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SPLIT TOLERANCE INDUCED BY ORTHOTOPIC LIVER TRANSPLANTATION IN MICE¹

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Abstract

Spontaneous orthotopic liver allograft acceptance associated with microchimerism in mice induces tolerance to subsequent skin or heart transplants from the donor but not third-party animals. Despite *in vivo* hyporesponsiveness, *in vitro* MLC and CTL assays showed continuing antidonor reactivity. Cells isolated from recipients' spleens and grafted livers, when tested in MLC and CTL assays, were antidonor reactive out to 3 months to the same degree as splenocytes obtained from either naive or presensitized (with skin or heart) mice. Nevertheless, passive transfer of splenocytes or liver lymphocytes from liver tolerant mice, but not naive or sensitized donor strain mice, were able to prolong skin graft survival significantly in naive irradiated recipients. By using a strain combination in which the donor but not the recipient expressed the stimulatory endogenous super-Ag (Mls_f), it was possible to determine whether super-Ag-reactive T cells bearing V β 5 and V β 11 were deleted or anergic. Phenotypic analysis of cells isolated from recipients' spleens and grafted livers (up to 90 days after transplant), when compared with naive animals, showed no significant difference in V β 5 and V β 11 TCR expression. Additionally, when these isolated spleen cells were tested for antibody-mediated stimulation, both anti-V β 5 and V β 11 TCR mAb led to marked proliferation of cells obtained from naive and liver-transplanted recipients, but as expected, proliferation was very low in cells from naive donors. These results suggest that liver transplantation induces donor-specific tolerance *in vivo*, which may not be reflected in *in vitro* proliferative and cytotoxicity assays (split tolerance). Furthermore, this tolerance does not seem to be induced by clonal deletion or anergy of minor-lymphocyte-stimulating-antigen-reactive T cells in the recipients.

OLT in mice across major and minor histocompatibility barriers results in a high incidence of spontaneous graft acceptance without immunosuppression (1). While some strain combinations (B10 \rightarrow C3H) fare better (mean survival > 100 days) than others (BALB/c \rightarrow C3H; mean survival > 47 days), the underlying mechanism(s) for this extemporaneous liver graft acceptance in an unmodified recipient is not clear. However, this property is not unique to the liver, only much stronger, since kidneys (2, 3) and hearts (4) in some strain combinations in mice can also lead to induction of donor-specific tolerance. The present study was designed to analyze the functional status of lymphocytes isolated from various lymphoid and nonlymphoid organs of liver graft recipients, and to investigate the role of

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minor lymphocyte-stimulating (Mls)* super-Ag and V β usage in the induction of donor-specific tolerance after OLT. For the latter study, an I-E/Mls_F-positive liver was grafted into an I-E/Mls_F-negative recipient, which provided us with a direct approach to examine V β usage in the periphery.

MATERIALS AND METHODS

Animals

Inbred male mice of the C57BL/10 (B10), B10.BR, B10.D2, C3H/HeJ, and BALB/c strains were obtained from Jackson Laboratory, Bar Harbor, ME. Mice were maintained in pathogen-free facilities, provided with Purina rodent chow and tap water ad libitum, and used at 10–12 weeks of age.

Medium

Dulbecco's modified Eagle's medium ([DMEM] GIBCO, Grand Island, NY) supplemented with 2 mM L-glutamine (GIBCO), 0.55 mM L-arginine (GIBCO), 0.3 mM L-asparagine (GIBCO), 13.6 μ M folic acid (GIBCO), 100 U/ml penicillin (GIBCO), 100 μ g/ml streptomycin (GIBCO), 1 mM sodium pyruvate (Sigma Chemical Co., St. Louis, MO), 10 mM HEPES (Sigma), 5×10^{-5} M 2-ME (GIBCO), and 0.75% mouse serum were used (complete DMEM) for all cell cultures except cytotoxicity and T cell activation assays, where instead of mouse serum, 10% decomplexed FCS (GIBCO) was used (MLC-DMEM).

Surgical techniques

OLT was performed as described previously (5). Abdominal heterotopic heart transplantation was performed as described previously by Ono and Lindsey (6). Transplanted hearts were monitored daily by direct palpation, and rejection was defined as termination of palpable cardiac contractility. A full-thickness skin graft from the donor tail was placed on the dorsal side of the recipient's trunk according to a previously described technique (7). The graft was secured by silk sutures, and protected by dressing for 7 to 8 days. The rejection process was monitored by daily inspection until the skin was completely destroyed. All procedures were performed under methoxyflurane anesthesia.

Histology

Animals were killed on days 2, 7, 14, 28, and 84 after transplantation. Tissues were harvested and fixed in neutral buffered formalin, embedded in paraffin, sectioned, and stained with hematoxylin and eosin.

Preparation of cell suspensions

Aseptically harvested spleens and lymph nodes were gently teased with 25-gauge needles in culture medium, and subsequently filtered through a nylon mesh. Erythrocytes were lysed by treatment with 5 ml of red cell lysing buffer (Sigma) for 5 min, and washed twice with complete DMEM.

Free lymphomyeloid cells from the liver were prepared as described previously (8). Briefly, the portal vein was exposed and cannulated with a 21-gauge needle and the inferior vena cava was transected proximal to the renal veins. The liver was then perfused with 40 ml of warm Hank's solution, excised, and transferred to sterile culture medium. Perfused livers were disrupted on a 50-mesh stainless steel sieve with a sterile 20-ml syringe pestle, and the

* Abbreviations: DMEM, Dulbecco's modified Eagle's medium; Mls, minor lymphocyte-stimulating antigen.

resulting cell suspension was filtered through a nylon mesh. Further purification was achieved by centrifugation on a continuous 35% Percoll (Pharmacia Diagnostics, Piscataway, NJ) gradient. Cells in the pellet were recovered, treated for 5 min with RBC lysis buffer, and, after 2 washes, resuspended in appropriate volume of complete DMEM.

MLC

All mixed cell culture assays were performed in triplicate in round-bottomed microtiter plates (Corning, Corning, NY) using equal numbers (2×10^5 cells/well) of responder cells, and γ -irradiated (2000 cGy) stimulator cells in a total volume of 200 μ l. Controls included responder cells and irradiated stimulators incubated in medium alone and responders incubated with autologous stimulators. Cultures were incubated at 37°C in a humidified 10% CO₂ atmosphere for 2, 3, 4, and 5 days and pulsed with 1 μ Ci/well of [³H]thymidine (NEN, Boston, MA) 18 hr before termination of the assay. Cells were harvested (Skatron, LKG, Wallac, Norway) onto glass fiber filter mats (Wallac Oy, Turku, Finland) and radioactivity was determined in a liquid scintillation counter (Betaplate 1205; Pharmacia LKB, Gaithersburg, MD). The results are expressed as the mean cpm \pm 1 SD of [³H]thymidine incorporation in triplicate cultures.

Cytotoxicity assays

Effector cells from spleens were prepared by incubating equal numbers (4×10^6 cells/well) of responders and γ -irradiated (2000 cGy) stimulators (syngeneic, donor, and third party) for 4 to 5 days in a 12-well tissue culture plate (Corning). On the contrary, freshly isolated free lymphomyeloid cells from the grafted livers were used as effectors. Target cells were prepared by incubating 4×10^6 spleen cells for 48 hr in MLC-DMEM containing Con A (Sigma) at 5 μ g/ml. The target cells were then labeled with Na₂ ⁵¹CrO₄ (NEN) and washed 3 times to remove excess ⁵¹Cr, and 4×10^3 cells/well were placed in V-bottomed 96-well tissue culture plates (Corning). Serial 3-fold dilutions of effector cells were added at a maximum E:T ratio of 100:1 in a total volume of 200 μ l/well, and the plates were incubated for 4 to 5 hr at 37°C in 10% CO₂. At the end of this incubation, plates were centrifuged at 500 \times g for 10 min, and 100 μ l of supernatant from each well were removed for counting. Spontaneous ⁵¹Cr release from target cells was determined by incubating targets with medium alone. The percentage of specific ⁵¹Cr release was calculated according to the formula:

$$\% \text{ specific } ^{51}\text{Cr} \text{ release} = \frac{(\text{cpm}_{\text{experimental}} - \text{cpm}_{\text{spontaneous}})}{(\text{cpm}_{\text{maximum}} - \text{cpm}_{\text{spontaneous}})} \times 100$$

The results are expressed as mean cpm \pm 1 SD of % specific ⁵¹Cr release in triplicate cultures.

Flow cytometric analysis

Cells isolated from spleens and grafted livers were phenotyped by single-color immunofluorescence labeling with the following FITC-conjugated rat anti-mouse mAb (Pharmingen, San Diego, CA): anti-Thy-1 (IgG_{2c}, pan T cell antigen), anti-CD4 (IgG_{2b}, L3T4, Th cells), anti-CD8 (IgG_{2a}, Ly-2, cytotoxic/suppressor T cells), and anti-CD45R (IgG_{2a}, B220, pre-B and B lymphocytes). FITC-conjugated, isotype-matched irrelevant mAb were used as appropriate negative controls. For surface marker analysis, 5×10^5 cells were incubated with 100 μ l (1:100 dilution) of appropriate mAb for 60 min at 4°C. The cells were washed twice, resuspended in PBS containing 1% paraformaldehyde, and analyzed using a FACScan flow cytometer (Becton-Dickinson, Mountain View, CA).

Cells isolated from spleens, lymph nodes, thymus, and grafted livers were also analyzed for relative frequency of various subsets of TCR, by double immunofluorescent staining with the following mAb (PharMingen): phycoerythrin-conjugated anti-Thy-1 (rat IgG_{2c}) and anti-CD3- ϵ (hamster IgG), FITC-conjugated or biotinylated anti-V β 3 TCR (hamster IgG), anti-V β 5.1, 5.2 TCR (mouse IgG₁), anti-V β 8.1,8.2 TCR (mouse IgG_{2a}), and anti-V β 11 TCR (rat IgG_{2b}). Cells (5×10^5 cells/tube) were first incubated for 60 min at 4°C with 100 μ l of phycoerythrin-conjugated anti-Thy-1 mAb (1:100 dilution). The cells were then washed twice and 100 μ l of FITC-conjugated or biotinylated anti-CD3 or anti- β 3 TCR mAb were added at 1:100 dilution. After 60 min of incubation at 4°C, the cells were washed and only those cells that were stained with biotinylated mAb were further incubated for 30 min at 4°C with 100 μ l of streptavidin-conjugated FITC (1:100 dilution, Jackson ImmunoResearch Laboratory, West Grove, PA). All cells were washed, resuspended in 1% paraformaldehyde, and finally analyzed on a FACScan flow cytometer (Becton-Dickinson). Fluorochrome-conjugated, isotype-matched irrelevant mAb were used as an appropriate negative control.

Adoptive transfer assays

All adoptive transfer assays were performed in the strain combination B10 \rightarrow C3H. Cells were isolated from spleens of naive animals (C3H), from C3H recipients of B10 liver or skin (4 weeks after transplant), and from transplanted B10 \rightarrow C3H livers (4 weeks after transplant). They were finally resuspended at 2×10^7 cells/ml in Hank's solution and 500 μ l (i.e., 10^7 cells/animal) were infused through the penile vein into sublethally irradiated (650 cGy) naive C3H recipients (syngeneic). These cell transplant recipients were simultaneously challenged with a full-thickness skin graft from a B10 donor.

TCR cross-linking assay

T cell proliferation was induced by cross-linking TCR with mAb directed against T cell surface antigens: CD3, V β 3, V β 5, V β 8, and V β 11 TCR. The assay was performed as described previously (35). Briefly, 100 μ l of relevant antibody solution (10 μ g/ml) were incubated overnight at 37°C in 96-well U-bottomed microtiter plates (Corning). At the end of this incubation period, plates were washed with sterile Hank's solution, and 2×10^5 spleen cells were then added to each well in a final volume of 200 μ l of MLC-DMEM. The cells were cultured for 3 to 5 days at 37°C in 10% CO₂, with [³H]thymidine (1 μ Ci/well) added for the last 16–18 hr of culture. Plates were harvested onto glass filter mats and the [³H]thymidine incorporation was determined by liquid scintillation spectrophotometry. Negative controls were cells cultured with MLC-DMEM alone. Positive controls were cells cultured with either Con A (5 μ g/ml) or LPS (40 μ g/ml). Results are expressed as mean cpm \pm 1 SD of triplicate samples.

RESULTS

Characterization of cellular infiltrate

All allotransplanted livers underwent a brisk rejection episode that resolved spontaneously within 4–5 weeks. However, despite heavy cellular infiltration, little hepatocellular necrosis was observed. Phenotyping of free lymphomyeloid cells isolated from normal (B10 and C3H) and grafted livers (C3H \rightarrow C3H and B10 \rightarrow C3H) (2–90 days after transplant) showed a pre-dominance of lymphocytes (Table 1). Normal B10 and C3H livers had approximately equal numbers of B and T cells, whereas after transplantation, the majority of isolated cells from allografted but not syngrafted livers were T lymphocytes (Table 1). Of the T cell subsets, CD8⁺ cells far outnumbered CD4⁺ cells in the allografted livers (Table 1). On the contrary, the phenotype of splenocytes isolated from liver, heart, or skin recipients was similar to naive controls (data not shown).

Proliferative responses of cells isolated from spleens and grafted livers

As early as 8 days after liver transplantation, the recipients' (C3H) spleen cells showed alloreactivity toward γ -irradiated donor (B10) and third-party (BALB/c) splenocytes (Fig. 1B), while sustaining nonresponsiveness to autologous lymphocytes (Fig. 1B). This reactivity was maintained during the entire observation period (tested on days 8, 14, 30, and 90) after liver transplantation (Fig. 1D). A similar lymphoproliferative response was also observed in spleen cells isolated from skin and heart transplant recipients (B10 \rightarrow C3H, 90 days after transplant) who had rejected their grafts within 14 days (Fig. 2). Spleen cells from long-term liver graft recipients (> 100 days after transplant) in 2 other strain combinations (B10.BR \rightarrow B10 and B10.BR \rightarrow B10.D2) also responded normally to donor and third-party, but not to autologous or naive syngeneic lymphocytes (data not shown). Free lymphomyeloid cells isolated from grafted livers (up to 90 days after transplant) were also nonresponsive to self and naive syngeneic lymphocytes, but responded normally to donor and third-party stimulation (data not shown).

Cytotoxic activity of cells isolated from spleens and grafted livers

An elevated level of cytotoxic killing was seen in stimulated effectors (C3H spleen cells) on days 8, 14, 30, and 90 after B10 liver transplantation (Fig. 3B and 3D). Lysis of third-party (BALB/c) targets was as effective as donor, whereas killing of syngeneic targets was low, ranging between 2% and 6% (Fig. 3, B and D). Similar observations were also made for spleen cells isolated after skin or heart transplantation (data not shown). Freshly isolated lymphocytes from spleens of liver graft recipients and grafted livers were also tested for their cytotoxic capacity. Cells isolated from grafted livers (8–90 days after transplant), but not spleens, showed high alloreactive cytolysis of both donor and third-party targets, with minimal lysis of syngeneic targets (Table 2).

Effects of transplanting cells from tolerant to naive animals

Unmodified C3H mice rejected B10 skin grafts in 13 ± 0.7 days (Table 3). Skin grafts were rejected in 18 ± 1 days if, at the time of skin transplant, naive spleen cells (10^7 cells/animal) were infused into γ -irradiated syngeneic mice. Furthermore, infusion of spleen cells from skin-sensitized mice (B10 \rightarrow C3H) failed to prolong skin graft survival beyond normal controls (17 ± 1 days), whereas reconstitution of γ -irradiated animals with spleen cells from liver recipients (4 weeks > OLT) or cells from grafted livers (4 weeks after transplant) delayed skin graft rejection far beyond normal controls (Table 3).

Effect of liver transplantation on V β TCR usage in the periphery

To investigate what role clonal deletion of super-Ag-reactive T cells might play in inducing tolerance after liver transplantation, lymphocytes from spleens, lymph nodes, and thymus of naive and liver-grafted animals were examined for V β TCR usage. Peripheral T cells of B10.BR (Mls $^+$ -positive) donors do not express V β 5 (0.94%) and V β 11 (1.0%) segments of TCR, whereas T cells of B10 (Mls $^-$ -negative) recipients expressed relatively high levels of V β 5 (6.4%) and V β 11 (5.5%) TCR (Fig. 4). However, both animals express comparable levels of V β 3 ($3.25 \pm 3\%$) and V β 8 ($16 \pm 1\%$) TCR, which serve as positive controls. While examining spleen cells from liver transplant recipients (2–90 days after transplant), we found that donor super-Ag-reactive T cells expressing V β 5 and V β 11 TCR were not deleted (Fig. 4). Cells isolated from recipients' lymph nodes, thymus, and grafted liver also expressed normal levels of V β 5 and V β 11 TCR (data not shown). Similar results were also obtained in rejecting B10.BR \rightarrow B10 skin graft recipients (6–15 days after transplant). Expression of V β 3 and V β 8 TCR in transplanted animals was comparable to naive controls (Fig. 4).

Antibody-mediated proliferation of peripheral T cells after liver transplantation

If purging of donor super-Ag-reactive T cells is not the basis for tolerance induction, could it be that these cells are rendered functionally inert (clonal “anergy” or “silencing”) by donor cells migrating out of transplanted livers? To test this hypothesis, spleen cells isolated from naive and transplanted recipients were stimulated with a panel of anti-V β TCR mAb. Cells isolated from naive B10 (recipient) showed a proliferative response when cultured with anti-V β 5 and anti-V β 11 TCR mAb, whereas B10.BR (donor) did not respond beyond background controls. Both B10 and B10.BR responded equally well to anti-CD3- ϵ (data not shown), anti-V β 3, and anti-V β 8 TCR mAb (Fig. 5). Cells isolated from spleens of liver graft recipients (2–90 days after transplant) responded in consonance with naive B10 by proliferating vigorously to anti-CD3- ϵ , anti-V β 5, and anti-V β 11 TCR mAb stimulation (Fig. 5), suggesting that donor super-Ag-reactive T cells in the recipient are functionally active.

DISCUSSION

We have demonstrated that liver transplantation in mice across major and minor histocompatibility barriers resulted in spontaneous graft acceptance and induction of donor-specific tolerance (1). Systemic tolerance to the donor was illustrated by the observation that other donor organs, such as heart and skin, were protected from rejection by the grafted liver, whereas third-party organs were rejected within the normal time course (1). Moreover, hepatic tolerogenicity was so robust that an ongoing skin graft rejection was reversed by subsequent liver transplant, thus reverting a state of sensitization to one of tolerance (1). Similar observations have also been made by Kamada et al., who showed that in the DA \rightarrow PVG rat strain combination, liver was only rejected if the recipient was presensitized against donor antigens (9-12). Despite the liver’s potentially strong tolerogenic capacity, this attribute is not uniquely ascribed to them, since kidneys (2,3) and hearts (4) in some strain combinations in mice can also lead to spontaneous graft acceptance and induction of donor-specific tolerance.

In addition to specific systemic hyporesponsiveness to the donor, these liver graft recipients also exhibited the establishment of microchimerism (1), which we have reported previously is an inevitable outcome of all successful whole organ transplantation (13-18). However, this *in vivo* nonreactivity to the donor was not maintained *in vitro*, since cells isolated from the spleens of long-term liver graft recipients (up to 90 days after transplantation) when tested in an MLC, responded vigorously to irradiated donor as well as to third-party stimulators, whereas response to syngeneic stimulators was very low. These cells also exhibited elevated levels of cytotoxicity both toward the donor and third party lymphocytes and cultured bile duct cells. This cytotoxic activity was evident at an E:T ratio as low as 3:1. The autologous cell lysis was very low (> 6%), suggesting that this killing was not due to lymphokine-activated killer cells, but rather due to the generation of specific cytotoxic cells early after liver transplantation. This dichotomy between *in vivo* hyporesponsiveness and *in vitro* alloreactivity has been referred to as “split tolerance” (19, 20). These findings are in agreement with those in rats (9,10, 12) and in humans (21), further supplementing our own assertion that *in vitro* observations are not always a true prediction of the *in vivo* immune status of the recipient.

Several attempts have been made to associate the phenotypic profile of graft-infiltrating cells with immunological responses to the allografts. Of particular interest are the phenotypes of cells infiltrating the graft and their *in vivo* and *in vitro* immunological status. A relative increase in CD8⁺ T cells with the reversal of the CD4 to CD8 ratio has been demonstrated in the cells infiltrating acutely rejecting rat cardiac allografts (22) and human (23) and rat renal (24) allografts. In our study, we also found a similar reversal of the CD4 to CD8 ratio 1 week after allogeneic but not syngeneic liver transplantation, which was

sustained throughout the observation period (12 weeks after transplant). Furthermore, despite significant and continuously persistent donor and third-party MLC and CTL alloreactivity, these liver graft-infiltrating cells appeared to be innocuous *in vivo*, as there was little morphological evidence of hepatocellular necrosis.

In adoptive transfer assays in rats, Kamada et al. demonstrated that transfusion of thoracic duct lymphocytes from liver-grafted animals into irradiated syngeneic recipients resulted in prolongation of skin grafts of the donor, but not third-party, strains (9-11,25). Similar observations were also made in our experiments in mice, where we found that donor but not third-party skin graft survival was significantly prolonged in naive syngeneic sublethally irradiated mice that were reconstituted with cells isolated from either spleens or livers of long-term liver allograft recipients. On the contrary, donor skin graft survival was not prolonged in irradiated syngeneic animals that were reconstituted with splenocytes from skin-sensitized mice.

Clonal deletion of immature T cells in the thymus bearing Mis-reactive TCR subsets is an established mechanism for the induction of self-tolerance (26-29). The relationship among clonal deletion, chimerism, and donor-specific tolerance induction has also been reported by Streilein and co-workers in an I-E-disparate neonatal transplantation model (30-32). Furthermore, clonal deletion or anergy of mature T cells in the periphery has also been proposed as an alternative mechanism for the induction of tolerance to self or alloantigens (33-37). These findings are in agreement with those of Kamada, who found that liver allograft acceptance and the subsequent induction of donor-specific tolerance in rats was associated with elimination of donor-reactive CD8⁺ T cells in the recirculating lymphocyte pool of the recipients (38). However, using Mls_f-positive (B10.BR, V β 5, V β 11 TCR-negative) mice as donors and Mls_f-negative (B10, V β 5, V β 11 TCR-positive) animals as recipients, a strain combination in which liver allografts are accepted spontaneously, resulting in the establishment of multilineage microchimerism (including the seeding of putative super-Ag-expressing donor B cells and dendritic cells in the recipient's thymus), and the subsequent induction of donor-specific tolerance (1), no difference in V β 5 and V β 11 TCR expression and antibody-mediated proliferative responses was observed in splenocytes, lymph node cells, and liver graft-infiltrating cells of naive and experimental animals at any time after transplantation (2-90 days after transplant). These observations are similar to earlier reports by Salaun et al. (39), who showed that the induction of specific tolerance in nude mice that were reconstituted with donor-strain thymic epithelium at birth was not due to clonal deletion or anergy of super-Ag-reactive T cells in the recipients.

In summary, our studies demonstrate that spleen and liver graft-infiltrating cells in recipients of liver allografts, though hyporesponsive *in vivo*, exhibit a strong donor and third-party alloreactivity *in vitro* (split tolerance). The complete absence of autologous/syngeneic reactivity suggests that this cytolytic response is not due to engendering of lymphokine-activated killer cells, but probably due to the generation of specific cytotoxic T cells after liver grafting. The significantly prolonged acceptance of donor, but not third-party, skin grafts by irradiated animals that were reconstituted with spleen or liver graft-infiltrating cells obtained from long-term liver allograft recipients is an affirmation that these cells are capable of transferring tolerance to naive syngeneic animals. Furthermore, clonal deletion or anergy of Mis-reactive T cells is not the basis for tolerance induction after liver transplantation in mice. Finally, liver-induced tolerance to the donor is probably not mediated by any suppressor factors in the recipient's serum (manuscript in preparation). A possible explanation of this paradox might be inherent in the donor bone-marrow-derived cells, which after whole organ transplantation are known to migrate from the allograft into the recipient, leading to the establishment of chimerism, which is thought to be the first step toward subsequent induction of donor-specific tolerance (1, 18).

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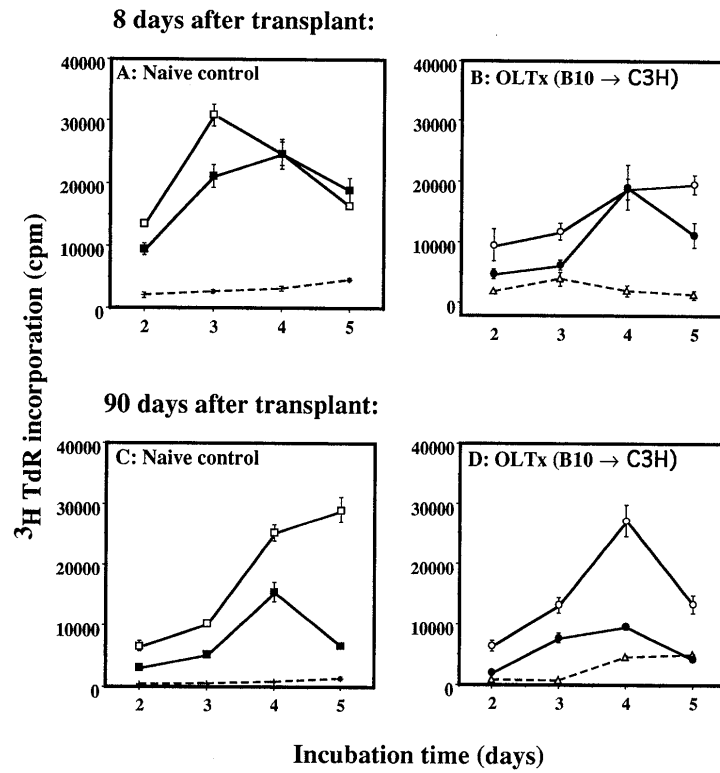


Figure 1. MLC. Spleen cells isolated from either naive C3H (A and C) or C3H recipients of B10 liver, 8 (B) and 90 days (D) after OLT, were used as responders. γ -Irradiated spleen cells from syngeneic (C3H [+, Δ]), donor (B10 [\square , \circ]), and third-party (BALB/c [\blacksquare , \bullet]) strains were used as stimulators. [^3H]Thymidine incorporation is expressed as mean cpm \pm 1 SD of triplicate cultures.

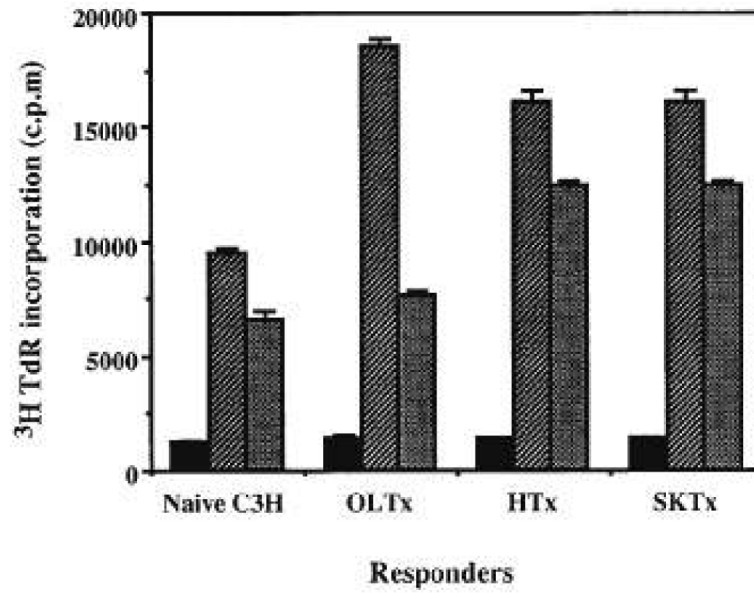


Figure 2. MLC. Responders were isolated from spleens of naive C3H or C3H recipients of B10 liver, heart, and skin (90 days after transplant), and stimulated for 96 hr with γ -irradiated splenocytes from (■) naive syngeneic, (▨) donor, and (▩) third-party (BALB/c) animals. The results are expressed as mean cpm \pm 1 SD of tritiated thymidine incorporation by triplicate cultures.

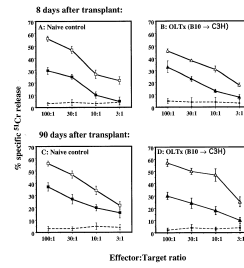


Figure 3.

CTL assay. Spleen cells (effectors) from naive and transplanted recipients were cultured for 4 days with γ -irradiated splenocytes from syngeneic, donor, or third-party strains. Lytic activity was determined by incubating decreasing doses of stimulated effectors with a constant dose of naive ^{51}Cr -labeled targets. All effectors were stimulated with target-strain-specific naive cells. Experimental groups (E:T): (+) naive C3H:C3H, (\square) naive C3H:B10, (\blacksquare) naive C3H:BALB/c, (X) C3H > OLTx:C3H, (Δ) C3H > OLTx:B10, and (\blacktriangle) C3H > OLTx:BALB/c. The results are mean % specific ^{51}Cr release \pm 1 SD of triplicate cultures.

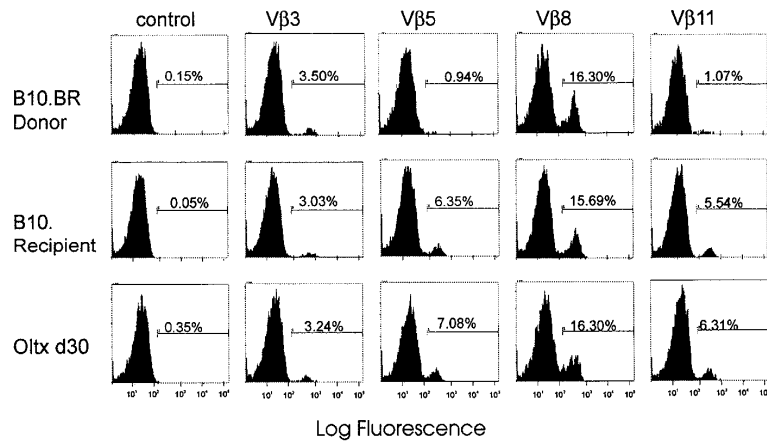


Figure 4. FACScan analysis of V β usage. The frequency of V β 3, V β 5, V β 8, and V β 11 TCR was examined in the T cells isolated from the spleens of naive B10.BR (donor), naive B10 (recipient), and recipients of B10.BR livers (30 days after transplant). Isotype-matched irrelevant mAb was used as a negative control.

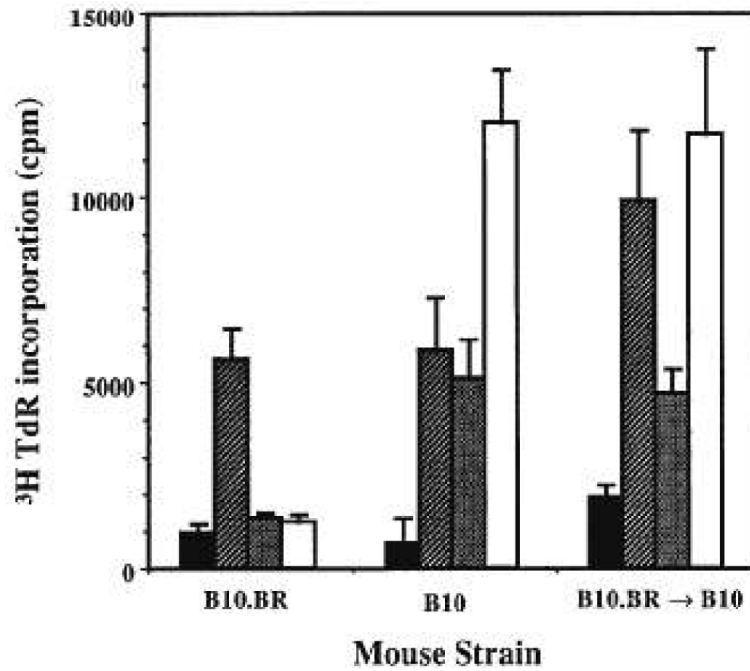


Figure 5.

Antibody-mediated T cell activation. Spleen cells isolated from naive B10.BR (donor), naive B10 (recipient), and recipients of B10.BR livers (90 days after transplant), were activated by mAb directed against (▨) Vβ3, (▩) Vβ5, and (□) Vβ11 segments of TCR. Spleen cells cultured in media alone (■) were used as controls. [³H]Thymidine uptake is expressed as mean cpm ± 1 SD of triplicate cultures.

Phenotype of cells isolated from livers of naive (C3H and B10) animals, and syngeneic (C3H → C3H) and allogeneic (B10 → C3H) transplanted livers

Table 1

Naive (n=3)	Liver cell yield (x10 ⁶)		B cells (%)		T cells (%)		CD4 ⁺ (%)		CD8 ⁺ (%)		CD4:CD8 ratio	
	B10	C3H	B10	C3H	B10	C3H	B10	C3H	B10	C3H	B10	C3H
	0.7±0.3	0.5±0.4	31±4	38±5.1	48±6.2	43±3.9	41±5.4	29±4.3	27±3.5	14±3.1	2.07	2.01
Time postTx ^a	syn	allo	syn	allo	syn	allo	syn	allo	syn	allo	syn	allo
day 2	1.4±0.6	2.5±1.6	26±4.2	18±9	43±7.5	47±8.1	37±0.5	27±8.8	11±1.4	20±14	1.35	3.36
1 wk	2.8±2.1	10±3.6	39±5.1	13±5	41±14	56±27	31±5.7	20±9.1	13±2.5	31±5.7	2.38	0.67
4 wk	0.5±0.3	9.8±3.1	18±8.5	20±7	40±3	73±18	33±7.1	19±2.5	13±2.5	53±21	2.54	0.36
12 wk	ND ^b	3.8±0.9	ND	28±9.5	ND	61±12	ND	18±3.5	ND	34±14	ND	0.53

^a n=4 at each time point after transplantation.

^b ND, not done.

Table 2

Direct cytotoxicity of freshly isolated cells from spleens and livers of liver graft recipients (B10 → C3H), tested against syngeneic (C3H), donor (B10) and third-party (BALB/c) targets^a

Time postTx	Animal	Percent specific ⁵¹ Cr release (mean cpm ± 1 SD)											
		Liver graft-derived cells (E:T ratio)						Spleen-derived cells (E:T ratio)					
		C3H		B10		BALB/c		C3H		B10		BALB/c	
		100:1	100:1	30:1	100:1	100:1	100:1	100:1	100:1	100:1	100:1	100:1	100:1
1-2 Weeks	a	10±3	37±3.3	16±2.5	10±3.2	12±2.3	1±0	15±1.5					
	b	2±1.1	56±5.4	26±4.9	10±2.1	1±0	1±0	1±0					
4 Weeks	a	ND ^b	36±2.9	20±2.6	ND	1±0	2±1	5±1.8					
	b	1±0	30±3	18±2.9	1±0	7±2.7	3±1.5	1±0					
8-12 Weeks	a	9 ± 3	35±3.9	12±4.2	8±2.7	12±2.6	14±1.9	4±2					
	b	1±0	20±3.3	10±3.6	3±1.3	1±0	1±0	3±1.2					

^a⁵¹Cr-labeled Con A blasts were used as targets.

^bND, not done.

Table 3

Adoptive transfer of 10⁷ cells isolated from spleens of either naive, skin, or liver graft recipients (B10 → C3H), or from transplanted livers to naive syngeneic recipients, which were subsequently challenged with B10 skin graft

Cells transfused	n	γ -Irradiation (650 cGy)	Skin graft survival mean days \pm 1 SD
Control group			
None	4	No	13 \pm 0.7
Naive C3H spleen cells	8	Yes	18 \pm 1.0
Experimental group ^a			
>OLT			
Spleen cells	7	Yes	27 \pm 0.5
Liver-derived cells	3	Yes	27 \pm 0.6
>Skin transplant			
Spleen cells	3	Yes	17 \pm 1.5

^aThirty days after transplant.