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EDITORIAL

Intracellular chloride regulates the G₁/S cell cycle progression in gastric cancer cells

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Abstract

Recent studies show that ion channels/transporters play important roles in fundamental cellular functions. Several reports indicating the important roles of Cl channels/transporters on cell proliferation suggest that the intracellular chloride concentration ([Cl⁻]_i) regulated by them would be one of critical messengers. We investigated whether the [Cl⁻]_i controls cell proliferation and cell cycle progression in human gastric cancer cells. Our studies indicated that furosemide, a blocker of $Na^{+}/K^{+}/2Cl^{-}$ cotransporter (NKCC), diminished cell growth by delaying the G1-S phase progression in gastric cancer cells with high expression and activity of NKCC. Furthermore, we found that the culture in the low Cl⁻ medium (replacement of Cl⁻ by NO₃⁻) decreased the [Cl⁻] and inhibited cell growth of gastric cancer cells and that this inhibition of cell growth was due to cell cycle arrest at the G₀/G₁ phase caused by diminution

of CDK2 and phosphorylated Rb. The culture of cells in the low Cl⁻ medium significantly increased expressions of p21 mRNA and protein. In addition, the low Cl⁻ medium induced phosphorylation of mitogen activated protein kinases (MAPKs). Treatment with an inhibitor of p38 or JNK significantly suppressed p21 upregulation caused by culture in a low Cl⁻ medium and rescued gastric cancer cells from the low Cl⁻-induced G₁ cell cycle arrest. These findings revealed that the [Cl⁻]_i affects the cell proliferation *via* activation of MAPKs through upregulation of p21 in gastric cancer cells. Our results suggest that the [Cl⁻]_i regulates important cellular functions in gastric cancer cells, leading to the development of novel therapeutic strategies.

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Key words: Intracellular chloride; Cell proliferation; Cell cycle; Gastric cancer; Cl⁻ channel; Cl⁻ transporter

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INTRODUCTION

Over the past two decades, many researchers have reported that ion channels and transporters play important roles in fundamental cellular functions. Particularly, their physiological roles in cell proliferation have been considered since cell volume changes, which require the participation of ion movement across the cell membrane,



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are indispensable in cell cycle progression. Recently, the roles of ion transporters have been studied in cancer cells^[1-3] and various types of ion transporters have been found in cancers of digestive organs. Some types of K⁺ channels, such as voltage-gated Kv1.3 channel, voltage-gated HERG channel and KCNK9 channel, have been reported to be expressed at extremely higher levels in colonic carcinoma specimens than in normal colon tissues^[4-6]. In gastric cancer, the voltage-gated HERG channel has revealed cancer-limited expression and its blocker diminishes the G₁ to S phase transition^[7]. Furthermore, increased mRNA levels of voltage-gated Cav1.2 Ca²⁺ channels and Ca²⁺-conducting channels (TRPM8) have been reported in colorectal adenocarcinoma^[8,9].

Several reports indicate that CI channels/transporters play an important role in gastrointestinal cancer cells. For instance, transcriptional downregulation of Ca²⁺activated 2Cl channels (CLCA1 and CLCA2) genes is detected in colorectal tumor samples^[10]. Sarosi et al^[11] have reported that the Cl/HCO3 exchanger influences the proliferation of Barrett's esophageal adenocarcinoma cells through changes of intracellular pH. In addition, we previously found that mRNA and the functional expression levels of $Na^+/K^+/2Cl^-$ cotransporter (NKCC) were higher in poorly-differentiated type gastric adenocarcinoma cells than in differentiated cells^[12]. These reports indicate that the transepithelial CI transport plays an important role in cell proliferation of gastrointestinal cancer cells. From this viewpoint, we investigated whether the intracellular chloride concentration ([Cl]i) regulates cell proliferation and cell cycle progression in human gastric cancer cells.

INTRACELLULAR CHLORIDE AND SIGNAL TRANSDUCTION

Our study^[13] indicates that the intracellular chloride could act as a signal to regulate mRNA expression of ion channel in renal epithelial A6 cells, suggesting that many physiological functions are associated with the change of the [Cl]:. Similarly, Heimlich *et al*¹⁴] have shown that an alteration of the [Cl]i plays an important role in the activation of signaling molecules upstream of the mitochondria, specifically impairing the intrinsic apoptotic pathway. In a mouse macula densa cell line, low chloride stimulates prostaglandin E2 release and COX-2 expression through activation of mitogen activated protein kinases (MAPKs)^[15]. These reports suggest that the changes in the [CI]: might act as a regulator of various types of intracellular enzymes. We previously showed that furosemide, a blocker of NKCC, diminished cell growth by delaying the G1-S phase progression in gastric cancer cells with high expression and activity of NKCC^[12] (Figure 1). NKCC is one of the important transporters controlling the [CI]i via uptake of Cl into the intracellular space and, therefore, furosemide decreases the $[CI]_{1}^{[16]}$ (Figure 1). Based on these findings, we hypothesized that the [CI]: would be one of critical messengers regulating cell proliferation and

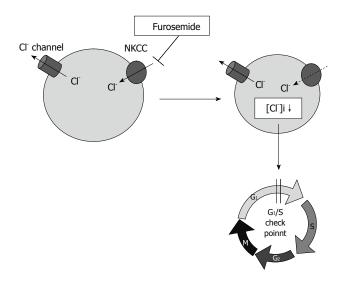


Figure 1 Na⁺/K⁺/2Cl⁻ cotransporter controls the intracellular chloride concentration via uptake of Cl⁻ into the intracellular space. Furosemide, a blocker of Na⁺/K⁺/2Cl⁻ cotransporter, delays the G₁-S phase progression by decreasing the intracellular chloride concentration ([Cl⁻]) in gastric cancer cells.

investigated whether the [Cl]i regulates cell cycle progression in human gastric cancer cells.

CELL CYCLE PROGRESSION AND [CI]; IN GASTRIC CANCER CELLS

We directed our interest to the roles of the [Cl]i in cell proliferation and cell cycle progression of gastric cancer cells. We applied media containing various chloride concentrations to human gastric cancer MKN28 cells and measured the [Cl]i at 48 h after the application. The [CI]i of gastric cancer cells incubated in the normal medium was around 30 mmol/L. When cells were incubated in the low Cl⁻ medium (replacement of Cl by NO₃) for 48 h, the [Cl]i decreased to around 0 mmol/L. Furthermore, the [Cl]i of cells cultured in the media containing various chloride concentrations was proportionally dependent on the chloride concentration of the cultured medium^[17,18]. These findings indicated that our experimental system using the low CI medium can be used as a model of the [Cl]i regulation (Figure 2). The proliferation rate in MKN28 cells was significantly diminished by the culture in the low CI medium compared with that in a normal one. In addition, analysis of cell proliferation of MKN28 cells cultured in the media containing various chloride concentrations indicated that the rate of cell proliferation depends on the extracellular chloride concentration^[18]. These results revealed that the [Cl]_i plays a key role in proliferation of gastric cancer cells. Cell cycle analysis revealed that the population of MKN28 cells staying in the G₀/G₁ phase was significantly increased and that cells staying in the S or G2/M phase were reduced by the culture in the low CI medium, suggesting that the decrease of the [CI]i shows an inhibitory effect on the proliferation of gastric cancer cells by mainly diminishing the transition from the G_1 phase to the S phase^[18] (Figure 3).

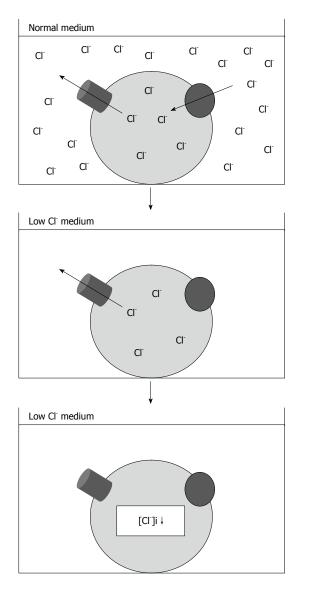


Figure 2 Experimental method for regulation of the intracellular chloride concentration of cultured cells. The intracellular chloride concentration ([CI]) of gastric cancer cells is decreased by the culture in the low CI medium, which were prepared by substituting, respectively, NaCI and KCI with NaNO₃ and KNO₃.

[CI]; CONTROLS THE G1/S CELL CYCLE CHECK POINT BY REGULATING THE EXPRESSION OF p21 IN GASTRIC CANCER CELLS

We analyzed the expression of cell cycle-associated proteins involved in G₁-S phase transition to determine the mechanisms by which the decrease of the [Cl]_i inhibited the proliferation of MKN28 cells. The culture in the low Cl⁻ medium significantly decreased phosphorylation of Rb. The expression of CDK2 protein, which is located at upstream of Rb in a signal pathway, was significantly downregulated by the culture in the low Cl⁻ medium. Furthermore, the low Cl⁻ medium elevated expression of p21^[18]. These results suggest that the [Cl]_i of gastric cancer cells acts on the transition from the G₁ phase to

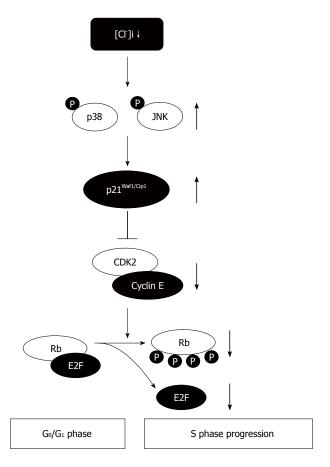


Figure 3 Roles of the intracellular chloride concentration in cell cycle progression of gastric cancer cells. The intracellular chloride concentration ([CI]) affects the cell proliferation *via* activation of p38 and/or JNK cascades through upregulation of the p21. The decrease of the [CI] causes the Ga/G1 phase arrest by diminishing expression of CDK2 and phosphorylated Rb due to an increase in expression of p21.

the S phase by regulating the expression of p21 and its downstream proteins in signal pathways (Figure 3). We found similar effects of the [CI]ⁱ on cell cycle associated proteins in human prostatic cancer cells and mouse osteoblast cells^[16,19].

[CI]; REGULATES p21 THROUGH THE ACTIVATION OF MAPKs IN GASTRIC CANCER CELLS

Generally, the induction of p21 is known to be dependent on tumor suppressor protein, p53, leading us to an idea that the upregulation of p21 induced by the decrease of the [CI]_i is due to activation of p53. However, the total expression and phosphorylation of p53 protein were not affected by application of the low CI⁻ medium^[18], indicating that the upregulation of p21 induced by the decrease of the [CI]_i was not dependent on activation of p53.

Therefore, we determined that MAPKs are involved in the p21 upregulation and cell cycle arrest induced by reduction of the [Cl]. Culture of MKN28 cells in the low Cl medium significantly induced phosphorylation of MAPKs (ERK, p38, and JNK)^[20]. Treatment with an inhibitor of p38 or JNK significantly suppressed p21 upregulation caused by culture in the low CI⁻ medium and rescued MKN28 cells from the low CI⁻ induced G₁ cell cycle arrest, whereas treatment with an ERK inhibitor had no significant effect on p21 expression or the growth of MKN28 cells in the low CI⁻ medium^[20]. These results suggest that the [CI]_i affects the cell proliferation *via* activation of p38 and/or JNK cascades through upregulation of the p21 in gastric cancer cells (Figure 3).

CONCLUSION

We showed that: (1) chloride has significant effects on cell cycle progress in human gastric cancer cells; (2) the decrease of the [CI]_i causes the G₀/G₁ phase arrest by diminishing expression of CDK2 and phosphorylated Rb due to an increase in expression of p21; (3) the [CI]_i has significant effects on the activity of MAPKs cascades; and (4) the activation of p38 and JNK by a low CI⁻ condition leads to growth inhibition *via* an increase of p21 expression. Our results suggest that the [CI]_i regulates intracellular signaling cascades participating in the control of proliferation in gastric cancer cells. A deeper understanding of these mechanisms may lead to the discovery of the [CI]_i as an important mediator in tumor development and as a novel therapeutic target for gastric cancer.

REFERENCES

- 1 **Russell JM**. Sodium-potassium-chloride cotransport. *Physiol Rev* 2000; **80**: 211-276
- 2 Kunzelmann K. Ion channels and cancer. J Membr Biol 2005; 205: 159-173
- 3 Schönherr R. Clinical relevance of ion channels for diagnosis and therapy of cancer. J Membr Biol 2005; 205: 175-184
- 4 Abdul M, Hoosein N. Voltage-gated potassium ion channels in colon cancer. Oncol Rep 2002; 9: 961-964
- 5 Lastraioli E, Guasti L, Crociani O, Polvani S, Hofmann G, Witchel H, Bencini L, Calistri M, Messerini L, Scatizzi M, Moretti R, Wanke E, Olivotto M, Mugnai G, Arcangeli A. herg1 gene and HERG1 protein are overexpressed in colorectal cancers and regulate cell invasion of tumor cells. *Cancer Res* 2004; 64: 606-611
- 6 Kim CJ, Cho YG, Jeong SW, Kim YS, Kim SY, Nam SW, Lee SH, Yoo NJ, Lee JY, Park WS. Altered expression of KCNK9 in colorectal cancers. *APMIS* 2004; **112**: 588-594
- 7 Shao XD, Wu KC, Hao ZM, Hong L, Zhang J, Fan DM. The potent inhibitory effects of cisapride, a specific blocker for human ether-a-go-go-related gene (HERG) channel, on gastric cancer cells. *Cancer Biol Ther* 2005; **4**: 295-301
- 8 Wang XT, Nagaba Y, Cross HS, Wrba F, Zhang L, Guggino

SE. The mRNA of L-type calcium channel elevated in colon cancer: protein distribution in normal and cancerous colon. *Am J Pathol* 2000; **157**: 1549-1562

- 9 Tsavaler L, Shapero MH, Morkowski S, Laus R. Trp-p8, a novel prostate-specific gene, is up-regulated in prostate cancer and other malignancies and shares high homology with transient receptor potential calcium channel proteins. *Cancer Res* 2001; **61**: 3760-3769
- 10 Bustin SA, Li SR, Dorudi S. Expression of the Ca2+-activated chloride channel genes CLCA1 and CLCA2 is downregulated in human colorectal cancer. DNA Cell Biol 2001; 20: 331-338
- 11 Sarosi GA, Jaiswal K, Herndon E, Lopez-Guzman C, Spechler SJ, Souza RF. Acid increases MAPK-mediated proliferation in Barrett's esophageal adenocarcinoma cells via intracellular acidification through a Cl-/HCO3- exchanger. *Am J Physiol Gastrointest Liver Physiol* 2005; 289: G991-G997
- 12 Shiozaki A, Miyazaki H, Niisato N, Nakahari T, Iwasaki Y, Itoi H, Ueda Y, Yamagishi H, Marunaka Y. Furosemide, a blocker of Na+/K+/2Cl- cotransporter, diminishes proliferation of poorly differentiated human gastric cancer cells by affecting G0/G1 state. J Physiol Sci 2006; 56: 401-406
- 13 Niisato N, Eaton DC, Marunaka Y. Involvement of cytosolic Cl- in osmoregulation of alpha-ENaC gene expression. Am J Physiol Renal Physiol 2004; 287: F932-F939
- 14 **Heimlich G**, Cidlowski JA. Selective role of intracellular chloride in the regulation of the intrinsic but not extrinsic pathway of apoptosis in Jurkat T-cells. *J Biol Chem* 2006; **281**: 2232-2241
- 15 Yang T, Park JM, Arend L, Huang Y, Topaloglu R, Pasumarthy A, Praetorius H, Spring K, Briggs JP, Schnermann J. Low chloride stimulation of prostaglandin E2 release and cyclooxygenase-2 expression in a mouse macula densa cell line. J Biol Chem 2000; 275: 37922-37929
- 16 Hiraoka K, Miyazaki H, Niisato N, Iwasaki Y, Kawauchi A, Miki T, Marunaka Y. Chloride ion modulates cell proliferation of human androgen-independent prostatic cancer cell. *Cell Physiol Biochem* 2010; 25: 379-388
- 17 Miyazaki H, Shiozaki A, Niisato N, Marunaka Y. Physiological significance of hypotonicity-induced regulatory volume decrease: reduction in intracellular Cl- concentration acting as an intracellular signaling. *Am J Physiol Renal Physiol* 2007; 292: F1411-F1417
- 18 Miyazaki H, Shiozaki A, Niisato N, Ohsawa R, Itoi H, Ueda Y, Otsuji E, Yamagishi H, Iwasaki Y, Nakano T, Nakahari T, Marunaka Y. Chloride ions control the G1/S cell-cycle checkpoint by regulating the expression of p21 through a p53-independent pathway in human gastric cancer cells. *Biochem Biophys Res Commun* 2008; **366**: 506-512
- 19 Maki M, Miyazaki H, Nakajima K, Yamane J, Niisato N, Morihara T, Kubo T, Marunaka Y. Chloride-dependent acceleration of cell cycle via modulation of Rb and cdc2 in osteoblastic cells. *Biochem Biophys Res Commun* 2007; 361: 1038-1043
- 20 Ohsawa R, Miyazaki H, Niisato N, Shiozaki A, Iwasaki Y, Otsuji E, Marunaka Y. Intracellular chloride regulates cell proliferation through the activation of stress-activated protein kinases in MKN28 human gastric cancer cells. J Cell Physiol 2010; 223: 764-770

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