Erratum

Co-regulation of β -lactam resistance, alginate production and quorum sensing in *Pseudomonas* aeruginosa

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The legend and labels for Fig. 1 on page 151 are incorrect. The last line of the legend should read one milliunit (mU) of β -lactamase is defined as 1 nmol nitrocefin hydrolysed min⁻¹ (µg protein)⁻¹, not one Miller unit of β -lactamase is defined as 1 nmol nitrocefin hydrolysed min⁻¹ (µg protein)⁻¹. The *y*-axis of Fig. 1 should be labelled β -lactamase activity (mU), not β -lactamase activity (Miller units).

The labels for Fig. 2 on page 151 are incorrect. The *y*-axis of Fig. 2 should be labelled β -galactosidase activity (Miller units), not β -lactamase activity (Miller units).

The labels for Fig. 3 on page 152 are incorrect. The *y*-axis of Fig. 3 should be labelled β -galactosidase activity (Miller units), not β -lactamase activity (Miller units). The correct figures are reproduced below.



Fig. 1. β -Lactamase expression in the Alg⁺ PDO*ampR* mutant. Assays were performed using the parent strain Alg⁺ PDO300, the mutant Alg⁺ PDO*ampR* and Alg⁺ PDO*ampR* (pAmpR) in the absence (-) and presence (+) of an inducer. The plasmid pAmpR carries the wild-type *ampR* gene on a broad-host-range low-copy-number plasmid pME6030 (Heeb *et al.*, 2000). Fresh cultures of OD₆₀₀ 0.6–0.8 were induced with 100 µg benzylpenicillin ml⁻¹ for 3 h before harvesting. Assays were performed on sonicated lysate using nitrocefin as a chromogenic substrate. Assays were performed in triplicate. One milliunit (mU) of β -lactamase is defined as 1 nmol nitrocefin hydrolysed min⁻¹ (µg protein)⁻¹.



Fig. 2. The effects of the *ampR* mutation on *algT/U* transcription. The promoter fusion $P_{algT/U}$ -*lacZ* was introduced into Alg⁻ PAO1, Alg⁻ PAO*ampR*, Alg⁺ PDO300 and Alg⁺ PDO*ampR*. Induction was carried out using 500 µg benzylpenicillin ml⁻¹ and the β -galactosidase activity was determined in Miller units after 30 min incubation. The basal level of expression was detected in the promoterless *lacZ* vector, pLP170.



Fig. 3. The effects of the *ampR* mutation on QS *las* and *rhl* gene transcription. The alteration in the transcription of the QS systems in Alg⁻ PAO1 (hatched bars), Alg⁻ PAO*ampR* (grey bars), Alg⁺ PDO300 (white bars) and Alg⁺ PDO*ampR* (black bars) was monitored using four transcriptional fusions, P_{*lasl*}-*lacZ*, P_{*lasR*}-*lacZ*, P_{*rhlR*}-*lacZ*. The promoterless *lacZ* vector has a low basal level of activity of <20 Miller units.