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Determinants of Protection among High Risk HIV Seronegative Persons – an overview

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Abstract

Both clinical experience and a growing medical literature indicate that there are persons who have been exposed to HIV infection who have remained uninfected. While in some instances this may represent good fortune, cohorts of uninfected persons have been reported where risks for infection are thought to be high. In these cohorts a variety of characteristics have been proposed as mediating protection but to date only the 32 base pair deletion in the CCR5 gene that results in complete failure of cell surface expression of this co-receptor has been associated with high level protection from HIV infection. With this in mind, there are likely numerous other factors that may individually or in combination provide some level of protection from acquisition of HIV infection. As some of these factors are likely incompletely protective or inconsistently active, identifying them with confidence will be difficult. Nonetheless, clarifying the determinants of protection

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against HIV infection is a high priority that will require careful selection of high risk uninfected cohorts to which targeted studies of plausible mediators and broad screening for unexpected determinants of protection should be applied.

Keywords

HIV infection; Exposed Seronegatives; High Risk Seronegatives; Interferon; Restriction factors; CCR5

As outlined by presentations throughout this meeting, there have been numerous reports of persons who were presumably exposed to HIV infection yet who remain uninfected. In light of the progressive spread of the HIV epidemic and the failure to date of most of our prevention strategies, it is increasingly important to evaluate potential determinants of this protection. This stated, it may be very difficult to identify these determinants for reasons we will outline in this brief overview.

Why do some presumably HIV exposed persons seem to remain uninfected?

As only a small minority of sexual exposures to persons who are HIV infected result in transmission of infection [1], undoubtedly some exposed persons who remain uninfected—have just been fortunate. But one could nonetheless divide the universe of potentially protective mechanisms into those characterized by diminished intrinsic susceptibility to infection, protective adaptive immune defenses, protective innate immune defenses and another category that we will call protection mediated by incidental events.

Caveats

As we start to explore these potential determinants of protection in various cohorts, several caveats are in order. First, the risks for HIV acquisition and factors that might prevent acquisition vary according to the route of transmission (e.g. parenteral versus mucosal) and among mucosal transmissions, vary according to the nature of the mucosal surface (vaginal/rectal/penile/oral/neonatal oral) [1, 2]. As a possible example of this, the -336T polymorphisms in promoter for the C type lectin DC SIGN seems to be associated with modest relative protection against parenteral but not mucosal acquisition of HIV infection [3].

What are the factors that may protect these persons? At the outset we must recognize the possibility (if not probability) there may be more than one factor that contributes to protection from infection, that protection by any one factor may be only partial and that combinations of factors may collaborate to provide higher level protection from infection [4]. Complicating this exploration even more is the recognition that protection may not be consistent over time, affected by recognized factors such as hormonal environment [5] or other factors not yet fully understood [4, 6] so that an individual apparently protected for one period of high risk exposure, may for a variety of reasons, lose that relative protection at other times. Thus identification of mechanisms that may protect against acquisition of infection will not be an easy task.

Absence of the HIV co-receptor CCR5 provides high-level resistance to HIV acquisition

Somewhere, perhaps in Europe, about a thousand years ago [7], or even much earlier [8, 9], a 32 base pair deletion in the open reading frame for the chemokine receptor CCR5 was passed on in the germ line. This mutation results in a frameshift such that several downstream amino acids differ from the wild type sequences until a premature stop codon terminates translation. Thus a truncated nonfunctional protein is generated. Persons

homozygous for this mutation are highly protected from HIV infection [10–12]. The haplotypic structure suggests that this mutation has occurred only once in human history. But the allele frequency for CCR5 Δ 32 ranges from 5 to 15% in European populations and currently the allele is in an apparent Hardy Weinberg equilibrium indicating that at least in recent times there was no major survival advantage or disadvantage to any of the three possible CCR5 genotypes. Yet with an allele frequency as high as 15% or more, there may have been a powerful selection pressure for its persistence in humans. Plague and poxviruses have been accused of providing this selection pressure but to date, no compelling evidence supporting these possibilities have been presented. Likewise, there is no indication to date that there had been a remote interaction between Europeans and HIV or a related lentivirus. Relative protection may be conferred by the heterozygous state, particularly when accompanied by other polymorphisms in the other CCR5 allele [13, 14]. As noted above, protection may be relative and additive. Several reports have suggested that predicted variation in (or sensitivity to [15]) levels of beta chemokine ligands that can block or sequester CCR5 may also determine the relative risk [16] or protection [17] from acquisition of HIV infection. To date though, only complete absence of surface CCR5 expression has been shown capable of providing high level protection from HIV infection.

Innate Defenses against HIV acquisition

Innate defenses play critical roles in early defense against microbial invasion and must hold small enemies at bay either blocking their tissue entry entirely or attenuating the magnitude of their growth until adaptive defenses can provide a targeted antimicrobial defense. Conceivably, variation in their expression or function could contribute to heterogeneous risks for HIV acquisition. Thus the physicochemical characteristics of mucus as well as the intercellular barriers to fluid and particulate transport provide protection against tissue penetration of viruses. Variabilities in the characteristics of these elements (such as pH for example) might play a role in protection against HIV acquisition (reviewed in [18]). These surfaces also contain antimicrobial defense molecules such as defensins for example that have antiretroviral activity [19] and are variably expressed in human populations with gene repeat frequencies ranging from 2 to 12 in any one individual [20]. Type I interferons comprise a large family of innate antiviral defense molecules that can be induced by a variety of microbial elements including viruses and their genetic sequences [21]. Type I interferons bind to a common receptor expressed on numerous cell types and activate hundreds of cellular genes including genes encoding certain proteins with antiviral activity [22] yet at mucosal sites of challenge with the simian immunodeficiency virus, the kinetics of interferon induction may be too delayed to provide sufficient protection from experimental infection [23]. It's not unreasonable to propose that there may be population variability in the induction, regulation and activities of such a complex defense network and to this point, a polymorphism near the gene for a type III interferon (interferon lambda-3, IL28B) has been found linked to variability in the outcome of hepatitis C virus infection both spontaneously and with treatment using a type I interferon plus ribavirin [24, 25].

The regulation of the activities of cellular RNaseL (up) and Eukaryotic initiation factor 2a (EIF2a)(down) by type I interferons paralyze translation of both cellular and viral RNAs [21]. Type I interferons also can activate other cellular elements such as APOBEC 3G, Trim-5a and tetherin that more selectively restrict the replication of lentiviruses such as HIV. The importance of APOBEC3G and tetherin in antiviral defense is underscored by the existence of HIV proteins, specifically VIF and VPU respectively that block their antiviral activities [26, 27]. Although human Trim-5a does not block HIV replication very effectively, certain non-human primate homologues do [28] and small changes in the human sequences result in potent restriction of HIV replication [29]. In one small study, higher levels of TRIM-5a RNA were found in unseparated peripheral blood mononuclear cells

from high risk persons who escaped HIV infection than in persons who did not [30]. A single nucleotide polymorphism in the APBOEC3G gene (C49693T) has been associated in one study with an increased risk for HIV acquisition [31]. Conceivably, other genetic variants in this restriction factor confer some degree of protection from infection. Do some persons apparently protected from HIV acquisition have intracellular restriction factors or combinations of restriction factors that are more highly expressed, more effective or relatively resistant to the activities of their viral inhibitors? This remains an area worthy of further investigation. At the same time, knockdown studies have identified hundreds of cellular elements that are utilized for HIV propagation [32] and while different studies have not always identified the same elements, it is plausible that differences in the expression or function of one or more of these elements also could contribute to relative resistance to HIV infection.

Natural killer cells are capable of lysing viral infected cells, particularly cells that have diminished expression of class I MHC - a defense strategy utilized by many viruses including HIV to escape recognition by cytolytic T cells [33]. Epidemiological analyses indicate that genetic differences in the expression of particular killer immunoglobulin receptors (KIR), typically expressed on NK cells, in conjunction with their HLA ligands determine differential courses of HIV infection [34]. Furthermore, elevated NK cell activity, and elevated KIR3DS1 transcript expression, has been associated with reduced risk of infection in a cohort of intravenous drug users, suggesting that these innate effector cells may play a role in preventing parenteral HIV acquisition [35, 36]. Is it possible that heterogeneities in NK cell regulation determine relative protection from HIV infection?

Adaptive Immunity and Protection from HIV infection

Several groups [37–40], but not all groups have found T cells recognizing HIV peptides in exposed seronegative persons and it has been proposed that these adaptive defenses might confer some level of protection against HIV acquisition. To our mind, this is a difficult proposal to support as vaccine studies in non human primates have often generated more robust T cell responses than the responses found in these exposed seronegative humans [41–43] yet these more robust responses have not succeeded in protecting challenged animals from infection. On the other hand the high level viral exposure designed to infect all challenged animals in experimental settings may overwhelm modest immune responses that might protect against the lower levels of virus exposure that at risk human subjects typically experience. This stated, it is our opinion that these responses, when found, more likely represent evidence of exposure to HIV than determinants of protection.

Exposed seronegative persons by definition do not have systemic antibodies reactive with HIV proteins. Several groups have however identified HIV-reactive antibodies in genital secretions of exposed seronegatives that in some settings have had neutralizing activity [44] But antibodies recognizing viral surface proteins could plausibly provide some level of protection such as for example, by blocking the free movement of virus in mucosal fluid (Tom Hope, personal communication).

Can environmental determinants drive protective mechanisms?

There is reasonable evidence that environmental factors may determine risk for or protection against HIV acquisition. Thus ulcerative infections of the genitourinary tract are associated with increased risk of HIV acquisition [45]. Likewise, bacterial vaginosis is associated with increased risk of infection [46] and whether this is a consequence of inflammation or replacement of the lactobacilli that are thought to be protective or both is not clear. As different colonizing microbes can differentially activate the spectrum of innate immune receptors that are widely distributed throughout the mucosal epithelium and on host defense

cells, it is not unreasonable to propose that the microbiome at mucosal sites could plausibly but temporally and inconsistently determine the relative susceptibility to and protection from HIV infection. This proposal is testable but not simply, and if correct, it will complicate efforts to sort out the determinants of HIV protection. Likewise some viruses including certain ubiquitous herpesviruses may attenuate HIV replication and could conceivably mediate some level of protection (reviewed in [47]).

Should we focus on exposed seronegatives?

As noted above, the risk of HIV acquisition with any mucosal exposure is estimated to be low thus calculating the magnitude of risk among exposed seronegatives is difficult. This adds another layer of complexity to studies aimed at identifying protective factors among exposed seronegatives as including in the study population persons who are just “lucky” and who do not possess the putative protective elements dilutes the study’s power to be informative. For this reason, it is very important to study individuals demonstrably at high risk for HIV infection yet who remain uninfected a group we prefer to call “High Risk Seronegatives” (HRSN).

Risk of HIV infection in men with Hemophilia

During the early years of the AIDS epidemic, HIV infection was broadly transmitted to the hemophilia population through administration of pooled concentrates of plasma derived clotting Factor VIII and clotting Factor IX [48, 49]. Each vial of concentrate was derived from pooled plasmas of thousands of donors throughout the United States who donated before the AIDS agent was identified and before the blood supply could be protected through donor screening. By accessing infusion records from participants in the NCI-sponsored multicenter hemophilia cohort study (MHCS), Barb Kroner was able to demonstrate that the risk of HIV acquisition was directly related to the type (higher with Factor VIII than with Factor IX) and intensity of exposure to these concentrates [50]. Among persons who were moderately or heavily treated before 1985, the risk of HIV infection was approximately 95%. We elected to study the 5% of these heavily treated men who did not acquire HIV infection despite intensive exposure. We found that 7 of these 43 HRSN hemophiliacs were homozygous for the CCR5 d32 mutation, an estimated 16 fold concentration of this uncommon genotype that validates identification of this cohort as HRSN [51]. An additional 7 were heterozygous for this allele – compatible with larger cohort studies suggesting that partial or low level protection from HIV infection might be conferred by the CCR5 d 32 heterozygote state [13, 14]. So what are the other potential contributors to protection? In unpublished work, we did not find evidence of T cell reactivity with HIV antigens and we could not find neutralizing antibodies in serum. Density of CCR5 was not low either on circulating CD4+ T cells or on circulating monocytes. Expression of CCR5 binding beta chemokines MIP-1 α , MIP-1 β and RANTES was normal in several culture conditions; their cells were not demonstrably resistant to infection with CCR5 tropic HIV *in vitro*. But one interesting characteristic that distinguished these patients from unselected healthy controls not at risk for HIV infection was that their cells were less readily activated by exposure to multiple concentrations of lectin (phytohemagglutinin) and interleukin-2 [51]. This quiescent phenotype seems to be a consistent observation among exposed seronegatives and high risk seronegatives as both gay men who did not seroconvert in the Amsterdam cohort study had fewer activated cells than did gay men who later seroconverted [52] and similar findings were reported among high risk seronegatives in the Pumwani sex cohort [53]. HIV more readily infects activated cells that typically express higher levels of chemokine receptors that not only can serve as co-receptors for HIV but also may serve to amplify the initial rounds of infection at mucosal sites [54]. Activated T cells may also express less active forms of endogenous inhibitors of HIV replication such as

APOBEC 3G [55]. It is therefore plausible to propose that activation state could be a factor that determines relative risk or protection from HIV acquisition.

Where to go from here?

Aside from the well-defined protection from infection that is provided by the CCR5 Δ 32 homozygous state, there are numerous proposed and plausible, but no clearly proven determinants of protection against HIV acquisition. Identifying these factors is a high priority as provision or induction of these elements could prove useful in HIV prevention strategies. Identifying these factors will be challenging as protection could be partial, inconsistent, relative and overcome with sufficient challenge or other alteration in susceptibility. Important protective factors also might provide protection against HIV acquisition at some routes but not others. Thus it is especially important that cohort studies assembled to explore the prevalence of these potentially protective factors be homogeneous at the start and be rigorously defined to include only persons at the highest risk for HIV acquisition.

Summary

There is compelling epidemiologic evidence supporting the existence of additional factors that protect persons from acquisition of HIV infection. Identifying these factors is a high scientific priority but for the reasons outlined above, this will not be a simple task. Success in this endeavor will require the establishment of cohorts of persons unambiguously at high risk for but who have escaped HIV infection. Effective recruitment of these cohorts will require attention to local and social sensitivities. To these cohorts must be applied targeted studies of plausible protective mediators as well as genetic screens to dissect other determinants of protection that are not yet suspected.

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References

1. Royce RA, Sena A, Cates W Jr, Cohen MS. Sexual transmission of HIV. *N Engl J Med*. 1997; 336:1072–8. [PubMed: 9091805]
2. Cohen MS, Shugars DC, Fiscus SA. Limits on oral transmission of HIV-1. *Lancet*. 2000; 356:272. [PubMed: 11071180]
3. Martin MP, Lederman MM, Hutcheson HB, et al. Association of DC-SIGN promoter polymorphism with increased risk for parenteral, but not mucosal, acquisition of human immunodeficiency virus type 1 infection. *J Virol*. 2004; 78:14053–6. [PubMed: 15564514]
4. Margolis L, Shattock R. Selective transmission of CCR5-utilizing HIV-1: the 'gatekeeper' problem resolved? *Nat Rev Microbiol*. 2006; 4:312–7. [PubMed: 16541138]
5. Smith SM, Baskin GB, Marx PA. Estrogen protects against vaginal transmission of simian immunodeficiency virus. *J Infect Dis*. 2000; 182:708–15. [PubMed: 10950763]
6. Kaul R, Rowland-Jones SL, Kimani J, et al. Late seroconversion in HIV-resistant Nairobi prostitutes despite pre-existing HIV-specific CD8+ responses. *J Clin Invest*. 2001; 107:341–9. [PubMed: 11160158]
7. Stephens JC, Reich DE, Goldstein DB, et al. Dating the origin of the CCR5-Delta32 AIDS-resistance allele by the coalescence of haplotypes. *Am J Hum Genet*. 1998; 62:1507–15. [PubMed: 9585595]

8. Hummel S, Schmidt D, Kremeyer B, Herrmann B, Oppermann M. Detection of the CCR5-Delta32 HIV resistance gene in Bronze Age skeletons. *Genes Immun.* 2005; 6:371–4. [PubMed: 15815693]
9. Sabeti PC, Walsh E, Schaffner SF, et al. The case for selection at CCR5-Delta32. *PLoS Biol.* 2005; 3:e378. [PubMed: 16248677]
10. Dean M, Carrington M, Winkler C, et al. Genetic restriction of HIV-1 infection and progression to AIDS by a deletion allele of the *CKR5* structural gene. Hemophilia Growth and Development Study, Multicenter AIDS Cohort Study, Multicenter Hemophilia Cohort Study, San Francisco City Cohort, ALIVE Study. *Science.* 1996; 273:1856–62. [PubMed: 8791590]
11. Huang Y, Paxton WA, Wolinsky SM, et al. The role of a mutant CCR5 allele in HIV-1 transmission and disease progression. *Nat Med.* 1996; 2:1240–3. [PubMed: 8898752]
12. Zimmerman PA, Buckler-White A, Alkhatib G, et al. Inherited resistance to HIV-1 conferred by an inactivating mutation in CC chemokine receptor 5: studies in populations with contrasting clinical phenotypes, defined racial background, and quantified risk. *Mol Med.* 1997; 3:23–36. [PubMed: 9132277]
13. Tang J, Shelton B, Makhatadze NJ, et al. Distribution of chemokine receptor CCR2 and CCR5 genotypes and their relative contribution to human immunodeficiency virus type 1 (HIV-1) seroconversion, early HIV-1 RNA concentration in plasma, and later disease progression. *J Virol.* 2002; 76:662–72. [PubMed: 11752157]
14. Trearichi EM, Tumbarello M, de Gaetano Donati K, et al. Partial protective effect of CCR5-Delta 32 heterozygosity in a cohort of heterosexual Italian HIV-1 exposed uninfected individuals. *AIDS Res Ther.* 2006; 3:22. [PubMed: 16999868]
15. Saez-Cirion A, Versmisse P, Truong LX, et al. Persistent resistance to HIV-1 infection in CD4 T cells from exposed uninfected Vietnamese individuals is mediated by entry and post-entry blocks. *Retrovirology.* 2006; 3:81. [PubMed: 17092330]
16. Colobran R, Adreani P, Ashhab Y, et al. Multiple products derived from two CCL4 loci: high incidence of a new polymorphism in HIV+ patients. *J Immunol.* 2005; 174:5655–64. [PubMed: 15843566]
17. Gonzalez E, Kulkarni H, Bolivar H, et al. The influence of CCL3L1 gene-containing segmental duplications on HIV-1/AIDS susceptibility. *Science.* 2005; 307:1434–40. [PubMed: 15637236]
18. Lederman MM, Offord RE, Hartley O. Microbicides and other topical strategies to prevent vaginal transmission of HIV. *Nat Rev Immunol.* 2006; 6:371–82. [PubMed: 16639430]
19. Quinones-Mateu ME, Lederman MM, Feng Z, et al. Human epithelial beta-defensins 2 and 3 inhibit HIV-1 replication. *Aids.* 2003; 17:F39–48. [PubMed: 14571200]
20. Hollox EJ, Armour JA, Barber JC. Extensive normal copy number variation of a beta-defensin antimicrobial-gene cluster. *Am J Hum Genet.* 2003; 73:591–600. [PubMed: 12916016]
21. Pichlmair A, Reis e Sousa C. Innate recognition of viruses. *Immunity.* 2007; 27:370–83. [PubMed: 17892846]
22. Der SD, Zhou A, Williams BR, Silverman RH. Identification of genes differentially regulated by interferon alpha, beta, or gamma using oligonucleotide arrays. *Proc Natl Acad Sci U S A.* 1998; 95:15623–8. [PubMed: 9861020]
23. Abel K, Rocke DM, Chohan B, Fritts L, Miller CJ. Temporal and anatomic relationship between virus replication and cytokine gene expression after vaginal simian immunodeficiency virus infection. *J Virol.* 2005; 79:12164–72. [PubMed: 16160143]
24. Ge D, Fellay J, Thompson AJ, et al. Genetic variation in *IL28B* predicts hepatitis C treatment-induced viral clearance. *Nature.* 2009; 461:399–401. [PubMed: 19684573]
25. Thomas DL, Thio CL, Martin MP, et al. Genetic variation in *IL28B* and spontaneous clearance of hepatitis C virus. *Nature.* 2009; 461:798–801. [PubMed: 19759533]
26. Malim MH. APOBEC proteins and intrinsic resistance to HIV-1 infection. *Philos Trans R Soc Lond B Biol Sci.* 2009; 364:675–87. [PubMed: 19038776]
27. Neil SJ, Zang T, Bieniasz PD. Tetherin inhibits retrovirus release and is antagonized by HIV-1 Vpu. *Nature.* 2008; 451:425–30. [PubMed: 18200009]
28. Stremlau M, Owens CM, Perron MJ, Kiessling M, Autissier P, Sodroski J. The cytoplasmic body component TRIM5alpha restricts HIV-1 infection in Old World monkeys. *Nature.* 2004; 427:848–53. [PubMed: 14985764]

29. Stremlau M, Perron M, Welikala S, Sodroski J. Species-specific variation in the B30. 2(SPRY) domain of TRIM5alpha determines the potency of human immunodeficiency virus restriction. *J Virol.* 2005; 79:3139–45. [PubMed: 15709033]
30. Sewram S, Singh R, Kormuth E, et al. Human TRIM5alpha expression levels and reduced susceptibility to HIV-1 infection. *J Infect Dis.* 2009; 199:1657–63. [PubMed: 19388851]
31. Valcke HS, Bernard NF, Bruneau J, Alary M, Tsoukas CM, Roger M. APOBEC3G genetic variants and their association with risk of HIV infection in highly exposed Caucasians. *AIDS.* 2006; 20:1984–6. [PubMed: 16988524]
32. Brass AL, Dykxhoorn DM, Benita Y, et al. Identification of host proteins required for HIV infection through a functional genomic screen. *Science.* 2008; 319:921–6. [PubMed: 18187620]
33. Schwartz O, Marechal V, Le Gall S, Lemonnier F, Heard JM. Endocytosis of major histocompatibility complex class I molecules is induced by the HIV-1 Nef protein. *Nat Med.* 1996; 2:338–42. [PubMed: 8612235]
34. Martin MP, Gao X, Lee JH, et al. Epistatic interaction between KIR3DS1 and HLA-B delays the progression to AIDS. *Nat Genet.* 2002; 31:429–34. [PubMed: 12134147]
35. Ravet S, Scott-Algara D, Bonnet E, et al. Distinctive NK-cell receptor repertoires sustain high-level constitutive NK-cell activation in HIV-exposed uninfected individuals. *Blood.* 2007; 109:4296–305. [PubMed: 17272507]
36. Scott-Algara D, Truong LX, Versmisse P, et al. Cutting edge: increased NK cell activity in HIV-1-exposed but uninfected Vietnamese intravenous drug users. *J Immunol.* 2003; 171:5663–7. [PubMed: 14634071]
37. Bernard NF, Yannakis CM, Lee JS, Tsoukas CM. Human immunodeficiency virus (HIV)-specific cytotoxic T lymphocyte activity in HIV-exposed seronegative persons. *J Infect Dis.* 1999; 179:538–47. [PubMed: 9952359]
38. Fowke KR, Dong T, Rowland-Jones SL, et al. HIV type 1 resistance in Kenyan sex workers is not associated with altered cellular susceptibility to HIV type 1 infection or enhanced beta-chemokine production. *AIDS Res Hum Retroviruses.* 1998; 14:1521–30. [PubMed: 9840285]
39. Goh WC, Markee J, Akridge RE, et al. Protection against human immunodeficiency virus type 1 infection in persons with repeated exposure: evidence for T cell immunity in the absence of inherited CCR5 coreceptor defects. *J Infect Dis.* 1999; 179:548–57. [PubMed: 9952360]
40. Wasik TJ, Bratosiewicz J, Wierzbicki A, et al. Protective role of beta-chemokines associated with HIV-specific Th responses against perinatal HIV transmission. *J Immunol.* 1999; 162:4355–64. [PubMed: 10201969]
41. Buge SL, Murty L, Arora K, et al. Factors associated with slow disease progression in macaques immunized with an adenovirus-simian immunodeficiency virus (SIV) envelope priming-gp120 boosting regimen and challenged vaginally with SIVmac251. *J Virol.* 1999; 73:7430–40. [PubMed: 10438833]
42. Sun Y, Bailer RT, Rao SS, et al. Systemic and mucosal T-lymphocyte activation induced by recombinant adenovirus vaccines in rhesus monkeys. *J Virol.* 2009; 83:10596–604. [PubMed: 19656883]
43. Zhao J, Pinczewski J, Gomez-Roman VR, et al. Improved protection of rhesus macaques against intrarectal simian immunodeficiency virus SIV(mac251) challenge by a replication-competent Ad5hr-SIVenv/rev and Ad5hr-SIVgag recombinant priming/gp120 boosting regimen. *J Virol.* 2003; 77:8354–65. [PubMed: 12857905]
44. Kaul R, Trabattoni D, Bwayo JJ, et al. HIV-1-specific mucosal IgA in a cohort of HIV-1-resistant Kenyan sex workers. *Aids.* 1999; 13:23–9. [PubMed: 10207541]
45. Galvin SR, Cohen MS. The role of sexually transmitted diseases in HIV transmission. *Nat Rev Microbiol.* 2004; 2:33–42. [PubMed: 15035007]
46. Sewankambo N, Gray RH, Wawer MJ, et al. HIV-1 infection associated with abnormal vaginal flora morphology and bacterial vaginosis. *Lancet.* 1997; 350:546–50. [PubMed: 9284776]
47. Lisco A, Vanpouille C, Margolis L. War and peace between microbes: HIV-1 interactions with coinfecting viruses. *Cell Host Microbe.* 2009; 6:403–8. [PubMed: 19917495]

48. Menitove JE, Aster RH, Casper JT, et al. T-lymphocyte subpopulations in patients with classic hemophilia treated with cryoprecipitate and lyophilized concentrates. *N Engl J Med.* 1983; 308:83–6. [PubMed: 6401196]
49. Lederman MM, Ratnoff OD, Evatt BL, McDougal JS. Acquisition of antibody to lymphadenopathy-associated virus in patients with classic hemophilia (factor VIII deficiency). *Ann Intern Med.* 1985; 102:753–7. [PubMed: 2986506]
50. Kroner BL, Rosenberg PS, Aledort LM, Alvord WG, Goedert JJ. HIV-1 infection incidence among persons with hemophilia in the United States and western Europe, 1978–1990. Multicenter Hemophilia Cohort Study. *J Acquir Immune Defic Syndr.* 1994; 7:279–86. [PubMed: 8106967]
51. Salkowitz JR, Purvis SF, Meyerson H, et al. Characterization of high-risk HIV-1 seronegative hemophiliacs. *Clin Immunol.* 2001; 98:200–11. [PubMed: 11161976]
52. Koning FA, Otto SA, Hazenberg MD, et al. Low-level CD4+ T cell activation is associated with low susceptibility to HIV-1 infection. *J Immunol.* 2005; 175:6117–22. [PubMed: 16237108]
53. Card CM, McLaren PJ, Wachihi C, Kimani J, Plummer FA, Fowke KR. Decreased immune activation in resistance to HIV-1 infection is associated with an elevated frequency of CD4(+)CD25(+)FOXP3(+) regulatory T cells. *J Infect Dis.* 2009; 199:1318–22. [PubMed: 19301980]
54. Li Q, Estes JD, Schlievert PM, et al. Glycerol monolaurate prevents mucosal SIV transmission. *Nature.* 2009; 458:1034–8. [PubMed: 19262509]
55. Chiu YL, Soros VB, Kreisberg JF, Stopak K, Yonemoto W, Greene WC. Cellular APOBEC3G restricts HIV-1 infection in resting CD4+ T cells. *Nature.* 2005; 435:108–14. [PubMed: 15829920]