Review

The vascular endothelial growth factor family; proteins which guide the development of the vasculature

MARC G. ACHEN AND STEVEN A. STACKER

Angiogenesis Laboratory, Ludwig Institute for Cancer Research, Royal Melbourne Hospital, Victoria, Australia

Received for publication 16 June 1998 Accepted for publication 18 June 1998

Summary. The development of the vascular tree during embryogenesis involves vasculogenesis, angiogenesis and tissue-specific differentiation of endothelium which gives rise to many different vessel types. These processes are physiologically complex and are therefore difficult to study in vitro. However, the discovery of endothelial cell-specific receptors and cognate ligands has led to the generation of transgenic and knockout mouse models which have shed light on the molecular mechanisms that regulate the development of blood and lymphatic vessels during embryogenesis. Such mouse models have demonstrated that members of the vascular endothelial growth factor (VEGF) family of proteins and the VEGF receptors are critical regulators of vasculogenesis, angiogenesis and endothelial cell differentiation. The availability of purified VEGF family members and of inhibitors of these growth factors may provide a means to modulate blood vessel growth for the treatment of cancer, retinopathies and diseases of ischemia.

Keywords: angiogenesis, vasculogenesis, embryogenesis, tumour development, VEGF

Vascular endothelial growth factor (VEGF), a secreted glycoprotein, was purified and cloned in the 1980s based on its activities as an inducer of vascular permeability (Senger *et al.* 1983; Connolly *et al.* 1989; Keck *et al.* 1989) and as a mitogen for endothelial cells (Ferrara & Henzel 1989; Leung *et al.* 1989). Since then it has become clear that VEGF plays a crucial role in the blood vessel growth integral to embryogenesis and tumour development. More recently, numerous proteins have been identified which are closely related in structure to VEGF and which are also thought to be involved in vascular development. These proteins constitute the

Correspondence: Marc G. Achen, Angiogenesis Laboratory, Ludwig Institute for Cancer Research, Post Office Box 2008, Royal Melbourne Hospital, Victoria 3050, Australia. Fax: +61 3 9341 3107; E-mail: Marc.achen@ludwig.edu.au

© 1998 Blackwell Science Ltd

VEGF family. Here we summarize what is known about the structure and biological functions of VEGF family members. It is clear that the VEGF family of vascular growth and differentiation factors, and inhibitors thereof, offer great potential for manipulation of blood vessel growth in patients.

Blood vessel growth in the embryo and adult

The formation of blood vessels in the embryo occurs by two distinct processes, vasculogenesis or angiogenesis (for review see Risau 1997). Vasculogenesis is the *in situ* differentiation, from mesoderm, of angioblasts (endothelial cell precursors which have not formed a lumen) and association of these cells to form blood vessels (Risau & Flamme 1995). The primary vascular plexus

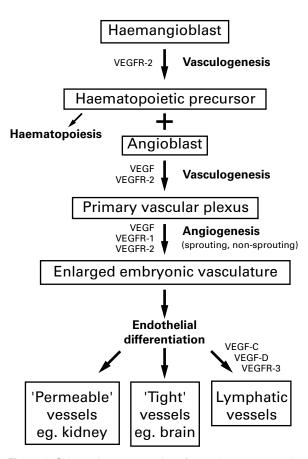


Figure 1. Schematic representation of some key processes in vascular development during embryogenesis. The order of events during embryogenesis proceeds from the top of the figure to the bottom. The steps of vasculogenesis shown are those which occur in the yolk sac, where both early haematopoiesis and endothelial cell differentiation occur side-by-side in the blood islands (Risau & Flamme 1995). The haemangioblast is a hypothesized cell type which gives rise to both haematopoietic and endothelial cell precursors (Choi *et al.* 1998). VEGF family members and receptors thought to be involved in steps of vascular development are shown. The scheme presented here is a simplification as vasculogenesis and angiogenesis occur simultaneously in some organs. The list of vessel types arising from endothelial cell differentiation shown here is far from complete.

of the early embryo is established by vasculogenesis (Figure 1). In contrast, angiogenesis is the formation of blood vessels from preexisting vessels and is primarily responsible for the development of blood vessels during later embryogenesis and adult life. Two types of angiogenesis have been described: sprouting of vessels from preexisting vessels and non-sprouting angiogenesis (intussusception) which involves splitting of vessels to generate greater numbers of vessels (Patan *et al.* 1996).

Once blood vessels have been established in the

embryo, endothelial cells undergo tissue-specific changes to generate numerous types of functionally distinct vessels as organs differentiate (Risau 1995). This process gives rise to a variety of endothelial cell types with distinct morphologies and biochemical characteristics. The diversity of the endothelium is exemplified by lymphatic endothelial cells which have a discontinuous or even partially absent basement membrane (Leak 1970), the endothelial cells of the peritubular capillaries in the kidney which are highly fenestrated and brain endothelial cells which are linked by tight junctions to form the blood-brain barrier.

In the adult, angiogenesis is tightly controlled - under normal circumstances it occurs almost exclusively in the female reproductive system. However, angiogenesis can be activated in the adult in response to tissue damage and is critical in certain pathological conditions such as tumourigenesis, rheumatoid arthritis and diabetic retinopathy (for review see Folkman & Shing 1992). Recent findings suggest that angiogenesis is not the only mechanism responsible for blood vessel growth in adults, as circulating endothelial precursor cells have been isolated from human peripheral blood which can differentiate into endothelial cells and be incorporated into newly growing vessels at sites of angiogenesis (Asahara et al. 1997). Thus endothelial cell differentiation, a hallmark of vasculogenesis, can contribute to vessel growth in adults.

The processes essential for development of the vasculature are physiologically complex. For example, angiogenesis involves not only endothelial cell proliferation, but also degradation of the extracellular matrix, cell migration, cell-cell adhesion, lumen formation and recruitment of pericytes and smooth muscle cells (Risau 1997). Therefore blood vessel development must require endothelial cells to respond to a variety of extracellular signals that activate receptors responsible for growth and differentiation. Studies carried out over the past five years have conclusively demonstrated that members of the VEGF family of growth factors are prominent among the extracellular signalling molecules that guide vascular development (Carmeliet *et al.* 1996; Ferrara *et al.* 1996; Jeltsch *et al.* 1997).

Tumour angiogenesis

Many solid tumours are capable of inducing angiogenesis – this serves to provide the tumour with nutrients for growth and appears to be critical for the generation of metastases. The concept has emerged that tumour cells may produce factors which either induce or inhibit angiogenesis and that the onset of angiogenic activity is

determined by the balance of these factors (Folkman & Shing 1992). It is now known that VEGF is critical for supporting, and perhaps initiating, angiogenesis in many tumours (Kim *et al.* 1993; Saleh *et al.* 1996).

Numerous therapeutic approaches for treatment of cancer, which target tumour angiogenesis, are currently under development. The potential attractions of such approaches are (i) selective toxicity due to the paucity of angiogenesis in the adult; (ii) assured access of drugs to target endothelial cells; (iii) the genetic stability of the target endothelial cells which means that drug resistant variants are unlikely to arise.

The VEGF family: general structural features

The VEGF family, members of which were originally defined on the basis of similarity of primary structure to VEGF, consists of VEGF (Leung et al. 1989), VEGF-B (Olofsson et al. 1996a), VEGF-C (Joukov et al. 1996), VEGF-D (Achen et al. 1998) and placenta growth factor (PIGF) (Maglione et al. 1991). These glycoproteins are members of a structural superfamily of growth factors containing a cystine knot motif which also includes platelet-derived growth factor BB (PDGF-BB) and transforming growth factor $\beta 2$ (TGF $\beta 2$) (McDonald & Hendrickson 1993). Crystallographic studies of numerous cystine knot growth factors, including VEGF (Muller et al. 1997a; Muller et al. 1997b), revealed that the six conserved cysteine residues of the motif contribute to a three-dimensional fold involving an unusual clustering of three cystine bridges. These bridges are intertwined in such a way as to resemble a knot. VEGF is the only member of the VEGF family for which the crystal structure has been determined. The crystal structure of VEGF is most similar to that of PDGF-BB (Muller et al. 1997a).

The VEGF family: specific characteristics and biological functions

VEGF

Structure. VEGF, also known as Vascular Permeability Factor (VPF), is a 34–46 kD homodimeric glycoprotein which is a highly specific mitogen for vascular endothelial cells, is capable of inducing angiogenesis (Leung *et al.* 1989), is a potent inducer of vascular permeability (Senger *et al.* 1983; Keck *et al.* 1989) and is a survival factor for newly formed blood vessels (Alon *et al.* 1995; Benjamin & Keshet 1997). VEGF monomers are linked together by two disulphide bridges to form the homodimer (Pötgens *et al.* 1994; Muller *et al.* 1997a). The known human VEGF isoforms are 121, 145, 165, 189 and 206 amino acids in length and are generated by alternative splicing of VEGF RNA derived from a single gene (Leung et al. 1989; Houck et al. 1991; Tischer et al. 1991; Poltorak et al. 1997). VEGF₁₆₅ is the predominant isoform secreted by many normal and transformed cells, however, transcripts for VEGF₁₂₁ and VEGF₁₈₉ are detected in most tissues that express the VEGF gene (Houck et al. 1991). In contrast, VEGF₁₄₅ and VEGF₂₀₆ are more restricted in expression (Houck et al. 1991; Poltorak et al. 1997). The four largest isoforms of VEGF bind to heparin and heparan sulphate proteoglycans whereas VEGF₁₂₁ does not (Houck et al. 1992; Poltorak et al. 1997). The affinities of the VEGF isoforms for heparin affect their bioavailability; VEGF₁₂₁ is secreted as a freely soluble protein, VEGF₁₆₅ is secreted, however, a proportion of the protein remains associated with the cell surface or extracellular matrix (ECM), and VEGF₁₈₉ and VEGF₂₀₆ are almost completely bound to the ECM (Houck et al. 1992; Park et al. 1993).

Function during embryogenesis. Embryonic vascular development is dependent on VEGF as the formation of vessels in mouse embryos heterozygous for a disrupted VEGF gene was aberrant and resulted in embryonic lethality (Carmeliet *et al.* 1996; Ferrara *et al.* 1996). VEGF deficiency impaired numerous steps of early vascular development including vasculogenesis, angiogenesis and formation of large vessels. A lethal phenotype for a heterozygous animal is highly unusual for inactivation of an autosomal gene and indicates stringent dosedependent regulation of vascular development by VEGF.

Three high affinity receptors for VEGF have been identified, VEGFR-1 (also known as Flt1) (De Vries et al. 1992), VEGFR-2 (also known as Flk1 and KDR in mouse and man, respectively) (Figure 2) (Quinn et al. 1993) and neuropilin-1 (Soker et al. 1998). VEGFR-1 and VEGFR-2 are cell surface receptor tyrosine kinases which are localized on endothelial cells during embryonic development. Neuropilin-1, in addition to binding VEGF₁₆₅, is a receptor that mediates the chemorepulsive activity of the collapsin/semaphorins, a large family of transmembrane and secreted glycoproteins that function in repulsive growth cone and axon guidance in the developing embryo (He & Tessier-Lavigne 1997; Kolodkin et al. 1997). The mechanism of signalling by neuropilin-1 and the physiological significance of this protein as a VEGF receptor is at present unclear.

The co-ordinated patterns of expression of the genes for VEGF, VEGFR-1 and VEGFR-2 suggest that these proteins participate in paracrine systems which regulate vascular development during embryogenesis (Breier *et al.* 1992; Jakeman *et al.* 1993; Millauer *et al.* 1993;

^{© 1998} Blackwell Science Ltd, International Journal of Experimental Pathology, 79, 255-265

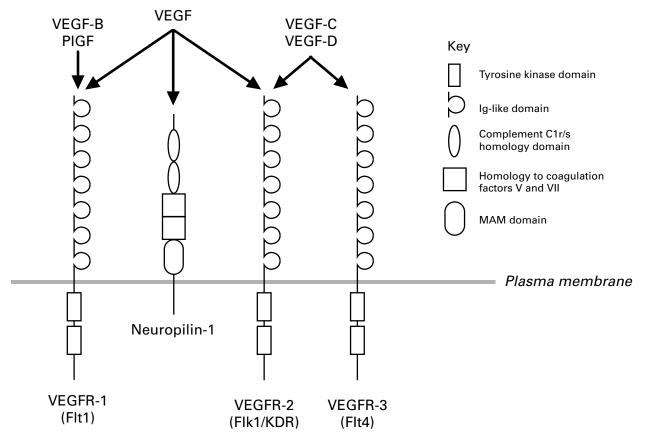


Figure 2. Interactions between VEGF family members and VEGF receptors. VEGF family members are shown at the top and arrows indicate the receptor specificities of these ligands. The similar structures of the extracellular domains of VEGFR-1, VEGFR-2 and VEGFR-3, each consisting of 7 immunoglobulin (lg)-like domains, are apparent. The structures and proposed functions of the domains of the extracellular portion of neuropilin-1 are discussed by He & Tessier-Lavigne (1997). It is not known if neuropilin-1, a recently identified receptor for VEGF, binds to VEGF-B, PIGF, VEGF-C or VEGF-D.

Peters et al. 1993; Breier et al. 1995). Consistent with such a hypothesis were the findings that these two VEGF receptors are essential for vascular development. Embryos lacking VEGFR-2, which die at approximately embryonic day 9, have drastically reduced numbers of haematopoietic precursors and angioblasts, indicating that this receptor plays a crucial role in vasculogenesis (Shalaby et al. 1995). In contrast, VEGFR-1 is not required for vasculogenesis as embryos lacking this receptor produce both haematopoietic and endothelial cells, however, assembly of endothelial cells into functional blood vessels is abnormal leading to death at approximately embryonic day 8.5 (Fong et al. 1995). It has been proposed that VEGFR-1 signalling may regulate endothelial cell-cell or cell-matrix interactions during vascular development (Fong et al. 1995).

Interestingly, the delayed endothelial cell differentiation which results from VEGF deficiency was not as extreme as the aborted endothelial cell differentiation resulting from VEGFR-2 deficiency. This observation suggests that other ligands for VEGFR-2 may be essential for normal vasculogenesis. Alternative VEGFR-2 ligands are discussed in following sections.

Role in tumour formation. VEGF is an inducer of tumour angiogenesis (Kim *et al.* 1993; Saleh *et al.* 1996) and is thought to be crucial for the angiogenesis which supports the female reproductive system (Ferrara *et al.* 1998). *VEGF* gene expression is upregulated within solid tumours, in cells adjacent to necrotic regions, probably in response to hypoxia (Shweiki *et al.* 1992; Plate *et al.* 1993a, b). Secretion of VEGF in central, hypoxic regions of a tumour establishes a concentration gradient of this mitogen across the tumour which serves to attract the growth of blood vessels. Blood vessels in the vicinity of the tumour can respond to VEGF because tumours

induce expression of VEGFR-1 and VEGFR-2 in the endothelial cells of nearby vessels (Plate et al. 1993a). The paracrine system involving VEGF and its receptors has been blocked in tumour models by use of neutralizing VEGF antibodies (Kim et al. 1993), overexpression of a dominant negative mutant of VEGFR-2 to block receptor activation (Millauer et al. 1994; 1996) and expression of VEGF antisense RNA in tumour cells to block VEGF synthesis (Saleh et al. 1996). In each case, vascularization of most tumours tested was severely impaired leading to drastic reductions in the rate of tumour growth. However, some tumours did not respond to approaches designed to block the VEGF/VEGFR-2 system, suggesting that these tumours utilize other angiogenic factors and/or receptors to induce angiogenesis (Millauer et al. 1996). Clinical trials are currently underway to test numerous compounds which block VEGF action as antitumour agents.

The induction of *VEGF* gene expression by hypoxia in tumour cells involves both an increase in the rate of gene transcription, mediated by the transcription factor hypoxia-inducible factor-1 (Forsythe *et al.* 1996), and enhancement of the stability of VEGF mRNA (Ikeda *et al.* 1995). These mechanisms may serve to stimulate angiogenesis during embryogenesis as well as tumour development. Interestingly, VEGF mRNA is stabilized in the absence of hypoxia by inactivation of the von Hippel-Lindau protein (pVHL), a tumour surpressor protein (Gnarra *et al.* 1996; Iliopoulos *et al.* 1996). Inactivation of pVHL can lead to formation of haemangioblastomas.

Therapeutic angiogenesis. The capacity of VEGF to induce angiogenesis suggests that this protein could be used to augment collateral vessel formation as an alternative to reconstructive surgery for treatment of disorders involving inadequate tissue perfusion. This notion is supported by the finding that treatment with $VEGF_{165}$ or DNA encoding $VEGF_{165}$ augmented collateral vessel formation in a rabbit model of acute limb ischemia (Takeshita *et al.* 1994; Tsurumi *et al.* 1997).

PIGF

PIGF is a disulphide-linked, homodimeric glycoprotein which exhibits approximately 46% identity in amino acid sequence to VEGF (Maglione *et al.* 1991), binds and activates VEGFR-1 but not VEGFR-2 (Figure 2) (Park *et al.* 1994; Sawano *et al.* 1996) and can form heterodimers with VEGF (DiSalvo *et al.* 1995; Cao *et al.* 1996a). Three isoforms of PIGF have been characterized (Maglione *et al.* 1993; Cao *et al.* 1997), only one of which, PIGF-2,

binds to heparin (Hauser & Weich 1993; Park *et al.* 1994; Cao *et al.* 1997). The multiple forms of PIGF are generated by alternative splicing of PIGF RNA derived from a single gene (Maglione *et al.* 1993; Cao *et al.* 1997). The *PIGF* gene is predominantly expressed in placenta (Maglione *et al.* 1993). PIGF homodimers are generally thought to be very poorly mitogenic for endothelial cells *in vitro* and extremely weak inducers of angiogenesis and vascular permeability in comparison to VEGF (Park *et al.* 1994; DiSalvo *et al.* 1995; Cao *et al.* 1996b; Kurz *et al.* 1998). However, PIGF-1 homodimers have been described as strongly angiogenic and mitogenic for endothelial cells in one report (Ziche *et al.* 1997).

In addition to forming homodimers, PIGF can form heterodimers with VEGF which are angiogenic and mitogenic for endothelial cells, although not as potently so as VEGF homodimers (DiSalvo *et al.* 1995; Cao *et al.* 1996a, b). Synthesis of VEGF/PIGF heterodimers *in vivo* would require colocalization of expression of the *VEGF* and *PIGF* genes. Such colocalization occurs in the trophoblastic giant cells associated with the parietal yolk sac at early stages of embryogenesis (Achen *et al.* 1997). Despite characterization of PIGF bioactivities and patterns of gene expression, the biological function of this VEGF family member is still unclear. Analysis of mutant mice deficient in PIGF may be required to define PIGF function.

VEGF-B

VEGF-B, also known as VEGF-related factor (VRF), is approximately 44% identical in amino acid sequence to VEGF, binds to VEGFR-1 (Dr B. Olofsson *et al.* unpublished observation) (Figure 2), forms disulphide-linked homodimers and exists as two isoforms consisting of 167 and 186 amino acids which arise due to alternative RNA splicing (Grimmond *et al.* 1996; Olofsson *et al.* 1996a, b). These isoforms differ in sequence only in the C-terminal region. VEGF-B₁₆₇ binds to heparin and remains predominantly cell-associated whereas VEGF-B₁₈₆ is freely secreted from cells (Olofsson *et al.* 1996a, b). It has been reported that VEGF-B₁₆₇ is mitogenic for endothelial cells *in vitro* (Olofsson *et al.* 1996a).

The VEGF-B gene is strongly expressed in the developing heart during embryogenesis and in adult cardiac and skeletal muscle (Grimmond *et al.* 1996; Olofsson *et al.* 1996a, b). The pattern of VEGF-B gene expression overlaps with that for VEGF gene expression in numerous tissues, which is noteworthy because both isoforms of VEGF-B can form heterodimers with VEGF (Olofsson *et al.* 1996a, b). VEGF-B gene expression is

^{© 1998} Blackwell Science Ltd, International Journal of Experimental Pathology, 79, 255-265

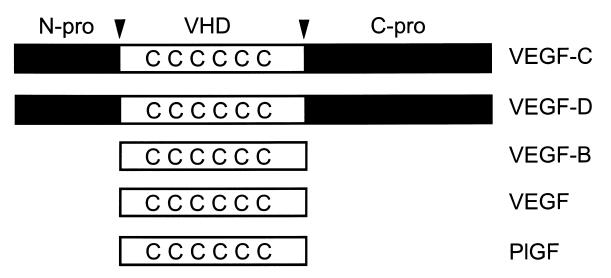


Figure 3. Schematic alignment of the primary structures of VEGF family members. VHD denotes the VEGF homology domain. Nand C-terminal regions of VEGF-C and VEGF-D, which are proteolytically cleaved from the VEGF homology domain, are designated N-pro and C-pro, respectively. Arrowheads denote the sites of proteolytic cleavage in VEGF-C and VEGF-D. Conserved cysteine residues in the VEGF homology domain, which constitute the cystine knot motif, are marked C. The positions of these residues are not shown accurately. The shortest isoforms of VEGF-B, VEGF and PIGF, which each consist essentially of a VEGF homology domain, are depicted here. Longer isoforms of each of these proteins exist which have extensions near their C-termini.

not induced by hypoxia (Enholm *et al.* 1997). It has been proposed that VEGF-B may play a role in regulating the vascularization of adult and embryonic tissues, in particular of muscle (Olofsson *et al.* 1996a).

VEGF-C

VEGF-C, also known as VEGF-related protein (VRP), was originally reported as a ligand for the tyrosine kinase VEGFR-3 (Flt4) (Joukov et al. 1996; Lee et al. 1996), a receptor similar in domain structure to VEGFR-1 and VEGFR-2 (Galland et al. 1993), but which does not bind VEGF or PIGF (Figure 2) (Pajusola et al. 1994; Lee et al. 1996). The amino acid sequence of VEGF-C has a central region, designated the VEGF homology domain, which is related to other members of the VEGF family, contains the cystine knot motif and exhibits approximately 30% identity to VEGF. In addition, the VEGF-C sequence has N-terminal and C-terminal extensions which are not present in VEGF, PIGF or VEGF-B (Figure 3) (Joukov et al. 1996; Lee et al. 1996). VEGF-C induces vascular permeability and is mitogenic for endothelial cells in vitro, although less potently so than VEGF (Joukov et al. 1997). VEGF-C gene expression is induced by numerous proinflammatory cytokines, but not by hypoxia (Enholm et al. 1997; Ristimaki et al. 1998).

The VEGF-C receptor VEGFR-3 is expressed in endothelial cell precursors in day 8.5 mouse embryos and later in development is expressed in venous and lymphatic endothelium (Kaipainen et al. 1995). The pattern of VEGF-C gene expression in relation to that for the VEGFR-3 gene during the sprouting of the lymphatic endothelium suggests that these molecules constitute a paracrine system which regulates angiogenesis of the lymphatic vasculature during embryonic development (Kukk et al. 1996). This hypothesis was supported by the findings that VEGF-C is lymphangiogenic in the avian chorioallantoic membrane (CAM) model (Oh et al. 1997) and that overexpression of VEGF-C in the skin of transgenic mice caused lymphatic, but not vascular, endothelial proliferation and vessel enlargement (Jeltsch et al. 1997). The observations that VEGF-C is lymphangiogenic and that VEGF-C gene expression is induced by proinflammatory cytokines suggests that such cytokines can modulate lymphatic vessel growth via VEGF-C and thereby regulate the composition and pressure of interstitial fluid and facilitate lymphocyte trafficking (Ristimaki et al. 1998).

VEGF-C is also a ligand for VEGFR-2 (Figure 2) (Joukov *et al.* 1996), but the functional significance of this potential interaction *in vivo* is unknown. VEGF-C at high concentrations induces a very mild angiogenic response in the CAM, which may be mediated by VEGFR-2, however, the lymphangiogenic response to VEGF-C is far more striking (Oh *et al.* 1997). A role for

VEGF-C in vasculogenesis in the early embryo has been proposed based on the hypothesis that VEGF-C may be responsible for inducing the VEGFR-2-mediated endothelial cell differentiation which occurs in VEGF-deficient mice (Eichmann *et al.* 1998). VEGF-C does not bind to VEGFR-1 (Lee *et al.* 1996; Joukov *et al.* 1997).

The biosynthesis of VEGF-C involves proteolytic processing that gives rise to a mature, secreted protein which essentially consists of the VEGF-homology domain (Figure 3) (Joukov et al. 1997). Therefore VEGF-C is initially synthesized as a precursor protein with the N- and C-terminal amino acid extensions mentioned above being propeptides. Proteolytic processing regulates the receptor specificity of VEGF-C, as stepwise proteolytic processing generates several VEGF-C forms with increasing activity towards VEGFR-3, whereas only fully processed VEGF-C activates VEGFR-2 (Joukov et al. 1997). VEGF-C forms mostly noncovalent homodimers in contrast to VEGF, PIGF and VEGF-B which form disulphide-linked dimers (Joukov et al. 1997). This finding was surprising given that the two cysteine residues of VEGF, which are involved in the intersubunit disulphide bonds, are conserved in VEGF-C (Joukov et al. 1997; Muller et al. 1997b).

VEGF-D

VEGF-D was first reported as a 'c-fos-induced growth factor' (FIGF) (Orlandini et al. 1996), but was subsequently designated VEGF-D based on the functional characteristics of the protein (Achen et al. 1998). VEGF-D is closely related to VEGF-C in primary structure, having a central VEGF homology domain and long N-and C-terminal extensions which are not found in other VEGF family members (Figure 3). VEGF-D and VEGF-C share 31% amino acid identity. The VEGF homology domain of VEGF-D is more similar to that of VEGF-C (61% identity in amino acid sequence) than to the other VEGF family members. Intriguingly, the C-terminal region of VEGF-D is rich in cysteine residues, many of which are located such that they resemble the spacing of the repeat units (CysX₁₀CysXCysXCys) which are found in the Balbiani ring 3 protein (BR3P), a major cysteine-rich protein synthesized in the larval salivary glands of the midge Chironomus tentans (Dignam & Case 1990; Achen et al. 1998). VEGF-D may interact with membrane-bound proteins via the cysteine residues as such intermolecular interactions have been proposed for BR3P (Paulsson et al. 1990). BR3P-like cysteine repeats are also found in the C-terminal region of VEGF-C (Joukov et al. 1996; Lee et al. 1996). VEGF-D exhibits similar receptor-binding specificities to VEGF-C, as it binds to and activates both VEGFR-2 and VEGFR-3 (Figure 2) (Achen *et al.* 1998). As is the case for VEGF-C, the capacity of VEGF-D to bind to these receptors is associated with the VEGF homology domain. The biosynthesis of VEGF-D involves similar proteolytic processing to that which generates mature VEGF-C (Stacker and Achen, manuscript in preparation). The similarities in structure, processing and receptor specificities between VEGF-C and VEGF-D demonstrate the existence of a subfamily of the vascular endothelial growth factors which has VEGF-C and VEGF-D as founding members.

The distributions of the VEGF-D receptors VEGFR-2 and VEGFR-3 on vascular and lymphatic endothelial cells during embryonic development suggest that VEGF-D may play a role in attracting the growth of these vessels into developing tissues. The finding that VEGF-D is mitogenic for endothelial cells *in vitro* is consistent with this hypothesis. In the light of these observations, it is noteworthy that the *VEGF-D* gene is strongly expressed in the developing lung during embryogenesis (Yamada *et al.* 1997). In the adult human, VEGF-D transcripts are found in many tissues but are most abundant in lung and heart (Yamada *et al.* 1997; Achen *et al.* 1998).

The expression of the gene for VEGF-D is induced by the transcription factor c-fos (Orlandini *et al.* 1996). Constitutive, ubiquitous expression of c-fos in transgenic mice induced formation of osteosarcomas (Grigoriadis *et al.* 1995) and tumours generated from c-fos-deficient cells failed to undergo malignant progression (Saez *et al.* 1995). Such experiments have indicated an essential role for c-fos in malignant tumour progression. It will be important to determine whether high levels of *VEGF-D* gene expression occur in tumours that overexpress c-fos and whether or not VEGF-D can contribute to the malignant tumour phenotype by promoting vascular and/or lymphatic angiogenesis.

Viral VEGFs

VEGF-like molecules have been described from the orf virus, a member of the Poxvirus family which infects sheep, goats and humans (Lyttle *et al.* 1994). Infection with orf virus causes a pustular dermititis which is characterized by capillary proliferation, dilation and swelling of the dermus (Groves *et al.* 1991). Two forms of VEGF-like molecules have been identified from orf viruses, OV-VEGF2 and OV-VEGF7, which are most closely related in primary structure to VEGF. OV-VEGF2 and OV-VEGF7 exhibit 29% and 23% amino

^{© 1998} Blackwell Science Ltd, International Journal of Experimental Pathology, 79, 255-265

acid identity with human $VEGF_{121}$, respectively. These proteins possess a VEGF homology domain in which all six cysteine residues of the cystine-knot motif are conserved.

The pathology of orf virus infection may well be explained by the activity of the VEGF-like molecules, as the infection appears to involve both induction of endothelial cell proliferation and enhancement of vascular permeability, well-characterized activities of VEGF. This hypothesis is supported by the finding that lesions resulting from infection by orf virus mutants in which the VEGF-like gene has been deleted are substantially less vascularised than are lesions associated with wild type orf virus (Drs L. Savory, S. B. Fleming, D. J. Lyttle & A. A. Mercer, personal communication). In addition, it has been shown that the VEGF-like proteins activate at least one of the VEGF receptors (Drs S. A. Stacker, L. Savory, S. B. Fleming & A. A. Mercer, unpublished observation).

Conclusion

Over the past 20 years many of the molecular mechanisms which direct the development of the haematopoietic system have been characterized (Metcalf 1989). This work has led to a deep understanding of how the diversity of blood cell lineages is generated. We are beginning to see a similar situation emerge for the development of the endothelium as key regulators of vasculogenesis, angiogenesis and lymphangiogenesis are identified. Members of the VEGF family are prominent among these regulators. Much work remains to be done, in particular defining the differences between embryonic and adult angiogenesis and determining the mechanisms which guide the formation of large vessels as opposed to microvessels and the differentiation of tissue-specific forms of endothelial cells. Nevertheless, our current knowledge of the biological functions of VEGF family members may be sufficient to allow us to manipulate the growth of blood vessels in tissue to the benefit of patients in the clinic. To this end, VEGF family members and inhibitors thereof are now in phase I/II clinical trials.

The capacity to inhibit blood vessel growth in the clinic will be of importance for the treatment of cancer, rheumatoid disease and retinopathies. Stimulation of blood vessel growth will be useful for treatment of myocardial and lower limb ischemia and will facilitate wound healing, skin grafting and tissue engineering. The emerging understanding of the molecular mechanisms which regulate development of the vascular tree will have a major impact on modern medicine.

Acknowledgements

We thank many colleagues who made experimental results available to us for this review prior to publication elsewhere.

References

- ACHEN M.G., GAD J.M., STACKER, S.A. & WILKS, A.F. (1997) Placenta growth factor and vascular endothelial growth factor are co-expressed during early embryonic development. *Growth Factors* **15**, 69–80.
- ACHEN M.G., JELTSCH M., KUKK E., MÄKINEN T., VITALI A., WILKS A.F., ALITALO, K. & STACKER, S.A. (1998) Vascular endothelial growth factor D (VEGF-D) is a ligand for the tyrosine kinases VEGF receptor 2 (Flk-1) and VEGF receptor 3 (Flt-4). *Proc. Natl. Acad. Sci. USA* **95**, 548–553.
- ALON T., HEMO I., ITIN A., PE'ER J., STONE, J. & KESHET, E. (1995) Vascular endothelial growth factor acts as a survival factor for newly formed retinal vessels and has implications for retinopathy of prematurity. *Nature Med.* **1**, 1024–1028.
- Asahara T., Murohara T., Sullivan A., Silver M., Van Der Zee R., Li T., Witzenbichler B., Schatteman, G. & Isner, J.M. (1997) Isolation of putative progenitor endothelial cells for angiogenesis. *Science* **275**, 964–967.
- BENJAMIN, L.E. & KESHET, E. (1997) Conditional switching of vascular endothelial growth factor (VEGF) expression in tumors: Induction of endothelial cell shedding and regression of hemangioblastoma-like vessels by VEGF withdrawal. *Proc. Natl. Acad. Sci. USA* **94**, 8761–8766.
- BREIER G., ALBRECHT U., STERRER, S. & RISAU, W. (1992) Expression of vascular endothelial growth factor during embryonic angiogenesis and endothelial cell differentiation. *Development* **114**, 521–532.
- BREIER G., CLAUSS, M. & RISAU, W. (1995) Coordinate expression of vascular endothelial growth factor receptor-1 (flt-1) and its ligand suggests a paracrine regulation of murine vascular development. *Dev. Dynamics* **204**, 228–239.
- CAO Y., CHEN H., ZHOU L., CHIANG M., ANAND-APTE B., WEATHERBEE J.A., WANG Y., FANG F., FLANAGAN, J.G. & TSANG M.L. (1996a) Heterodimers of placenta growth factor/vascular endothelial growth factor. Endothelial activity, tumor cell expression, and high affinity binding to Flk-1/KDR. *J. Biol. Chem.* 271, 3154–3162.
- CAO Y., LINDEN P., SHIMA D., BROWNE, F. & FOLKMAN J. (1996b) In vivo angiogenic activity and hypoxia induction of heterodimers of placenta growth factor/vascular endothelial growth factor. *J. Clin. Invest.* **98**, 2507–2511.
- CAO Y., JI W.R., QI P., ROSIN, A. & CAO, Y. (1997) Placenta growth factor: identification and characterization of a novel isoform generated by RNA alternative splicing. *Biochem.*. *Biophys. Res. Commun.* 235, 493–498.
- CARMELIET P., FERREIRA V., BREIER G., POLLOFEYT S., KEICKENS L., GERTENSTEIN M., FAHRIG M., VANDENHOECK A., HARPAL K., EBER-HARDT C., DECLERQ C., PAWLING J., MOONS L., COLLEN D., RISAU, W. & NAGY, A. (1996) Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. *Nature* 380, 435–439.
- CHOI K., KENNEDY M., KAZAROV A., PAPADIMITRIOU, J.C. & KELLER, G. (1998) A common precursor for hematopoietic and endothelial cells. *Development* **125**, 725–732.

- CONNOLLY D.T., OLANDER J.V., HEUVELMAN D., NELSON R., MONSELL R., SIEGEL N., HAYMORE B.L., LEIMGRUBER, R. & FEDER, J. (1989) Human vascular permeability factor. Isolation from U937 cells. *J. Biol. Chem.* **264**, 20017–20024.
- DE VRIES C., ESCOBEDO J.A., UENO H., HOUCK K., FERRARA, N. & WILLIAMS, L.T. (1992) The fms-like tyrosine kinase, a receptor for vascular endothelial growth factor. *Science* 255, 989–991.
- DIGNAM, S.S. & CASE, S.T. (1990) Balbiani ring 3 in Chironomus tentans encodes a 185-kDa secretory protein which is synthesized throughout the fourth larval instar. *Gene* **88**, 133–140.
- DISALVO J., BAYNE M.L., CONN G., KWOK P.W., TRIVEDI P.G., SODERMAN D.D., PALISI T.M., SULLIVAN, K.A. & THOMAS, K.A. (1995) Purification and characterization of a naturally occurring vascular endothelial growth factor.placenta growth factor heterodimer. J. Biol. Chem. 270, 7717–7723.
- EICHMANN A., CORBEL C., JAFFREDO T., BRÉANT C., JOUKOV V., KUMAR V., ALITALO, K. & L.E.DOUARIN, N.M. (1998) Avian VEGF-C: cloning, embryonic expression pattern and stimulation of the differentiation of VEGFR2-expressing endothelial cell precursors. *Development* **125**, 743–752.
- ENHOLM B., PAAVONEN K., RISTIMÄKI A., KUMAR V., GUNJI Y., KLEFSTROM J., KIVINEN L., LAIHO M., OLOFSSON B., JOUKOV V., ERIKSSON, U. & ALITALO, K. (1997) Comparison of VEGF, VEGF-B, VEGF-C and Ang-1 mRNA regulation by serum, growth factors, oncoproteins and hypoxia. *Oncogene* 14, 2475–2483.
- FERRARA N., CARVER-MOORE K., CHEN H., DOWD M., LU L., O'SHEA K.S., POWEL-BRAXTON L., HILLAN, K.J. & MOORE, M.W. (1996) Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature* **380**, 439–443.
- FERRARA N., CHEN H., DAVIS-SMYTH T., GERBER H.-P., NGUYEN T.-N., PEERS D., CHISHOLM V., HILLAN, K.J. & SCHWALL, R.H. (1998) Vascular endothelial growth factor is essential for corpus luteum angiogenesis. *Nature Med.* **4**, 336–340.
- FERRARA, N. & HENZEL, W.J. (1989) Pituitary follicular cells secrete a novel heparin-binding growth factor specific for vascular endothelial cells. *Biochem. Biophys. Res. Commun.* 161, 851–858.
- FOLKMAN, J. & SHING, Y. (1992) Angiogenesis. J. Biol. Chem. 267, 10931–10934.
- Fong G.-H., Rossant J., GERTSENSTEIN, M. & BREITMAN, M.L. (1995) Role of the Flt- receptor tyrosine kinase in regulating the assembly of vascular endothelium. *Nature* **376**, 66–70.
- FORSYTHE J.A., JIANG B.-H., IYER N.V., AGANI F., LEUNG S.W., KOOS, R.D. & SEMENZA, G.L. (1996) Activation of vascular endothelial growth factor gene transcription by hypoxiainducible factor.-1. *Mol. Cell. Biol.* **16**, 4604–4613.
- GALLAND F., KARAMYSHEVA A., PEBUSQUE M., BORG J., ROTTAPEL R., DUBREUIL P., ROSNET, O. & BIRNBAUM, D. (1993) The *FLT4* gene encodes a transmembrane tyrosine kinase related to the vascular endothelial growth factor receptor. *Oncogene* **8**, 1233–1240.
- GNARRA J.R., ZHOU S., MERRILL M.J., WAGNER J.R., KRUMM A., PAPAVASSILIOU E., OLDFIELD E.H., KLAUSNER, R.D. & LINEHAN, W.M. (1996) Post-transcriptional regulation of vascular endothelial growth factor mRNA by the product of the VHL tumor suppressor gene. *Proc. Natl. Acad. Sci. USA* 93, 10589–10594.
- GRIGORIADIS A.E., WANG, Z.Q. & WAGNER, E.F. (1995) Fos and bone cell development: lessons from a nuclear oncogene. *Trends. Genet.* **11**, 436–441.

- GRIMMOND S., LAGERCRANTZ J., DRINKWATER C., SILINS G., TOWNSON S., POLLOCK P., GOTLEY D., CARSON E., RAKAR S., NORDENSKJÖLD M., WARD L., HAYWARD, N. & WEBER, G. (1996) Cloning and characterization of a novel human gene related to vascular endothelial growth factor. *Genome Res.* 6, 124–131.
- GROVES R.W., WILSON-JONES, E. & MACDONALD, D.M. (1991) Human orf and milkers' nodule: a clinicopathologic study. *J. Am. Acad. Dermatol.* **25**, 706–711.
- HAUSER, S. & WEICH, H. (1993) A heparin-binding form of placenta growth factor (PIGF-2) is expressed in human umbilical vein endothelial cells and in placenta. *Growth Factors* **9**, 259–268.
- He, Z. & TESSIER-LAVIGNE, M. (1997) Neuropilin is a receptor for the axonal chemorepellent semaphorin III. *Cell* **90**, 739–751.
- HOUCK K.A., FERRARA N., WINER J., CACHIANES G., L.I.B. & LEUNG, D.W. (1991) The vascular endothelial growth factor family: identification of a fourth molecular species and characterization of alternative splicing of RNA. *Mol. Endocrinol.* 5, 1806–1814.
- HOUCK K.A., LEUNG D.W., ROWLAND A.M., WINER, J. & FERRARA, N. (1992) Dual regulation of vascular endothelial growth factor bioavailability by genetic and proteolytic mechanisms. *J. Biol. Chem.* 267, 26031–26037.
- IKEDA E., ACHEN M.G., BREIER, G. & RISAU, W. (1995) Hypoxiainduced transcriptional activation and increased mRNA stability of vascular endothelial growth factor in C6 glioma cells. *J. Biol. Chem.* 270, 19761–19766.
- ILIOPOULOS O., LEVY A.P., JIANG C., KAELIN J.R.W.G. & GOLDBERG, M.A. (1996) Negative regulation of hypoxia-inducible genes by the von Hippel-Lindau protein. *Proc. Natl. Acad. Sci. USA* **93**, 10595–10599.
- JAKEMAN L.B., ARMANINI M., PHILLIPS, H.S. & FERRARA, N. (1993) Developmental expression of binding sites and messenger ribonucleic acid for vascular endothelial growth factor suggests a role for this protein in vasculogenesis and angiogenesis. *Endocrinology* **133**, 848–859.
- JELTSCH M., KAIPAINEN A., JOUKOV V., MENG X., LAKSO M., RAUVALA H., SWARTZ M., FUKUMURA D., JAIN, R.K. & ALITALO, K. (1997) Hyperplasia of lymphatic vessels in VEGF-C transgenic mice. *Science* **276**, 1423–1425.
- JOUKOV V., PAJUSOLA K., KAIPAINEN A., CHILOV D., LAHTINEN I., KUKK E., SAKSELA O., KALKKINEN, N. & ALITALO, K. (1996) A novel vascular endothelial growth factor, VEGF-C, is a ligand for the Flt-4 (VEGFR-3) and KDR (VEGFR-2) receptor tyrosine kinases. *EMBO J.* **15**, 290–298.
- JOUKOV V., SORSA T., KUMAR V., JELTSCH M., CLAESSON-WELSH L., CAO Y., SAKSELA O., KALKKINEN, N. & ALITALO, K. (1997) Proteolytic processing regulates receptor specificity and activity of VEGF.-C. *EMBO J.* **16**, 3898–3911.
- KAIPAINEN A., KORHONEN J., MUSTONEN T., VAN HINSBERGH V.W., FANG G.H., DUMONT D., BREITMAN, M. & ALITALO, K. (1995) Expression of the fms-like tyrosine kinase 4 gene becomes restricted to lymphatic endothelium during development. *Proc. Natl. Acad. Sci. USA* 92, 3566–3570.
- KECK P.J., HAUSER S.D., KRIVI G., SANZO K., WARREN T., FEDER, J. & CONNOLLY, D.T. (1989) Vascular permeability factor, an endothelial cell mitogen related to PDGF. *Science* 246, 1309–1312.
- KIM K.J., LI B., WINER J., ARMANINI M., GILLETT N., PHILLIPS, H.S. & FERRARA, N. (1993) Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumour growth in vivo. *Nature* **362**, 841–844.

^{© 1998} Blackwell Science Ltd, International Journal of Experimental Pathology, 79, 255-265

- KOLODKIN A.L., LEVENGOOD D.V., ROWE E.G., TAI Y.-T., GIGER, R.J. & GINTY, D.D. (1997) Neuropilin is a semaphorin III receptor. *Cell* **90**, 753–762.
- KUKK E., LYMBOUSSAKI A., TAIRA S., KAIPAINEN A., JELTSCH M., JOUKOV, V. & ALITALO, K. (1996) VEGF-C receptor binding and pattern of expression with VEGFR-3 suggests a role in lymphatic vascular development. *Development* **122**, 3829–3837.
- KURZ H., WILTING J., SANDAU, K. & CHRIST, B. (1998) Automated evaluation of angiogenic effects mediated by VEGF and PIGF homo- and heterodimers. *Microvasc. Res.* 55, 92–102.
- LEAK, L.V. (1970) Electron microscopic observations on lymphatic capillaries and the structural components of the connective tissue–lymph interface. *Microvasc. Res.* **2**, 361–391.
- LEE J., GRAY A., YUAN J., LUOH S.M., AVRAHAM, H. & WOOD, W.I. (1996) Vascular endothelial growth factor-related protein: a ligand and specific activator of the tyrosine kinase receptor Flt4. *Proc. Natl. Acad. Sci. USA* **93**, 1988–1992.
- LEUNG D.W., CACHIANES G., KUANG W.J., GOEDDEL, D.V. & FERRARA, N. (1989) Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science* **246**, 1306–1309.
- LYTTLE D.J., FRASER K.M., FLEMING S.B., MERCER, A.A. & ROBINSON, A.J. (1994) Homologs of vascular endothelial growth factor are encoded by the poxvirus orf virus. *J. Virol.* **68**, 84–92.
- MAGLIONE D., GUERRIERO V., VIGLIETTO G., DELLI B.O.V.I.P. & PERSICO, M.G. (1991) Isolation of a human placenta cDNA coding for a protein related to the vascular permeability factor. *Proc. Natl. Acad. Sci. USA* **88**, 9267–9271.
- MAGLIONE D., GUERRIERO V., VIGLIETTO G., FERRARO M., APRELIKOVA A., ALITALO K., DEL VECCHIO S., LEI K., CHOU, J.Y. & PERSICO, M.G. (1993) Two alternative mRNAs coding for the angiogenic factor, placenta growth factor (PIGF), are transcribed from a single gene of chromosome 14. *Oncogene* 8, 925–931.
- McDONALD, N.Q. & HENDRICKSON, W.A. (1993) A Structural Superfamily of Growth Factors Containing a Cystine Knot Motif. *Cell* **73**, 421–424.
- METCALF, D. (1989) The molecular control of cell division, differentiation commitment and maturation in haemopoietic cells. *Nature* **339**, 27–30.
- MILLAUER B., LONGHI M.P., PLATE K.H., SHAWVER L.K., RISAU W., ULLRICH, A. & STRAWN, L.M. (1996) Dominant-negative inhibition of FIk-1 suppresses the growth of many tumor types *in vivo. Cancer Res.* 56, 1615–1620.
- MILLAUER B., SHAWVER L.K., PLATE K.H., RISAU, W. & ULLRICH, A. (1994) Glioblastoma growth inhibited in vivo by a dominant negative Flk-1 mutant. *Nature* **367**, 576–579.
- MILLAUER B., WIZIGMANN-VOOS S., SCHNÜRCH H., MARTINEZ R., MøLLER N.P., RISAU, W. & ULLRICH, A. (1993) High affinity VEGF binding and developmental expression suggest Flk-1 as a major regulator of vasculogenesis and angiogenesis. *Cell* 72, 835–846.
- MULLER Y.A., CHRISTINGER H.W., KEYT, B.A. & DE VOS A.M. (1997a) The crystal structure of vascular endothelial growth factor (VEGF) refined to 1.93Å resolution: multiple copy flexibility and receptor binding. *Structure* **5**, 1325–1338.
- MULLER Y.A., LI B., CHRISTINGER H.W., WELLS J.A., CUNNINGHAM, B.C. & DE VOS A.M. (1997b) Vascular endothelial growth factor: Crystal structure and functional mapping of the kinase domain receptor binding site. *Proc. Natl. Acad. Sci.* USA 94, 7192–7197.

- OH S., JELTSCH M.M., BIRKENHÄGER R., MCCARTHY J.E.G., WEICH H.A., CHRIST B., ALITALO, K. & WILTING, J. (1997) VEGF and VEGF-C: specific induction of angiogenesis and lymphangiogenesis in the differentiated avian chorioallantoic membrane. *Dev. Biol.* 188, 96–109.
- OLOFSSON B., PAJUSOLA K., KAIPAINEN A., VON EULER G., JOUKOV V., SAKSELA O., ORPANA A., PETTERSSON R.F., ALITALO, K. & ERIKSSON U. (1996a) Vascular endothelial growth factor B, a novel growth factor for endothelial cells. *Proc. Natl. Acad. Sci. USA* **93**, 2576–2581.
- OLOFSSON B., PAJUSOLA K., VON EULER G., CHILOV D., ALITALO, K. & ERIKSSON U. (1996b) Genomic organization of the mouse and human genes for vascular endothelial growth factor B (VEGF-B) and characterization of a second splice isoform. *J. Biol. Chem.* **271**, 19310–19317.
- ORLANDINI M., MARCONCINI L., FERRUZZI, R. & OLIVIERO, S. (1996) Identification of a c-fos-induced gene that is related to the platelet-derived growth factor/vascular endothelial growth factor family. *Proc. Natl. Acad. Sci. USA* **93**, 11675–11680.
- PAJUSOLA K., APRELIKOVA O., PELICCI G., WEICH H., CLAESSON-WELSH, L. & ALITALO, K. (1994) Signalling properties of FLT4, a proteolytically processed receptor tyrosine kinase related to two VEGF receptors. *Oncogene* 9, 3545–3555.
- PARK J.E., KELLER, G..-A. & FERRARA, N. (1993) The vascular endothelial growth factor (VEGF) isoforms: Differential deposition into the subepithelial extracellular matrix and bioactivity of ECM-bound VEGF. *Mol. Biol. Cell* 4, 1317–1326.
- PARK J.E., CHEN H.H., WINER J., HOUCK, K.A. & FERRARA, N. (1994) Placenta growth factor. Potentiation of vascular endothelial growth factor bioactivity, in vitro and in vivo, and high affinity binding to Flt-1 but not to Flk-1/KDR. *J. Biol. Chem.* 269, 25646–25654.
- PATAN S., HAENNI, B. & BURRI, P.H. (1996) Implementation of intussusceptive microvascular growth in the chicken choriollantoic membrane. 1. Pillar formation by folding of the capillary wall. *Microvasc. Res.* **51**, 80–98.
- PAULSSON G., LENDAHL U., GALLI J., ERICSSON, C. & WIESLANDER, L. (1990) The Balbiani ring 3 gene in *Chironomus tentans* has a diverged repetitive structure split by many introns. *J. Mol. Biol.* 211, 331–349.
- PETERS K.G., DE VRIES, C. & WILLIAMS, L.T. (1993) Vascular endothelial growth factor receptor expression during embryogenesis and tissue repair suggests a role in endothelial differentiation and blood vessel growth. *Proc. Natl. Acad. Sci. USA* **90**, 8915–8919.
- PLATE K.H., BREIER G., MILLAUER B., ULLRICH, A. & RISAU W. (1993a) Up-regulation of vascular endothelial growth factor and its cognate receptors in a rat glioma model of tumor angiogenesis. *Cancer Res.* 53, 5822–5827.
- PLATE K.H., BREIER G., WEICH, H.A. & RISAU W. (1993b) Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas *in vivo*. *Nature* **359**, 845–848.
- POLTORAK Z., COHEN T., SIVAN R., KANDELIS Y., SPIRA G., VLODAVSKY I., KESHET, E. & NEUFLED, G. (1997) VEGF₁₄₅, a secreted vascular endothelial growth factor isoform that binds to extracellular matrix. *J. Biol. Chem.* **272**, 7151–7158.
- PÖTGENS A.J.G., LUBSEN N.H., VAN ALTENA M.C., VERMEULEN R., BAKKER A., SCHOENMAKERS J.G.G., RUITER, D.J. & DE WAAL R.M.W. (1994) Covalent dimerization of vascular permeability factor/vascular endothelial growth factor is essential for its biological activity. Evidence from cys to ser mutations. *J. Biol. Chem.* **269**, 32879–32885.

- QUINN T.P., PETERS K.G., DE VRIES C., FERRARA, N. & WILLIAMS, L.T. (1993) Fetal liver kinase 1 is a receptor for vascular endothelial growth factor and is selectively expressed in vascular endothelium. *Proc. Natl. Acad. Sci. USA* **90**, 7533–7537.
- RISAU, W. (1995) Differentiation of endothelium. FASEB J. 9, 926–933.
- RISAU, W. (1997) Mechanisms of angiogenesis. *Nature* **386**, 671–674.
- RISAU, W. & FLAMME, I. (1995) Vasculogenesis. Ann. Rev. Cell Dev. Biol. 11, 73–91.
- RISTIMAKI A., NARKO K., ENHOLM B., JOUKOV, V. & ALITALO, K. (1998) Proinflammatory cytokines regulate expression of the lymphatic endothelial mitogen vascular endothelial growth factor.-C. *J. Biol. Chem.* **273**, 8413–8418.
- SAEZ E., RUTBERG S.E., MUELLER E., OPPENHEIM H., SMOLUK J., YUSPA, S.H. & SPIEGELMAN, B.M. (1995) c-fos is required for malignant progression of skin tumors. *Cell* 82, 721–732.
- SALEH M., STACKER, S.A. & WILKS, A.F. (1996) Inhibition of growth of C6 glioma cells *in vivo* by expression of antisense vascular endothelial growth factor sequence. *Cancer Res.* 56, 393–401.
- SAWANO A., TAKAHASHI T., YAMAGUCHI S., AONUMA, M. & SHIBUYA, M. (1996) Flt-1 but not KDR/Flk-1 tyrosine kinase is a receptor for placenta growth factor, which is related to vascular endothelial growth factor. *Cell Growth Differentiation* 7, 213–221.
- SENGER D.R., GALLI S.J., DVORAK A.M., PERRUZZI C.A., HARVEY, V.S. & DVORAK, H.F. (1983) Tumor cells secrete a vascular permeability factor that promotes accumulation of ascites fluid. *Science* **219**, 983–985.
- SHALABY F., ROSSANT J., YAMAGUCHI T.P., GERTSENSTEIN M., WU X.F., BREITMAN, M.L. & SCHUH, A.C. (1995) Failure of

blood-island formation and vasculogenesis in flk-1-deficient mice. *Nature* **376**, 62–66.

- SHWEIKI D., ITIN A., SOFFER, D. & KESHET, E. (1992) Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. *Nature* **359**, 843–845.
- SOKER S., TAKASHIMA S., MIAO H.Q., NEUFELD, G. & KLAGSBRUN, M. (1998) Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor. *Cell* **92**, 735–745.
- TAKESHITA S., ZHENG L.P., BROGI E., KEARNEY M., PU L.Q., BUNTING S., FERRARA N., SYMES, J.F. & ISNER, J.M. (1994) Therapeutic angiogenesis. A single intraarterial bolus of vascular endothelial growth factor augments revascularization in a rabbit ischemic hind limb model. J. Clin. Invest. 93, 662–670.
- TISCHER E., MITCHELL R., HARTMAN T., SILVA M., GOSPODAROWICZ D., FIDDES, J.C. & ABRAHAM, J.A. (1991) The human gene for vascular endothelial growth factor. Multiple protein forms are encoded through alternative exon splicing. *J. Biol. Chem.* 266, 11947–11954.
- TSURUMI Y., KEARNEY M., CHEN D., SILVER M., TAKESHITA S., YANG J., SYMES, J.F. & ISNER, J.M. (1997) Treatment of acute limb ischemia by intramuscular injection of vascular endothelial growth factor gene. *Circulation* **96** (9, Suppl.), II-II3828.
- YAMADA Y., NEZU J., SHIMANE, M. & HIRATA, Y. (1997) Molecular cloning of a novel vascular endothelial growth factor, VEGF.-D. *Genomics* **42**, 483–488.
- ZICHE M., MAGLIONE D., RIBATTI D., MORBIDELLI L., LAGO C.T., BATTISTI M., PAOLETTI I., BARRA A., TUCCI M., PARISE G., VINCENTI V., GRANGER H.J., VIGLIETTO, G. & PERSICO, M.G. (1997) Placenta growth factor-1 is chemotactic, mitogenic, and angiogenic. *Lab. Invest.* **76**, 517–531.