
The nucleotide sequence of the *Escherichia coli fus* gene, coding for elongation factor G

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ABSTRACT

We have determined the nucleotide sequence of the *Escherichia coli fus* gene, which codes for elongation factor G. The protein product of the sequenced gene contains 703 amino acids, with a predicted molecular weight of 77,444. The *fus* gene shows the nonrandom pattern of codon usage typical of ribosomal proteins and other proteins synthesized at a high level. We have identified several potential promoter sequences within the gene. One of these sequences may correspond to the secondary promoter for expression of the downstream *tufA* gene (encoding elongation factor Tu) whose activity has been described previously (1,2). A comparison of the nucleotide and amino acid sequences of elongation factors G and Tu reveals a limited but significant homology between the two proteins within the 150 amino acid residues at their amino-terminal ends.

INTRODUCTION

Elongation factors G, Tu, and Ts (EF-G, EF-Tu and EF-Ts, respectively) play essential roles during protein biosynthesis in *Escherichia coli*. In addition to having a close functional relationship with ribosomes, these proteins are encoded by genes whose expression is tightly coordinated with the expression of ribosomal protein genes (3). We have analyzed the expression of *tufA* and *fus*, encoding EF-Tu and EF-G, respectively (1,4). Both of these genes map in the *str* operon, which also contains genes for two ribosomal proteins, S12 and S7 (5). Interestingly, EF-Tu is also synthesized from the *tufB* gene, which maps elsewhere on the *E. coli* chromosome (5). The nucleotide sequences of both *tufA* and *tufB* have been published (6,7). As a prerequisite to extending our studies on the expression of the *str* operon, we have now determined the sequence of the *fus* gene.

MATERIALS AND METHODS**Bacterial and phage strains**

Bacteriophages M13mp8 and M13mp9 and their host strain JM103 were

obtained from BRL. The M13 phages were propagated as described by the BRL manual.

Enzymes and reagents

Restriction endonucleases were obtained from BRL, New England Biolabs, or Boehringer-Mannheim. Mung bean nuclease was obtained from P-L Biochemicals. T4 DNA ligase was purified by E. Lifson in this laboratory. Reagents for DNA sequencing were purchased as a kit from BRL.

Construction of M13 clones

M13 clones for DNA sequencing were constructed by subcloning DNA from plasmid pLL145 (Fig. 1), a pBGP120 (8) derivative carrying a 3.2kb KpnI fragment from fus3 (9) that contains the intact fus gene (1,4). Clones Z11, Z19, Z20, Z21, Z22, and Z24 were constructed by inserting various portions of the 3.2kb KpnI fragment onto M13mp8 or M13mp9 using the restriction enzymes indicated in Fig. 1. [For Z11, the KpnI generated staggered end was converted to a blunt end with mung bean nuclease and ligated to a blunt end HincII site on the M13 vector.] All other clones were derived by deleting, "flipping", and/or subcloning various portions of the original subclones.

DNA sequence determination and analysis

The DNA sequences were obtained from the M13 clones using the dideoxynucleotide chain termination method of Sanger (10) as described by the BRL sequencing manual. Sequencing samples were routinely run on 40 cm long 8% polyacrylamide gels. To extend the sequence of Z21, we also analyzed this sample on a 5% polyacrylamide 87 cm long "King Kong" gel. The sequences of most of the M13 clones were determined from at least two independent sequencing reactions and, in most cases, the sequence was confirmed by sequencing an independently isolated clone containing overlapping DNA sequences or the same sequence in the opposite orientation. The final sequence was constructed as shown in Fig. 1.

The fus sequence was analyzed using programs designed by J. Pustell at the Biological Laboratories at Harvard University (11,12).

RESULTS AND DISCUSSION

Determination of the fus gene sequence

Figure 2 shows the complete 2115 base pair sequence of the fus gene, plus the 5' flanking region up to the UGA termination codon of the gene for r-protein S7, which lies immediately upstream of fus in the str operon. Using the strategy shown in Fig. 1, we determined the sequence of the S7-fus intercistronic region and all of the fus sequence, except for the final 22

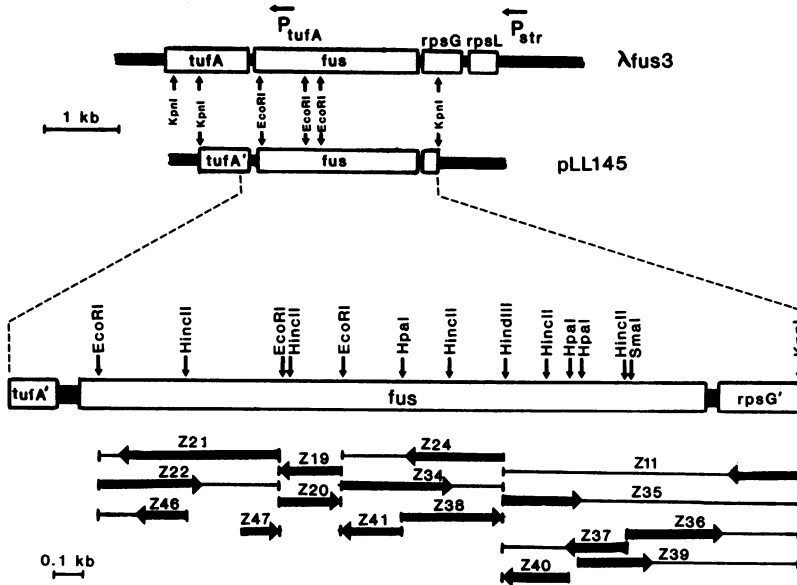


Figure 1.

Strategy for sequencing *fus*. The top of the figure shows the genetic organization of the *str* operon and the *KpnI* fragment from the operon carried on pLL145. P_{str} designates the position of the major promoter of the *str* operon; P_{tufA} , the approximate position of the secondary promoter for *tufA* expression (1). Subfragments cloned from pLL145 onto M13 vectors are shown below. Arrows indicate the direction and thick bars indicate the extent of sequencing obtained from each subclone.

codons at the 3' end which were sequenced previously by Yokota *et al.* (6). The sequence of the S7-*fus* intercistronic region and the first 279 nucleotides of the *fus* gene was reported previously by Post and Nomura (13); our sequencing data are consistent with their results.

Amino acid sequence of the *fus* gene product

The *fus* DNA sequence predicts that the gene product, EF-G, contains 703 amino acids (not including the Met encoded by the initiation codon) and has a molecular weight of 77,444. The amino acid sequence shown in Fig. 2 is in good agreement with the published amino acid sequence of EF-G (14), which contains 701 amino acids. The most serious discrepancy between the published amino acid sequence and the sequence predicted from the nucleotide sequence is in the cysteine content of the protein. According to Ovchinnikov *et al.* (14), EF-G contains 5 cysteine residues, 2 of which reportedly are involved in a disulfide bond. However, the nucleotide sequence predicts only three

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-30		-20		-10		1		10		20		30							
*		*		*		*		*		*		*							
TGA	ACG	CCT	AAA	AGA	TAA	ACG	AGG	AAA	CAA	ATG	GCT	CGT	ACA	ACA	CCC	ATC	GCA	CGC	TAC
											Ala	Arg	Thr	Thr	Pro	Ile	Ala	Arg	Tyr
			40			50		60		70		80		90					
			*			*		*		*		*		*					
CGT	AAC	ATC	GGT	ATC	AGT	GCG	CAC	ATC	GAC	GCC	GGT	AAA	ACC	ACT	ACT	ACC	GAA	CGT	ATT
Arg	Asn	Ile	Gly	Ile	Ser	Ala	His	Ile	Asp	Ala	Gly	Lys	Thr	Thr	Thr	Thr	Glu	Arg	Ile
			100			110		120		130		140		150					
			*			*		*		*		*		*					
CTG	TTC	TAC	ACC	GGT	GTA	AAC	CAT	AAA	ATC	GGT	GAA	GTT	CAT	GAC	GGC	GCT	GCA	ACC	ATG
Leu	Phe	Tyr	Thr	Gly	Val	Asn	His	Lys	Ile	Gly	Glu	Val	His	Asp	Gly	Ala	Ala	Thr	Met
			160			170		180		190		200		210					
			*			*		*		*		*		*					
GAC	TGG	ATG	GAG	CAG	GAG	CAG	GAA	CGT	GGT	ATT	ACC	ATC	ACT	TCC	GCT	GCG	ACT	ACT	GCA
Asp	Trp	Met	Glu	Gln	Glu	Gln	Glu	Arg	Gly	Ile	Thr	Ile	Thr	Ser	Ala	Ala	Thr	Thr	Ala
			220			230		240		250		260		270					
			*			*		*		*		*		*					
TTC	TGG	TCT	GGT	ATG	GCT	AAG	CAG	TAT	GAG	CCG	CAT	CGC	ATC	AAC	ATC	ATC	GAC	ACC	CCG
Phe	Trp	Ser	Gly	Met	Ala	Lys	Gln	Tyr	Glu	Pro	His	Arg	Ile	Asn	Ile	Ile	Asp	Thr	Pro
			280			290		300		310		320		330					
			*			*		*		*		*		*					
GGG	CAC	GTT	GAC	TTC	ACA	ATC	GAA	GTA	GAA	CGT	TCC	ATG	CGT	GTT	CTC	GAT	GGT	GCG	GTA
Gly	His	Val	Asp	Phe	Thr	Ile	Glu	Val	Glu	Arg	Ser	Met	Arg	Val	Leu	Asp	Gly	Ala	Val
			340			350		360		370		380		390					
			*			*		*		*		*		*					
ATG	GTT	TAC	TGC	GCA	GTT	GGT	GGT	GTT	CAG	CCG	CAG	TCT	GAA	ACC	GTA	TGG	CGT	CAG	GCA
Met	Val	Tyr	Cys	Ala	Val	Gly	Gly	Val	Gln	Pro	Gln	Ser	Glu	Thr	Val	Trp	Arg	Gln	Ala
			400			410		420		430		440		450					
			*			*		*		*		*		*					
AAC	AAA	TAT	AAA	GTT	CCG	CGC	ATT	GCG	TTC	GTT	AAC	AAA	ATG	GAC	CGC	ATG	GGT	GCG	AAC
Asn	Lys	Tyr	Lys	Val	Pro	Arg	Ile	Ala	Phe	Val	Asn	Lys	Met	Asp	Arg	Met	Gly	Ala	Asn
			460			470		480		490		500		510					
			*			*		*		*		*		*					
TTC	CTG	AAA	GTT	GTT	AAC	CAG	ATC	AAA	ACC	CGT	CTG	GGC	GCG	AAC	CCG	GTT	CCG	CTG	CAG
Phe	Leu	Lys	Val	Val	Asn	Gln	Ile	Lys	Thr	Arg	Leu	Gly	Ala	Asn	Pro	Val	Pro	Leu	Gln
			520			530		540		550		560		570					
			*			*		*		*		*		*					
CTG	GCG	ATT	GGT	GCT	GAA	GAA	CAT	TTC	ACC	GGT	GTT	GTT	GAC	CTG	GTG	AAA	ATG	AAA	GCT
Leu	Ala	Ile	Gly	Ala	Glu	Glu	His	Phe	Thr	Gly	Val	Val	Asp	Leu	Val	Lys	Met	Lys	Ala
			580			590		600		610		620		630					
			*			*		*		*		*		*					
ATC	AAC	TGG	AAC	GAC	GCT	GAC	CAG	GGC	GTA	ACC	TTC	GAA	TAC	GAA	GAT	ATC	CCG	GCA	GAC
Ile	Asn	Trp	Asn	Asp	Ala	Asp	Gln	Gly	Val	Thr	Phe	Glu	Tyr	Glu	Asp	Ile	Pro	Ala	Asp
			640			650		660		670		680		690					
			*			*		*		*		*		*					
ATG	GTT	GAA	CTG	GCT	AAC	GAA	TGG	CAC	CAG	AAC	CTG	ATC	GAA	TCC	GCA	GCT	GAA	GCT	TCT
Met	Val	Glu	Leu	Ala	Asn	Glu	Trp	His	Gln	Asn	Leu	Ile	Glu	Ser	Ala	Ala	Glu	Ala	Ser

700 * 710 * 720 * 730 * 740 * 750 *
 GAA GAG CTG ATG GAA AAA TAC CTG GGT GGT GAA GAA CTG ACT GAA GCA GAA ATC AAA GGT
 Glu Glu Leu Met Glu Lys Tyr Leu Gly Gly Glu Glu Leu Thr Glu Ala Glu Ile Lys Gly

760 * 770 * 780 * 790 * 800 * 810 *
 GCT CTG CGT CAG CGC GTT CTG AAC AAC GAA ATC ATC CTG GTA ACC TGT GGT TCT GCG TTC
 Ala Leu Arg Gln Arg Val Leu Asn Asn Glu Ile Ile Leu Val Thr Cys Gly Ser Ala Phe

820 * 830 * 840 * 850 * 860 * 870 *
 AAG AAC AAA GGT GTT CAG GCG ATG CTG GAT GCG GTA ATT GAT TAC CTG CCA TCC CCG GTT
 Lys Asn Lys Gly Val Gln Ala Met Leu Asp Ala Val Ile Asp Tyr Leu Pro Ser Pro Val

880 * 890 * 900 * 910 * 920 * 930 *
 GAC GTA CCT GCG ATC AAC GGT ATC CTG GAC GAC GGT AAA GAC ACT CCG GCT GAA CGT CAC
 Asp Val Pro Ala Ile Asn Gly Ile Leu Asp Asp Gly Lys Asp Thr Pro Ala Glu Arg His

940 * 950 * 960 * 970 * 980 * 990 *
 GCA AGT GAT GAC GAG CCG TTC TCT GCA CTG GCG TTC AAA ATC GCT ACC GAC CCG TTT GTT
 Ala Ser Asp Asp Glu Pro Phe Ser Ala Leu Ala Phe Lys Ile Ala Thr Asp Pro Phe Val

1000 * 1010 * 1020 * 1030 * 1040 * 1050 *
 GGT AAC CTG ACC TTC TTC CGT GTT TAC TCC GGT GTG GTT AAC TCT GGT GAT ACC GTA CTG
 Gly Asn Leu Thr Phe Phe Arg Val Tyr Ser Gly Val Val Asn Ser Gly Asp Thr Val Leu

1060 * 1070 * 1080 * 1090 * 1100 * 1110 *
 AAC TCC GTG AAA GCT GCA CGT GAG CGT TTC GGT CGT ATC GTT CAG ATG CAC GCT AAC AAA
 Asn Ser Val Lys Ala Ala Arg Glu Arg Phe Gly Arg Ile Val Gln Met His Ala Asn Lys

1120 * 1130 * 1140 * 1150 * 1160 * 1170 *
 CGT GAA GAG ATC AAA GAA GTT CGC GCG GGC GAC ATC GCT GCT GCT ATC GGT CTG AAA GAC
 Arg Glu Glu Ile Lys Glu Val Arg Ala Gly Asp Ile Ala Ala Ala Ile Gly Leu Lys Asp

1180 * 1190 * 1200 * 1210 * 1220 * 1230 *
 GTA ACC ACT GGT GAC ACC CTG TGT GAC CCG GAT GCG CCG ATC ATT CTG GAA CGT ATG GAA
 Val Thr Thr Thr Gly Asp Thr Leu Cys Asp Pro Asp Ala Pro Ile Ile Leu Glu Arg Met Glu

1240 * 1250 * 1260 * 1270 * 1280 * 1290 *
 TTC CCT GAG CCG GTA ATC TCC ATC GCA GTT GAA CCG AAA ACC AAA GCT GAC CAG GAA AAA
 Phe Pro Glu Pro Val Ile Ser Ile Ala Val Glu Pro Lys Thr Lys Ala Asp Gln Glu Lys

1300 * 1310 * 1320 * 1330 * 1340 * 1350 *
 ATG GGT CTG GCT CTG GGC CGT CTG GCT AAA GAA GAC CCG TCT TTC CGT GTA TGG ACT GAC
 Met Gly Leu Ala Leu Gly Arg Leu Ala Lys Glu Asp Pro Ser Phe Arg Val Trp Thr Asp

1360 * 1370 * 1380 * 1390 * 1400 * 1410 *
 GAA GAA TCT AAC CAG ACC ATC ATC GCG GGT ATG GGC GAA CTG CAC CTC GAC ATC ATC GTT
 Glu Glu Ser Asn Gln Thr Ile Ile Ala Gly Met Gly Glu Leu His Leu Asp Ile Ile Val

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      1420      1430      1440      1450      1460      1470
      *        *        *        *        *        *
GAC CGT ATG AAG CGT GAA TTC AAC GTT GAA GCG AAC GTA GGT AAA CCG CAG GTT GCT TAC
Asp Arg Met Lys Arg Glu Phe Asn Val Glu Ala Asn Val Gly Lys Pro Gln Val Ala Tyr

      1480      1490      1500      1510      1520      1530
      *        *        *        *        *        *
CGT GAA ACT ATC CGC CAG AAA GTT ACC GAT GTT GAA GGT AAA CAC GCG AAA CAG TCT GGT
Arg Glu Thr Ile Arg Gln Lys Val Thr Asp Val Glu Gly Lys His Ala Lys Gln Ser Gly

      1540      1550      1560      1570      1580      1590
      *        *        *        *        *        *
GGT CGT GGT CAG TAT GGT CAT GTT GTT ATC GAC ATG TAC CCG CTG GAG CCG GGT TCA AAC
Gly Arg Gly Gln Tyr Gly His Val Val Ile Asp Met Tyr Pro Leu Glu Pro Gly Ser Asn

      1600      1610      1620      1630      1640      1650
      *        *        *        *        *        *
CCG AAA GGC TAC GAG TTC ATC AAC GAC ATT AAA GGT GGT GTA ATC CCT GGC GAA TAC ATC
Pro Lys Gly Tyr Glu Phe Ile Asn Asp Ile Lys Gly Gly Val Ile Pro Gly Glu Tyr Ile

      1660      1670      1680      1690      1700      1710
      *        *        *        *        *        *
CCG GCC GTT GAT AAA GGT ATC CAG GAA CAG CTG AAA GCA GGT CCG CTG GCA GGC TAC CCG
Pro Ala Val Asp Lys Gly Ile Gln Glu Gln Leu Lys Ala Gly Pro Leu Ala Gly Tyr Pro

      1720      1730      1740      1750      1760      1770
      *        *        *        *        *        *
GTA GTA GAC ATG GGT ATT CGT CTG CAC TTC GGT TCT TAC CAT GAC GTT GAC TCC TCT GAA
Val Val Asp Met Gly Ile Arg Leu His Phe Gly Ser Tyr His Asp Val Asp Ser Ser Glu

      1780      1790      1800      1810      1820      1830
      *        *        *        *        *        *
CTG GCG TTT AAA CTG GCT GCT TCT ATC GCC TTT AAA GAA GGC TTT AAG AAA GCG AAA CCA
Leu Ala Phe Lys Leu Ala Ala Ser Ile Ala Phe Lys Glu Gly Phe Lys Lys Ala Lys Pro

      1840      1850      1860      1870      1880      1890
      *        *        *        *        *        *
GTT CTG CTT GAG CCG ATC ATG AAG GTT GAA GTA GAA ACT CCG GAA GAG AAC ACC GGT GAC
Val Leu Leu Glu Pro Ile Met Lys Val Glu Val Glu Thr Pro Glu Glu Asn Thr Gly Asp

      1900      1910      1920      1930      1940      1950
      *        *        *        *        *        *
GTT ATC GGT GAC TTG AGC CGT CGT CGT GGT ATG CTC AAA GGT CAG GAA TCT GAA GTT ACT
Val Ile Gly Asp Leu Ser Arg Arg Arg Gly Met Leu Lys Gly Gln Glu Ser Glu Val Thr

      1960      1970      1980      1990      2000      2010
      *        *        *        *        *        *
GGC GTT AAG ATC CAC GCT GAA GTA CCG CTG TCT GAA ATG TTC GGA TAC GCA ACT CAG CTG
Gly Val Lys Ile His Ala Glu Val Pro Leu Ser Glu Met Phe Gly Tyr Ala Thr Gln Leu

      2020      2030      2040      2050      2060      2070
      *        *        *        *        *        *
CGT TCT CTG ACC AAA GGT CGT GCA TCA TAC ACT ATG GAA TTC CTG AAG TAT GAT GAA GCG
Arg Ser Leu Thr Lys Gly Arg Ala Ser Tyr Thr Met Glu Phe Leu Lys Tyr Asp Glu Ala

      2080      2090      2100      2110
      *        *        *        *
CCG AGT AAC GTT GCT CAG GCC GTA ATT GAA GCC CGT GGT AAA TAA
Pro Ser Asn Val Ala Gln Ala Val Ile Glu Ala Arg Gly Lys ---

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cysteines. The two missing residues are those reported to be in the S-S bond: the DNA sequence predicts a Gly instead of Cys at nucleotides 889-891, and a Thr instead of Cys at nucleotides 1186-1188 (Fig. 2).

Other differences predicted by the nucleotide sequence include an Asn instead of Asp at nucleotides 886-888, a Glu instead of Gln at nucleotides 1876-1878, 1936-1938, and 1969-1971, an Ile instead of Val at nucleotides 1726-1728, a Lys instead of His at nucleotides 1780-1782, and a His instead of Lys at nucleotides 1750-1752. In addition, the DNA sequence predicts an additional three amino acids, Asp-Asp-Gly, at nucleotides 898-906, not found in the published amino acid sequence, and does not predict a Gln between nucleotides 1986 and 1987 that is present in the previously published sequence.

We have double-checked our nucleotide sequence in regions where we have detected a discrepancy between our results and those reported by Ovchinnikov *et al.* (14), and are confident that our data are correct. Some of the differences may be due to errors in the original amino acid sequence or, possibly, to strain specific differences in the *fus* sequence between the cells used in the amino acid analysis and the cells from which the DNA for cloning was derived. Alternatively, the discrepancies could have resulted from mutations which arose during the construction of the *fus* clones. We would like to point out in this connection, however, that the source of the sequenced DNA, pLL145, contains a *fus* gene from which a functional EF-G protein is synthesized, as defined by its ability to convert a fusidic acid resistant host to fusidic acid sensitivity (4). Also, the EF-G derived from pLL145 comigrates with the chromosomally derived EF-G on a two-dimensional O'Farrell gel (4). In any event, given the size of the *fus* gene and its protein product, there are surprisingly few discrepancies between the published amino acid sequence and the sequence predicted by our DNA sequence analysis.

Codon frequencies

Table 1 shows the frequency of codon usage in *fus*. The pattern of codon

Figure 2.

Nucleotide sequence of *E. coli fus*. The DNA strand with the *fus* mRNA polarity is given, along with the deduced amino acid sequence. The nucleotide positions are numbered beginning with the initiation codon for *fus*. The sequence through nucleotide 279 was reported previously by Post and Nomura (13); the nucleotide sequence from 2047 through 2115 was reported previously by Yokota *et al.* (6).

Table 1. Comparison of codon usage in the *E. coli* fus, tufA/tufB and ribosomal protein genes.

Codons	EF-G	EF-Tu	r-proteins	Codons	EF-G	EF-Tu	r-proteins
TTT Phe	4	2	17	TAT Tyr	4	3	7
TTC Phe	20	26	38	TAC Tyr	15	17	23
TTA Leu	0	0	4	TAA ---	1	2	13
TTG Leu	1	0	7	TAG ---	0	0	1
CTT Leu	1	2	7	CAT His	6	3	9
CTC Leu	3	1	9	CAC His	9	19	11
CTA Leu	0	0	1	CAA Gln	0	0	18
CTG Leu	38	53	108	CAG Gln	24	16	44
ATT Ile	9	6	37	AAT Asn	0	0	15
ATC Ile	41	52	80	AAC Asn	27	14	52
ATA Ile	0	0	0	AAA Lys	37	35	134
ATG Met	22	20	54	AAG Lys	7	11	50
GTT Val	38	46	88	GAT Asp	10	8	35
GTC Val	0	1	25	GAC Asp	32	41	51
GTA Val	19	21	59	GAA Glu	50	60	95
GTG Val	3	6	21	GAG Glu	12	13	28
TCT Ser	15	14	39	TGT Cys	2	2	4
TCC Ser	8	6	26	TGC Cys	1	4	4
TCA Ser	2	0	2	TGA ---	0	0	0
TCG Ser	0	0	2	TGG Trp	6	2	3
CCT Pro	3	0	13	CGT Arg	29	41	84
CCC Pro	1	0	2	CGC Arg	7	5	47
CCA Pro	2	2	7	CGA Arg	0	0	3
CCG Pro	27	38	45	CGG Arg	0	0	1
ACT Thr	14	25	48	AGT Ser	3	0	3
ACC Thr	21	31	41	AGC Ser	1	1	13
ACA Thr	3	3	4	AGA Arg	0	0	1
ACG Thr	0	1	6	AGG Arg	0	0	0
GCT Ala	26	24	113	GGT Gly	46	38	107
GCC Ala	5	2	27	GGC Gly	11	41	65
GCA Ala	16	11	68	GGA Gly	1	0	1
GCG Ala	20	17	45	GGG Gly	1	2	7

The values for EF-Tu are the combined frequencies for tufA (6) and tufB (7). The values for r-proteins include all 10 r-protein genes from the spc operon (16) plus the genes for L11, L1, L10, and L7/L12 (15). The initiation codons are not included.

usage shows the same, highly non-random, pattern observed for r-proteins and EF-Tu (see, e.g., 6,7,15,16). Since EF-G is synthesized in equimolar amounts with ribosomal proteins, this result is consistent with the hypothesis that

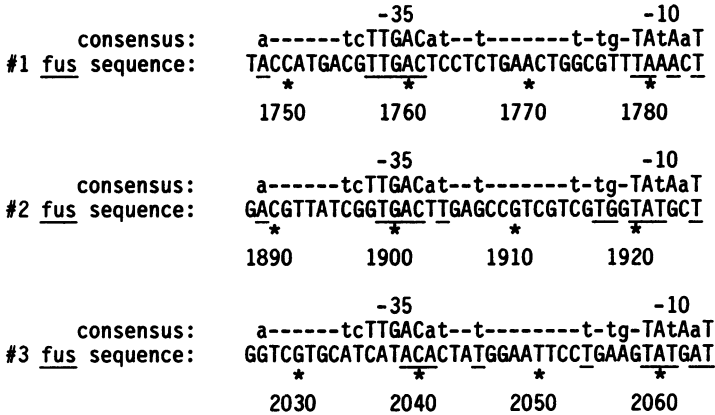


Figure 3.

Potential promoter sequences within the fus gene. Shown are three sequences within the structural gene for EF-G which resemble the consensus promoter sequence (19). The bases with exact homology are underlined. The numbering system for fus sequences corresponds to the system used in Fig. 2.

the genes for proteins synthesized at very high rates, such as ribosomal proteins and translation elongation factors, preferentially use codons recognized by the most abundant of the isoaccepting species of tRNA (15,17).

Possible secondary promoter sequences

One strong incentive for sequencing the fus gene was to facilitate our analysis of a secondary promoter in the str operon, apparently mapping within the structural gene for EF-G (1,2). Analysis of the nucleotide sequence has turned up several sequences (Fig. 3) which contain "-35" and "-10" regions typical of E. coli promoters (18,19). Two of these sequences, #1 and #2, are located within a 600 base pair EcoRI fragment internal to the fus gene. Hybridization experiments measuring the transcription rates from various regions of the str operon in cells carrying multicopy plasmid pLL145 (Fig. 1) indicate that the secondary promoter is either within the 600 base pair EcoRI sequence or just upstream of this region (J. M. Zengel, unpublished experiments). Therefore, both of these sequences, particularly #1 because of its strong homology with the consensus -35 sequence, are reasonable candidates for the secondary promoter.

Yokota et al. (6) pointed out a possible Pribnow box at the very end of the fus gene, just downstream from the EcoRI site at nucleotide 2047. At the time, the upstream sequence containing the putative -35 region was not known.

Now that we have completed the fus sequence, the -35 region can be analyzed. As shown in Fig. 3, this potential secondary promoter, #3, has a relatively good Pribnow box, but a relatively weak -35 region. Since our hybridization results (described above) suggest that the promoter is further upstream, it seems unlikely that this promoter contributes significantly to the expression of the tufA gene. In any event, experiments are in progress to map more precisely the secondary promoter.

Homology between EF-G and EF-Tu

A comparison of the amino acid sequence of the amino-terminal region of EF-G with the amino acid sequence of elongation factor Tu (EF-Tu) revealed a region of homology between the two elongation factors (20,21). Jones et al. (21) have suggested that at least the regions comprising amino acid residues 25 to 150 of EF-G and EF-Tu have evolved from a common ancestral protein by gene duplication. We have searched for homology between EF-Tu and EF-G, both at the nucleotide level and at the amino acid level, using the homology matrix program developed by Pustell and Kafatos (12). Our analysis revealed no significant regions of homology between EF-Tu and EF-G, with the exception of the region of weak homology already pointed out (20,21). However, by introducing two additional "gaps" in the alignment of the amino acid sequences, we were able to extend this region of homology to include the first 25 amino acids at the amino-terminal ends of the two proteins. Figure 4 shows the comparison of the amino acid and nucleotide sequences in the regions of homology, aligned for maximum amino acid homology. Note that short stretches of strong homology are flanked by regions of essentially no homology and that a number of gaps have been introduced in the EF-Tu and EF-G sequences to maximize the homology (Fig. 4). These factors plus our failure to identify any significant homology elsewhere in the two proteins makes it difficult to evaluate the evolutionary relationship between the two elongation factors.

Figure 4.

Comparison of the amino acid sequences of EF-G and EF-Tu. The corresponding nucleotide sequences of fus and tufA are also shown. The numbers refer to the amino acid sequences. Amino acids with exact homology are shown in boldface, upper case letters. Exact nucleotide homologies are indicated with asterisks. Regions of extensive sequence homology are enclosed in blocks. The amino acid alignment beginning with residue 32 of the two proteins is essentially the same as that pointed out previously by Jones et al. (21).

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