The nucleotide sequence of a nematode vitellogenin gene

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ABSTRACT

The nematode, Caenorhabditis elegans, contains a family of six genes that code for vitellogenins. Here we report the complete nucleotide sequence of one of these genes, vit-5. The gene specifies a mRNA of 4869 nucleotides, including untranslated regions of 9 bases at the 5' end and 51 bases at the 3' end. Vit-5 contains four short introns totalling 218 bp. The predicted vitellogenin, yp17OA, has a molecular weight of 186,430. At its N terminus it is clearly related to the vitellogenins of vertebrates. However, the vit-5-encoded protein does not contain a serine-rich sequence related to the vertebrate vitellin, phosvitin. In fact, the amino acid composition of the nematode protein is very similar to that of the vertebrate protein without phosvitin. Vit-5 has a highly asymmetric codon choice dictionary. The favored codons are different from those favored in other organisms, but are characteristic of highly expressed C_0 elegans genes. The strong selection against rare codons is not as great near the 5' end of the gene; rare codons are 15 times more frequent within the first 54 bp than in the next 4.8 kb.

INTRODUCTION

Four distinct yolk proteins have been identified in the nematode, Caenorhabditis elegans (1). Two of these, ypl70A and yp170B, are translated as polypeptides estimated to be 170,000 daltons (2). These vitellogenins undergo little if any processing before they are packaged into yolk platelets. In contrast, the two smaller yolk proteins, yp115 and yp88, are cut from a single precursor polypeptide, VIT180 (3). ypl7OA and B and VIT 180 are synthesized only in the intestine of the adult hermaphrodite worm, from which they are cotranslationally secreted into the body cavity $(3, 4)$. There, VIT180 is cleaved and the yolk proteins are subsequently taken up into developing oocytes.

The vitellogenins are encoded by a small family of genes (5, 6). ypl70A is specified by $vit-3$, $vit-4$, and $vit-5$. These three genes are at least 95% homologous to one another at the nucleotide sequence level. Vit-5 mRNA is much more abundant than that from vit-3 and vit-4 (5; Cane, unpublished observations). ypl70B is specified by $vit-2$. Vit-1 is a pseudogene, about

95% identical to vit-2 (7). The vit-1,2 sub-family is about 70% homologous to the vit-3,4,5 sub-family. VIT180 is encoded by a very distantly related family member, vit-6 (6).

We have recently reported the nucleotide sequences of the regions surrounding the 5' ends of five of the C. elegans vitellogenin genes (7). Our results indicated that these genes were surprisingly closely related to the vitellogenin genes of vertebrates. Like the nematode genes, the vertebrate genes are expressed in a tissue of endodermal origin, the liver. Like ypl70A and ypl70B, the vertebrate vitellogenins are transported intact to the oocytes. However, unlike the nematode vitellogenins, the vertebrate proteins are cleaved in the oocyte to form vitellins: phosvitin and the lipovitellins (8). In this paper we report the complete nucleotide sequence of vit-5, the first member of the vertebrate/nematode vitellogenin gene family to be sequenced. Our data indicate that although the nematode gene is closely related to the vertebrate gene near the 5' end, the region encoding the serine-rich vertebrate protein, phosvitin, is not present in vit-5.

RESULTS AND DISCUSSION

Characteristics of vit-5

The complete nucleotide sequence of vit-5, along with 261 bp of 5' flanking and 27 bp of 3' flanking DNA, is shown in Fig. 1. The location of the 5' end of the mRNA was determined by primer extension using a synthesized 15 bp oligonucleotide primer, accompanied by dideoxy-sequencing. A single, unequivocal extension product was found (data not shown). Similar experiments were performed with vit-2, vit-4, and vit-6. In each case a single extension product, the same distance 3' from a canonical TATA box sequence was found. The 3' end of the gene was located by sequencing a 3' terminal cDNA clone containing a 50 base poly(A) stretch. The gene contains four short introns, totaling 218 bp. The presence of the two introns near the 3' end of the gene was directly demonstrated by sequencing cDNA clones which overlap this region. The presence of the other two introns is inferred: they contain canonical splice sites at their 5' and 3' boundaries (Table 1); they contain stop codons in all three reading frames; and they are at least 70% A + T. Excision of these proposed introns at the positions indicated allows translation utilizing primarily the highly favored codons typical of the C. elegans vitellogenin genes (see below).

We presume that translation begins at the first AUG in each mRNA. This assumption is supported by: 1) The presence of a highly hydrophobic amino

AAC AAB CCA SAB CTC CBT ATB BCT BCT CTT TBB ABA ATB ATB CAC ACC ATT CCA BAA GAA CCA BTT CTT BCT CAC ATC BTT TCC CAA ATB +1926 ^N KP EL RHNA ^A ^L ^W ^R NhH ^T ^I ^P ^E ^E ^P VLA ^H ^I ^V ^S BN BAA AAC 6AA TCC AAC CAA CAC BTT BCT 6CC TTC ACC TAC CAC BTC CTC CBC CAA TTC TAC AAA TCC ACC AAC CCA TBC TAC CAA CAA ITB +2016 E N E S N B U V A A FT Y U V I B B E Y K S T N P C Y B B I BCI BTT CST TBC TCT RAB ATC CIT CTC TTC ACC CST TAT CAA CCA CAA BAA CAB ATB CTC TCC ACC TAC TCC CAA CTT CCA CTT TTC AAC +2106 A V R C S K I L L F T R Y B P B E B M L S T Y S B L P L F TCT BAS 166 CTC TCC 66A BTI CAA TTC BAC TTC 6CC ACC ATT TIC BAB AA6 AAC BCT TTC TTB CCA AAB BAA BTT CAA BCA TCA TTB BAA +2196 S EWL S & V Q F D F A T I F E K N A F L P K E V Q A S L E ACC BTC TTC BBR 66A AAC TB6 AAC AAA TAC TIC BCT CAA BIT BBA TTC TCT CAA CAB AAC TTT BAB CAA BTC AIC CTC AAB ACC CTC BAA 42286 ^T VF 6B6NWkN ^K Y FAB9VB6F SBB N FEB9V ^I LIKTILE AAA CIT TCT CIT TAC 66A AA6 CAA TCT 6AT 6AA CTC C6T TCC C6T 6TC CAA TCT 66A ATC CAA AT6 CTT CAA 6A6 ATT 6TC AA6 AA6 +2376 K L S L Y 6 K B S D E L R S R R V B S 6 I B M L B E I V K K ATB AAC ATC CST CCA CGT BTC CAA CAA ACC BAT TCI CAA AAT BCT CAC BCI BIT TIC TAdC'CIT CBC TAd AAG GAB RIB BAC TAC ATC BIT +2466 ^M ^N IT ^R ^P ^R ^V BB ITD SB9N A HA VF Y LR YK ENhDY ^I ^V CIT CCA ATT BAC ATB BAA ACT ATT BAC ACT CIT BTT BAB AAG TAT BTC ABA AAC BBA 6AG TIT BAC ATC ATC CTC CTC CTC ACT TIC TTG +2556 L P ¹ D N E T ^I D ^I L V E K Y V R N B E F D .1 K S I L T F L ACC AAC SAC ICC AAB TIC BAB CTT CAC CGT BCT CTC TIC TIC TAC BASGBCT BAA CBC ABA ATT CCA ACA ACC AlT BGSA RIB CCA CTC ACC +2646 ^I ND SK ^F ^E LHR ^A ^L ^F ^F YE AE ^R ^R ^I ^P TT ^I ^B NPL ^T ATT TCT BBA AAB ATB CCA ACT ATC CTC TCT ATC AAC BBA AAB BIT TCA ATT BAB CTC BAB AAB CTT GBA BCT CBT CTT BTT CTT BAT ATC +2736 ^S 6K ^N PT ^I ^L SI NB6K VS ^I E LE KIB6A RL VL D ^I BIT CCA ACT BIT 6CC ACC ACC CAC BIC ACT BAG ATB CCB CIT CIB TAT CCA BIC All BAA CAA BBA B7C AAG TCA CTT CAR ICT BCT CST +2826 ^V ^P TIVA I ^T HVYT ENhPLL YPVY ^I EBB6V ^K SLBS A R CTC CAC ACT CCA ¹¹⁶ ABA TIC BAA TCA ACT BIT BAA ¹¹⁶ AAG MB MAC ACT CTC BAA ATC ACT CAC AAG ITT GTT BIC CCA GAG MAC AAB +2916 ^L ^H TP LR FE SIT ^E ^L KK ^N TLE IT ^H KF ^V YP EN ^K AAG ACC ACT 6TT TCC 6TT CAT ACC C6C CCA 6TT 6CT TTC ATC C6T 6TT CCA AAG AAC CAA GAC TCT 6AA TAT 6TT 6A6 6CT 6AA 6A6 AA6 +3006 KITT ^V ^S VH ^I ^R ^P ^V AF ^I ^R VPK N9BDS ^E ^Y ^V EA EE ^K ACT ATT TCC CAC TCA CAA TAC CAA ATB TCT ACT BAA BAB ATT BAT CBT CAA TAT BAB ACC TTT BBA CTC ABA ATC AAT BCC CAA BBA AAT +3096 119 ^H SB9 YB9 ^N S TE ElI DR ^B ^Y ET FBI ^R ^I ^N ^A ^B ^B ^N 611 CII fC CAA 166 ACT CIT CCA ATG BIT 116 ATB ACT BAA CAA BAT TIC BAG TAC ACT CII BAA MAC AAA AAC CGI CCA 611 BAG TIC +3186 VILSBNIIT ^P HVLN ^T EB0D ^F EY TLEN KN ^R PVYEF ACA GCT CGC BTC ACT ATT BSA AAC CTC GAG AAG ACT BAT CIT ICC BAG ATC AAB TTC BAC AAG ATC TIC BAA AAA BAA TTC BAC CIT GAG +3276 ^I ARV TII BRN L EKITDILS E ^I ^K FD ^K ^I ^F ^E ^K EF DILE AAC AAC GAA TCT GAG AAC CGC CGC CAA TAC TTC CAC AAG ATG ATC CGT GAG ATT CAA TCT GAG CAA GGA TTC AAG AAC CTC ATC ACC CTC +3366 N N E S E N R R B Y F H K M I R E I B S E B B F K N L I T L RAG CTI BAR 6CC CCA CAR CAR RIB TAC 166 ARC ACT BAR CTI CGT ACC GTC 161 BAC RAAA 1GB AId COT RIG TGC RAG BIT GAB RIG BAT +3456 K L E A P Q Q M Y W N T E L R T V C D K W I R M C K V E M D SCT CGC CGC TCT CCA AT6 GA6 CAC 6A6 AAC AAA 6AA T66 ACT CTT CGT ACT GA6 CTT CTT GCT 6CC CGC CCA CAA AT6 CCA TCC TCC +3546 A R R S P M E H E N K E W T L R T E L L A A R P Q M P S S L CGT CAA CTT CGT GAB CAA CCA CAC CGT GA6 GTT CAA CTC GCA TTC AAT GCC AAG TGG GGA TCA TCA AAG AAG AGC GA6 ATC ACA GTC AAT +3636 R B L R E B P H R E V B L A F N A K W B S S K K S E I T V N GCT CAR CTC BAR CAR ICC AC6GBAR CAR RAG RAG TIC ATC CGC MAC ATC BAG CST GAG TAC RAG 66A All CCA SGA TAd BAA CII TTB RTC +3726 ARBILE9S ^T EBK KF IR ^N IE RE ^Y KG6IP EY ELL ^I

AA6 6CT 6CT CST CTT AAC CAA 6TC AAT 6TC TCT 6A6 TAC AAG CTC CACC CAA CAGE ATT CAFE CAGE ATT TTC TCC C6C ATT TTC 6ACC CTT +3816
K A A R L N Q V N V V S E Y K L T P Q S E Y T F S R I F D L ATC AAB 6CA TAC AAC TTC T66 ACT 6TT TCT 6A6 AA6 CST 6TC CAA aaC 6A6 AaT CGC C6C 6TT 6TT CTT CAA CTT TCT 6TT BA6 CCA CTT +3906 I K A Y N F N T V S E K R V Q N E N R R V V L Q L S V F P L TCC C6C CAA TCA CAT 6AA CAT 6AC CAT CA6 ACT CCA 6AA CAA BAA 6TT BAG TT6 aA6 AAT 6CT CST ATT CCA c6A 6TC 6TT CTC CCA ACT +3996 ^S ^R Q ^S ^H ^E H ^D ^H ^Q ^T ^P ^E ^Q ^E ^V ^E ^L ^K ^N ^A ^R ^I P ^R ^V V ^L ^P ^T ATT BCT CST ABT 6CC AT6 TTC CAA CAA ACC T66 BAA AAB ACC 66A 6CC ACC T6C AA6 6TT 6AC CAA TCT GAB BTT TCT ACC TTT CAC AAC +4OB6 ^I A ^R ^S ^A 1 ^F GQ TT ^E ^K ^T ⁶ A ^T ^C ^K V D ^Q ^S ^E ^V ^S ^T ^H ^N BTB ATC TAC CBC BCT CCA CTC ACC ACC TBC TAC TCT CTT BTT BCC AA6 6AT TGC TCT BAA CA6 CCA AGA TTC BCT 6TT CTT BCC AA6 AA6 +4176 ^V ^I ^Y ^R ^A ^P ^L T ^C ^Y ^S ^L ^Y ^A ^K ^D ^C ^S ^E ^Q ^P ^R ^F ^A ^V ^L ^A ^K ^K ATC AAC AAB AAC TCT 6A6 BAG CTT CTC 6TT AA6 6Tt 6tC CBC CST GAG 6AA 6AA ATT BITI6B AM AAa6 TCT AC BAT AA6 TTC CTT 6TC +4266 I ^N ^K ^N ^S ^E ^E ^L ^L V ^K ^V ^V ^R ^R ^E ^E ^E ^I ^Y ^V ^K ^K ^S D ^D ^K ^F ^L ^V AAG 6TT SAC 66A AAG AAG 6TT AAC CCA ACT 6AA CTT 6AA CAA TAC AA gtaagctttaactctaaatcacatgaattttttctaatctcgtttttcag T ATC +4366 K V D 6 K K V N P T F I F O Y N BAA ATT CTT BBA BAT AAC CTT ATT BTT ATT CGT CTT CCA CAA 66A BAG 6TT CST TTC BAT 66A TAC ACT 6TC AAG ACC MAC ATG CCA TCC +4456 ^E ^I ^L ⁶ ^D ^N ^L ^I ^V ^I ^R ^L ^P ^Q ⁶ ^E ^V ^R ^F ^D ⁶ ^Y ^T ^V ^K ^T ^N A ^P ^S BTT GCT TCA CAA AAC CAA CTT TBC 66A CTT T6C 66A AAC aaT 6AC 66T 6A6 ABa 6AC AAT GA6 TTC ATG ACC 6CT GAC AAC TAC BAA ACT +4546 V *A S Q N Q L C 6 L C 6 N N D 6 E R D N E F 8. T A D N Y E T BAG BAT 6TT BAG 6AA TTC CAC C66 TCT TAC CTT CTC AA6 AAT BAB 6AA T6C 6A6 TTT GAG TTC 6AC CGC ATC TCC 6A6 AAB AA6 AAC TAC +4636 E D V E E F H R S Y L L K N E E C E F E F D R ^I S E K K N Y A6A AAC AAA T66 aaC AA 6AA BA6 AM M6 TCC 6AC TAC BA6 A6C ABC TCC 6AC TAC 6A6 A6C AAC TAC BAT ⁴⁶AG ⁶GAA ACT BAA BAG +4726 R N K N R E E K K S D Y E S S S D Y E S N Y D E K E T E E ⁶ gttagtctaaggctgaatgatgtgtagttttaacaaaatacatatttttcag AA CTC 6TC AA6 M6 ACC CTC ATC AA6 BAG TTC TCC aMC C6C 6TC T6C +4826 E L V K K T L ^I K E F S N R V C TTC TCC ATC 6A6 CCA 6TC TCT 6A6 T6C CBC CBT 66A CTC 6AA TCC 6A6 AA6 ACT TCC AAC AA6 AA6 ATC C6T TTC ACT T6C AT6 CCA C6T +4916 ^F ^S ^I ^E ^P ^V ^S ^E ^C ^R ^R ⁶ ^L ^E ^S ^E ^K ^T ^S ^N ^K ^K ^I ^R ^F ^T ^C N ^P ^R CAC A66 CAA 6AA CST AST CST TTT CTT CAA 66A A6C TCT 6A6 CAA ACT 6TT 6CC GAB 116 BTC 6AT TTC CCA 6TC TCC TTC 6TT 6A6 TCT +5006 ^H ^R Q ^E ^R ^S ^R ^F ^L ^Q ⁶ ^s ^s ^E ^Q ^T V ^a ^E ^L V ^D F ^P ^V ^S F ^Y ^E ^S 6TC AA6 ATC CCA ACC 6CC T6C 6TT 6CC TAT TA6 ATTTCATAT6TTTTATTAATTTTCTATTAAATAAA6CATTTTCACTA6aatgattttttttctcacacttctaga +5114 ^V ^K ^I ^P ^T A ^C ^V A ^Y

Figure 1. The nucleotide sequence of vit-5 and predicted amino acid sequence of ypl70a. The sequence was determined by the method of Sanger (20). Restriction fragments of genomic clone 2017 containing all of vit-5 (5) were subcloned into pUC8 from which random Sau3a fragments were cloned into mp8 and mp9 for sequencing. Sequences were aligned by comparison of restriction sites found by sequencing, with restriction maps of the sub-clones. Alignments were confirmed by sequencing predicted fragments which overlapped the boundaries of adjacent clones. Both strands were sequenced over most of the length of the gene. Portions of cDNA clone 1728 (5) were sequenced to confirm the predicted intron 3 and 4 boundaries. Numbering is from the first base in the mRNA, determined by premer extersion accompained by dideoxy sequencing. Introns and 5' and 3' untranslated region is double-underlined. Rare codons (as defined in the test) are underlined.

Intron/Exon Border Sequences					
	Intron 1 AAG GTATTCTTTTCAG GTT				
$\mathbf{2}$	AAG GTAACCCTCTTAG GTT				
3	CAA GTAAGCTTTTCAG TAT				
4	AGG GTTAGTTTTTCAG AAC				
C. elegans Consensus*	$AG GTAAG$ TTTTCAG $ _G^A$				

Table ¹

*The C. elegans consensus is taken from a compilation of 29 sequenced C. elegans intron borders (Blumenthal, unpublished).

acid sequence at the N-terminus of the proposed protein, which is probably the signal sequence responsible for cotranslational secretion. 2) The sequences of this region of the other vitellogenin genes show that all of the proteins would start with a very similar signal sequence if the first AUG in each mRNA is used for translation initiation (7). 3) This signal sequence would be homologous to signal sequences at the N-termini of vertebrate vitellogenins (9). 4) The AUG at the proposed site is preceded by G at position -3 and followed by A at position +4, and so should be favored for initiation (10). 5) The proposed site for initiation precedes the only long, open reading frame, one with a very asymmetric codon usage typical of abundantly-expressed C. elegans genes (11-14).

The 5' untranslated region of vit-5 is exceptionally short, only nine bases long. The other members of the vit-l-vit-5 group also have unusually short 5' untranslated regions, 9-11 bases long (7). Translation terminates at a UAG codon which is followed by an AAUAAA sequence 30 bases further on and the 3' end of the mRNA 18 bases later. Hence, of the 4869 bases in the vit-5 mRNA, only 60 are untranslated.

The vit-5 introns range in length from 47 to 70 base pairs. Only 218 base pairs of the total gene length of 5087 base pairs are in introns. Thus vit-5 is a very compact gene. Very little RNA is spliced out and almost all that remains is translated. This gene structure is typical of the C. elegans genes sequenced thus far (11-14). In contrast, the members of the vitellogenin gene family of vertebrates are about 25 kb in length and contain 33 introns (15, 16).

Codon Usage

The codon usage is very asymmetric (Table 2). In the 1603 codons of

	Amino			Amino			Amino			Amino	
Codon	Acid	Residues	Codon	Acid	Residues	Codon	Ac1d	Residues	Codon	Acid	Residues
UUU	F	11	UCU	s	51	UAU	Y	8	UGU	C	1
UUC	F	59	UCC	S	38	UAC	Y	45	UGC	C	19
			UCA	s	13						
UUA	L	ı	UCG	S	3	CAU	H	8	UGG	W	14
UUG	L	18	AGU	S	4	CAC	H	25			
cuu	L	76	AGC	s	9				CGU	$\mathbb R$	45
CUC	L	44				CAA	Q	90	$_{\rm ccc}$	R	34
CUA	L	0	CCU	P	0	CAG	Q	10	CGA	R	3
CUG	L	$\overline{2}$	$_{\rm ccc}$	P	0				CGG	R	ı
			CCA	P	61	AAU	N	17	AGA	R	14
AUU	I	40	$_{\rm ccc}$	p	2	AAC	N	56	AGG	R	ı
AUC	I	51									
AUA	I	1	ACU	T	43	AAA	K	16	GGU	G	2
			ACC	T	49	AAG	K	108	$_{\rm GGC}$	G	$\mathbf 0$
AUG	N	35	ACA	T	5				GGA	G	41
			ACG	T	ı	GAU	D	27	GGG	G	1
GUU	V	68				GAC	D	31			
GUC	٧	46	CCU	A	46						
GUA	V	1	$_{\rm{ccc}}$	A	28	GAA	E	66			
GUG	V	$\overline{2}$	GCA	A	5	GAG	E	106			
			CCG	A							

Table 2

vit-5, 19 codons are used three times or less, including four that are not used at all (CUA, CCU, CCC, GGC) and nine others that are used only once (UUA, AUA, GUA, ACG, GCG, UGU, CGG, AGG, GGG). In general, third position purines are rare. For instance, 114 out of 117 valine codons have third position pyrimidines. However, there are exceptions to this rule: 61 out of 63 proline codons are CCA; and 41 out of 44 glycine codons are GGA. This degree of asymmetry has been observed previously only in single-celled organisms (17, 18). An examination of published sequences of other abundantly expressed C. elegans genes, including actin (14), myosin (12), collagen (13), and the major sperm protein (11), reveals the same highly asymmetric codon usage.

Interestingly, the 19 rare codons occur at a much higher frequency near the beginning of the gene. While only 23 of 1603 total codons, or 1.4%, are rare codons, four of the first 18 codons, or 22 percent are rare codons, and two of the four occur nowhere else in the protein. We have also sequenced portions of the other vitellogenin genes (unpublished observations) and observed the same phenomenon: the same codons are rare, and each gene has several rare codons within the first 18 (a total of 16 rare codons in this region of the 5 vitellogenin genes for which sequence data is available). We do not know why rare codons are more acceptable near the 5' end of the gene. Assuming that rare codons are decoded by rare tRNAs and that selection is for rapid translation of abundantly expressed genes as has been shown to be the case in other systems (17, 18), we hypothesize there is no strong selection

Nucleic Acids Research

	Amino Acid Composition of Vitellogenins	rante o		
	yp170A	Xenopus	<u>Chick</u>	
$Asp + Asn$	131	148	154	
Thr	98	87	79	
Ser	118	171	236	
$Glu + Gln$	272	229	179	
Pro	63	84	79	
Gly	44	84	84	
Ala	80	135	126	
Val	117	102	102	
Met	35	42	39	
Ile	92	81	89	
Leu	141	141	138	
Tyr	53	50	48	
Phe	70	62	43	
His	33	60	55	
Lys	124	113	138	
Arg	98	87	118	
Cys	20	10	ND	
Trp	14	ND	ND	

Table 3

The amino acid composition of the nematode vitellogenin ypl70a is inferred from the DNA sequence of vit-5 (Fig. 1). The amino acid compositions of Xenopus and chicken vitellogenins were determined by amino acid analysis and are taken from Wiley and Wallace (21) and Wang et al. (22), respectively.

for rapid translation of the region of the mRNA encoding the signal sequence. Indeed the ubiquity of rare codons in this region of the five genes sequenced suggests there may be selection for slow translation of the region. Perhaps slow translation allows interaction between the nascent polypeptide and some component required for secretion.

Characteristics of ypl70A

Vit-5 encodes a polypeptide of 186,430 daltons. The predicted amino acid composition is shown in Table 3. The protein is very high in glutamic acid and lysine, as expected for a vitellogenin. Table 3 also presents the amino acid compositions of vitellogenins from Xenopus and from chicken (vitellogenin II) for comparison. On the whole the predicted amino acid composition of ypl70A is quite similar to the vertebrate compositions, but there are some notable differences. ypl70A has more glutamic acid + glutamine and less glycine, alanine and histidine. Most interestingly, yp170A has much less serine than do the vertebrate vitellogenins. In chicken vitellogenin, 123 of 236 serine residues are contained in the phosvitin region of the precursor (19). We have searched the ypl70A sequence for a phosvitin-like region and failed to find it. (In the "core" region of chickin phosvitin, 80 of 99 residues are serine). Thus it appears that phosvitin is missing from yp170A, even though at the N-terminus the protein is clearly related to the chicken vitellogenin (7), and overall, the amino acid compositions of the two proteins are quite similar (Table 3). Since almost all of the phosvitin sequence of chicken is contained in a single exon (19), it seems likely that this exon is missing from the C. elegans gene.

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