The nucleotide sequence of a nematode vitellogenin gene

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ABSTRACT

The nematode, Caenorhabditis elegans, contains a family of six genes that code for vitellogenins. Here we report the complete nucleotide sequence of one of these genes, vit-5. The gene specifies a mRNA of 4869 nucleotides, including untranslated regions of 9 bases at the 5' end and 51 bases at the 3' end. Vit-5 contains four short introns totalling 218 bp. The predicted vitellogenin, ypl70A, has a molecular weight of 186,430. At its N terminus it is clearly related to the vitellogenins of vertebrates. However, the vit-5-encoded protein does not contain a serine-rich sequence related to the vertebrate vitellin, phosvitin. In fact, the amino acid composition of the nematode protein is very similar to that of the vertebrate protein without phosvitin. Vit-5 has a highly asymmetric codon choice dictionary. The favored codons are different from those favored in other organisms, but are characteristic of highly expressed C. elegans genes. The strong selection against rare codons is not as great near the 5' end of the gene; rare codons are 15 times more frequent within the first 54 bp than in the next 4.8 kb.

INTRODUCTION

Four distinct yolk proteins have been identified in the nematode, <u>Caenorhabditis elegans</u> (1). Two of these, yp170A and yp170B, are translated as polypeptides estimated to be 170,000 daltons (2). These vitellogenins undergo little if any processing before they are packaged into yolk platelets. In contrast, the two smaller yolk proteins, yp115 and yp88, are cut from a single precursor polypeptide, VIT180 (3). yp170A and B and VIT 180 are synthesized only in the intestine of the adult hermaphrodite worm, from which they are cotranslationally secreted into the body cavity (3, 4). There, VIT180 is cleaved and the yolk proteins are subsequently taken up into developing oocytes.

The vitellogenins are encoded by a small family of genes (5, 6). ypl70A is specified by <u>vit-3</u>, <u>vit-4</u>, and <u>vit-5</u>. These three genes are at least 95% homologous to one another at the nucleotide sequence level. <u>Vit-5</u> mRNA is much more abundant than that from <u>vit-3</u> and <u>vit-4</u> (5; Cane, unpublished observations). ypl70B is specified by <u>vit-2</u>. <u>Vit-1</u> is a pseudogene, about

95% identical to <u>vit-2</u> (7). The <u>vit-1,2</u> sub-family is about 70% homologous to the <u>vit-3,4,5</u> sub-family. VIT180 is encoded by a very distantly related family member, <u>vit-6</u> (6).

We have recently reported the nucleotide sequences of the regions surrounding the 5' ends of five of the <u>C. elegans</u> vitellogenin <u>genes</u> (7). Our results indicated that these genes were surprisingly closely related to the vitellogenin genes of vertebrates. Like the nematode genes, the vertebrate genes are expressed in a tissue of endodermal origin, the liver. Like yp170A and yp170B, the vertebrate vitellogenins are transported intact to the oocytes. However, unlike the nematode vitellogenins, the vertebrate proteins are cleaved in the oocyte to form vitellins: phosvitin and the lipovitellins (8). In this paper we report the complete nucleotide sequence of <u>vit-5</u>, the first member of the vertebrate/nematode vitellogenin gene family to be sequenced. Our data indicate that although the nematode gene is closely related to the vertebrate gene near the 5' end, the region encoding the serine-rich vertebrate protein, phosvitin, is not present in <u>vit-5</u>.

RESULTS AND DISCUSSION

Characteristics of vit-5

The complete nucleotide sequence of vit-5, along with 261 bp of 5' flanking and 27 bp of 3' flanking DNA, is shown in Fig. 1. The location of the 5' end of the mRNA was determined by primer extension using a synthesized 15 bp oligonucleotide primer, accompanied by dideoxy-sequencing. A single, unequivocal extension product was found (data not shown). Similar experiments were performed with vit-2, vit-4, and vit-6. In each case a single extension product, the same distance 3' from a canonical TATA box sequence was found. The 3' end of the gene was located by sequencing a 3' terminal cDNA clone containing a 50 base poly(A) stretch. The gene contains four short introns, totaling 218 bp. The presence of the two introns near the 3' end of the gene was directly demonstrated by sequencing cDNA clones which overlap this region. The presence of the other two introns is inferred: they contain canonical splice sites at their 5' and 3' boundaries (Table 1); they contain stop codons in all three reading frames; and they are at least 70% A + T. Excision of these proposed introns at the positions indicated allows translation utilizing primarily the highly favored codons typical of the C. elegans vitellogenin genes (see below).

We presume that translation begins at the first AUG in each mRNA. This assumption is supported by: 1) The presence of a highly hydrophobic amino

 - 261 acttcattcaagcaatcattcagaaaaactta - 261 acttcattcaagcaatcattcagaaaaactta - taacggtcacatatacgtcgaccacgatatgcacgttgaattagaagagtgacagctgtactctgatcgctacctgaacgacgacgtcagtttcagtttcagttgaatgcggtgacag + tcagattgacattgactttatcgaagaagaagtagagatggatg	-111
ATE AAG <u>TCE ATA</u> ATC ATT GCC TCT CTT GTC GCC <u>TT6 GCC</u> ATT GCC GCC TCA <u>CC6</u> GCT CTT GAC CGT ACC TTC TCT CCA AAG AGC GAA TAC + M K S I I I A S L V A L A I A A S P A L D R T F S P K S E Y	99
GTC TAC AAA TTT GAC 66A CTT CTT CTC TCT 66A CTC CCA ACC <u>C6A</u> TCT TCC 6AT 9CT TCC CAA ACC <u>CT6</u> ATT TCT T6C C6T ACC C6T CTT + V Y K F D 6 L L L S 6 L P T R S S D A S Q T L I S C R T R L	189
CAA GCT GTT GAT GAT CGT TAC ATT CAT CTT CAA TTG ACT GAT ATT CAA TAC TCT GCT TCC CAC ATT CCA CAA TCT GAG CAA TGG CCA AAG + Q A V D D R Y I H L Q L T D I Q Y S A S H I P Q S E Q W P K	•279
ATC GAA TCT ITG GAG CAA CGT GAG CTT TCC GAT GAG TTC AAG GAG CTT CTT GAG CTT CCA TTC CGT GCT CAA ATC AGA AAT GGA CTT ATT + I E S L E @ R E L S D E F K E L L E L P F R A @ I R N G L I	-369
TCT BAG ATC CAA TTC TCT TCC GAA GAT GCC GAG TGG TCC AAG AGC GCC AAA AGA <u>TCG</u> ATT CTC AAT CTC TTC TCT CTC CGC AAG TCA GCT + S E I Q F S S E D A E W S K N A K R S I L N L F S L R K S A	459
CCA GTI GAT GAG ATG AAC CAA GAT CAG AAA GAT ATG GAA TCC GAC AAG GAT ICT GTI TIC TIC AAT GTI CAT GAA AAG ACC ATG GAA GGA + P V D E M N Q D Q K D M E S D K D S V F F N V H E K T M E G	-549
GAC TGC <u>CGA</u> AGT CGC <u>TTA</u> CAC ATT 6TT CAA 6A6 66A 6A6 6AG AAG ACC ATC TAC ACC AAA TCT 6TC AAC TTC 6AC AAA TGC ATC ACT C6C CCA + D C R S R L H I V Q E G E K T I Y T K S V N F D K C I T R P	+639
BAG ACT BCT TAT <u>BGT</u> CTT CGT TTT BGA TCT BAG TGC CAA BAG BAA TGC BAG AGG BAG BGG CAA TTT GTT AAG CCA CAA ACT GTC TAC ACC TAC ↓ E T A Y G L R F G S E C K E C E K E G Q F V K P Q T V Y T Y	+72 9
TFKNEKLQESEVHSVYTLNVN6QEVVKSET	+819
CGC GCC AAG GTC ACT TTC GTC GAG GAG AGC AAG AGC AGA GAG GAG AGC AGA GAG gtattctatttcaaaatatattttacgggcttttgatagctaactct + R A K V T F V E E S K I N R E I K K	
aacagcagcacacaagttttcag GTT TCT 66A CCA AA6 6A6 6A6 ATC 6TC TAC TCC AT6 6AA AAC 6A6 AA6 CTT ATT 6A6 CAA TTC TAC CAA CAA ↔ V S 6 P K E E I V Y S M E N E K L I E Q F Y Q Q	
66A GAC AAG GCT GAG GTC AAC CCA TTC AAG GCT ATC GAG ATG GAG GAG GAG GTT GAG GAA CTT CAA GAA ATC TTC CGC CAA ATT CAA GAG → 6 D K A E V N P F K A I E N E Q K V E Q L Q E I F R Q I Q E 9 F R Q I Q E	
ÇAC GAG CAG AAC ACC CCA GAA ACT GTC CAC CIT ATT GCC CGC GCC GTC CGC ATG TTC CGC ATG TGC ACC ATC GAA GAA CTC AAG AAG GTT + H E Q N T P E T V H L I A R A V R M F R M C T I E E L K K V	
CAC ACC ACC ATC TAC ACT AAG GCC GAG AAG AAG gtaaccaataccattcataatcatcatgtctctcttag GTT CAA CTT GTC ATT GAA ACC 4 H T T I Y T K A E K K V Q L V I E T V Q L V I E T	
SIAVAGTKNTIQHLIHHFEKKSITPLRAAE	+1386
LLKSVØETLYPSEHIADLLIØLAØ S PLSEK	+1476
Y E P L R Q S A W L A A 6 S V V R 6 F A S K T Q D L P L I R	+1566
CCA BCA TCT AGA CAA ACC AAG GAA AAG TAC GTT CGC GTC TTC ATG CAA CAC TTC CGT AAC GCT GAC TAC GAG AAG GTT CTT GCT P A S R Q T K E K Y V R V F N Q H F R N A D S T Y E K V L A CTT AAG ACC CTT GGA AAC GCC GGA ATC GAC CTC TCC GTC TAT GAG CTT GTC CAG ATT ATC CAA GAT CCA CGT CAA CCA CTT TCC ATC CGC	
LKTL6NA6IDLSVYELVQIIQDPRQPLSIR	+1836

AAC AAG CCA SAG CTC CGT ATG GCT CTT TGG AGA ATG ATG CAC ACC ATT CCA GAA GAA CCA GTT CTT GCT CAC ATC GTT TCC CAA ATG +1926 N K P E L R M A A L W R H M H T I P E E P V L A H I V S Ø H GAA AAC GAA TEC AAC CAA CAC GTT GET GET TEC ACC TAC CAC GTC CTC CGE CAA TTE TAC AAA TEC ACC CAA CCA TGE TAC CAA CAA TTG +2016 ENESNQHVAAFTYHVLRQFYKSTNPCYQQL SET SET DE TEC TEC ANS ATE CTT CTC TEC ACE CET TAT CAN CEN CAN GAN CAN ATE CTC TEC AND TAC TEC CAN CIT CEN CIT TEC AND +2106 A V R C S K I L L F T R Y Q P Q E Q M L S T Y S Q L P L F N TCT GAG TGG CTC TCC 6GA GTT CAA TTC GAC TTC GCC ACC ATT TTC GAG AAG AAC GCT TTC TTC CCA AAG GAA GTT CAA GCA TCA TTG GAA +2194 SEWLS6VQFDFATIFEKNAFLPKEVQASLE ACC GTC TTC 66A 66A AAC T66 AAC AAA TAC TTC 6CT CAA GTT 66A TTC TCT CAA CAG AAC TTT 6A6 CAA GTC ATC CTC AA6 ACC CTC 6AA +2286 TVF66NWNKYFAQV6FSQQNFEQVILKTLE AAA CTT TCT CTT TAC 66A AA6 CAA TCT 6AT 6AA CTC C6T TCC C6T C6T 6TC CAA TCT 66A ATC CAA AT6 CTT CAA 6A6 ATT 6TC AA6 AA6 +2376 K L S L Y 6 K Q S D E L R S R R V Q S 6 I Q N L Q E I V K K ATE AAC ATC CET CCA CET ETC CAA CAA ACC EAT TCT CAA AAT ECT CAC ECT ETT TTC TAC CTT CEC TAC AAE BAE ATE EAC ATC ETT +2466 MNIRPRV99TDS9NAHAVFYLRYKEHDYTV CIT CCA ATT GAC ATG GAA ACT ATT GAC ACT CIT GIT GAG AAG TAT GIC AGA AAC GGA GAG TIT GAC ATC AAA TCC CIC CIC ACT TIC TIG +2556 L P I D M E T I D T L V E K Y V R N 6 E F D .I K S L L T F L ACC AAC GAC TEC AAG TIC GAG CIT CAC CGT GET CTC TTC TAC GAG GET GAA CGC AGA ATT CCA ACA ACC ATT GGA ATG CCA CTC ACC +2646 TNDSKFF!HRA!FFYFAFRRIPTTIGMP!T ATT TCT SGA AAG ATG CCA ACT ATC CTC TCT ATC AAC 66A AAG GTT TCA ATT GAG CTC GAG AAG CTT GGA GCT CGT CTT GTT CTT GAT ATC +2736 ISGKMPTILSINGKVSIELEKLGARLVLDI BTT CCA ACT BTT BCC ACC ACC CAC GTC ACT BAB ATB CCB CTT CTB TAT CCA BTC ATT BAA CAA BBA BTC AAB TCA CTT CAA TCT BCT CBT +2826 V P T V A T T H V T E M P L L Y P V I E Q G V K S L Q S A R CIC CAC ACT CCA TTE ABA TTC GAA TCC ACT GTT GAA TTE AAG AAG AAG AAC ACT CTC GAA ATC ACT CAC AAG TTT GTT GTC CCA GAG AAC AAG +2916 LHTPLRFESTVELKKNTLEITHKFVVPENK AAG ACC ACT STT TCC STT CAT ACC CSC CCA STT SCT TTC ATC CST STT CCA AAG AAC CAA SAC TCT SAA TAT STT SAS SCT SAA SAS AAS +3004 K T T V S V H T R P V A F I R V P K N Q D S E Y V E A E E K ACT ATT TCC CAC TCA CAA TAC CAA ATE TCT ACT BAA BAB ATT BAT CAT CAA TAT BAB ACC TTT BBA CTC ABA ATC AAT BCC CAA BBA AAT +3096 TISHSQYQMSTEEIDRQYETF6LRINAQ6 STT CTT TCC CAA T66 ACT CTT CCA AT6 STT TT6 AT6 ACT 6AA CAA BAT TTC 6A6 TAC ACT CTT 6AA AAC AAA AAC C6T CCA GTT 6A6 TTC +3186 V L S Q W T L P H V L H T E Q D F E Y T L E N K N R P V E F ACA GCT CGC GTC ACT ATT GGA AAC CTC GAG AAG ACT GAT CTT TCC GAG ATC AAG TTC GAC AAG ATC TTC GAA AAA GAA TTC GAC CTT GAG +3276 T A R V T I G N L E K T D L S E I K F D K I F E K E F D L E AAC AAC GAA TOT GAG AAC CGC CGC CAA TAC TTC CAC AAG ATG ATC CGT GAG ATT CAA TOT GAG CAA GGA TTC AAG AAC CTC ATC ACC CTC +3366 N N E S E N R R Q Y F H K M I R E I Q S E Q 6 F K N L I T L AAG CTT GAA GCC CCA CAA ATG TAC TGG AAC ACT GAA CTT CGT ACC GTC TGT BAC AAA TGG ATC CGT ATG TGC AAG BTT GAG ATG GAT +3456 KLEAP & & MYWNTELRTVCDKWIRHCKVEHD SCT CGC CGC TCT CCA ATG GAG CAC GAG AAC AAA GAA TGG ACT CTT CGT ACT GAG CTT CGT GCT CGC CGA CAA ATG CCA TCA TCC CTC +354A A R R S P H E H E N K E W T L R T E L L A A R P Q H P S S L COT CAA CTT COT GAG CAA CCA CAC COT GAG GTT CAA CTC OCA TTC AAT GCC AAG TGG GGA TCA TCA AAG AAG AAG AGC GAG ATC ACA GTC AAT +3636 R Q L R E Q P H R E V Q L A F N A K N 6 5 5 K K S E I T V N SET CAA ETE BAA CAA TEE ACA GAA CAA AAG AAG TTE ATE CEE AAC ATE GAG EGT GAG TAE AAG GGA ATT EEA GAG TAE GAA ETT TTE ATE +3726 A Q L E Q S T E Q K K F I R N I E R E Y K 6 I P E Y E L L I

AND GET GET CAT CAT CAA GET ANT GET GET TET GAG TAC AND CIC ACC CCA CAG TET GAA TAC ACT TET CCC CGC ATT TET GAC CET +3816 KAARLNOVNVVSEYKLTPOSEYTFSRIFDL ATE AAG GCA TAC AAC TIC TGG ACT GTT TCT GAG AAG CGT GTC CAA AAC GAG AAT CGC CGC GTT GTT CTT CAA CTT TCT GTT GAG CCA CTT +3906 I KAYNFNTVSEKRVONEN RRVVLOLSVEPL TCC CGC CAA TCA CAT GAA CAT GAC CAT CAG ACT CCA GAA CAA GAA GTT GAG ATT GAG AAT GCT CGT ATT CCA CGA GTC CTC CCA ACT +3996 S R Q S H E H D H Q T P E Q E V E L K N A R I P R V L P T ATT GET CET AGT GEC ATE TIC CAA CAA ACC TEG GAA AAG ACC GEA BEC ACC TEC AAG ETT GAC CAA TET BAG ETT TET ACC TIT CAC AAC +4086 IARSAMF Q T WEKT 6 AT C K V D Q S E V S T T H N STG ATC TAC CGC GCT CCA CTC ACC ACC TGC TAC TCT GTT GCC AAG GAT TGC TCT GAA CAG CCA AGA TTC GCT GTT CTT GCC AAG AAG +4176 V IYRAPLTTCYSLVAKDCSEQPRFAVLAKK ATC AAC AAG AAC TCT GAG GAG CTT CTC GTT AAG GTT GTC CGC CGT GAG GAA GAA ATT GTT GTG AAG AAG TCT GAC GAT AAG TTC CTT GTC +4266 IN K N S E E L L V K V V R R E E E I V V K K S D D K F L V AAG GIT GAC 66A AAG AAG AT AAC CCA ACT GAA CIT GAA CAA TAC AA gtaagctttaactctaaatcaatgaatttttctaatctcgtttttcag T ATC +4366 K V D G K K V N P T E L E Q Y N GAA ATT CTT GGA GAT AAC CTT ATT GTT ATT CGT CTT CCA CAA GGA GAG GTT CGT TTC GAT GGA TAC ACT GTC AAG ACC AAG ATC CT +4456 EIL 6 D N L I V I R L P Q 6 E V R F D 6 Y T V K T N M P S STT GCT TCA CAA AAC CAA CTT TGC 66A CTT TGC 66A AAC AAT GAC 66T 6AG AGA GAC AAT 6AG TTC ATG ACC 6CT 6AC AAC TAC 6AA ACT +4546 VAS RNRLCGLCGNNDGERDNEFHTADNYET BAB GAT GTT GAG GAA TTC CAC CGG TCT TAC CTT CTC AAG AAT GAG GAA TGC GAG TTT GAG TTC GAC CGC ATC TCC GAG AAG AAG AAC TAC +4636 EDVEEFH RSYLLKNEECEFEFDRISEKKNY AGA AAC AAA TGE AAC AGA GAA GAG AAG AAG TCC GAC TAC GAG AGC TCC GAC TAC GAG AGC AAC TAC GAT GAG AAG GAA ACT GAA GAG +4726 R N K W N R E E K K S D Y E S S S D Y E S N Y D E K E T E E 6 gttagtctaaggctgaatgatgtgtgtgtgtgtgtagttttaacaaaatacatatttttcag AA CTC GTC AAG AAG ACC CTC ATC AAG GAG TTC TCC AAC CGC GTC TGC +4826 ELVKKTLIKEFSNRVC TTC TCC ATC GAG CCA GTC TCT GAG TGC CGT GGA CTC GAA TCC GAG AAG ACT TCC AAG AAG ATC CGT TTC ACT TGC ATG CCA CGT +4916 FSIEPVSECRRGLESEKTSNKKIRFTCMPR CAC AGE CAA GAA CGT AGT CGT TIT CIT CAA GGA AGC TCT GAG CAA ACT GTT GCC GAG TTG GTC GAT TTC CCA GTC TCC TTC GTT GAG TCT +5006 H R Q E R S R F L Q G S S E Q T V A E L V D F P V S F V E S GTC AAG ATC CCA ACC GCC TGC GTT GCC TAT TAG ATTICATATGTTTTATAATTITCAATAAAGCATTTTCACTAGaatgatttttttttctcacacttctaga +5114 V K I P T A C V A Y

Figure 1. The nucleotide sequence of $\underline{vit-5}$ and predicted amino acid sequence of yp170a. The sequence was determined by the method of Sanger (20). Restriction fragments of genomic clone 2017 containing all of $\underline{vit-5}$ (5) were subcloned into pUC8 from which random Sau3a fragments were cloned into mp8 and mp9 for sequencing. Sequences were aligned by comparison of restriction sites found by sequencing, with restriction maps of the sub-clones. Alignments were confirmed by sequencing predicted fragments which overlapped the boundaries of adjacent clones. Both strands were sequenced over most of the length of the gene. Portions of cDNA clone 1728 (5) were sequenced to confirm the predicted intron 3 and 4 boundaries. Numbering is from the first base in the mRNA, determined by premer extersion accompained by dideoxy sequencing. Introns and 5' and 3' untranslated region is double-underlined. Rare codons (as defined in the test) are underlined.

<u>1</u>	Intron/Exon Border Sequences
Intron 1	AAG GTATTCTTTTCAG GTT
2	AAG GTAACCCTCTTAG GTT
3	CAA GTAAGCTTTTCAG TAT
4	AGG GTTAGTTTTTCAG AAC
C. elegans Consensus [*]	$AG GTAAGTTTTCAG _{G}^{A}$

Table 1

"The C. elegans consensus is taken from a compilation of 29 sequenced C. elegans intron borders (Blumenthal, unpublished).

acid sequence at the N-terminus of the proposed protein, which is probably the signal sequence responsible for cotranslational secretion. 2) The sequences of this region of the other vitellogenin genes show that all of the proteins would start with a very similar signal sequence if the first AUG in each mRNA is used for translation initiation (7). 3) This signal sequence would be homologous to signal sequences at the N-termini of vertebrate vitellogenins (9). 4) The AUG at the proposed site is preceded by G at position -3 and followed by A at position +4, and so should be favored for initiation (10). 5) The proposed site for initiation precedes the only long, open reading frame, one with a very asymmetric codon usage typical of abundantly-expressed C. elegans genes (11-14).

The 5' untranslated region of vit-5 is exceptionally short, only nine bases long. The other members of the vit-l-vit-5 group also have unusually short 5' untranslated regions, 9-11 bases long (7). Translation terminates at a UAG codon which is followed by an AAUAAA sequence 30 bases further on and the 3' end of the mRNA 18 bases later. Hence, of the 4869 bases in the vit-5 mRNA, only 60 are untranslated.

The vit-5 introns range in length from 47 to 70 base pairs. Only 218 base pairs of the total gene length of 5087 base pairs are in introns. Thus vit-5 is a very compact gene. Very little RNA is spliced out and almost all that remains is translated. This gene structure is typical of the C. elegans genes sequenced thus far (11-14). In contrast, the members of the vitellogenin gene family of vertebrates are about 25 kb in length and contain 33 introns (15, 16).

Codon Usage

The codon usage is very asymmetric (Table 2). In the 1603 codons of

	Amino			Amino			Amino			Amino	
Codon	Acid	Residues	Codon	Acid	Residues	Codon	Acid	Residues	Codon	Acid	Residues
ບບບ	F	11	UCU	S	51	UAU	Y		UGU	c	1
UUC	F	59	UCC	S	38	UAC	Y	45	UGC	С	19
			UCA	S	13						
UUA	L	1	UCG	S	3	CAU	н	8	UGG	W	14
UUG	L	18	AGU	S S	4	CAC	н	25			
CUU	L	76	AGC	S	9				CGU	R	45
CUC	L	44				CAA	Q	90	CGC	R	34
CUA	L	0	CCU	P	0	CAG	Q	10	CGA	R	3
CUG	L	2	CCC	P	0				CGG	R	1
			CCA	P	61	AAU	N	17	AGA	R	14
AUU	I	40	CCG	P	2	AAC	N	56	AGG	R	1
AUC	I	51									
AUA	Ī	1	ACU	Т	43	AAA	K	16	GGU	G	2
	-		ACC	Т	49	AAG	ĸ	108	GGC	G	0
AUG	м	35	ACA	T	5				GGA	G	41
	••		ACG	Ť	ī	GAU	D	27	GGG	Ğ	1
GUU	v	68		-	-	GAC	D	31		-	-
GUC	v	46	GCU	A	46		-				
GUA	v	1	GCC	Ă	28	GAA	E	66			
GUG	v	2	GCA	Â	5	GAG	Ĕ	106			
000	•	2	GCG	Â	1	0110	-				

Table 2 Codon Usage in vit-5

vit-5, 19 codons are used three times or less, including four that are not used at all (CUA, CCU, CCC, GGC) and nine others that are used only once (UUA, AUA, GUA, ACG, GCG, UGU, CGG, AGG, GGG). In general, third position purines are rare. For instance, 114 out of 117 valine codons have third position pyrimidines. However, there are exceptions to this rule: 61 out of 63 proline codons are CCA; and 41 out of 44 glycine codons are GGA. This degree of asymmetry has been observed previously only in single-celled organisms (17, 18). An examination of published sequences of other abundantly expressed <u>C</u>. <u>elegans</u> genes, including actin (14), myosin (12), collagen (13), and the major sperm protein (11), reveals the same highly asymmetric codon usage.

Interestingly, the 19 rare codons occur at a much higher frequency near the beginning of the gene. While only 23 of 1603 total codons, or 1.4%, are rare codons, four of the first 18 codons, or 22 percent are rare codons, and two of the four occur nowhere else in the protein. We have also sequenced portions of the other vitellogenin genes (unpublished observations) and observed the same phenomenon: the same codons are rare, and each gene has several rare codons within the first 18 (a total of 16 rare codons in this region of the 5 vitellogenin genes for which sequence data is available). We do not know why rare codons are more acceptable near the 5' end of the gene. Assuming that rare codons are decoded by rare tRNAs and that selection is for rapid translation of abundantly expressed genes as has been shown to be the case in other systems (17, 18), we hypothesize there is no strong selection

Nucleic Acids Research

<u>Amino A</u>	Table cid <u>Composition</u>		nins	
	<u>yp170A</u>	Xenopus	Chick	
Asp + Asn	131	148	154	
Thr	98	87	79	
Ser	118	171	236	
Glu + Gln	272	229	179	
Pro	63	84	79	
Gly	44	84	84	
Ala	80	135	126	
Val	117	102	102	
Met	35	42	39	
Ile	92	81	89	
Leu	141	141	138	
Tyr	53	50	48	
Phe	70	62	43	
His	33	60	55	
Lys	124	113	138	
Arg	98	87	118	
Сув	20	10	ND	
Trp	14	ND	ND	

Table 3

The amino acid composition of the nematode vitellogenin yp170a is inferred from the DNA sequence of vit-5 (Fig. 1). The amino acid compositions of Xenopus and chicken vitellogenins were determined by amino acid analysis and are taken from Wiley and Wallace (21) and Wang et al. (22), respectively.

for rapid translation of the region of the mRNA encoding the signal sequence. Indeed the ubiquity of rare codons in this region of the five genes sequenced suggests there may be selection for slow translation of the region. Perhaps slow translation allows interaction between the nascent polypeptide and some component required for secretion.

Characteristics of yp170A

Vit-5 encodes a polypeptide of 186,430 daltons. The predicted amino acid composition is shown in Table 3. The protein is very high in glutamic acid and lysine, as expected for a vitellogenin. Table 3 also presents the amino acid compositions of vitellogenins from Xenopus and from chicken (vitellogenin II) for comparison. On the whole the predicted amino acid composition of ypl70A is quite similar to the vertebrate compositions, but there are some notable differences. ypl70A has more glutamic acid + glutamine and less glycine, alanine and histidine. Most interestingly, ypl70A has much less serine than do the vertebrate vitellogenins. In chicken vitellogenin, 123 of 236 serine residues are contained in the phosvitin region of the precursor (19). We have searched the yp170A sequence for a phosvitin-like region and failed to find it. (In the "core" region of chickin phosvitin, 80 of 99 residues are serine). Thus it appears that phosvitin is missing from yp170A, even though at the N-terminus the protein is clearly related to the chicken vitellogenin (7), and overall, the amino acid compositions of the two proteins are quite similar (Table 3). Since almost all of the phosvitin sequence of chicken is contained in a single exon (19), it seems likely that this exon is missing from the <u>C. elegans</u> gene.

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