Interacdon of Z form poly(dG-dC).poly(dG-dC) with divalent metal ions: localization of the binding sites by I.R. spectroscopy

J.A.Taboury, P.Bourtayre, J.Liquier and E.Taillandier*

Laboratoire de Spectroscopie Biomoléculaire, UER de Médecine, Université Paris XIII, 74, rue Marcel Cachin, 93000 Bobigny, France

Received ⁵ April 1984; Accepted ¹ May 1984

ABSTRACT

The secondary structures of poly(dG-dC).poly(dG-dC) in the presence of alcaline, alcaline earth and first row transition metal ions (Na+, Mq^{2+} , Co^{2+} , Ni^{2+}) are investigated by infrared spectroscopy. The conformational transitions are studied as a function of the hydration of the polynucleotide and counter-ion nature and content. The use of selectively deuterated poly(dG-dC).poly(dG-dC) on the 8-carbon of guanines allows to show that a direct interaction occurs between the N7 site of guanines and the transition metal ions C_0 ⁺ and N_1 ⁺⁺. In the case of M_9 ⁻⁺, for high ion/nucleotide ratios, the interaction occurs essentially at the level of the phosphate groups. This interaction leads to a modification of the left-handed conformation. Based on the IR spectroscopy results, an explanation is proposed for the different efficiencies of these various ions to induce the $B \rightarrow Z$ transition.

INTRODUCTION

In synthetic polynucleotides, the existence of left-handed helixes, first suggested by U.V. circular dichroism experiments (1), has been confirmed by X ray crystal diffraction studies (2,3) and characterized by other various techniques: $31P$ and $1H$ NMR (4-7), Raman (8,9) and IR spectroscopy (9,10). In native systems, the existence of such conformations has been evidenced by indirect immunological techniques (12-14). This novel conformation is now extensively studied, because of its possible biological importance: hypothesis have been proposed concerning the role played by Z DNA in fundamental processes such as regulation of gene expression (15). Among the various parameters involved in the $B \stackrel{\epsilon}{\rightarrow} Z$ transition of poly(dG-dC). poly(dG-dC) in solution, the nature and the concentration of various ions has been previously studied by U.V. absorbance and circular dichroism (1,16-19). On the other hand, the important role played in vivo and in vitro by metal ions in processes involving nucleic acids has been investigated for many years (for review, see ref. 20) ; in particular, different binding modes have been proposed: interaction between the ions and the phosphate

© IRL Press Limited, Oxford, England.

groups, either directly or via a water bridge, interactions with electron donor sites of the bases,... (21,22).

The IR spectra of sodium salt poly(dG-dC).poly(dG-dC) have been partly described (10,11,23). In the present work, we have investigated by IR spectroscopy the conformation effect of Mg^{2+} , Co^{2+} and Ni^{2+} on poly(dG-dC).poly(dG-dC), and the possible binding sites of these ions. The use of a polynucleotide, specifically deuterated on the 8-carbon of guanines (poly(dD8G-dC).poly(dD8G-dC)), allows to localize a structurally important interaction of the transition metal ions with the N7 site of this base.

MATERIALS AND METHODS

Poly(dG-dC).poly(dG-dC) (lot 719-76), 2'-deoxyguanosine 5'-phosphate (dGMP) and neutral 2'-deoxycytidine (dC) were purchased from P.L. Biochemicals. Specific deuteration of the guanines was obtained by incubation in D_2 O at 80°C for 3 hours (24-25). Samples were deposited on ZnSe windows and gently dried so as to give homogeneous films. The desired amount of ions is obtained by diffusion of a droplet of the corresponding solution (NiCl₂, CoBr₂, MgCl₂, at pH 7), followed by slow evaporation. Films are placed in cells with controlled relative humidity (H₂0 or D₂0) (between 32 and 100% RH). The hydration of the complexes is determined directly on the IR spectra of the samples. Ion/nucleotide ratios were varied between 1 and 5. The Perkin-Elmer 180 double-beam spectrophotometer is coupled to a Hewlett-Packard 9825 A calculator allowing systematic data treatment such as baseline and water contribution corrections, scaled spectrum substraction,...

RESULTS

Z conformation of poly(dD8G-dC).poly(dD8G.dC) in the presence of divalent ions

At high RH (> 76%), and whatever the ions (Co²⁺, Ni²⁺, Mg²⁺), the I.R. spectra reflect a Z type double helix conformation of the polynucleotide. The spectral regions corresponding to the absorptions of the bases, of the sugar-base glycosidic linkage and of the phosphodiester backbone show the characteristic features of the left-handed structure.

In the 1750 cm^{-1} -1550 cm^{-1} region, the drastic modification of the base pair stacking which occurs during the $B \rightarrow Z$ transition induces important shifts of the bands corresponding to the double bond vibrations

Figure 1. Infrared spectra of poly(dD8G-dC).poly(dD8G-dC) in D20 in the base double bond stretching vibration region : a/ Na+ B form ; b/ Na'
Z form ; c/ Ni²+ (or Co²⁺, or Mg²⁺), 1 M²⁺ per nucleotide, Z form.

of the bases. In $\mathsf{D}_2\mathsf{O}_2$, two bands are observed in the poly(dD8G-dC). poly(dD8G-dC) - Na+ spectrum, located at 1681 cm 1 and 1649 cm 1 in the B conformation (Fig. 1a). They are found respectively at 1666. cm^{-1} and 1634 $cm⁻¹$ in the Z conformation (Fig. 1b). In the presence of divalent metal ions, the spectrum recorded in D_2 O shows two absorptions at the same latter wavenumbers (Fig. 1c). In the case of spectra recorded in H_20 , the modification of the base pair stacking in the sodium salt polymer is reflected by a shift of the 1710 cm ⁺ band to 1692 cm ⁺ (Fig. 2). In the presence of divalent ions, this absorption is also detected at 1692 cm^{-1} (results not shown). Thus, the absence of the 1710 cm $^{-}$ band (in H₂O), and the presence of a doublet at 1666 cm⁻¹-1634 cm⁻¹ (in D₂0) can be correlated to a Z type base stacking of the poly(dD8G-dC).poly(dD8G-dC) in the presence of divalent metal ions.

In the 1550 cm^{-1} -1250 cm^{-1} region (Fig. 3), that is to say in the spectral region corresponding mainly to the in plane vibrations of the

Figure 2. IR spectra of poly(dD8G-dC).poly(dD8G-dC).Na+: bottom: B form 98X% RRT, top: Z form (86% RH).

Figure 3. IR spectra of poly(dD8G-dC).poly(dD8G-dC) glycosidic linkage vibration region. Ion/nucleotide
a/ Na+, b/ Mg²⁺ ; c/ Ni²⁺ ; d/ Co²⁺. in the sugar base and ratio = 1. RH > 76%

Figure 4.IR spectra of out of plane vibrations of poly(dD8G-dC).poly(dD8G-dC) in D₂O : a/Na+, B form ; b/ Co^{z+} (l per nucleotide), Z form ; c/ Ni^{z+} (l per nucleotide), Z form.

bases, the spectra of the poly(dD8G-dC).poly(dD8G-dC)- M^{2+} are characterized by the presence of the 1409 cm^{-1} , 1321 cm^{-1} and 1264 cm^{-1} bands, which are also observed for the poly(dD8G-dC).poly(dD8G-dC)-Na⁺ in the Z form (Fig. 3a). However, a new spectroscopic feature is detected in this region in the case of the interaction with the transition metal ions Co^{2+} and Ni^{2+} : we have previously shown that the absorption located at 1374 cm^{-1} in the B form and 1354 cm^{-1} in the Z form of poly(dG-dC).poly(dG-dC)-Na⁺ involves the dG residue in an anti and a syn conformation respectively (26) ; this band, which remains at 1354 cm^{-1} in the case of Mg²⁺ (Fig. 3b), is split into two components at 1354 cm⁻¹ and 1343 cm⁻¹ in the case of Co²⁺ or Ni²⁺ (Fig. 3, c and d). This, and also the effect on the 1467 cm^{-1} band (which will be discussed in the second part of this work), suggests a strong interaction of the transition metal ions with the guanines.

In the region below 800 cm^{-1} (spectral region of the out of plane base vibrations) guanine and cytosine contributions are superimposed at 778 cm^{-1} in the case of the B conformation (Fig. 4a). The spectra of the

Figure 5. IR spectrum of poly(dD8G-dC).poly(dD8G-dC) in the presence of Mg $^{\mathsf{L}\mathsf{+}}$ $(5$ per nucleotide), $RH = 32\%$).

poly(dD8G-dC).poly(dD8G-dC)-M²⁺ complexes present, in D₂0, two absorptions located at 778 cm⁻¹ and 784 cm⁻¹ (Fig. 4 b and c). This doublet is characteristic of a Z conformation of the alternating G-C polymer ; thanks to the effect of the C-5-methylation of the cytosine, these two bands are attributed to out of plane vibrations of cytosines and guanines respectively (26,27).

Concerning the deoxyribose and the phosphodiester backbone vibrations, three absorptions of the poly(dD8G-dC).poly(dD8G-dC)- M^{2+} complexes can also be correlated with the Z conformation ; two intense bands at 925 cm^{-1} and 1014 cm^{-1} , and a weaker one at 1124 cm^{-1} (spectra not shown).

All these spectroscopic evidences show that in highly concentrated phases (films above 76% RH), the divalent ions Mg^{2+} , Co²⁺ and Ni²⁺ stabilize the left-handed conformation of the poly(dD8G-dC).poly(dD8G-dC).

Decreasing the relative humidity below 58% leads to IR spectra depending on the amount and nature of divalent counter-ion. Fig. ⁵ presents the IR spectrum of poly(dD8G-dC).poly(dD8G-dC) in the presence of high Mg²⁺/nucleotide ratio (above 5), recorded at low relative humidity (32%). Such a spectrum which has never been observed with any of the transition metals studied (Co^{2+}, Ni^{2+}) , has been interpreted as reflecting a new structure in the left-handed family, called Z_{0} (10,28). The most striking

changes are observed at the level of the phosphate absorptions. The strong direct interaction of Mg²⁺ with the PO₂⁻ groups is evidenced by a drastic shift of the $v_{\sf as}$ and $v_{\sf s}$ PO₂ absorptions from 1213 cm⁻¹ to 1267 cm⁻¹ (54 cm 1) and from 1088 cm⁻¹ to 1122 cm⁻¹ (34 cm⁻¹) respectively. In the case of Co^{2+} and Ni²⁺, for the same high metal ion/nucleotide ratios and in the same extreme dehydration conditions, we have only observed indications of a destabilization of the double helix: presence of a single band around 1650 cm^{-1} in D₂0 (while two absorption bands are always present in the B and the Z forms in this region), and shift of the 1297 cm^{-1} cytosine band to 1282 cm⁻¹ (while it is not affected by the presence of \cot^{2+} or Ni²⁺ when the Z structure is well formed). This last result supports the hypothesis of an interaction between these two ions and the cytosine at very low hydration. Localization of the divalent metal ions interaction sites

Different nucleophilic sites of the polynucleotide are potential candidates for an interaction with divalent cations: phosphates, carbonyl groups of the bases, N7 site of guanines. The use of selectively deuterated polymers allows us to put forward an eventual interaction with this latter site : the IR spectrum of this deuterated polynucleotide presents an absorption located at 1467 cm^{-1} which involves the motion of the N7 of guanines and moreover which is not affected by the $B \rightarrow Z$ conformational transition induced by $Na⁺$ ions. The modifications of the position and the relative intensity of this band will therefore be a probe for specific interactions with the N7 of guanines.

The spectra of the Z conformations of poly(dG-dC).poly(dG-dC) and poly(dD8G-dC).poly(dD8G-dC) in the region of the base absorptions are presented Fig. 6a. The new band in the spectrum of the deuterated compound, observed at 1467 cm^{-1} , can be attributed by comparison with the spectra of films of dGMP, dD8GMP (Fig. 6c) and dC obtained from neutral pH solutions, and with the computed spectrum corresponding to a 1/1 ratio of dC and dD8GMP presented Fig. 6b. The contributions of the cytosines and of the guanines which are superimposed in the spectrum of the poly(dG-dC).poly(dG-dC) at 1496 cm^{-1} can be easily distinguished by the deuteration on the C8 of guanines : such a deuteration shifts the dGMP band from 1482 cm^{-1} to 1460 cm^{-1} (Fig. 6c). In the case of the polymer, the 1467 cm^{-1} band can therefore be attributed to a vibration of the selectively deuterated guanine. Raman spectroscopy studies of rGMP have shown that the line located at 1487 cm^{-1} and attributed to the 8 C-H deformation coupled to the guanine ring brea-

Figure 6. Effect of selective deuteration of guanine : a/ IR spectra of $\frac{p \cdot \overline{p} \cdot \overline{p}}{p \cdot \overline{p} \cdot \overline{p} \cdot \overline{p}}$ (dG-dC).poly (dG-dC).poly d(D8G-dC) (----);
b/ IR spectrum of dD8G (----), dC (-----), and recalculated spectrum with a l/l ratio of dD8G/dC (-----) ; c/ IR spectra of 2'-deoxyguanosine dG(----) and $dDSG$ $($ ------ $)$.

thing involving the N7 motion is similarly shifted to 1464 cm^{-1} by selective deuteration of the C8 site (25). Thus the modifications of the 1467 cm^{-1} absorption of the IR spectrum of poly(dD8G-dC).poly(dD8G-dC) will reflect specific interactions with the N7 of guanines.

The ionsused in our study affect differently this absorption as can be seen Fig. 3. The transition metal ions Co^{2+} and Ni²⁺ shift the 1467 $cm⁻¹$ band to higher frequencies, the absorption is now detected as a shoulder at 1473 cm⁻¹ (Fig. 3 c and d). At high hydration, Mg^{2+} remains effectless on this absorption. However in the conditions corresponding to the existence of the Z_0 conformation, a slight shift of this band to the higher wavenumbers (1470 cm⁻¹) is detected (Fig. 5). In this case we must nevertheless beware of effects not specifically correlated with an interaction on the N7 site and due to the structural modifications of the whole phosphodiester backbone.

Another potentially important interaction site for the metal ions must be expected at the level of the phosphate groups. At high relative humidity the binding of the M^{2+} ions induces only slight modifications in the stretching vibrations of the PO₂. The v_{e} PO₂ absorption band located around 1087 cm⁻¹ is not significantly shifted ; however the band becomes broader and its relative intensity is drastically decreased. The v_{as} PO₂ absorption around 1215 cm⁻¹ in the Z form of poly(dD8G-dC).poly(dD8G-dC)-Na⁺ is slightly shifted $(-5cm^{-1})$ upon addition of divalent ions (spectra not shown). This reflects the absence of specific site binding on the phosphate groups ; the ions retain their normal water molecule shell and reduce the effective charge on the polyelectrolyte ("territorial binding"). However, as already discussed, at low relative humidity and high M^2 /nucl. ratio, a very strong direct interaction with the phosphate groups has been evidenced for Mq^{2+} .

Concerning the possibility of an interaction with the carbonyl groups of the bases, the spectra presented Fig. 1 (samples in D_20) show that no chelation is detected neither with the transition metal ions Co^{2+} and Ni^{2+} , nor with the alcaline-earth Mg²⁺, as the doublet at 1666 cm⁻¹-1634 cm⁻¹ is observed at the same position as with the alcaline ion N_a^+ .

DISCUSSION

we have shown by IR spectroscopy that the Na⁺, Mg²⁺, Co²⁺ and Ni^{2+} stabilize in condensed hydrated phases the poly(dD8G-dC).poly(dD8G-dC) in the Z conformation. The specific deuteration of the 8-carbon of guanines allows to show that depending on the chemical properties of the metal ions (alcaline, alcaline-earth ions and first row transition metal ions), differences exist in the interaction with the N7 site of the guanine.

In the IR spectra recorded with Co^{2+} and Ni²⁺ as counter-ions the 1467 cm^{-1} band involving the N7 motion in the ring breathing coupled to the 8-CD deformation is shifted to a higher frequency. Correlated with this displacement the contribution to the 1354 cm^{-1} absorption of the dG residu in a syn conformation (26) is observed at a lower frequency (1343 cm⁻¹). From these results, it is clear that the 3d7 and 3d8 electronic configuration of Co^{2+} and Ni^{2+} have a great affinity for the electron donating site of N7 of guanines. This implies that the perturbation is sufficient to modify the electronic distribution around the N7 and to induce changes in the local force constants. In the case of magnesium ion, two different effects are detected depending on the hydration of the complex. For a high water content, the

1467 cm^{-1} band remains unshifted, while at low RH it is slightly displaced. X ray diffraction studies (29) have shown that in the d(C-G)3 crystallized in presence of Mg^{2+} , the metal ion is localized close to the N7 site. However as the Mg²⁺ is an alcaline-earth metal ion, the electronic perturbation expected on the N7 nucleophile site will be weaker than that due to a transition metal ion. Thus the +3 cm^{-1} shift of the 1467 cm^{-1} band at low RH compared to the + 6 cm⁻¹ in the case of Co²⁺ and Ni²⁺ can be interpreted as a loss of a water molecule between Mq^{2+} and the N7 site while this molecule would be present at higher hydration. We must notice however that the direct chelation of the Mg²⁺ to the N7 site, which is not observed at high relative humidity, is not necessary to stabilize the left-handed helix.

Thus we conclude from the different effects of the Co^{2+} , Ni²⁺ and Mg $^{2+}$ on the vibrational mode situated at 1467 $\rm cm^{-1}$ that the metal-ligand interaction constant is stronger between Co^{2+} or Ni^{2+} and the N7 of guanines than between Mg^{2+} and the same site. This is in good agreement with data previously obtained which proposed a similar order of metal ions binding ability in case of native DNA (30) and an electrostatic site binding of Mg^{2+} to phosphate groups (31).

The difference in the spectroscopic effect induced by $Co²⁺$ and $Ni²⁺$ compared to the effect of Mg²⁺ on the 1467 cm⁻¹ band seems to be well correlated with the efficiency of the metal ions to induce the B to Z transition in solution (1,16,18,32) and allows us to propose an explanation for the particular role of the nucleophile site in the mechanism of this transition. In conditions in which the B conformation in solution is mainly observed, the thermal agitation through the DNA respiration can modify locally the dG conformation into a syn geometry without breaking the Watson-Crick base pairing (2,7,33). The lifetime of this configuration may be very short but sufficient to expose, and thus increase the accessibility, of the N7 site of the guanine which will greatly favour the fixation of a metal ion to it. As our IR results show, the Co^{2+} and Ni²⁺ ions interact in a much more drastic way with the guanine than does the Mq^{2+} . So it can be assumed that in solution the former ions will stabilize longer the dG residu in a sun conformation than Mg^{2+} . Therefore a smaller amount of transition metal ions will be needed to induce a cooperative transition to the Z conformation. Thus our IR results give a satisfactory explanation of the quite different concentration of metal ions needed to induce the B to Z transition in solution.

The initiation of a biologically important event by the conformational change of DNA induced by metal-binding, may happen in vivo by local changes in the concentration of other ions, including proteins. Thus regulation may be achieved by small and therefore metabolically controled, changes in metal concentration.

*To whom correspondence should be addressed

REFERENCES

- 1. Pohl, F.M. and Jovin, T.M. (1972) J. Mol. Biol. 67, 375-396.
- 2. Wang, A.H.-J, Quigley, G.J., Kolpak, F.J., Crawford, J.L., Van Boom, J.H., Van der Marel, G.A. and Rich, A. (1979) Nature 282, 680-686.
- 3. Drew, H., Takano, T., Tanaka, S., Itakura, K. and Dickerson, R.E. (1980) Nature 286, 567-573.
- 4. Patel, D.J., Canuel, L.L. and Pohl, F.M. (1979) Proc. Natl. Acad. Sci. U.S.A. 76, 2508-2511.
- 5. Cohen, J.S., Wooten, J.B. and Chatterjee, C.L. (1981) Biochem. 20, 3049-3055.
- 6. Feigon, J., Wang, A.H-J., Van der Marel, G.A., Van Boom, J.H. and Rich, A. (1984) 12, 1243-1263.
- 7. Tran Dinh, S., Taboury, J., Neumann, J.M., Huynh-Dinh, T., Genissel, B., Longlois d'Estantot, B. and Igolen, J. (1984) Biochem., (in press).
- 8. Pohl, F.M., Ranade, A., Stockburger, M. (1973) Bioch. Biophys. Acta 335, 85-92.
- 9. Thamann, T.J., Lord, R.C., Wang, A.H.-J and Rich, A. (1981) Nucleic Ac. Res. 9, 5443-5457.
- 10. Taillandier, E., Taboury, J., Liquier, J., Sautiere, P. and Couppez, M. (1981) Biochimie 63, 895-898.
- 11. Pilet, J. and Leng, M. (1982) Proc. Natl. Acad. Sci., U.S.A. 79, 26-30.
- Nordheim, A., Pardue, M.L., Lafer, E.M., Möller, A., Stollar, B.D. and Rich, A. (1981) Nature 294, 417-421.
- 13. Lemeunier, F., Derbin, C., Malfoy, B., Leng, M. and Taillandier, E. (1982) Exp. Cell. Res. 141, 508-513.
- 14. Viegas-Pecquignot, E., Derbin, C., Lemeunier, F. and Taillandier, E. (1982) Ann. Genet. 25, 218-222.
- 15. Rich, A. (1982) in Cold Spring Harbor Symposium on Quantitative Biology, Vol. 1, p. 1.
- 16. Behe, M. and Felsenfeld, G. (1981) Proc. Natl. Acad. Sci., U.S.A. 78, 1619-1623.
- 17. Zacharias, W., Larson, J.E., Klysik, J., Stirdivant, S.M. and Wells,R.D. (1982) J. Biol. Chem. 257, 2775-2782.
- 18. Van de Sande, J.H., McIntosh, L.P. and Jovin, T.M. (1982) EMBO 1, 777-782.
- 19. Liquier, J., Bourtayre, P., Pizzorni, L., Sournies, F., Labarre, J.F. and Taillandier, E. (1984) Anticancer Res. (in the press).
- 20. Eichorn, G.L. (1973) Inorg. Biochem. 2, 1210-1240.
- Rose, D.M., Bleam, M.L., Record, M.J.Jr. and Bryant, R.G. (1980) Proc. Natl. Acad. Sci.,U.S.A., 77, 6289-6292.
- 22. Granot, J., Feigon, J. and Kearns, D.R. (1982) Biopolymers 21, 181-201.
- 23. Taillandier, E., Liquier, J. and Taboury, J.A. (1984) Advances in

Infrared and Raman Spectroscopy (in the press).

- 24. Thomas, G.J.Jr. and Livramento, J. (1975) Biochem. 14, 5210-5218.
- 25. Lane, M.J. and Thomas, G.J.Jr. (1979) Biochem. 18, 3839-3846
- 26. Ghomi, M., Taboury, J.A. and Taillandier, E. (1984) Biochimie (in the press).
- 27. Chinsky, L., Jolles, B., Laigle, A., Turpin, P.Y., Taboury, J.A. and Taillandier, E. (1984) Biopolymers (in the press).
- 28. Taboury, J.A. and Taillandier, E. (1984) Magnesium (in the press).
- 29. Wang, A.H-J., Quigley, G.J., Kolpak, F.J., Van der Marel, G.,
- Van Boom, J.H. and Rich, A. (1981) Science 211, 171-176. 30. Zimmer, C., Luck, G. and Triebel, H. (1974) Biopolymers 13, 425-453.
- 31. Clement, R.M., Sturm J. and Daune, M.P. (1973) Biopolymers 12, 405-421.
- 32. Van de Sande, J.H. and Jovin, T.M. (1982) EMBO J. 1, 115-120.
- 33. Harvey, S.C. (1983) Nucleic Ac. res. 11, 4867-4878.