Ring contraction of hydroporphinoid to corrinoid-complexes

(corrin synthesis/vitamin B_{12} biosynthesis/sulfide contractions/nucleophilic carbenes/photochemistry)

VITTORIO RASETTI*, ANDREAS PFALTZ*, CHRISTOPH KRATKY[†], AND ALBERT ESCHENMOSER^{*}

*Laboratory of Organic Chemistry, Eidgenössische Technische Hochschule Zürich, Universitätstrasse 16, CH-8092 Zürich, Switzerland; and †Institute of Physical
Chemistry, University of Graz, Heinrichstrasse 28, A-8010 Graz,

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ABSTRACT Crystalline nickel(II) and dicyanocobalt(III) com-

COOH COOH plexes of a racemic 1,2,2,7,7,12,12,17,17,20-decamethyl-20-hydroxy-1,2,3,7,8,12,13,20-octahydro-17H-porphyrin rearrange to \bigoplus_{HOOC} the corresponding complexes of racemic 19-acetyl-1,2,2,7,7,12,-

12,17,17-nonamethyl-trans-corrin on melting (approximately 290°C H₃C 20

and 260°C, respectively). The nickel(II) 19-acetylcorrinate formed H₃C 20

and in this way is shown to deacetylate to racemic nickel(II) 1,2,2,7,7,and 260°C, respectively). The nickel(II) 19-acetylcorrinate formed

in this way is shown to deacetylate to racemic nickel(II) 1,2,2,7,7,-

12,12,17,17-nonamethyl-trans-corrinate on treatment with 2 M
 $2.12,17,17-$ nonamet Example and 260°C, respectively). The nickel (II) 19-acetyl-1,2,2,7,7,12,-

Hooc 12.Hooking complexes of racemic 19-acetyl-1,2,2,7,7,12,-

12,17,17-nonamethyl-trans-corrin on melting (approximately 290°C

and 260°C, respe chemical models for the elusive ring contraction step in vitamin ϵ_{COM} B₁, biosynthesis.

The crucial and still unknown step in the biosynthesis of vitamin B_{12} from uroporphyrinogen (1, 2) is the generation of the characteristic A/D-ring junction of the corrinoid ligand. A major stride towards a solution of this problem was made in 1979 when it was discovered (3-5) that the third methyl group in the consecutive methyl group transfers from adenosylmethionine to hydroporphinoid intermediates most surprisingly becomes attached at the meso position C-20 between rings A and D (cf. structure ¹ in Fig. 1), only to leave the scene again, probably together with carbon atom C-20, at some unknown later stage of the uroporphyrinogen \rightarrow corrin transformation. It was this important finding combined with the results of recent chemical studies on the mechanistic spectrum of A/D -secocorrin \rightarrow corrin ring closures (6, 7) that led us to envisage and to test in a model system the chemical feasibility and characteristics of the hypothetical dihydrocorphinol^{$\ddagger \rightarrow$} corrin rearrangement depicted in Fig. 2. §

In a hexahydroporphinoid structure of type 2 protonation of the enaminoid double bond ofring D generates the structure 2a containing an α -tert-hydroxyiminium function, which in principle should be prone to undergo two isomeric rearrangements, one ofwhich constitutes a ring contraction to the 19-acetylcorrin structure 3. Such a transformation is formally analogous to the known semibenzoin rearrangement of α -tert-hydroxyketones and is in fact-as is best recognized in its stepwise mechanistic variant (see structure 2b)-very closely related to our previously described (7) A/D -secocorrin \rightarrow corrin cycloisomerization of 1-methylidene-19-formyl-1, 19-secocorrinates to 19-formylcorrinates.

A process of type $2 \rightarrow 3$ implies the isomerization of an allylic alcohol system to an unconjugated carbonyl function and may well, therefore, be thermodynamically feasible in the free ligand; on the other hand, in metal complexes the position of a corresponding equilibrium may reflect the preference of a given metal ion for either the hydroporphinoid or the (smaller) corrinoid coordination sphere. Kinetically, the rearrangement is expected to respond to acid/base catalysis and it may also offer a thermally inducible pathway via an intramolecular proton shift

FIG. 1. Hypothetical dihydro derivative of established vitamin B_{12} precursor 20-methylsirohydrochlorin (3-5).

from the tertiary hydroxyl group to the nucleophilic enamine carbon in ring D.

Fig. 3 summarizes the synthesis of the model structure 12, specifically prepared in order to put the above-mentioned expectations to an experimental test. In its first phase the synthesis makes use of previously described intermediates 4-10 (9, 10) and proceeds to the unusual and crucial photochemical cyclization step $10 \rightarrow 11$. This process is assumed to imply an attack of a photochemically generated nucleophilic carbene function at position C-19 of ring D onto the nitrile group (which is held in close proximity to the carbenoid sp^2 lone pair by the complexing cadmium ion), followed by a proton transfer from C-18 to the negatively charged nitrogen of the newly formed ketiminate function (see $10a \rightarrow 10b \rightarrow 10c$, in Fig. 3). After chromatographic hydrolysis of the ketimine group of 10c on silica gel and acidolytic decomplexation with trifluoroacetic acid, crystalline ¹¹ is isolated in up to 60% yield. A Grignard reaction with CH3MgI gives 12 as only one of the two possible diastereomers; the reaction is therefore assumed to proceed under steric control, and the configuration of 12 is tentatively assigned as indicated.

A variant of the synthesis of ¹² (see Fig. 4) uses as rings A/B precursor the bicyclic acetylthiolactam 13 available from 4 by reaction with $CH₃MgI$, followed by (ketimine) hydrolysis and introduction of sulfur at the lactam function with the "Lawesson reagent" (11). A sequence of reactions analogous to the one described above converts 13 into a (noncrystallized) chloro-cadmium complex 14. This is cyclized photochemically, either as such or, preferably, as the lithium complex 14a to give

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Abbreviations: DBU, 1,5-diazabicyclo[5.4.0]undec-5-ene; RT, room temperature.

^{*} For the hexahydroporphinoid ligand system named "corphin" see ref. 8.

[§] Any extrapolation of structure ¹ to a potential biosynthetic intermediate of structural type 2 has to obey only two *a priori* constraints: methylation at C-1 preceding methylation at C-17 and decarboxylation preceding methylation at C-12.

FIG. 2. Hypothetical dihydrocorphinol \rightarrow corrin rearrangement.

two diastereomeric products, which can be separated and were isolated in the form of their crystalline nickel(II) (perchlorate) complexes 15; the main diastereomer proved to be identical with the nickel(II) complex prepared from 12 derived from 11.

The reaction sequence described in Fig. 4 had been carried out in the hope that the zwitterionic intermediate 15a in the photochemical cyclization of 14 or 14a would undergo directly ring contraction to the 19-acetylcorrinate (see discussion of Fig. 2). This, however, was not the case, the proton transfer leading from 15a to 15 apparently being faster than the rearrangement

under the conditions of the photochemical ring closure.

) analysis (see Fig. 6). Worthy of mention are the quite unusual Ring contraction to 19-acetylcorrinate does occur, however, when crystalline samples of both the nickel(II) (chloride) complex 15 and the dicyanocobalt(III) complex 16 are heated for a few minutes above their melting points (approximately 290'C and 260°C, respectively) under exclusion of oxygen (yields up to 40%) (see Fig. 5). Mass, ultraviolet/visible, infrared, and 'H NMR spectra of the (crystalline) products (see Experimental Data) fully support their structural assignments as racemic nickel(II) and dicyanocobalt(III) 19-acetyl-1,2,2,7,7, 12, 12,- 17,17-nonamethyl-trans-corrinates 17 and 18, respectively. For the nickel corrinate 17 this assignment is corroborated by x-ray chemical shifts of the CH3CO- methyl singlets in the 'H NMR spectra of 17 and 18 (2.79 and 2.58 ppm relative to tetramethylsilane); model considerations suggest, and the results of the xray analysis (see Fig. 6) support the contention, that the sterically least-crowded position of this methyl group is the one pointing towards the metal ion.

> Deformylation of nickel(II) 19-formylcorrinates by hydroxide ions has been demonstrated previously (7); deacetylation of the nickel(II) 19-acetyl-corrinate 17 also proceeds under similar conditions, though, understandably, much more sluggishly (days instead of hours at ⁷⁰'C with ² M KOH). The crystalline product of deacetylation was shown to be racemic nickel(II) 1,2,2,7,7,12,12,17,17-nonamethylcorrinate perchlorate 19 by direct comparison (ultraviolet/visible, infrared, 'H NMR, mass spectroscopy, thin-layer chromatography) with a previously synthesized (9) authentic sample.

It remains to be learned whether the ring contraction/deace-

FIG. 3. Synthesis of 12. THF, tetrahydrofuran; DBU, 1,5-diazabicyclo[5.4.0]undec-5-ene.

FIG. 4. Variant of the synthesis of 12.

tylation sequence can also be effected in the experimentally more demanding metal-free model series. Encouragement for the pursuit of such studies comes from recent results that Arigoni and Battersby and their colleagues describe in the accompanying papers (12-14), which show that one acetic acid equivalent is extruded during the biosynthetic ring contraction process. The special appeal of the dihydrocorphinol \rightarrow corrin rearrangement as a model for this process resides in that it assigns to the methyl group at C-20 a chemically transparent function in the ring contraction and, furthermore, that it would allow corrin biosynthesis to proceed from the beginning to the end at a single oxidation level (7).

EXPERIMENTAL DATA

Reaction conditions

Percentages in parentheses indicate product yields. Numbers indicate molar ratios of reagents or solvents. RT, room temperature. $4 \rightarrow 10$: see refs. 7 and 9. $10 \rightarrow 11$: (i) $h\nu$ (Hg) in dry

FIG. 5. Ring contraction and deacetylation.

FIG. 6. Crystal structure of 17. The molecule is projected into the plane of atoms N-21-N-22-N-23 and tilted by 21° to minimize overlap. The radii of circles around atom positions are arbitrary and increase in the order $C < N < O < N$ i.

H₄furan, 15°C; (ii) CF₃COOH/CH₂Cl₂, RT (ca. 60%). $11 \rightarrow 12$: (i) DBU, H₄furan; (ii) CH₃MgI, ether, 33°C (ca. 50%). $12 \rightarrow 16$: (i) 15 $Co(OAc)_2$ ⁻4H₂O, CH₃CN, 60°C; (ii) KCN, air (ca. 80%). $16 \rightarrow 18$: ca. 265°C under argon, 3 min (ca. 30%). $4 \rightarrow 13$: (i) 4.5 CH3MgI, diethyl ether, RT, work up with aqueous 0.1 M HCl, 2 hr, RT (60% yield of acetyllactam, mp 128° C); (ii) 0.55 p-anisylthionophosphine sulfide (11) in H₄furan, 20°C, 1 hr (90%) . 13 + 8 \rightarrow 14: (i) 6 DBU, CH₃CN, RT; (ii) 2.5 $CdCl_2 \cdot 2H_2O$, 2.0 $(C_6H_5)_3P$, NaHCO₃, CH₃OH, RT (40-55%). $14 \rightarrow 14a \rightarrow 15$ (X = ClO₄⁻): (i) 10 KCN, CH₃OH, RT; (ii) excess LiClO₄, solid NaOH, H₄furan, RT; (iii) $h\nu$ (Hg/Pyrex), toluene, 5-10 $^{\circ}$ C; (iv) 10 NiCl₂ \cdot 6H₂O, acetate buffer, CH₃OH, 60 $^{\circ}$ C, work up with 0.1 M HClO_4 , chromatographic separation (silica gel, NaClO₄) of two diastereomers (5:1) (40%). 15 (X = Cl) \rightarrow 17: melt (295°C) 5 min, under argon, work up with 0.1 M HClO₄ (40%). 17 \rightarrow 19: 2 M KOH in C₂H₅OH/H₂O (9:1), 70°C, 20 days in evacuated tube (ca. 20%).

Spectral data

Data are given in the following order: ultraviolet/visible $(\lambda_{\text{max}}$ in nm, $\log \varepsilon (M^{-1} \text{ cm}^{-1})$; spectra taken in C₂H₅OH), infrared ($\bar{\nu}$ in cm⁻¹; CHCl₃), ¹H NMR (δ in ppm, δ tetramethylsilane = 0 ppm, C^2HCl_3), molecular weight as determined by mass spectroscopy. Only selected infrared and NMR data are given. sh, shoulder; s, singlet; M^+ , parent ion.

11. (mp 183°C): $\lambda_{\text{max}} = 230 (3.98), 261 (4.31), 292 (4.29), 322$ $(\text{sh}/4.23)$, 339 $(\text{sh}/4.36)$, 353 (4.41) , 504 (4.11) ; $\tilde{\nu} = 3270$, 1775/ 1730, 1665, 1620, 1565; $\delta = 7.18/5.62$ (2s/2H), 5.68 (s/2H) (4 $=CH$ —); M⁺ (- CF₃COOH) = 458.

12. (mp 194°C): $\lambda_{\text{max}} = 264 (4.15), 324 (4.48), 335 (4.48), 349$ $(\text{sh}/4.37), 382 (\text{sh}/3.92), 406 (\text{sh}/3.75), 504 (4.09); \tilde{\nu} = 3595/$ 3455/3270, 1660, 1617, 1565; $\delta = 6.18/5.57/5.51/5.49$ (4s/4 =CH-), 1. 75/1.50/1.42/1.39/1.37/1.34/1.31/1.27/1.22/1.18 $(10s/10 \text{ CH}_3)$; M⁺ (- CF₃COOH) = 474.

13. (mp 128°C): $\lambda_{\text{max}} = 245 (4.15), 332 (4.33), 410 (3.49); \tilde{\nu} =$ 1700, 1655, 1570; δ (CH₃CO) = 2.32; M⁺ = 292.

14. $\lambda_{\text{max}} = 276 (4.40), 346 (4.49), 550 (4.08); \bar{\nu} = 1700, 1628,$ 1600, 1572; $\delta = 5.20/5.03/5.00$ (3s/3 = CH-), 2.17 (s/ CH₃CO).

15. (X = ClO₄⁻ main diastereomer, mp 240°C): $\lambda_{\text{max}} = 284$ (4.02), 338 (4.34), 348 (sh/4.28), 396 (sh/3.75), 420 (sh/3.80), 467 (4.00) ; $\bar{\nu} = 3595/3460$, 1637, 1568, 1515; $\delta = 6.23/6.12/6.04/$ 5.77 (4s/4 = CH--), 1.97/1.44/1.39/1.30/1.20/1.05 (6s/6 CH₃), 1.34/1.29 (2s/4 CH₃); M^+ (- HClO₄) = 530 (⁵⁸Ni).

16. (mp 248°C): $\lambda_{\text{max}} = 304$ (3.91), 346 (sh/3.95), 362 (sh/ 4.27), 380 (4.41), 422 (sh/3.43), 553 (3.85); $\bar{\nu} = 3590$, 2120, 1637, 1573, 1520; $\delta = 6.11/5.47/5.41/5.40$ (4s/4 = CH--), 1.89/

1.50/1.35/1.30/1.28/1.26/1.25/1.19/1.16/1.02 (10s/10 CH₃); M^+ (- 2CN) = 532.

17. Racemic nickel(II) 19-acetyl-1,2,2,7,7,12,12,17,17-nonamethylcorrinate perchlorate (mp 172°C): $\lambda_{\text{max}} = 245(4.17), 254$ $(\frac{\sin(4.07)}{270} \cdot \frac{270 \cdot (\frac{\sin(3.94)}{308} \cdot \frac{308 \cdot (\frac{\sin(4.30)}{4.30})}{323 \cdot (4.38)} \cdot 433 \cdot (4.02)}$, 454 $(\text{sh}/3.95)$; $\tilde{\nu} = 1707, 1628, 1582, 1554, 1510, \delta = 6.30/6.04/5.94$ $(3s/3=CH-), 2.79 (s/CH₃CO), M⁺ (- HClO₄) = 530 (°Ni).$

X-ray analysis of 17 (see Fig. 6) (crystal from methyl acetate/ hexane): orthorombic, $Pna2_1$, $a = 20.58$ Å, $b = 11.44$ Å, $c =$ 14.68 Å, Z = 4. There were 4508 reflections (sin $\vartheta/\lambda \le 0.54$) measured at -168° C, of which 2268 were observed $[|F| >$ $2.5\sigma(|F|)$, $R = 0.15$ (208 parameters refined). The asymmetric unit contains one disordered ClO₄⁻ ion and a disordered solvent molecule.

18. Racemic dicyanocobalt(III) 19-acetyl-1,2,2,7,7,12,12, 17,17-nonamethylcorrinate (mp 215°C): $\lambda_{\text{max}} = 274 (4.00), 290$ (3.91), 304 (3.93), 318 (3.90), 345 (sh/4.14), 363 (4.38), 415 $(3.56), 495 (sh/3.61), 526 (3.73), 567 (3.81); \tilde{\nu} = 2120 (CN), 1695$ (C=O), 1624, 1583, 1560, 1510; $\delta = 5.57$ (s/1H), 5.52 (s/2H) $(=CH\rightarrow)$, 2.58 (s/CH₃CO); M⁺ (- HCN, - CN) = 531.

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- 1. Scott, A. I. (1978) Acc. Chem. Res. 11, 29–36.
2. Battersby. A. R. & McDonald. E. (1978)
- Battersby, A. R. & McDonald, E. (1978) Bioorg. Chem. 7, 161-173.
- 3. Battersby, A. R., Matcham, G. W. J., McDonald, E., Neier, R., Thompson, M., Woggon, W.-D., Bykovsky, V. Y. & Morris, H. R.

(1979) J. Chem. Soc. Chem. Commun., 185-186.

- 4. Lewis, N. G., Neier, R., Matcham, G. W. J., McDonald, E. & Battersby, A. R. (1979) J. Chem. Soc. Chem. Commun., 541-542.
- 5. Muller, G., Gneuss, K. D., Kriemler, H.-P., Scott, A. I. & Irwin, A. J. (1979) J. Am. Chem. Soc. 101, 3655-3657.
- 6. Eschenmoser, A. (1976) Chem. Soc. Rev. 5, 377-410.
- 7. Pfaltz, A., Buhler, N., Neier, R., Hirai, K. & Eschenmoser, A. (1977) Helv. Chim. Acta 60, 2653-2672.
- 8. Johnson, A. P., Wehrli, P., Fletcher, R. & Eschenmoser, A. (1968) Angew. Chem. 80, 622-625; Angew. Chem. Int. Ed. EngL 7, 623-625.
- 9. Pfaltz, A., Hardegger, B., Muller, P. M., Farooq, S., Krautler, B.

& Eschenmoser, A. (1975) Helv. Chim. Acta 58, 1444-1450.

- 10. Pfaltz, A. (1978) Dissertation, Eidgenössische Technische Hochschule Zürich, No. 6184 (Juris, Zürich), pp. 78-95.
- 11. Scheibye, S., Pedersen, S. & Lawesson, S.-0. (1978) BulL Soc. Chim. BeIg. 87, 229-232.
- 12. Mombelli, L., Nussbaumer, C., Weber, H., Muller, G. & Arigoni, D. (1981) Proc. Nati Acad. Sci. USA 78, 11-12.
- 13. Nussbaumer, C., Imfeld, M., Worner, G., Muller, G. & Arigoni, D. (1981) Proc. NatL Acad. Sci. USA 78, 9-10.
- 14. Battersby, A. R., Bushell, M. J., Jones, C., Lewis, N. G. & Pfenninger, A. (1981) Proc. Nati Acad. Sci. USA 78, 13-15.