

NIH Public Access

Author Manuscript

J Am Soc Echocardiogr. Author manuscript; available in PMC 2012 November 1

Published in final edited form as:

J Am Soc Echocardiogr. 2011 November ; 24(11): 1276–1284.e5. doi:10.1016/j.echo.2011.08.002.

Segmental Analysis of Carotid Arterial Strain Using Speckle-Tracking

Eric Y. Yang, MD^{*,†,1}, Hisham Dokainish, MD, FACC, FASE^{‡,1}, Salim S. Virani, MD, FACC^{§,*,†}, Arunima Misra, MD, FACC^{\$,*}, Allison M. Pritchett, MD, FACC^{\$,*}, Nasser Lakkis, MD, FACC^{\$,*}, Gerd Brunner, PhD^{*}, Jaromir Bobek, RCIS^{\$,*}, Marti L. McCulloch, MBA, RDCS, FASE[†], Craig J. Hartley, PhD^{*}, Christie M. Ballantyne, MD, FACC, FACP, FNLA^{*,†}, Sherif F. Nagueh, MD, FACC, FAHA, FASE[†], and Vijay Nambi, MD, FACC, FASE, FNLA^{*,†,\$}

[†]The Methodist DeBakey Heart and Vascular Center, the Methodist Hospital – Houston, TX

^{\$}Ben Taub General Hospital, Houston TX

[‡]McMaster University – Ontario, CA

§Michael E. DeBakey Veterans Affairs Medical Center, Houston, TX

Abstract

Background—Increased arterial stiffness has been shown to be associated with aging and cardiovascular risk factors. Speckle-tracking algorithms are being used to measure myocardial strain. We evaluated if speckle-tracking could be used to measure carotid arterial wall strain (CAS) reproducibly in healthy volunteers and then examined if CAS was lesser in individuals with diabetes.

Methods—Bilateral electrocardiography-gated ultrasound scans of the distal common carotid arteries [D-CCA] (3 cardiac cycles, 14 MHz linear probe, mean 78.7 [Standard deviation (SD) 8.9]) frames per second were performed twice (2–4 days apart) on 10 healthy volunteers to test repeatability. Differences in CAS between healthy (n=20) and diabetic subjects (n=21) were examined. Peak CAS was measured in each of 6 equal segments and averages of all segments (i.e., global average), of the 3 nearest the probe, and of the 3 farthest from the probe (i.e., far wall average) were obtained.

Results—Global CAS (intraclass correlation coefficient [ICC]=0.40) and far wall average (ICC=0.63) had the greatest test-retest reliability. The global and far wall averaged CAS were lower in diabetics (4.29% [Standard Error (SE) 0.27%]; 4.30% [SE 0.44%], respectively) than in controls (5.48% [SE 0.29%], p=0.001; 5.58% [SE 0.44%], p=0.003, respectively). This difference persisted after adjustment for age, gender, race, and hemodynamic parameters.

Conclusions—Speckle-tracking for measuring carotid arterial wall strain is feasible and modestly reliable. Diabetic subjects had a lower carotid arterial wall strain obtained with speckle-tracking when compared with healthy controls.

Corresponding Author: Vijay Nambi, MD, 6565 Fannin St., STE B160 / MS-A601, Houston, TX 77030, Business Phone:(713) 798-7545, Fax: (713) 798-4210, vnambi@bcm.edu. ¹These authors contributed equally to the publication.

Introduction

Arterial stiffness, a mechanical property of the arterial wall, has been shown to be associated with cardiovascular risk factors and cardiovascular events. (1–7). Local arterial stiffness can be estimated using calculations based on changes in arterial diameter i.e. arterial distension during the cardiac cycle (4).

Echo-tracking devices have emerged as the most popular method for measuring vessel distension because of their higher resolutions (on the order of 0.001 mm) (8). However, echo-tracking is sensitive to tissue motion in directions other than that of the ultrasound insonation and to interference from other reflectors (e.g., speckles) within the arterial wall (9). Furthermore, the reproducibility of echo-tracking has been shown to be only similar to that of B-mode ultrasound derived changes in diameter (9).

Advances in echocardiography now permit quantification of myocardial stiffness via myocardial strain measures based on speckle-tracking (10). Strain is a percent or fractional measure of the spatial deformation of an object relative to its original size and has been validated in animal models as a surrogate measure of myocardial stiffness (10, 11). Speckle tracking is angle independent, has been shown to reduce error in myocardial strain measurements over tissue Doppler imaging, and hence may offer significant advantages (12, 13). We aimed to adapt the myocardial speckle-tracking technology and evaluate its feasibility for measuring carotid arterial strain.

We hypothesized that we could: a) measure carotid arterial strain reproducibly using the speckle-tracking technique; and b), that such a technique would detect differences associated with diabetes, a cardiovascular risk factor known to be associated with increased arterial stiffness. We chose to examine circumferential strain as it more closely approximates changes in luminal dimensions than radial strain, which represents change in wall thickness (14).

Methods

Study Population

Diabetic subjects (n=21) were recruited from individuals presenting to the non-invasive cardiac laboratory at the Ben Taub General Hospital (Houston, TX) for routine transthoracic echocardiography. A diagnosis of diabetes was confirmed using criteria from the American Diabetes Association 2010 Guidelines or active use of insulin and/or oral hypoglycemic therapies with a historical diagnosis (15). Healthy individuals without any previously known cardiovascular risk factors (n=20) were also recruited as a control comparison group (Table 1). All volunteers were recruited on presentation to the non-invasive cardiac laboratory, and thus were not asked specifically to refrain from food, caffeine, alcohol, tobacco, or vasoactive medications prior to the study. This scenario was considered to represent conditions encountered in a clinical setting.

Exclusion criteria included age <18 years, history of carotid stenting or carotid endarterectomy, history of radiation therapy to the neck, or a life expectancy less than 6 months. The study was approved by the institutional review board of the Baylor College of Medicine and the Harris County Hospital District. Written consent was obtained prior to study entry. All coronary heart disease risk factors as defined by the Adult Treatment Panel III guidelines (16) were noted for each subject.

Acquisition of Ultrasound Images

All subjects were interviewed and scanned by the same study investigator. The subjects were positioned supine with elevation of the head of the bed at a 30° incline. Electrocardiography (ECG)-gated B-mode bilateral carotid artery ultrasound scans were obtained (carotid artery pre-sets, Vivid 7 echocardiography machine; General Electric Healthcare, Waukesha, WI) over 3 cardiac cycles at 14 MHz with an M12L vascular transducer. Images were acquired at a mean 78.7 frames per second (fps) (SD 8.9 fps) and at a dynamic range of 72 dB. The gain was set to just below the presence of echoes in blood. The probe marker was always oriented to the left of the patient, regardless of the side imaged.

The probe was placed anteriorly over the carotid artery, and image sweeps from the proximal common carotid artery (CCA) to the carotid bifurcation were used to survey the CCA anatomy in transverse (cross-section). The transverse distal CCA, approximately one centimeter inferior to the carotid bulb, was used for our analysis. Subjects were instructed to perform a breath hold maneuver and to refrain from swallowing during the acquisition. The presence of significant motion artifact was noted on each scan, since motion would interfere with speckle-tracking and thus measured strain values. If motion artifacts occurred in a scan, image acquisition was repeated for that view when possible. Images were exported in Digital Imaging and Communications in Medicine (DICOM) format for offline analysis with an anonymization code unique to each subject.

Speckle-tracking Strain Analysis

Ultrasound images were uploaded to a computer workstation with a software package designed to measure myocardial strain using speckle-tracking algorithms (2D Cardiac Performance Analysis, TomTec, Munich, Germany). TomTec provided the analysis software but otherwise had no input into the study design, data accrual, analysis, or manuscript preparation. On images of the transverse distal CCA, the image frame at the end of electrical diastole (start of QRS wave) was identified, and the vessel on this frame was manually marked with a contour at the lumen-intima boundary (Figure 1A). The analysis tool was then run to obtain strain data.

The speckle-tracking algorithm divided a manually marked contour into 48 equidistant points and tracked the vessel intimal wall using a cross-correlation velocity vector imaging algorithm, which was accelerated with a Fast Fourier transform function and optimized for myocardial strain analysis (17). In principle, the algorithm would enable tracking of the smallest strain corresponding to the motion of a fraction of the US wavelength (on the order of microns) (18). Strain was calculated with reference to the end of electrical diastole of each cardiac cycle (Figure 1B). The raw data were exported to Excel (Microsoft, Redmond, WA) spreadsheets for further processing.

Post-processing of Speckle-tracking Strain Analysis

Since a myocardial analysis tool was used, the six segment convention of the myocardial short-axis views was adopted for identification of vessel wall segments (19). Anterior regional segments were identified as near wall segments; inferior regional segments, as far wall segments. The septal segment was identified as medial for the right CCA and lateral for the left, since the probe marker always pointed to the left, and similarly the lateral segment was identified as lateral for the right CCA but medial for the left. The anterior segment and the inferior segment themselves were identified as mid segments (i.e., near wall, mid; far wall, mid, respectively).

Superficial probe pressure may affect vessel wall segments near the probe. We observed medial-lateral echo dropout in our images (Supplemental Figure 1), and the reliability of strain measurements has been shown in phantom models to be lower in medial-lateral segments, likely due to heterogeneity in the density of ultrasonic lines/ echo dropout that can lead to unreliable strain measurements when the US beam is not aligned with direction of strain (20, 21). Thus, machine settings were optimized to achieve frame rates not too low to result in errors in lateral displacement and not too high to result in a decreased number of ultrasound scan lines. Further, we opted, *a priori*, to use the far wall as the primary segment for analysis. We also examined the global net average of all segments for comparison with traditional luminal measures. We later tested inter-visit repeatability and intra- and inter-visit reproducibility of the strain measures in each segment and of their averages.

Distensibility Measurement

For comparison of speckle-tracking derived strain with US based distensibility measurements, we measured the minimum and maximum of mean arterial diameters along 1 cm of the distal CCA on longitudinal views with an analysis system (Carotid Analyzer, Medical Imaging applications, LLC, Coralville, IA), which has been shown to have a mean difference of 0.02 pixel (95% confidence interval –2.98 to 3.02 pixels) between repeat measurements (22). Luminal strain was calculated by taking the difference between the two and dividing the minimum of mean arterial diameters. This method differs from previous methods that have used M-mode assessments of the carotid artery or aorta, by assessing diameters over a length of the CCA instead of a single beam axis.

Statistical Methods

Statistical analyses were performed using STATA 11 (STATA, College Station, TX). The Shapiro-Wilk test was used to determine non-normality of the data. Appropriate parametric and non-parametric tests (e.g., two-tailed two sample t-tests and Wilcoxon rank-sum tests) were employed to compare the characteristics and strain measures between the groups of diabetic and healthy controls. Values for each side were examined separately and as an average. Regression models were then used to adjust for covariates in the side-averaged strain comparisons (i.e., mean of left and right measures) with normalization of the dependent and predictor variables as needed. Two models were examined adjusting for participant characteristics and hemodynamic parameters since these may account for variations in carotid arterial strain, and both models included age, race, gender, heart rate, current smoking, and past history of smoking. The first model added systolic and diastolic blood pressures as covariates, and the second model added pulse pressure. These analyses were conducted with and without exclusion of image scans having motion artifacts.

Evaluation of Reproducibility

A separate set of healthy volunteers (n=10; mean age 34.1 [SE 3.2] years, 5 males, 5 females; 20 carotid arteries total) underwent US imaging as described in the above sections to allow testing for reproducibility. After a baseline scan, all the volunteers returned 2–4 days later for repeat imaging at approximately the same time of the day as the baseline scan. All carotid artery wall segments (e.g., lateral, mid, and medial segments of the near and far wall regions) were analyzed. One reader conducted the speckle-tracking strain analysis on the images acquired at both visits for inter-visit reproducibility analyses and repeated the analysis for the first visit for the intra-observer repeatability analyses. Another reader

analyzed the same set of images for the inter-observer reproducibility analyses. Intraclass correlation coefficients (ICC[2,1]) and coefficient of variations were used to assess intervisit repeatability and intra- and inter-reader reproducibility (23). Bland-Altman plots were also used to assess for bias (24).

Results

Reproducibility Analyses

Given the potential influence of heart rate and pulse pressure on strain values, we tested repeatability with and without consideration of these hemodynamic parameters. When segmental carotid arterial strain (CAS) values were examined for significant inter-visit differences, all wall segmental CAS and their averages showed no significant difference between visits (p > 0.05). This trend persisted after accounting for heart rate and pulse pressure (Supplemental Table 1).

For the primary repeatability study, test-retest analysis showed the greatest correlation for the far wall segments (ICC=0.63) and the global net averaged strain of all segments (intraclass correlation coefficient [ICC]=0.40) (Figure 2). The inter-visit coefficients of variation (CV) for the averages of the far wall segments and of all segments were 27% and 26%, respectively. When the strain to heart rate-pulse pressure product (i.e. heart rate times pulse-pressure) ratio was used, test-retest analysis showed the highest correlation again for far wall average (ICC=0.59) (Supplemental Table 2). These observations supported our *a priori* decision to examine the average strain of the 3 far wall segments for our primary analysis.

Intra- and inter-reader reproducibility was assessed for each segment, with agreement given as intraclass correlation coefficients (ICC's). The highest agreements for within-reader (intra-reader) and between-reader (inter-reader) correlations were again observed for the far wall average and the net average of all arterial wall segments (Figure 2). The intra-reader CV for these averages were 15% and 12%, respectively; and the inter-reader CV, 14% and 10%, respectively. Bland-Altman plots showed no systematic bias for all reproducibility measures and reasonable intra-reader and inter-reader limits of agreements (Supplemental Figure 2, Supplemental Table 3).

Comparison of Diabetics and Healthy Controls

The mean age of all individuals (diabetics and healthy volunteers) was 56.8 years. As a whole, the group consisted of 49% males and 29% Caucasians; had mean systolic, diastolic, and pulse pressures of 123.3 (SE 2.7) mm Hg, 74.5 (SE 1.6) mm Hg, and 48.8 (SE 1.7) mm Hg, respectively; and had a mean heart rate of 70.6 (SE 1.9) bpm (Table 1). No significant differences between the two groups were noted in baseline characteristics, except that diabetics were more likely to be hypertensive, on antihypertensive medications, current smokers and to have higher heart rates than controls (Table 1). Of the 21 diabetics, 9 were on beta-blockers, 2 on non-dihydropyridine calcium channel blockers, and 1 on amiodarone. No controls were on medications that affect heart rate. Despite repeated acquisitions, motion artifact was still present in scans of the right carotid artery in two diabetic volunteers. When the two individuals with motion artifacts were excluded, the baseline differences between the two groups remained similar.

Some participants (diabetics n=7, controls n=2) had atherosclerotic plaques at or above the level of the carotid artery bulb, but none had plaques in the carotid artery segments analyzed. No subject had a history of aortic valve disease, which may influence arterial hemodynamic measurements and potentially strain measurements, or any irregular heart rhythm such as atrial fibrillation.

Overall, side-averaged CAS values (i.e., individual mean of left and right measures together) were normally distributed among healthy controls (p > 0.05) but positively skewed among diabetics (p < 0.01) for all segments and their averages. Therefore, all comparison tests were non-parametric. CAS values were lower in diabetics than in controls for the far wall average (4.30% [SE 0.44%] v. 5.58% [SE 0.29%], p=0.003) and global net average (4.29% [SE 0.29%] v. 5.48% [SE 0.30%], p=0.001) (Figure 3). When each far wall segment was examined, CAS values continued to be significantly lower in diabetics than in controls for the far wall medial segment (p=0.18) (Figure 4). When scans with motion artifact were excluded, side-averaged CAS values continued to remain lower in diabetics (n=19) than in controls (n=20) for the global net average (3.95% [SE 0.18%] v. 5.48% [SE 0.30%], p=0.0002).

For the entire group (diabetics and healthy controls), the carotid arterial strain (CAS) values on the left were much lower than on the right for the global net average (p=0.003), far wall average (p=0.0007), and the far wall segments except for the far wall medial segment, which had no difference (p<0.0001 for far wall mid, p=0.005 for the far wall lateral, p=0.33 for the far wall medial) (Figure 3, Supplemental Table 4). After exclusion for the two individuals with motion artifacts, the lower left-sided strain values persisted.

There was no significant difference for CAS values derived from measures of luminal dimensions between diabetics (mean CAS of left and right sides 8.00% [SE 0.55%]) and healthy controls (mean CAS of left and right sides 7.14% [SE 0.51%]; p=0.32), even after exclusion of the 2 participants with motion artifact (diabetic CAS 8.25% [SE 0.58%] v. control CAS 7.14% [SE 0.51%], p=0.16).

Regression analyses were performed on log-transformed values of CAS to normalize the variables. On univariate analysis, diabetic status, current smoking status, heart rate were significant predictors of lower far wall average and global net average strain values; age and blood pressure measurements, notably pulse pressures (Supplemental Figure 3), were not (Supplemental Table 5). After exclusion of scans with motion artifact, this relationship between CAS and predictors persisted (Supplemental Table 5). When the comparisons were adjusted for covariates (Model 1 including age, gender, race, heart rate, current and past smoking, plus systolic and diastolic blood pressures; Model 2 including age, gender, race, heart rate, current and past smoking, plus pulse pressure), diabetic status was no longer significantly associated with speckle-tracking derived CAS values for both models. However, when the two participants with motion artifact were excluded, diabetic status became significantly associated with the global net average strain (p=0.01 for both models) and the far wall average strain (p=0.01 for the first model, p=0.02 for the second model) (Table 2). Overall, these results suggest that diabetes significantly and adversely impacts circumferential carotid arterial strain even after adjustments for covariates.

Discussion

In summary, we showed that measuring carotid arterial strain (CAS) was feasible and modestly reliable using a novel speckle-tracking based technique and that CAS values were lower in diabetics compared with non-diabetics. Furthermore, we found inter-visit, intrareader, and inter-reader agreements were consistently higher for the mean CAS of all segments and the far wall segments (Figure 2). The inter-visit ICC suggested moderate reliability based on criteria set forth by Fleiss (0.40 to <0.60 for moderate ICC) whereas the intra-/inter-reader ICC's were excellent (ICC \geq 0.75) (25). CAS values were lower in diabetics versus non-diabetics, and this difference persisted even after adjustments for covariates and appropriate exclusions (Table 2).

Measurements of arterial wall characteristics have become increasingly recognized as possible cardiovascular risk markers in large epidemiologic studies. The elastic property of large arterial vessels depend greatly on the elastin content (organized as elastic lamina) of the arterial media, and to a lesser extent on the collagen content of the vessel (26–28). Both components in turn are produced by the smooth muscle cell population of the same layer (i.e., arterial media) (26–28).

As large arteries age, smooth muscle cells degenerate and decrease in number through apoptosis, and the elastin lamina also degenerate and become fragmented (28). In contrast, the more rigid collagen component has been shown to proportionally increase (28). These processes lead to increased arterial stiffness, a process best labeled with the term arteriosclerosis (27, 28). These observations have been borne out in clinical studies, which have found increased pulse pressures, a marker of arterial stiffness, to increase with age (27, 29). Certain pathologic states (e.g., hypertension through medial hypertrophy; atherosclerosis through foam cell infiltration and plaque deposition) have also been recognized to contribute and potentially to accelerate arterial stiffening (2–5, 27, 28, 30, 31). Similarly, non-insulin dependent diabetes has been postulated to increase proliferation of smooth muscle cells through elevated insulin levels and to lead to advanced glycosylation end-products (AGEs) involving the collagen and elastin of the medial layer through hyperglycemia (32). Therefore, several of the cardiovascular risk factors seem to be associated with arterial stiffness.

Regional stiffness measured using pulse wave velocity (tonometry) has been shown to be associated with incident coronary heart disease (7). Although local carotid artery stiffness measures have shown strong associations with cardiovascular risk factors, their associations with incident cardiovascular events have been weaker/ non-significant (33, 34). Recent results from the Atherosclerosis Risk in Communities Study support a significant association between ultrasound-based measures of arterial stiffness and incident stroke events, independent of established atherosclerotic risk factors (35). Thus, local measures of arterial stiffness may also hold potential as cardiovascular risk markers.

Local arterial stiffness can be estimated non-invasively through the measurement of arterial distension using imaging (e.g., magnetic resonance imaging, ultrasound imaging) (4). Imaging-based distension measurements rely on the detection of changes in arterial dimensions over the cardiac cycle, which can be obtained from lumen-wall border tracking (i.e., lumen size) or from direct wall tracking (4, 36, 37). For ultrasound, direct vessel wall tracking has been accomplished with echo-tracking and more recently with tissue Doppler imaging (TDI) and with speckle-tracking (8, 38–40).

Echo-tracking has been well studied and validated for measuring arterial distension but has its limitations (4, 8). Although Hoeks et al, found the echo-tracking resolution to be less than 0.001 mm *in vitro*, they also noted the standard deviation of distension measured in a human test subject to be 0.043 mm (8). They attributed this finding to arterial motion and physiological variations, which may be evidence of a threshold beyond which higher resolutions do not improve distension measurements (8). Furthermore, the technology is fundamentally based on M-mode, or single-beam axes, limiting its ability to assess motion assessment in other axial directions (8). Recently, Kawaski et al, used TDI to measure strain rates on longitudinal views of the common carotid artery in patients with coronary heart disease (38). However, the TDI method is limited by angle dependence (38).

We sought to evaluate the application of speckle-tracking for direct assessment of carotid arterial wall strain and are among the first groups to do so (40, 41). Speckle-tracking is a 2-dimensional angle-independent method of tissue-tracking that is used for assessing

Bjallmark et al, employed speckle-tracking analysis of carotid strain in a population of younger (n=10) and older (n=10) individuals and showed increasing age to be associated with lower strain values (39), which our models support. However, Bjallmark et al, acquired images for only the right common carotid artery, did not report on the inter-visit repeatability of their measurements, and examined a smaller population sample. Our work differs by demonstrating the reliability and repeatability of speckle-tracking for measuring CAS in arterial wall segments and furthermore examines CAS in a population known to have stiffer arteries, i.e. diabetics.

Furthermore, we examined CAS measures using a lumen based assessment of distension and found no significant differences between the two groups suggesting that speckle tracking may be more sensitive. Although our lumen-based distension measure was not obtained with echo-tracking, studies by others have shown the reliability of distension measured using echo-tracking and of that using B-mode ultrasound imaging to be comparable (9, 37). Furthermore, our lumen based distensibility measurement evaluates a larger segment of the artery compared with echo tracking.

Finally, we found arterial strain measurements to be significantly higher in the right than in the left carotid artery for the two of the three far wall segments, the average of all three far wall segments, and the net average of all arterial wall segments. Differences in right versus left measurements have been reported in common carotid intima-media thickness measurements, with the right having lesser values than the left (43–45). In fact, in one study of participants with and without migraine headaches, arterial distension was also found to be significantly greater in the right than in the left for both groups of individuals. These findings are consistent with the findings in our study. These observations suggest that there may be differences in blood pressure, shear forces, and vascular anatomy specific in the same individual between the right and the left carotid arteries and merits further investigation.

Clinical perspective

Ultrasound imaging offers a non-invasive and safe method of evaluating the arterial wall. Carotid artery measures of atherosclerosis (namely carotid intima-media thickness and plaque presence) has already been shown to be useful in coronary heart disease risk prediction and as a surrogate marker to test efficacy of anti-atherosclerotic therapy (46, 47). Ultrasound based stiffness measures provide potentially yet another measure to improve our assessment of vascular health. Pulse pressure is often considered a good marker of arterial stiffness (48); and in general, when the arterial stiffness increases, in order to maintain a similar arterial distensibility (strain), the pulse pressure should also proportionally increase.

However, as noted in our analysis, there is a very poor correlation between pulse pressure and CAS as measured by speckle-tracking. The poor correlation between these two measures of stiffness may be explained by the fact that CAS measures only local carotid stiffness while pulse pressure reflects the stiffness of the entire vascular tree. Furthermore, given that the association between CAS and diabetes persisted after adjustment for pulse pressure, one could hypothesize that a simple pulse pressure measurement may not be an adequate surrogate for CAS, and perhaps both provide complementary information on vascular health. Whether local arterial strain reflects the advanced sequelae of pathologic processes or remodeling due to various medical interventions remains a potential area for

further clinical investigation. Clearly, much further work will be required before this technology will have any use in routine clinical practice.

Limitations

Our study had limitations. Brachial blood pressures were used instead of local carotid pressures, both of which have been shown to differ from each other and to differ in local remodeling effects within an individual (49, 50). However, it must be noted that in individuals with cardiovascular risk factors such as diabetes, the peripheral pulse pressure tends to be similar to the central pulse pressure while in healthy individuals the central pulse pressure tends to be lower (51). Despite this (i.e. likely greater central pulse pressures which in turn could increase arterial distension/ strain) diabetics in our study had lower arterial strain, suggesting that the arterial strain we measured by speckle tracking also reflects a stiffer arterial wall. The inter-visit intraclass correlation coefficients were modest and may have been due to variations in time from last meal, use of vasoactive substances (e.g., caffeine or alcohol), and diurnal variations. Additionally we used a system intended for myocardial analysis and used DICOM images instead of raw US data. Dedicated strain platforms for arterial wall imaging may further improve the repeatability and reproducibility of arterial strain measurements. We also acquired at higher frame rates than those used for myocardial analyses; however, the effects of higher frame rates on reproducibility (by potentially affecting spatial resolution) of strain measurements are unclear. Strain measurements were not compared with phantom or animal models using sonomicrometry crystals. Still, we tested the technique with a common clinical disease entity known to lead to increased arterial strain to lay the groundwork for the potential clinical vascular applications of the speckle-tracking technique.

The diabetic sample size was small, and we did not examine the association of strain values with the severity or duration of diabetes. We were also unable to adjust for glomerular filtration rates, as serum creatinine levels were not available on all subjects. Acoustic shadowing of the medial-lateral wall segments was observed and may have limited the reliability of strain measures in these segments. We analyzed strain measurements in only the distal common carotid artery, and differences in these measures may be present along the different carotid artery segments. We examined the carotid artery for only circumferential strain; other strain measures including longitudinal and radial strain should also be examined. Lastly, we adapted an ultrasound system and analysis package originally optimized for cardiac use to vascular applications.

Future Directions

The results of this study point to several potential avenues of investigation. Different scanning approaches need to be investigated to see if probe pressure does indeed influence strain measurements. Differences in strain values at different levels of the carotid vasculature should also be examined. Additional data on differences between left versus right sides for not only distensibility measures but also carotid intima-media thickness measures are still needed. Optimization of speckle-tracking systems for use with vascular applications will also be required.

Conclusion

Measurement of carotid arterial wall strain using speckle-tracking is feasible and reliable in the wall segments farthest from the ultrasound probe. These strain measures were significantly lower in diabetics even after adjustment for age, gender, race, and hemodynamic parameters, suggesting that this technique may be more sensitive to changes from early atherosclerosis than previous techniques based on luminal measurements.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We thank Berthold Klas and TomTec, Munich, Germany, for supplying the speckle-tracking strain analysis software and associated documentation. We thank the participants of the study for their contribution and Joanna Brooks, B.A., for editorial assistance. Dr. Yang was supported by a National Institutes of Health (NIH) / National Heart, Lung, and Blood Institute (NHLBI) T32 HL007812 training grant and an American Heart Association South Central Affiliate Postdoctoral Fellowship grant at different times during this study. Dr. Virani is supported by a Veterans Affairs Career Development Award. Dr. Nambi is supported by a NIH/NHLBI K23 HL096893 grant.

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Figure 1. Measuring circumferential carotid arterial strain using speckle-tracking

Contour generated after manual marking of intima-luminal border of the right distal common carotid artery (A). Strain values depicted by software system over 3 cardiac cycles (B). The top graph shows the change in circumferential strain over time at each of the 48 points along a closed loop contour. The graph below it illustrates a color map representation, with each of the 48 points represented on the y-axis; time, on the x-axis; red, positive strain values; and blue, negative values.



Reproducibility of Strain Measurements with Speckle-Tracking by Transverse Carotid Wall Segments

Figure 2. The reproducibility of carotid arterial strain measures using speckle-tracking Test-retest of image acquisition, intra-reader repeatability, and inter-reader reproducibility by intraclass correlation coefficients (ICC) type (2,1) of carotid arterial strain measures by arterial wall segments on a separate set of volunteers (n=10, 20 carotid scans).

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Figure 3. Comparison of peak circumferential carotid arterial strain in controls vs. diabetics by net average and far wall average values

Strain values listed are means with standard error bars. Tests of comparison were performed using the Wilcoxon rank sum test. (L=Left, R=Right, # p < 0.05, * p < 0.001)

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Figure 4. Comparison of peak carotid arterial strain in controls vs. diabetics by medial, mid, and lateral far wall segments

Strain values listed are means with standard error bars. Tests of comparison were performed using the Wilcoxon rank sum test. (L=Left, R=Right, NS=non-significant, # p < 0.05, * p < 0.001)

Table 1

Clinical characteristics of volunteers for the diabetic vs. control carotid strain comparison*

Variable	Controls (n = 20)	Diabetics (n = 21)	p-value
Age (years)	56.6 (1.9)	57.0 (1.8)	0.88
Male (%)	50.0	47.6	0.88
Caucasian (%)	30.0	28.6	0.92
African-American (%)	30.0	33.3	0.82
Hispanic (%)	25.0	28.6	0.80
Asian (%)	15.0	9.5	0.59
Systolic BP (mm Hg)	120.8 (2.7)	125.7 (4.5)	0.64
Diastolic BP (mm Hg)	73.8 (1.6)	75.2 (2.6)	0.86
Pulse Pressure (mm Hg)	47.0 (1.5)	50.5 (2.9)	0.68
Heart Rate (bpm)	63.3 (2.3)	77.6 (2.3)	0.0001
Hypertensive (%)	0	76.2	< 0.0001
Antihypertensive Medication (%)	0	85.7	< 0.0001
Current Smoker (%)	0	23.8	0.02
Past Smoker (%)	30.0	23.8	0.65

* All values are means or proportions (SE). Proportions were compared between controls and diabetics using two-sided tests of proportions; continuous variables were compared between controls and diabetics using two-sided Student's t-test or Wilcoxon rank sum test as appropriate.

BP = blood pressure

Table 2

Linear regression of carotid arterial strain measurement for each wall segment, normalized by log transformation, regressed against diabetic status, adjusted for covariates.* Blood pressures were inverted to normalize the variables.

	Beta-coefficient (95% CI) p-value	Net Avg.	Far Wall Avg.	Lateral Far Wall	Mid Far Wall	Medial Far Wal
+	Presence of diabetes (without image exclusions)	-0.146 (-0.332, 0.041) p=0.12	-0.176 (-0.426, 0.073) p=0.16	-0.316 (-0.692, 0.060) p=0.10	-0.158 (-0.478, 0.162) p=0.32	-0.071 (-0.332, 0.190) p=0.58
Aodel 1	Presence of diabetes (with image exclusion)	-0.226 (-0.405, -0.047) p=0.02	-0.284 (-0.521, -0.048) p=0.02	-0.470 (-0.840, -0.101) p=0.01	-0.182 (-0.508, 0.144) p=0.26	-0.190 (-0.440, 0.059) p=0.13
+	Presence of diabetes (without image exclusions)	-0.132 (-0.321, 0.057) p=0.17	-0.151 (-0.407, 0.106) p=0.24	-0.281 (-0.660, 0.099) p=0.14	-0.143 (-0.462, 0.176) p=0.37	-0.041 (-0.319, 0.238) p=0.77
Aodel 24	Presence of diabetes (with image exclusion)	-0.228 (-0.407, -0.050) p=0.01	-0.283 (-0.522, -0.044) p=0.02	-0.463 ($-0.832, -0.095$) p=0.02	-0.189 (-0.511, 0.133) p=0.24	-0.188 (-0.451, 0.075) p=0.16

 † Model 1 covariates included the common covariates plus systolic and diastolic blood pressures.