 1994;264:850-852.
127. Nathan BP, Chang KC, Bellosta S, et al. The inhibitory effect of apolipoprotein E4 on neurite outgrowth is associated with microtubule depolymerization. *J Biol Chem*. 1995;270:19791-19799.