

Published in final edited form as:

Chem Sci. 2010 June 11; 1(2): 202–205. doi:10.1039/C0SC00284D.

Synthesis enables a structural revision of the *Mycobacterium tuberculosis*-produced diterpene, edaxadiene†

Jillian E. Spangler, Cheryl A. Carson, and Erik J. Sorensen*

Department of Chemistry, Princeton University, Frick Chemical Laboratory, Princeton, NJ, 08544, USA

Abstract

A stereodivergent synthesis of the [3.3.1] bicyclic core of edaxadiene was completed utilizing a key intramolecular oxidative ketone allylation. Significant discrepancies between the spectroscopic data obtained for the synthetic construct and the natural isolate raised questions about the structural assignment of edaxadiene. A subsequent structural reassignment was validated by completion of a total synthesis of the correct structure of the natural product.

Tuberculosis is a pulmonary disease caused by the pathogen *Mycobacterium tuberculosis*. Roughly two billion people, or one-third of the world's population, are believed to be infected by the bacterium, resulting in over 1.5 million deaths each year.¹ Despite these grave statistics, details about the infectivity and virulence of *M. tuberculosis* are only partially understood.² Peters and co-workers recently disclosed their isolation and structural elucidation of a halimane-type diterpenoid, edaxadiene (**1**), a compound produced by *M. tuberculosis*.³ The genetic operon responsible for the production of edaxadiene is present only in pathogenic strains of mycobacteria.⁴ In addition, edaxadiene was demonstrated to be an *in vitro* inhibitor of macrophage maturation.³ These experimental observations suggest that production of edaxadiene is crucial for the infectivity and virulence of *M. tuberculosis*.

Our group has a growing interest in molecules related to the biology and treatment of tuberculosis.⁵ The biological importance as well as the scarcity of edaxadiene (**1**) led us to actively pursue a total synthesis of this intriguing natural product. Our goal was to produce sufficient quantities of this molecule to confirm its structural assignment and enable a search for its biomolecular target.

We recognized that at the heart of this challenge is the synthesis of a [3.3.1] bicyclic core bearing five contiguous stereogenic centers, including two all-carbon quaternary stereogenic centers. In addition, the relative configuration of the C₁₃ stereogenic center was not assigned during structural elucidation. Although edaxadiene (**1**) has conservation of the halimane skeleton (Fig. 1), formation of a unique C₇–C₁₃ bond locks the conformation of the B-ring and produces a highly unfavorable 1,3-diaxial interaction between the C₁₇ methyl group and C₁ methylene that is absent in related diterpenes.⁶

We envisioned that edaxadiene could arise from enone **3** by annulation of the leftmost ring onto the convex face of the [3.3.1] bicycle with subsequent deoxygenation of the C₆ ketone (**4**) to form the C₅–C₆ olefin (Scheme 1). We reasoned that this enone (**3**) could be fashioned

†Electronic supplementary information (ESI) available: Experimental procedures and spectroscopic data; interview with Erik Sorensen. See DOI: 10.1039/c0sc00284d

through an intramolecular ketone allylation of an enone of type **2** to forge the C₇–C₁₃ bond. Importantly, we anticipated that variation of olefin geometry and reaction conditions in this cyclization would provide access to both epimers at the unassigned C₁₃ stereogenic center.

Our efforts thus began with the development of a synthesis of enone **2**. The thermal Diels–Alder cycloaddition of tigraldehyde and a modified Rawal diene⁷ (**5**) provided silyl enol ether **6** (Scheme 2). Subsequent methylenation of the aldehyde provided alkene **7** in good yield on a large scale (>15 g). Regioselective hydroboration of the terminal olefin with 9-BBN furnished a *B*-alkyl borane that was subjected to a Suzuki–Miyaura cross-coupling reaction⁸ with (*Z*)-2-bromo-2-butene (**10a**) or (*E*)-2-bromo-2-butene (**10b**). The geometrical isomers **9a** and **9b** were obtained after deprotection of the enol ether and elimination of oxazolidinone with tetra-*n*-butylammonium fluoride.

After exploring multiple methods for constructing the C₇–C₁₃ bond to provide a [3.3.1] bicycle we found that oxidative cyclization of **9a** with a bimetallic system of manganese(III) acetate and copper(II) acetate in a mixture of acetic acid and benzene (3:1) provided the volatile [3.3.1] bicycle **12a** in moderate yield as a single stereoisomer (Scheme 3).⁹ This cyclization most likely proceeds through the intermediacy of an A(1,3) minimized intermediate (**11a**) to provide **12a**. Utilizing the same reaction conditions enone **9b** underwent cyclization to provide **12b**, the opposite C₁₃ epimer, albeit with a lower degree of diastereoselectivity. We were pleased to find that we could gain access to both C₁₃ epimers by controlling the geometry of the starting trisubstituted olefin.

It was at this point in our efforts that we began to note discrepancies between the reported ¹³C shifts of the [3.3.1] core of edaxadiene and our synthetic constructs. To address our scepticism of the proposed structure, we converted the diastereomeric bicycles **12a,b** into dienes **14a,b**, bearing the full [3.3.1] bicyclic core of structure **1**. Thus, stepwise reduction of each enone to the saturated alcohol (**13a** and **13b**) was followed by dehydration to the corresponding alkene (**14a** and **14b**) with Martin sulfurane in refluxing benzene.¹⁰ The unusually high temperature requirement for the dehydration is attributed to the equatorial disposition of the substrate alcohols. Although the high volatility and non-polarity of these compounds complicated their isolation, we were able to obtain sufficient quantities of each C₁₃ epimer for characterization *via* semi-preparative gas chromatography. The spectroscopic data for each C₁₃ epimer differed dramatically from the reported spectra of edaxadiene; most notably, the reported ¹³C-NMR chemical shift at C₁₃ was substantially different ($\Delta\delta$ of >30 ppm) from the ¹³C chemical shifts observed for bicycles **14a,b** (Fig. 2).

These significant differences between the ¹³C-NMR data of the isolated **1** and our synthetic bicycles (**14a** and **14b**) led to our reevaluation of the reported spectroscopic data for **1**. Our consideration of this data led us to propose compound **15** (Fig. 3) as the actual structure of edaxadiene. This proposal is consistent with the ¹³C-NMR data, most notably the deshielded nature of C₁₃. Observation of the parent [M – H₂O]⁺ ion by EI mass spectrometry is presumably due to the facile fragmentation of the tertiary allylic alcohol at C₁₃ of **15**. We further suspect that edaxadiene, as reported, is likely not a pure compound, but rather a diastereoisomeric mixture at the remote C₁₃ stereogenic center, as indicated by fine doubling of the olefinic protons in the ¹H spectrum of **1**. Our confidence in this reassignment was bolstered by a comparison of this structure to known terpenes in the literature, which indicated that this was a previously identified halimane diterpene, nosyberkol (**15**), isolated in 2004 from extracts of the Red Sea sponge *Raspailia* sp.¹¹ Furthermore nosyberkol (**15**, also referred to as isotuberculosinol) was previously speculated to be the product of the Rv3378c enzyme produced by *M. tuberculosis*.¹²

This proposed structural reassignment led us to develop a synthesis of nosyberkol (**15**) for structural verification.¹³ Our efforts began with an *exo*-selective Diels–Alder cycloaddition¹⁴ of diene **17** (available in two steps from 2,2-dimethylcyclohex-anone)¹⁵ and ethyl tiglate to give the desired cycloadduct as an inseparable mixture of diastereoisomers (2:1) (Scheme 4). Subsequent reduction of the esters and separation of the primary alcohols provided the desired C₁₀ epimer.¹⁶ Aldehyde **18** was obtained in good yield using the conditions of Parikh and Doering.¹⁷ Our attempts to perform a homologation of **18** through reaction with a phosphonate or phosphonium ylide led to no product formation; however, an aldol condensation with the sodium enolate of acetone and conjugate reduction of the resultant enone with Wilkinson’s catalyst¹⁸ allowed the desired homologation to ketone **20**. Our synthesis of nosyberkol was completed by the addition of vinylmagnesium bromide to the ketone to provide the desired compound as a 1.5:1 diastereomeric mixture at the tertiary allylic alcohol-bearing carbon. The spectroscopic data obtained for synthetic nosyberkol (**15**) were identical with those reported for both natural nosyberkol and edaxadiene (Fig. 2, see ESI[†]).¹⁹

We were also intrigued by the reported biomimetic conversion of tuberculosinol²⁰ (**16**) (Fig. 3) into edaxadiene (**15**) by treatment with a mixture of copper(II) chloride and *N,N'*-dicyclohexyl-carbodiimide (DCC).³ To examine this conversion we pursued a short synthesis of tuberculosinol from aldehyde **18**. Thus methylenation of the aldehyde provided diene **19** in good yield. Subsequent regioselective hydroboration with 9-BBN and a palladium-mediated cross-coupling with (*E*)-3-iodobut-2-en-1-ol²¹ (**21**) (Scheme 4) provided tuberculosinol (**16**). On exposure to catalytic copper(II) chloride tuberculosinol (**16**) was converted into nosyberkol (**15**) (Scheme 4). We found that addition of DCC is unnecessary for this transformation and propose that this is a Lewis acid-mediated allylic transposition.²² In addition, we observed elimination of the allylic alcohol to provide a mixture of dehydrated products (34% as an *E/Z* mixture at the C₁₂–C₁₃ olefin); however, formation of the proposed structure **1** was never observed.

In conclusion, a rapid and stereodivergent synthesis of the core of the originally proposed structure of edaxadiene (**1**) raised questions regarding its assignment. Re-evaluation of the spectral data led to the proposal that edaxadiene is actually nosyberkol (**15**), a known diterpene previously isolated from *Raspailia* sp. Furthermore, an independent synthesis of nosyberkol has unambiguously established this structural revision.²³ This synthesis of nosyberkol should provide sufficient quantities of this compound to elucidate its role in the infectivity and virulence of *M. tuberculosis*.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We gratefully acknowledge Princeton University, the National Science Foundation (Graduate Research Fellowship for J. E. S.), the Natural Sciences and Engineering Research Council of Canada (Postdoctoral Fellowship for C. A. C.), and the Merck Research Laboratory for supporting this work. For invaluable technical assistance during the course of this project we gratefully acknowledge Lotus Separations (preparative SFC) and John Eng (high resolution EI mass spectrometry).

[†]Electronic supplementary information (ESI) available: Experimental procedures and spectroscopic data; interview with Erik Sorensen. See DOI: 10.1039/c0sc00284d

Notes and references

1. World Health Organization. Fact Sheet. March. 2010 p. 104
2. Russell DG. Nat Rev Microbiol. 2007; 5:39–47. [PubMed: 17160001]
3. Mann FM, Xu M, Chen X, Ulton DB, Russell DG, Peters RJ. J Am Chem Soc. 2009; 131:17526–17527. [PubMed: 19583202] Mann FM, Pristic S, Hu H, Xu M, Coates RM, Peters RJ. J Biol Chem. 2009; 284:23574–23579. [PubMed: 19574210]
4. Pethe K, Swenson DL, Alonso S, Anderson J, Wang C, Russell DG. Proc Natl Acad Sci U S A. 2004; 101:13642–13647. [PubMed: 15340136]
5. Tilley SD, Reber KP, Sorensen EJ. Org Lett. 2009; 11:701–703. [PubMed: 19119873]
6. Breitmaier, E. Terpenes. J. Wiley-VCH; Germany: 2006.
7. Huang Y, Iwama T, Rawal VH. J Am Chem Soc. 2000; 122:7843–7844. Janey JM, Iwama T, Kozmin SA, Rawal VH. J Org Chem. 2000; 65:9059–9068. [PubMed: 11149852]
8. Miyaura N, Ishiyama T, Sasaki H, Ishikawa M, Satoh M, Suzuki A. J Am Chem Soc. 1989; 111:314–321.
9. Kates SA, Dombroski MA, Snider BB. J Org Chem. 1990; 55:2427–2436. Snider BB, McCarthy Cole B. J Org Chem. 1995; 60:5376–5377. McCarthy Cole B, Han L, Snider BB. J Org Chem. 1996; 61:7832–7847. [PubMed: 11667741] Snider BB. Tetrahedron. 1996; 52:6073–6084.
10. Martin JC, Arhart RJ. J Am Chem Soc. 1971; 93:4327–4329.
11. Rudi A, Aknin M, Gaydou E, Kashman Y. J Nat Prod. 2004; 67:1932–1935. [PubMed: 15568794]
12. Nakano C, Sato T, Hoshino T. Koen Yoshishu – Koryo, Terupen oyobi Seiyu Kagaku ni kansuru Toronkai. 2005; 49:247–249.
13. Unfortunately the published spectrum of nosyberkol (15) was provided in CDCl₃, whereas the published spectrum of the proposed edaxadiene (1) was provided in C₆D₆, preventing a direct spectral comparison.
14. de Miranda DS, de Conceição GJA, Zukerman-Schpector J, Guerrero MC, Schuchardt U, Pinto AC, Rezende CM, Marsaioli J. J Braz Chem Soc. 2001; 12:391–402.
15. Tanis SP, Abdallah YM. Synth Commun. 1986; 16:251–559.
16. We could further separate the two enantiomers of the *exo* primary alcohols to provide enantioenriched material (>99% ee) through purification with supercritical fluid chromatography (see ESI[†]).
17. Parikh JR, Doering WE. J Am Chem Soc. 1967; 89:5505–5507.
18. Ojima I, Kogure T. Tetrahedron Lett. 1972; 13:5035–5038.
19. The stereochemistry of C₁₃ was unassigned in the original isolation of nosyberkol (ref. 11). It is also unclear whether the original isolate was a single diastereoisomer at this stereogenic center.
20. Nakano C, Okamura T, Sato T, Dairi T, Hoshino T. Chem Commun. 2005:1016–1018.
21. Chen S, Horvath RF, Joglar J, Fisher MJ, Danishefsky SJ. J Org Chem. 1991; 56:5834–5845.
22. Arata K, Matsuura C. Chem Lett. 1989:1797. Yu W, Wen M, Yang L, Liu ZL. Chin Chem Lett. 2002; 13:495–496.
23. Snider and co-workers independently discovered that the original structural assignment for edaxadiene is actually nosyberkol. Their studies, which also include a synthesis of nosyberkol, were published in Organic Letters (see: Mangel N, Mann FM, Hillwig ML, Peters RJ, Snider BB. Org Lett. 2010; 12:2626–2629. [PubMed: 20462237]) after this manuscript was accepted for publication.

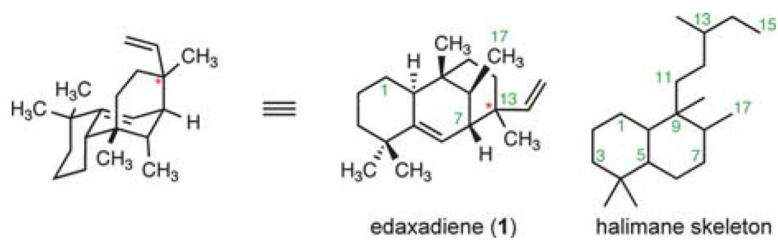


Fig. 1. Proposed structure of edaxadiene. Stereochemistry is undefined at the starred (*) carbon.

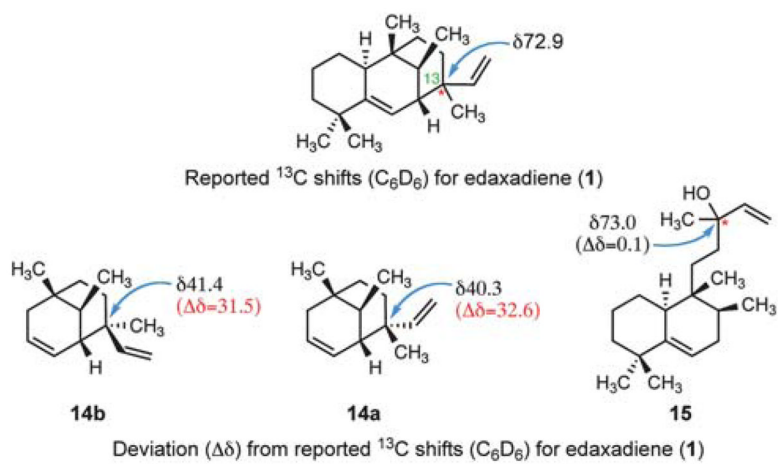


Fig. 2. Discrepancies in ^{13}C -NMR shifts that indicate the structure of edaxadiene has been incorrectly assigned.

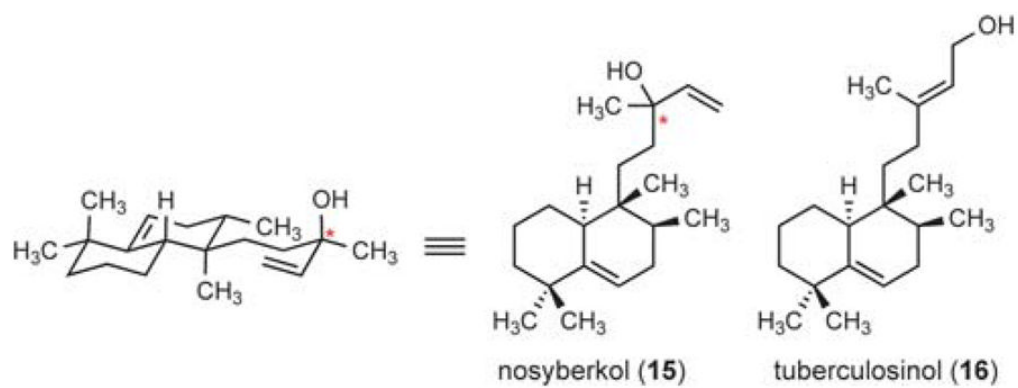
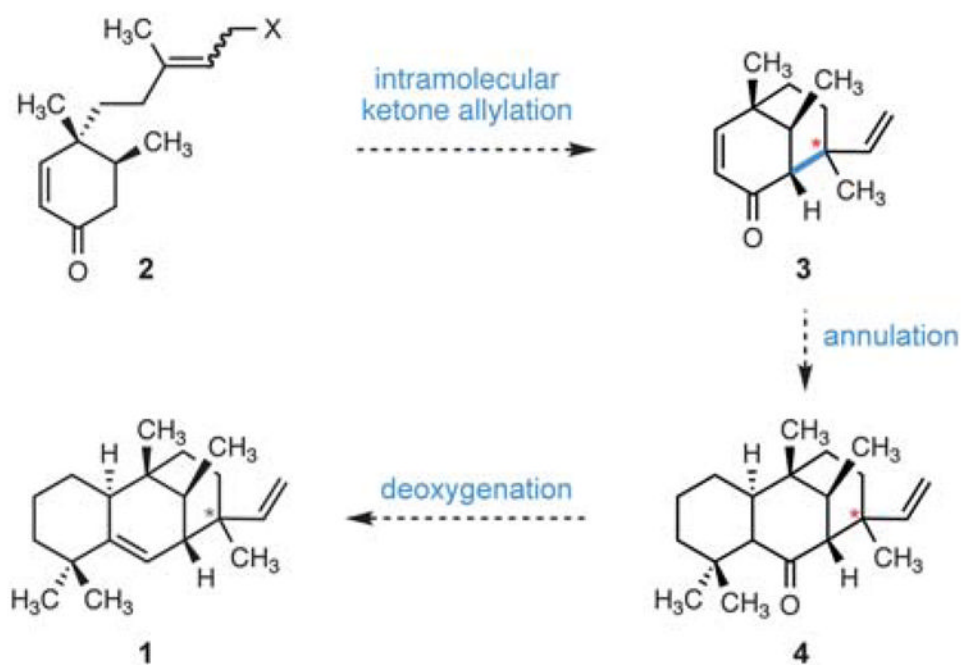
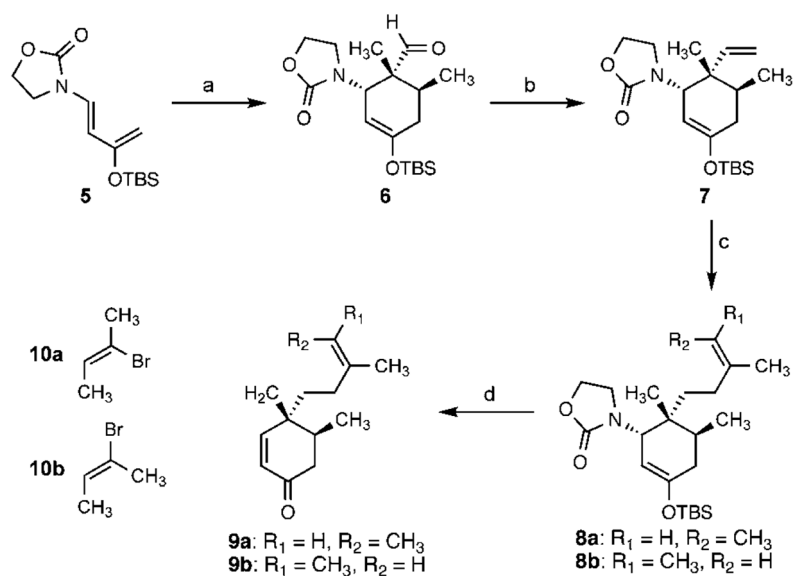


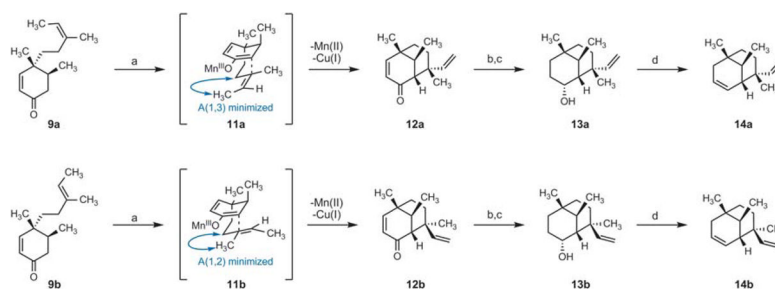
Fig. 3. Structures of nosyberkol and tuberculosinol. Stereochemistry is unassigned at the starred (*) carbon.

**Scheme 1.**

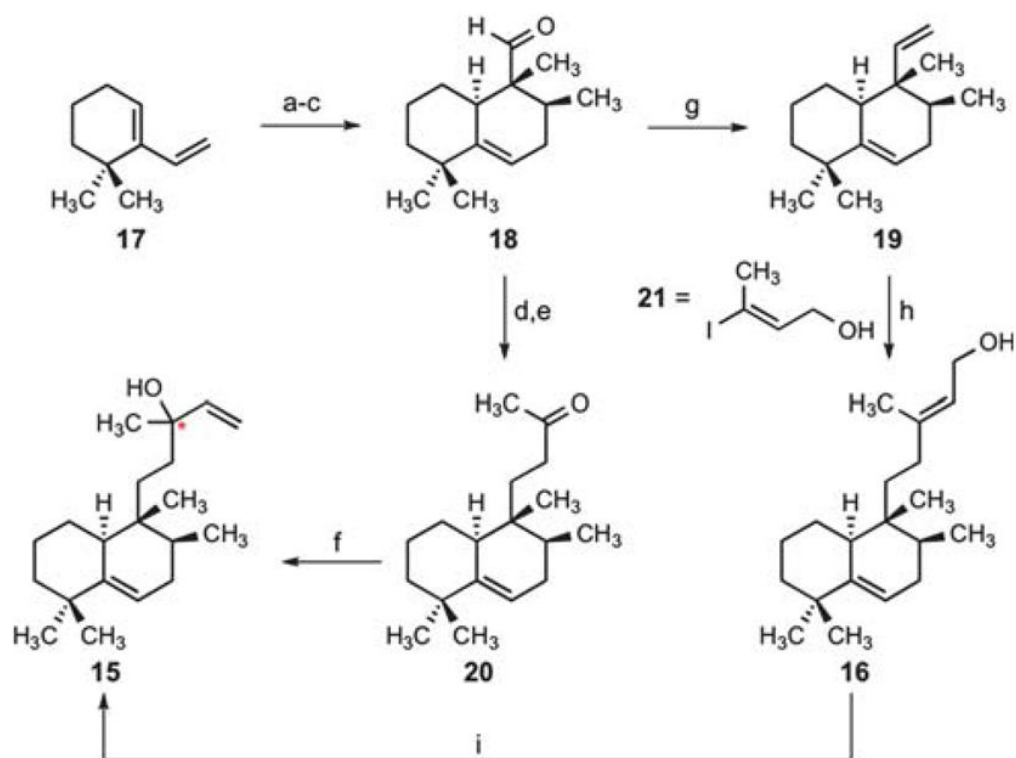
Proposed synthesis of **1** via an intramolecular ketone allylation. Stereochemistry is undefined at the starred (*) carbon.

**Scheme 2.**

Diels–Alder-mediated synthesis of enones **9a,b**. Conditions: (a) tiglaldehyde, toluene, 120 °C, 80%; (b) Ph₃PCH₃Br, *n*-BuLi, THF, 0 °C, 67%; (c) **8a**: 9-BBN, THF, reflux; then 10 mol% PdCl₂(dppf), aq. K₃PO₄, **10a**, DMF, 50 °C; **8b**: 9-BBN, THF, reflux; then 10 mol% PdCl₂(dppf), aq. K₃PO₄, **10b**, DMF, 50 °C; (d) **9a**: TBAF, THF, 86% (2 steps); **9b**: TBAF, THF, 100% (2 steps). 9-BBN = 9-bor-abicyclo(3.3.1)nonane; dppf = 1,1'-bis(diphenylphosphino)ferrocene; TBAF = tetra-*n*-butylammonium fluoride.

**Scheme 3.**

Synthesis of the [3.3.1] bicyclic core of **1** via a stereodivergent intramolecular oxidative ketone allylation. Conditions: (a) $\text{Mn}(\text{OAc})_3 \cdot 2\text{H}_2\text{O}$, $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$, 80–100 °C, AcOH–benzene (1:3), **12a**: 57%, **12b**: 51% (1.4:1 dr); (b) NaBH_4 , MeOH; (c) LiAlH_4 , Et_2O , 0 °C, **13a**: 87% (2 steps), **13b**: 73% (2 steps); (d) Martin sulfurane, benzene, 80 °C, **14a**: 23%, **14b**: 34%.

**Scheme 4.**

Syntheses of nosyberkol and tuberculosinol *via* an *exo*-selective Diels–Alder reaction.

Conditions: (a) ethyl tiglate, neat, 160 °C, 71% (2:1 *exo:endo*); (b) LiAlH_4 , THF, 40 °C, 56% (+24% *endo* isomer); (c) SO_3 ·pyridine, NEt_3 , CH_2Cl_2 –DMSO, 0 °C, 86%; (d) acetone, NaHMDS, THF, $-78 \rightarrow 23$ °C, 87%; (e) 10 mol% $\text{Rh}(\text{PPh}_3)_3\text{Cl}$, HSiEt_3 , CH_2Cl_2 , 40 °C, 83%; (f) vinylmagnesium bromide, THF, 0 °C, 93% (1.5:1 dr at the starred (*) carbon); (g) $\text{Ph}_3\text{PCH}_3\text{Br}$, KHMDS, 0 °C, THF, 91%; (h) 9-BBN, THF, 80 °C; then 10 mol% $\text{PdCl}_2(\text{dppf})$, Ph_3As , Cs_2CO_3 , **21**, DMF, 73%; (i) 20 mol% CuCl_2 , acetone, 20% (dr = 1:1 at the starred (*) carbon). 9-BBN = 9-borabicyclo(3.3.1)nonane; dppf = 1,1'-bis(diphenylphosphino)ferrocene.