
Locations and sequences of tobacco chloroplast genes for tRNA^{Pro}(UGG), tRNA^{Trp}, tRNA^{fMet} and tRNA^{Gly}(GCC): the tRNA^{Gly} contains only two base-pairs in the D stem

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ABSTRACT

The nucleotide sequences of tobacco chloroplast genes for tRNA^{Pro}(UGG), tRNA^{Trp}, tRNA^{fMet} and tRNA^{Gly}(GCC) have been determined. None of these genes contains an intron. One unusual feature is that the tRNA^{Gly} contains only two base-pairs (A-U, G-U) in the D stem. These four tRNA genes were located in the known physical map of tobacco chloroplast DNA. Hybridization analysis to chloroplast tRNA revealed that all four tRNA genes are transcribed in vivo.

INTRODUCTION

Chloroplasts contain their own tRNAs which are believed to be coded in their chloroplast genomes (1). Eleven, twelve, six, and sixteen tRNA genes have been sequenced so far in tobacco (2,3,4,5), maize (2,6), spinach (2) and Euglena (2,7,8) chloroplast DNA, respectively, and these show features of both prokaryotic and eukaryotic tRNA genes.

To analyze the organization and fine structures of tRNA genes of tobacco chloroplasts, we have determined nucleotide sequences of the chloroplast DNA fragments which hybridized to its chloroplast tRNAs (2,3,4). However, their locations on a physical map of tobacco chloroplast DNA have not been determined. In this report, we show the map locations and nucleotide sequences of tRNA^{Pro}(UGG), tRNA^{Trp}, tRNA^{fMet} and tRNA^{Gly}(GCC) genes of tobacco chloroplasts. The tRNA^{Gly}(GCC) deduced from the DNA sequence has only two base-pairs in the D stem.

MATERIALS AND METHODS

The recombinant plasmid pTS9, which contains a 5.7 kbp SalI fragment, and pTcS-2, which contains a 6.6 kbp EcoRI fragment, of Nicotiana tabacum (var. Bright Yellow 4) chloroplast DNA have been constructed as described (9). pTS9 DNA was digested with SalI and the 5.7 kbp fragment was separated by electrophoresis in a 5% polyacrylamide gel. pTcS-2 DNA was digested with

EcoRI and the 6.6 kbp fragment was separated as above. DNA sequence was determined by the method of Maxam and Gilbert (10). For Southern blot hybridization, DNA fragments were transferred to nylon membrane (Biodyne A Pall) and the membrane was immersed for 1hr at 65°C in the hybridization buffer (0.9 M NaCl, 50 mM sodium phosphate buffer (pH 7.7), 5 mM EDTA, 0.2% SDS, 0.1% Ficoll, 0.1% polyvinylpyrrolidone, 0.1% bovine serum albumin and 10 µg/ml denatured calf thymus DNA). Hybridization was carried out at 65°C for 16hr in the hybridization buffer containing [5' 32P]-labeled tobacco chloroplast tRNAs.

RESULTS AND DISCUSSION

tRNA^{Pro} and tRNA^{Trp} genes

The 5.7 kbp SaliI fragment (S9) of tobacco chloroplast DNA is located on a SaliI cleavage map of tobacco chloroplast DNA (11) as shown in Fig. 1. On digestion with BglII and MluI, the 5.7 kbp fragment yields 3.4, 1.1, 0.8 and 0.4 kbp sub-fragments (Fig. 2a). Only the 1.1 kbp BglII-MluI sub-fragment

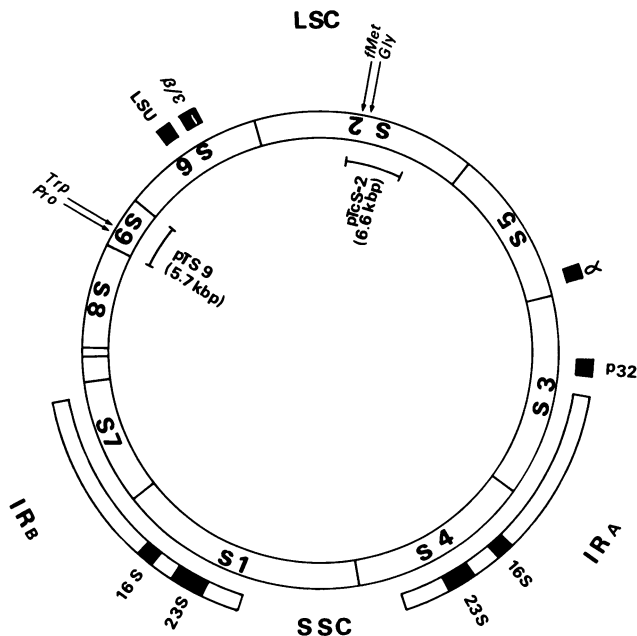


Fig. 1. Positions of the cloned fragments and the genes for tRNA^{Pro}, tRNA^{Trp}, tRNA^{fMet} and tRNA^{Gly} on the SaliI cleavage map of tobacco chloroplast DNA (11). Arrows, tRNA genes; IRA and IRB, inverted repeat sequence (5); LSC and SSC, large and small single copy regions.

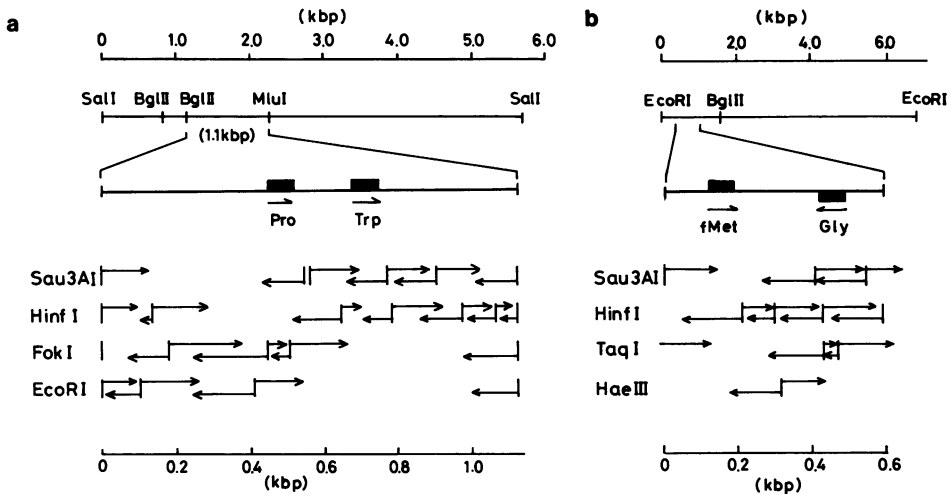


Fig. 2. Physical maps of the cloned 5.7 kbp SalI (a) and 6.6 kbp EcoRI (b) fragments from tobacco chloroplast DNA and the strategy for sequencing parts of them. Coding regions are shown by thick lines and arrows (\rightarrow) below them indicate the direction of transcription.

was hybridized with total tobacco chloroplast tRNA (data not shown). We then sequenced the 1.1 kbp sub-fragment by the sequence strategy shown in Fig. 2a. Figure 3a shows the nucleotide sequence of the 1131 bp BglII-MluI sub-fragment.

A tRNA^{Pro}(UGG) gene was found between positions 450 and 523 (Figs. 3a & 4a). The tRNA^{Pro}(UGG) deduced from the DNA sequence shows 93% sequence homology with spinach chloroplast tRNA^{Pro}(UGG) (12). *E. coli* tRNA^{Pro} sequence has not been reported (2,14). A tRNA^{Trp} gene (688 to 761) was found 165 bp downstream from the tRNA^{Pro} gene in the same orientation (Figs. 3a & 4b). The tRNA^{Trp} deduced from the DNA sequence shows 96%, 87% and 67% sequence homologies with spinach chloroplast (13), *Euglena* chloroplast (7) and *E. coli* (14) tRNA^{Trp}, respectively.

A "Pribnow box"-like sequence TATAAT (289 to 294) and a "-35 region"-like sequence TTGTAT (269 to 264) were found in the region upstream from the tRNA^{Pro} gene (Fig. 3a). Likewise in the region upstream from the tRNA^{Trp} gene a "Pribnow box"-like sequence TATCAT (598 to 603) and a "-35 region"-like sequences TTGGACA (577 to 583) were found (Fig. 3a). These tRNA genes may be transcribed separately. Inverted repeat sequences (427 to 448, 524 to 554, 666 to 685, and 762 to 795) were found in both flanking

a) Bg1 II
 ↓
 100 GATCTCGCAT TGA AAAACCT COTTCCTTTG TATTATTGT TGG AATAGAT AATACATATA TAATCAGATG CATATGTAGT TAAGGGCTGC TTAGAGTTGG
 200 ACACAGAATT CCTAATTC AA ATTGC AATGT ACAATGATTC GGG AATAAG AHTTTCCTTT TCTTAAAACA GAAAAAAA ATACATCCCC TTCTTTCTTA
 300 CC GAGAATTT CCGAAAAATA CTAATTGATT TTTACGACGG GTTCTGTATT TATATAATTT GTATATAATT GTATATAAGT ATTTTCTATA TAAATCTATA
 400 GAGAACTTT TTCCTTTTAT TTTAACTATT ATATGTTAAA TGAGACCAA AGACGAAAT GAGCAGAAA TTTGTATATA TCAATGGAGG AANTAACCAT
 500 GTGGAAAACA AGACAGGAAT TCTCTACAAT TACACTGTGG AACAAATTGA GGGATGTGGC GCAGCTTGGT AGCGCGTTTG TTTTGGGTAC AAAATGTCAC
 600 AGGTTCAAAAT CCTGTCAATCC CTA CCTATT TA CTGCTCAAAG GAGCAGTAA C GAGGGATCAA TTGAGATCC CTC A AATGG ACATAA TCTT TGATTTTAT
 700 CATGCTATTC TAGTATATCC ATACAAAATG TATTGGGGTT ATGAAAAGAA TCCTTTTGT TACAGCTTTA TCTATTCTGA TAAGAAA ECG CTCTTAGTTC
 800 AGTTCGGTAG AACGTGGTTC TCCAAAACC GATGTCGTAG GTTCAAATCC TACAGAGCGT GATTTTTTC TTCTTAGATC GAATTAGAAT AAAATAGATT
 900 CAATTCAGTAA CTTTCACAA C AATCGGAATT TGACCTCCTG TGCCGTGTGAA TTAAGAAAG GAGGAGTCA ATCAAAAAGA GATGTTAANT AATCAAAGGT
 1000 CCAACTGATC ACCACGCCTA TATTGTAAAT ATGCAGTTAC GAATAATCCA GCCAAAAGTAA TAGGAATTAG ACCTAACACG ATTCCAAATA GAAAAACTTC
 1100 AATCAATTTT ATTTGTTTGA AAGGAGAAA AAAGAGGTAA TATCGATCC TAAATATTA TCCGAAATTA CAGTAATCTC AATGACAATG AACAAGAATG
 GACAATGGT AACGTAAGTA AATATAGAAT A
 Mlu I ↓

b) Sau 3AI
 ↓
 100 GATCATAGAA GCCCCTTTAC CATTCTGTAT AAATGGGCTA TTCTATTTCG ACAGATAGGG TGGAGGGGG CATTTAAATCC TTGTTTATCT ATTAGTTTTC
 TATCTT CGGGAAATG GTAAGACATA TTTACCCGAT AAGATAACA TGTCTATCCC ACCTCCCAGC GTAAATFAGG AACAAATAGA TAATCAAAAG

200 AGTTCCTATC TTCGGCCGG GGTAGAGCAG TTTGGTAGCT CGCAAGGCTC ATAACTTGA GFTCACGGGT TCAATFCCIG TCTCCGCAC ATCTTGTFTT
 TCAAGAATAG AAGCCCGGCC CCACTCGTC AAACCATCGA CGTTCGGAG TATTGGAAC TCCAGTGCCA AGTTTAVGAC AGAGGCGTTG TAGAACAAA

300 GCCAAACTAT TTTAGGTTG ACTCTGTTAA CTAGTAATTA ATTCCCCTT TTCGCTTTT GGGGTGGAA GGA~~AAA~~AGAA AACGTAGGG AGGATAGAA
 CGGTTTGATA AAATCCCAAC TGAGACAATT GATCATTAT TAAGGGCGGA AAGCGAAAA CCCCCACCTT CCTTTTCTT TTGCATCCCC TCCCTATCTT

400 TCACTACACT ATCACGGCCA ACTATACCAA ATCCTTAATT TAAGGATATA TTTAATGCTA TTTATGAAAT TAAATAATA ATAAATFAGTA ATAAATTTAC
 AGTGATGTA TAGTCCCGGT TGATATGGTT TAGGAATTA ATTCCATPAT AAATFACGAT AAATACTTTA ATTTATTATT TATTTATCAT TATTTAATG

500 TTTATCTGG ATCTTGGCG GATAGCGGA ATCGAACCCG CATCTTCTCC TTGGCAAGA GAAATTTAC CATTCGACCA TATCCGCATT TTTTGTGTCT
AAATAGAACC TAGACECGC CTAFCGCCCT TAGCTTGGC GTAGAGAGG AACCTTTCT CTTAAATG GTAAGCTGT ATAGCGTAA AAAACAAGA

TGATACAAA TATGTACCCA CATATATGAT ATATAACCG ATCTTTTGG TGCAGTGCCG GGACACATAT TCTCTTCGA ACG↓
 ACTATGTGT ATACATGGGT GTATATACTA TATATTTGCC TAGAAAAAC ACFTCACGGC CCTGTGTATA AGAGAAGCCT TGCTAAG

Hinf I
 tRNA^{Gly}

Fig. 3. Nucleotide sequences of the regions containing tRNA^{Pro} (UGG) and tRNA^{Trp} genes (a), and tRNA^{fMet} and tRNA^{Gly} (GCC) genes (b). Coding regions are boxed. "Pribnow box"-like sequences and "-35 region"-like sequences are underlined. Horizontal arrows indicate inverted repeat sequences.

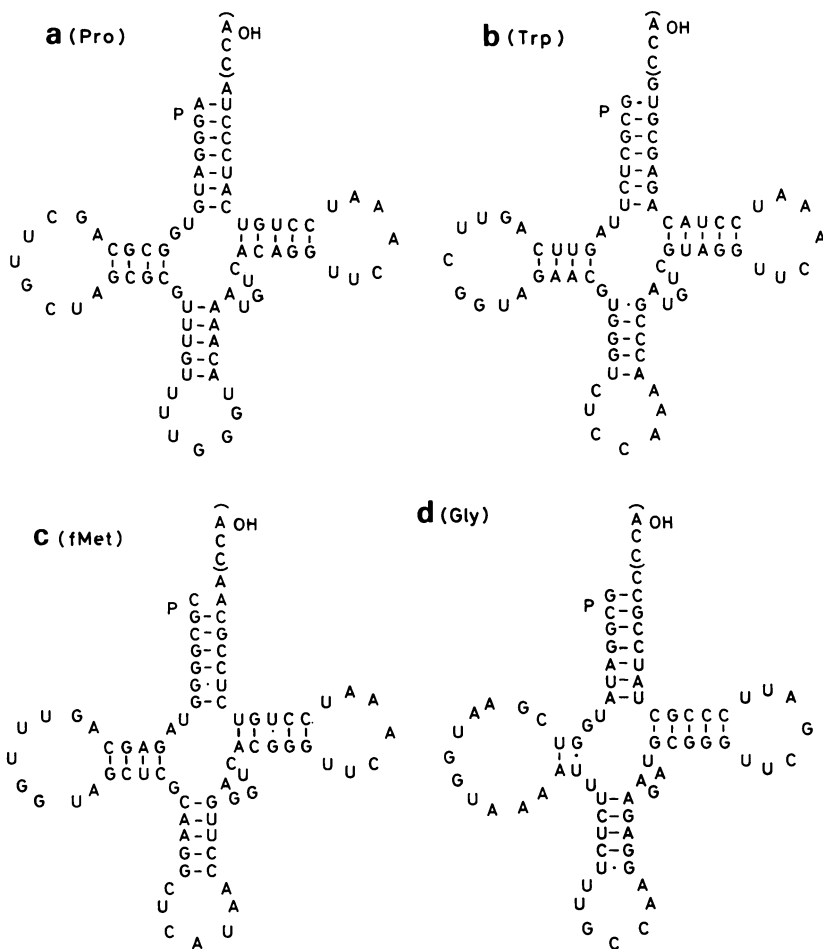


Fig. 4. Sequences of unmodified bases and cloverleaf structures of tobacco chloroplast tRNA^{Pro}(UUG) (a) tRNA^{Trp} (b), tRNA^{fMet} (c) and tRNA^{Gly}(GCC) (d) predicted from the DNA sequences.

regions of the tRNA^{Pro} and tRNA^{Trp} genes (Fig. 3a). These sequences may be related to tRNA processing and transcriptional termination.

tRNA^{fMet} and tRNA^{Gly} genes

The 6.6 kbp EcoRI fragment of tobacco chloroplast DNA is located in the SalI fragment 2 (S2) on the tobacco chloroplast DNA map (11) shown in Fig. 1. On digestion with BglII, the 6.6 kbp EcoRI fragment yields 5.0 and 1.6 kbp sub-fragments (Fig. 2b). The 1.6 kbp EcoRI-BglII sub-fragment was hybridized with total tobacco chloroplast tRNA (data not shown). We then

sequenced part of the 1.6 kbp sub-fragment by the sequence strategy shown in Fig. 2b. Figure 3b shows the nucleotide sequence of a 583 bp portion of it.

A tRNA^{Met} gene was found between positions 116 and 189 (Figs. 3b & 4c). It is likely to be an initiator tRNA^{Met} gene because its sequence is identical with that of spinach chloroplast initiator tRNA^{Met} (15). The tRNA^{fMet} deduced from the DNA sequence shows 84% and 83% sequence homologies with *Euglena* chloroplast (8) and *E. coli* (14) tRNA^{fMet}, respectively, but only 47% homology with the tobacco chloroplast elongator tRNA^{Met} (3). A tRNA^{Gly}(GCC) gene (417 to 487) was found 228 bp downstream from the tRNA^{fMet} gene in the opposite orientation (Figs. 3b & 4d). The tRNA^{Gly}(GCC) deduced from the DNA sequence shows 63%, 61% and 73% sequence homologies with *Euglena* chloroplast (8), *E. coli* (14) tRNA^{Gly}(GCC) and tobacco chloroplast tRNA^{Gly}(UCC) (4), respectively. The 3'-terminal CCA sequence is not found in any of the tRNA genes presented here.

A "Pribnow box"-like sequence TTAAAT (73 to 78) and a "-35 region"-like sequence TTGTACA (47 to 53) were found in the region upstream from the tRNA^{fMet} gene (Fig.3b). Likewise the region upstream from the tRNA^{Gly} gene contains a "Pribnow box"-like sequence TATCAT (531 to 526) and a "-35 region"-like sequence TGCACA (554 to 549) (Fig. 3b). Inverted repeat sequences (196 to 207, 255 to 278, and 387 to 401) were found in the regions downstream from the tRNA^{fMet} and tRNA^{Gly} genes. These sequences may be tRNA processing or transcriptional termination signals.

One unusual feature is that the tRNA^{Gly} deduced from the DNA sequence has only one Watson-Crick base-pair (A-U) and an additional G-U pair in the D stem (Fig. 4d). Transfer RNAs with such base-pairs in the D stem have not been reported, except for *Xenopus* mitochondrial tRNA^{Asn} and mammal mitochondrial tRNA^{Ser} which have no D stems (2,14). Wheat germ, rat and mouse tRNAs^{Ser} contain one A-U pair and two additional G-U pairs in their D stems and a calf tRNA^{Tyr} has two C-G pairs only in its D stem (2,14). The tobacco chloroplast tRNA^{Gly}(UCC) has been reported to contain a 691 bp intron in the D stem (4). Therefore the tobacco chloroplast tRNA^{Gly} species are quite unique in their D stems.

Based on fine physical mapping of the 5.7 kbp SalI and 6.6 kbp EcoRI fragments and their neighboring regions (data not shown), the tRNA^{Pro}, tRNA^{Trp} and tRNA^{fMet} are coded for on the same strand as the α and β genes for H⁺-ATPase (16, strand B) and the tRNA^{Gly} gene on the same strand as the LS gene for ribulose-1,5-bisphosphate carboxylase (17, strand A) of the

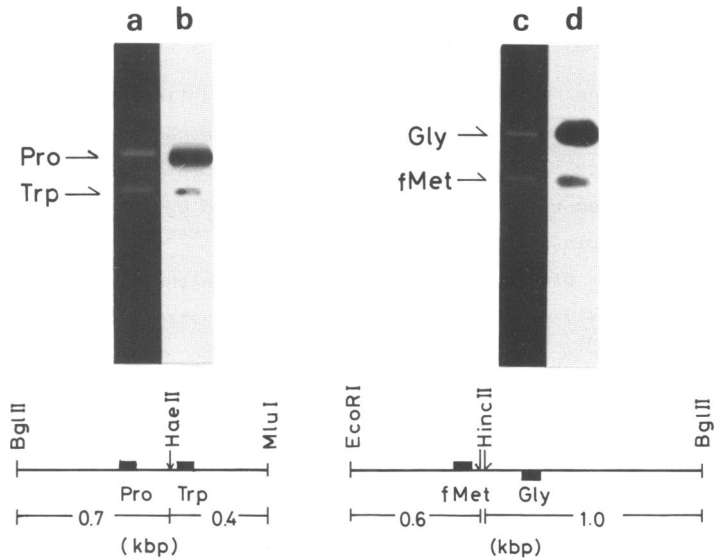


Fig. 5. 2% agarose gel electrophoresis of restriction fragments and autoradiographs of Southern blots hybridized with total tobacco chloroplast tRNAs. HaeII digest of the 1.1 kbp BglII-MluI sub-fragment from the 5.7 kbp SalI fragment (a). HincII digest the 1.6 kbp EcoRI-BglII sub-fragment from the 6.6 kbp EcoRI fragment (c). Autoradiographs (b,d).

tobacco chloroplast DNA (Fig. 1). The locations of these four tRNA genes are consistent with the tRNA gene map obtained by the tRNA/DNA hybridization studies (18).

Expression of tRNA genes

To examine whether these four tRNA genes are expressed, Southern hybridization was carried out. The 1.1 kbp BglII-MluI sub-fragment of the 5.7 kbp SalI fragment was cut with HaeII into two fragments, one containing the tRNA^{Pro} gene and the other containing the tRNA^{Trp} gene. The 1.6 kbp EcoRI-BglII sub-fragment of the 6.6 kbp EcoRI fragment was cut with HincII into two main fragments, one containing the tRNA^{fMet} gene and the other containing the tRNA^{Gly} gene. Total tobacco chloroplast tRNAs hybridized to these four fragments (Fig. 5), indicating that these tRNA genes are expressed in vivo.

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REFERENCES

1. Driesel, A.J., Crouse, E.J., Gordon, K., Bohnert, H.J., Herrmann, R.G., Steinmetz, A., Mubumbila, M., Keller, M., Burkard, G. and Weil, J.H. (1979) *Gene* 6, 285-306.
2. Sprinzl, M. and Gauss, D.H. (1984) *Nucleic Acids Res.* 12, r59-r132.
3. Deno, H., Kato, A., Shinozaki, K. and Sugiura, M. (1982) *Nucleic Acids Res.* 10, 7511-7520.
4. Deno, H. and Sugiura, M. (1984) *Proc. Natl. Acad. Sci. USA* 81, 405-408.
5. Sugita, M., Kato, A., Shimada, H. and Sugiura, M. (1984) *Mol. Gen. Genet.* 194, 200-205.
6. Krebbers, E., Steinmetz, A. and Bogorad, L. (1984) *Plant Mol. Biol.* 3, 13-20.
7. Hollingsworth, M. and Hallick, R.B. (1982) *J. Biol. Chem.* 257, 12795-12799.
8. Karabin, G.D. and Hallick, R.B. (1983) *J. Biol. Chem.* 258, 5512-5518.
9. Sugiura, M. and Kusuda, J. (1979) *Mol. Gen. Genet.* 172, 137-141.
10. Maxam, A.M. and Gilbert, W. (1977) *Proc. Natl. Acad. Sci. USA* 74, 560-564.
11. Seyer, P., Kowallik, K.V. and Herrmann, R.G. (1981) *Current Genetics* 3, 189-204.
12. Francis, M., Kashdan, M.A., Sprouse, H., Otis, L. and Dudock, B. (1981) *Nucleic Acids Res.* 10, 2755-2758.
13. Canaday, J., Guillemaut, P., Gloeckler, R. and Weil, J.H. (1981) *Nucleic Acids Res.* 9, 47-53.
14. Sprinzl, M. and Gauss, D.H. (1984) *Nucleic Acids Res.* 12, r1-r58.
15. Calagan, J.L., Pirtle, R.M., Pirtle, I.L., Kashdan, M.A., Vreman, H.J. and Dudock, B.S. (1980) *J. Biol. Chem.* 255, 9981-9984.
16. Shinozaki, K., Deno, H., Kato, A. and Sugiura, M. (1983) *Gene* 24, 147-155.
17. Shinozaki, K. and Sugiura, M. (1982) *Gene* 20, 91-102.
18. Bergmann, P., Seyer, P., Burkard, G. and Weil, J.H. (1984) *Plant Mol. Biol.* 3, 29-36.