# **Evaluation of the Acute Oral Toxicity Class of Trinuclear Chromium(III) Glycinate Complex in Rat**

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Received: 13 December 2010 / Accepted: 5 January 2011 /

Published online: 18 January 2011

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**Abstract** Chromium(III) is considered as an essential element playing a role in carbohydrate and lipid metabolism, and various chemical forms of this element are widely used in dietary supplements. A new trinuclear chromium(III) glycinate complex  $[Cr_3O(NH_2CH_2CO_2)_6(H_2O)_3]^+NO_3^-$  (CrGly), an analogue of Cr3 (trinuclear Cr(III) propionate complex) has been synthesized as a potential source of supplementary Cr. In this study, we evaluated the acute toxicity class of CrGly in Wistar rats applying the OECD 423 procedure. Male and female Wistar rats (n=12,  $6 \ \bigcirc$  and  $6 \ \bigcirc$ ) were given by gavage either a single dose of CrGly 2,000 mg/kg body mass (equals to 300 mg Cr(III)/kg body mass; in aqueous solution) or equivalent volumes of distilled water and fed *ad libitum* commercial *Labofeed B* diet, and observed carefully for 14 days, then sacrificed to collect blood and internal organs for biochemical and histologic examination. No death cases were detected. No abnormalities in animal behavior, body mass gains, gross organ histology, or blood morphology and biochemistry were observed. The results demonstrate that LD<sub>50</sub> of CrGly is greater than 2,000 mg/kg when administrated orally to rat; thus, this compound appears to belong to the fifth category in the GHS system or the fourth class ("unclassified") in the EU classification system.

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**Keywords** Chromium(III) glycinate complex · Acute oral toxicity · Rat

## Introduction

For 50 years, trivalent chromium (Cr) has been considered as an essential element playing an important role in carbohydrate and lipid metabolism and providing significant beneficial effects in the insulin system [1-4]. These effects appear to be related to an increase in insulin sensitivity, with the proposed mechanisms reviewed in detail by Vincent and Bennett [5]. However, the failure to identify the responsible biomolecules that bind Cr and their mode of action made researchers to readdress the essentiality of this element [6]. In short, the main controversy exists as to whether dietary supplementation with Cr should be routinely recommended in subjects without documented deficiencies. The majority of studies showed that supplementing the diet with nutritionally relevant quantities of Cr does not give any beneficial effects, while effects from Cr may arise at pharmacological doses [6, 7]. Recent data fail to demonstrate a significant improvement in carbohydrate metabolism in individuals with metabolic syndrome, impaired glucose tolerance, or consistently in individuals with type 2 diabetes after Cr supplementation. Wang et al. [8] suggested that only a subgroup of type 2 diabetic subjects respond positively to Cr supplementation and subject's phenotype appears to be very important when assessing the clinical response to Cr. Wang and Celafu [9] recently postulated that patient selection may be an important factor in determining clinical response to Cr (decreased glucose and improved insulin sensitivity), as it may be more likely in insulin-resistant individuals with type 2 diabetes who have more elevated fasting glucose and hemoglobin A(1c) levels.

Despite these controversies, over last decades Cr nutritional supplements become very popular for weight loss, muscle development, and diabetes. As Cr supplements various chemical forms of Cr(III), including Cr picolinate (CrPic), Cr histidinate (CrHis), Cr nicotinate (CrNic; also known as Cr niacinate (CrN)), and a Cr complex of D-phenylalanine (Cr(D-phe)<sub>3</sub>) have been used, patented, manufactured, and/or marketed worldwide.

The most popular chemical forms of Cr(III) used in dietary supplements are Cr(III) trispicolinate (CrPic) and Cr(III) nicotinate (CrNic). Over the last decades, sales of CrPic, as a nutritional supplement for weight loss and increasing muscle mass, were increasing, and the supplement was drawing interest for its potential use in alleviating symptoms of type 2 diabetes. The beneficial action of CrPic on glucose and lipid metabolism and its helpful role in the treatment of diabetes has been documented, but the mechanisms of action are not well understood. On the other hand, CrPic has some drawbacks as a nutritional supplement. It has very limited solubility in water (0.6 mM, at near neutral pH) and stability (decreases in lower pH until the complex hydrolyzes to release picolinic acid) [7, 10], which is responsible for its low gastrointestinal absorption rate as 1% [6]. The safety of Cr(III) compounds used for dietary supplements has been a matter of considerable debate over last decades. The discussion about the toxic effects produced by CrPic have been reviewed recently [11] and will not be repeated in this article. Because there is widespread human exposure to CrPic in dietary supplements, recently Stout et al. [12] conducted toxicity and carcinogenicity studies of this compound (in the monohydrate form) in male and female F344/N rats and B6C3F1 mice exposed in feed to concentrations of 0, 2,000, 10,000, or 50,000 ppm for 2 years. Exposure to CrPic did not induce biologically significant changes in survival, body mass, feed consumption, or nonneoplastic lesions in rats or mice. In male rats, however, a statistically significant increase in the incidence of preputial gland adenoma at 10,000 ppm was considered an equivocal finding. CrPic was not carcinogenic to female rats or to male or female mice.



Among the Cr compounds (other than CrPic and CrNic) proposed for dietary supplements and therapeutics, of particular interest is the Cr(III) propionate cation (CrProp),  $[Cr_3O(O_2CCH_2CH_3)_6(H_2O)_3]^+$ , known as  $Cr_3$  [2, 13–18]. This compound belongs to the trinuclear oxo-centered metal carboxylate assemblies of the general formula  $[Me_3O(O_2CR)_6L_3]^+$  (Me = trivalent 3d metal, R = aliphatic or aromatic group, L = monodentate ligand). CrProp was found to mimic the ability of low-molecular-weight chromium-binding substance or chromodulin to stimulate tyrosine kinase activity of the insulin receptor. Clodfelder et al. [18] reported that  $[Cr_3O(O_2CCH_2CH_3)_6(H_2O)_3]^+$  is absorbed with very high efficiency of 40–60%, while popular Cr supplements such as:  $CrCl_3$ , Cr(III) nicotinate, or CrPic are absorbed at only 0.5–1.3% of the gavaged dose. The difference in the degree of absorption is readily explained by the stability and solubility of the cation in the physiological milieu. The biological activity and safety of CrProp has been studied on various experimental models. This compound given at a wide range of dosages (up to 2,000 mg/kg body mass) to experimental animals was confirmed to be safe [19–24] and does not exert genotoxic effects [25, 26].

Having assumed that the trinuclear oxo-centered Cr carboxylate structure is the key element that determines high bioavailability and biological activity of a Cr(III) compound, we synthesized a novel Cr(III) complex with glycine (CrGly), an analogue of CrProp. Its fundamental physicochemical properties were determined with the use of thermoanalytical and spectroscopic methods and compared to that of well-known trinuclear chromium(III) complex with propionic acid. The new complex, similarly as previously studied CrProp, in contrast to CrPic and CrNic, is also well soluble in water and thermally stable until temperature of at least 220°C [27, 28].

The CrGly biological activity (together with other Cr(III) compounds, like chromium chloride, histidinate, lactate, acetate, and propionate) was already tested in vitro, using three experimental models: isolated rat liver membranes, cultured mouse C2C12 myoblasts, and 3T3-L1 preadipocytes [29]. It was found that the examined Cr (III) compounds, in contrast to the results of other lab experiments in vitro (which may be related to the specific experimental conditions applied in this experiment), did not elevate the binding of insulin to the liver membranes. Moreover, chromium histidinate, lactate, acetate, and propionate complexes diminished the specific binding of insulin. Simultaneously, chromium chloride, which did not significantly elevate insulin binding, increased the number of membrane accessible particles of the insulin receptors. However, it was accompanied by slightly diminished affinity of the receptor to the hormone. Chromium acetate and propionate significantly diminished the binding capacity of the low-affinity insulin receptor class. Investigations with the myoblast cell line C2C12 and preadipocyte cell line 3T3-L1 did not allow differentiation of the influence of the examined complexes on insulin binding. Immunodetection of phosphorylated forms of IRS-1 showed that the chromium compounds modulated the transduction of the insulin signal. Chromium glycinate, acetate, and propionate decreased the amount of IRS-1 phosphorylated at serine. Since it is generally thought that phosphorylation of serine in IRS-1 may moderate insulin action, the above-mentioned chromium complexes may, in this way, enhance insulin effects inside target cells. Phosphorylation of tyrosine in IRS-1, which acts as a stimulatory signal for further steps of insulin action, was elevated after the incubation of 3T3-L1 cells with insulin. Cr(III) did not additionally intensify this process. However, in the absence of insulin, chromium glycinate and acetate slightly elevated the level of IRS-1 phosphorylated at tyrosine. This fact may be important in vivo at low levels of insulin in blood. The results indicate that the action of chromium(III) complexes, including CrGly, involves a direct effect on the number of receptors accessible to insulin,



their affinity to the hormone, and the modulation of the signal multiplying proteins by their phosphorylation.

In the present study, we focused on the determination of the acute toxicity class of CrGly (as an analogue of CrProp), when administered orally in a single dose to Wistar rats. The study was performed in compliance with the Organization for Economic Cooperation and Development (OECD) Guidelines for Testing Chemicals [30].

# Material and Methods

#### Test Chemicals

Chromium(III) complex with glycine (CrGly) in the form of nitrate salt (chemical formula [Cr<sub>3</sub>O(NH<sub>2</sub>CH<sub>2</sub>CO<sub>2</sub>)<sub>6</sub>(H<sub>2</sub>O)<sub>3</sub>]<sup>+</sup>NO<sub>3</sub>•H<sub>2</sub>O, the theoretical content of Cr 25.7%), as an analogue of chromium(III) propionate (CrProp; chemical formula [Cr<sub>3</sub>O (O<sub>2</sub>CCH<sub>2</sub>CH<sub>3</sub>)<sub>6</sub>(H<sub>2</sub>O)<sub>3</sub>]<sup>+</sup>NO<sub>3</sub>) was synthesized in our laboratory according to the method described previously [28]. The product was recrystallized from water and then desiccated under vacuum in temperature of 100°C. The product has a form of purple crystals, well soluble in water. To ensure that molecular structure of synthesized complex corresponds with the structure of the well-known chromium(III) propionate complex (Cr<sub>3</sub>) propionic acid, some physicochemical properties of the new compounds were determined, including thermal stability was determined by using DTA and TGA methods (module DTA-50, Shimadzu). UV–VIS absorption spectra were obtained using Genesys 2 spectrophotometer. IR spectra were recorded in KBr using Specord M-80 spectrophotometer [27, 28]. The analytically determined content of elemental Cr in the compound (17.3%) was determined by the AAS method (spectrometer AAS-3 with BC correction, Zeiss, Germany).

#### Acute Oral Toxicity Study in Male and Female Rats

For determination of the Acute Toxicity Class of CrGly, The OECD 423 Guideline For Testing of Chemicals Acute Oral Toxicity—Acute Toxic Class Method [30] procedures were applied as an alternative to the oral LD<sub>50</sub> test [31]. Since the previous acute toxicity study demonstrated that LD<sub>50</sub> of CrProp is greater than 2,000 mg/kg when administrated orally to rat, it was hypothesized that its analogue with glycine (CrGly) might also have a low toxicity potential and mortality is unlikely, thus the test procedure applied with a starting dose of 2,000 mg/kg body mass (per three animals of each sex). According to the 19th paragraph of the OECD-GL 423 procedure (OECD-GL 423, adopted in 17 December 2001) [30], when available information suggests that mortality is unlikely at the highest starting dose level (2,000 mg/kg body weight), then a limit test should be conducted. The limit test is primarily used in situations where the experimenter has information indicating that the test material is likely to be non-toxic, i.e., having toxicity only above regulatory limit doses.

Male and female Wistar rats (n=12, 6  $\circlearrowleft$  and 6  $\circlearrowleft$ ; age 12–14 weeks old), obtained from Department of Toxicology Medical University of Poznan, Poland, were housed in the University-approved animal facility in rooms maintained at 22±2°C, with 55–60% humidity and 12-h photoperiod. After a 5-day adaptation to laboratory conditions, rats were divided into four equal groups (controls— $K \circlearrowleft$ ,  $K \circlearrowleft$  and treated— $T \hookrightarrow$  and  $T \circlearrowleft$ , three animals in each



group, equal body mass) and given by gavage either a single dosage of 2,000 mg CrGly (in aqueous solution)/kg body mass (treated groups, T and T and T , or equal volumes of distilled water (control groups, K and K and K and T and T groups groups and a stainless steel intubation cannula. The single dosage of CrGly equals approximately 420 mg Cr/kg body mass/rat. Rats had free access to distilled water and commercial *Labofeed B* diet composed of cereal grains (wheat, rye, soybeans, flax seeds), brewery yeast, potato protein, milk and whey powder, mineral, and vitamin mix. The chemical composition of this diet (calculated per dry mass) was as follows: protein (19.4%), fat (3.3%), dietary fiber (6.9%), crude ass (6.5%), carbohydrates (63.9%), and metabolic energy (12.5 MJ/kg). The dietary Cr content was determined by the AAS method (1.5 mg/kg diet)

Animals were observed individually after dosing at least once during the first 30 min, periodically during the first 24 h, with special attention given during the first 4 h, and daily thereafter, for a total of 14 days. All observations were systematically recorded with individual records being maintained for each animal. Individual body masses of animals were determined shortly before the test substance was administered and weekly thereafter. All the animals were sacrificed at the end of the observation period and subjected to a necropsy. All the procedures applied on animals were approved by the Local Bioethical Commission in Poznan (No. 48/2005).

# Data Collection

At the end of the test (14-day of experiment), animals, after 16-h starvation, were weighed and euthanized by intraperitoneal injection of thiopental (40 mg/kg body mass) and dissected to collect blood for morphological and biochemical studies and harvest internal organs (liver, kidney, spleen, heart) for microscopic examination.

# Laboratory Analyses

Blood hemoglobin (Hb) level was determined by the Drabkin's cyanohemoglobin method. Red blood count and related morphology indices were obtained by analytical hematology system CELLDYN-1700 [32].

Blood serum indices were determined by the following methods: glucose concentration by the UV photometric method [33] and total, LDL, HDL, and triacylglycerol concentrations by the colorimetric methods [34–36] using Olympus AU 560 equipment.

Activity of ALT, AST, and AP enzymes was measured by the kinetic methods [37], while urea concentration was measured by the kinetic method using urease and glutamine dehydrogenase [38]. Total protein concentration was measured by the colorimetric method using Cu<sup>2+</sup> ions [39], and creatinine concentration was measured by the Jaffe's kinetic method with picric acid [38]. The serum Fe concentration was determined by the colorimetric method with 2,4,6-tri(2-pyridylo)-5-triazine [40]. The serum Na and K concentrations were measured by the photometric method using CORMING-614 analyzer [41].

## Histology

A 3- to 5-mm section of the representative tissue (liver and kidney) was collected at the time of sacrifice and preserved in 10% buffered formalin. Sections were processed by standard histologic methods and embedded in paraffin. Slices were prepared and stained with hematoxylin and eosin [42].



# Data Analysis

All the analytical data are presented as means± SD, and observations for each rat were recorded and presented in tables, while histologic images of liver and kidney tissue were shown in figures.

#### **Results and Discussion**

As has been reported previously [11], the analysis of the toxicity is often a first step in the study of biological properties of the compounds. The oral acute toxic class method (ATC method) was developed as an alternative to replace the oral  $LD_{50}$  test. The ATC method is a sequential testing procedure using only three animals of one sex per step at any of the defined dose levels. Depending on the mortality rate of three, but never more than six, animals are used per dose level. This approach results in the reduction of numbers of animals used in comparison to the  $LD_{50}$  test by 40–70%. For the above reasons described previously [11], this method was applied to test the acute toxicity class for CrGly.

Male and female Wistar rats, after administration of CrGly (2,000 mg/kg body mass) and neutral carrier (distilled water) by gavage, were observed carefully during the 14-day experimental period. None of the treated animals died during this period. Also no pathological changes were noticed in skin, fur, and eyes, as well as in their behavior in comparison with the control groups.

After 14 days of observations, all the rats were sacrificed and dissected to collect blood from heart and remove internal organs for gross and histopathologic observations. No pathological changes were observed in liver, kidney, heart, spleen, ovaries and testes, and intestines, both in the treated male and female rats. Microscopic pictures (zoom ×100) of liver and kidney tissue of the treated male and female rats are shown in Figs. 1 and 2. Oral exposure to a single dosage of CrGly (2,000 mg/kg body mass) did not cause any observable pathological changes in liver and kidney tissue.

Table 1 shows the overall somatic indices, such as body and internal organ masses, in the treated and control rats. As can be seen, none of the studied parameters obtained for the treated with CrGly male and female rats were significantly different from those of the control group.

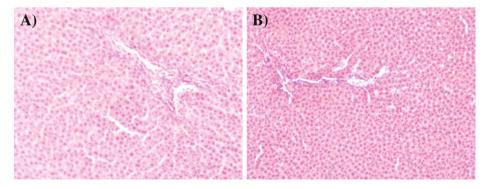


Fig. 1 Histopathological images of liver (zoom  $\times 100$ ): a control group and b tested group exposure to CrGly at dose 2,000 mg·kg<sup>-1</sup> body mass



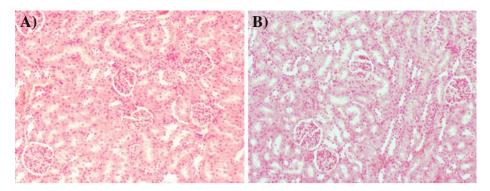


Fig. 2 Histopathological images of kidney (zoom  $\times 100$ ): a control group and b tested group exposure to CrGly at dose 2,000 mg·kg<sup>-1</sup> body mass

Table 2 presents serum biochemical indices, including glucose, total cholesterol, HDL cholesterol, tDL cholesterol, triacylglycerols, urea, creatinine, total protein concentrations, serum Na, K, and Fe concentrations, as well as the activities of the enzymes (ALT, AST, and AP), in the treated and control male and female rats. None of the above parameters were changed in male and female rats treated with CrGly, in comparison with the control groups.

**Table 1** Overall nutritional parameters of rats following exposure to 2,000 mg CrGly per kilogram body mass (mean±SD)

Parameter	Male		Student t test	Female	Student t test	
	Control	Tested CrGly 2,000 mg/kg b.m.	i test	Control	Tested CrGly 2,000 mg/kg b.m.	i test
Initial body mass (g)	306.2±8.2	306.2±4.3	NS	174.2±12.3	174.2±5.9	NS
Food intake (g)	$341 \pm 21$	$309 \pm 37$	NS	$280 \pm 43$	291±16	NS
Body mass gain (g)	$24.250 \pm 8.057$	$17.500\!\pm\!10.786$	NS	$10.250\pm2.500$	$8.250\pm2.872$	NS
Feeding efficiency ratio (g/100 g diet)	$7.031\pm2.040$	5.405±2.942	NS	$3.649\pm0.604$	2.822±0.899	NS
Final body mass (g)	$330 \pm 15$	$324 \pm 8$	NS	$184 \pm 12$	182±8	NS
Liver mass (g)	$9.527 \pm 0.158$	$9.327 \pm 0.328$	NS	$4.962\pm0.217$	$5.064 \pm 0.365$	NS
Relative liver mass (% b.m.)	2.886±0.096	$2.881 \pm 0.048$	NS	2.694±0.124	2.772±0.099	NS
Kidneys mass (g)	$2.247 \pm 0.097$	$2.224 \pm 0.107$	NS	$1.368 \pm 0.108$	$1.305 \pm 0.032$	NS
Relative kidneys mass (% b.m.)	$0.681 \pm 0.042$	$0.687 \pm 0.024$	NS	$0.741 \pm 0.033$	$0.716\pm0.032$	NS
Spleen mass (g)	$0.706 \pm 0.048$	$0.676 \pm 0.062$	NS	$0.420 \pm 0.056$	$0.421 \pm 0.036$	NS
Relative spleen mass (% b.m.)	0.214±0.023	$0.209\pm0.019$	NS	0.227±0.016	$0.231\pm0.027$	NS
Heart mass (g)	$0.945 \pm 0.069$	$0.940 \pm 0.036$	NS	$0.587 \pm 0.068$	$0.580 \pm 0.038$	NS
Relative heart mass (% b.m.)	0.286±0.012	0.290±0.008	NS	0.317±0.017	0.318±0.008	NS

NS differences statistically non-significant



Table 2 Blood serum indices of rats following exposure to 2,000 mg CrGly per kilogram body mass (mean±SD)

Parameter	Male			Female		Student
	Control group	Tested CrGly mg/kg b.m.	t test	Control group	Tested CrGly 2,000 mg/kg b.m.	t test
Glucose concentration (mg·dl <sup>-1</sup> )	98.5±15.9	84.8±2.4	NS	91.2±9.7	98.2±8.8	NS
Total cholesterol concentration (mg·dl <sup>-1</sup> )	94.5±6.4	90.8±8.9	NS	86.8±10.7	92.2±7.4	NS
LDL cholesterol concentration (mg·dl <sup>-1</sup> )	24.5±6.4	26.5±4.4	NS	22.0±5.5	25.2±8.8	NS
HDL cholesterol concentration (mg·dl <sup>-1</sup> )	65.2±10.0	58.8±4.3	NS	60.0±5.7	60.5±6.6	NS
TAG concentration (mg·dl <sup>-1</sup> )	25.5±2.6	$29.2 \pm 8.3$	NS	$26.0 \pm 2.4$	$34.0 \pm 6.5$	NS
ALT $(U \cdot dm^{-3})$	$34.8 \pm 6.6$	$31.8 \pm 3.2$	NS	$29.0 \pm 4.2$	$28.5 \pm 3.3$	NS
AST (U·dm <sup>-3</sup> )	94.5±5.3	$89.2 \pm 13.7$	NS	102.2±21.6	$119.2 \pm 46.6$	NS
ALP (U·dm <sup>-3</sup> )	146.0±16.2	131.8±22.7	NS	118.2±19.4	$107.3 \pm 4.3$	NS
Total protein concentration $(g \cdot dl^{-1})$	6.2±0.2	5.8±0.2	NS	6.4±0.2	$6.3 \pm 0.1$	NS
Creatinine concentration (mg·dl <sup>-1</sup> )	0.55±0.06	$0.52 \pm 0.05$	NS	$0.48 \pm 0.05$	$0.50 \pm 0.00$	NS
Urea concentration (mg·dl <sup>-1</sup> )	$38.5 \pm 8.7$	35.8±2.6	NS	34.8±5.2	$34.8 \pm 3.5$	NS
Na (mmol·dm <sup>-3</sup> )	139.2±2.1	$140.0 \pm 1.4$	NS	137.5±5.8	$138.5 \pm 0.6$	NS
K (mmol·dm <sup>-3</sup> )	$4.2 \pm 0.1$	$4.1 \pm 0.1$	NS	$3.9 \pm 0.1$	$4.0 {\pm} 0.4$	NS
Fe (μmol·dm <sup>-3</sup> )	19.8±1.5	14.8±3.5	NS	67.0±7.9	55.6±8.0	NS

NS differences statistically non-significant

Tables 3 and 4 show blood morphological and hematological indices, including WBC, RCB, LYMPH, PTL, MPV, PDW, hemoglobin, hematocrit, MCV, MCH, MCHC, and RDW values in the treated and control male and female rats. Most of these indices were not markedly different in animals treated with CrGly, except lower WBC (by 35%), LYMPH

**Table 3** Blood morphological indices of rats following exposure to 2,000 mg CrGly per kilogram body mass (mean±SD)

Blood index	Male		Student t test	Female	Student t test	
	Control	Tested CrGly 2,000 mg/kg b.m.	i test	Control	Tested CrGly 2,000 mg/kg b.m.	t test
WBC (10 <sup>9</sup> ·dm <sup>-3</sup> )	3.82 <sup>b</sup> ±0.56	$2.50^{a}\pm0.42$	p<0.05	1.48±1.02	1.40±0.38	NS
RBC (10 <sup>12</sup> ·dm <sup>-3</sup> )	$7.65 \pm 0.38$	$7.41 \pm 0.23$	NS	$7.46 \pm 0.18$	$7.74 \pm 0.47$	NS
LYMPH $(10^9 \cdot dm^{-3})$	$3.5^{b}\pm0.6$	$2.2^{a}\pm0.4$	p<0.05	$1.9 \pm 1.3$	$1.3 \pm 0.3$	NS
$PLT (10^9 \cdot dm^{-3})$	$1,182\pm104$	1,072±49	NS	994±264	1103±51	NS
MPV (fl)	$6.85 \pm 0.10$	$6.98 \pm 0.05$	NS	$6.92 \pm 0.28$	$6.88 \pm 0.15$	NS
PDW (fl)	$8.08 \pm 0.15$	$8.08 \pm 0.15$	NS	$8.08 \pm 0.35$	$8.18 \pm 0.26$	NS

Different letters indicate a statistically significant difference at p < 0.05

NS differences statistically non-significant, RBC red blood count



` /						
Blood index	Male		Student t test	Female		Student t test
	Control	Tested CrGly 2,000 mg/kg b.m.	i test	Control	Tested CrGly 2,000 mg/kg b.m.	i test
Hemoglobin (mmol·dm <sup>-3</sup> )	14.6±0.5	13.8±0.6	NS	14.4±0.2	14.5±0.6	NS
HCT (%)	$42.7 \pm 2.0$	$41.2 \pm 2.0$	NS	$42.3 \pm 1.0$	$43.1 \pm 1.7$	NS
MCV (fl)	$55.9 \pm 2.6$	55.6±1.1	NS	$56.8 \pm 1.4$	$55.8 \pm 1.2$	NS
MCH (pg)	$19.1 \pm 0.7$	$18.6 \pm 0.5$	NS	19.4±0.5	$18.7 \pm 0.4$	NS
MCHC $(g \cdot dl^{-1})$	$34.1^{b} \pm 0.4$	$33.4^a \pm 0.4$	p<0.05	$34.0 \pm 0.3$	33.6±0.3	NS
RDW (%)	12.9±0.9	$13.1 \pm 0.7$	NS	$12.7 \pm 0.4$	$12.8 \pm 0.8$	NS

**Table 4** Blood hematological indices of rats following exposure to 2,000 mg CrGly per kilogram body mass (mean±SD)

NS differences statistically non-significant

(by 38%), and MCHC (by 2%) values only in the case of male rats in comparison with the appropriate control rats. However, these changes were still contained within the physiological ranges established for healthy rats.

Chromium(VI) compounds generally are considered as more toxic than chromium (III) compounds. For example, the LD<sub>50</sub> for K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> (oral exposure) established in male and female rats (Fisher-344) were, 26 and 17 mg/ kg body mass, respectively, for CaCrO<sub>4</sub> in male and female rats (Sprague-Dawley), 249 and 107 mg/kg body mass, respectively [43].

The acute toxicity of a Cr(III) compound depends on its nature and solubility in water. Inorganic salts are more toxic than the organic ones. For example, LD<sub>50</sub> for chromium nitrate (Cr(NO<sub>3</sub>)<sub>3</sub>•9H<sub>2</sub>O) in male and female rats is 200 and 183 mg kg<sup>-1</sup> body mass, respectively, while for chromium(III) acetate (Cr(CH<sub>3</sub>COO)<sub>3</sub>•H<sub>2</sub>O), LD<sub>50</sub> was evaluated as low (LD<sub>50</sub>=2,365 mg kg<sup>-1</sup> body mass) [43]. The lower toxicity of chromium(III) acetate compared with chromium(III) nitrate may be related to its solubility, as chromium(III) acetate is less soluble in water than is chromium(III) nitrate.

Niacin-bound Cr(III) complex was found to be non-toxic for Sprague-Dawley male and female rats [44, 45], polynuclear Cr(III) complexes with natural ligands, including glycine, cysteine, glutathione, glutaminic, nicotinic, and asparaginic acids, such as: [Cr<sub>3</sub>(Glu)<sub>3</sub>Cys(OH)<sub>2</sub>(H<sub>2</sub>O)<sub>6</sub>], [Cr<sub>4</sub>Nic(GSH)<sub>2</sub>(OH)<sub>6</sub>(H<sub>2</sub>O)<sub>6</sub>], [CrNicCys(H<sub>2</sub>O)<sub>2</sub>], [Cr<sub>2</sub>(Asp)<sub>2</sub>(GSH)(OH)<sub>3</sub>H<sub>2</sub>O]•4H<sub>2</sub>O, [Cr<sub>2</sub>(Nic)<sub>2</sub>(OH)<sub>4</sub>]•4H<sub>2</sub>O, [Cr(Nic)<sub>2</sub>(OH)]•3H<sub>2</sub>O, mer-[Cr(Gly)<sub>3</sub>H<sub>2</sub>O]•H<sub>2</sub>O, and fac-[Cr(Gly)<sub>3</sub>H<sub>2</sub>O]•H<sub>2</sub>O, were found to be "non-toxic" (LD<sub>50</sub>>2,000 mg/kg body mass) for mice [46–48].

In our previous work, it was demonstrated that  $LD_{50}$  of CrProp is greater than 2,000 mg kg<sup>-1</sup> body mass, when administrated orally to rat; thus, this complex appears to belong to the fifth category in the GHS system or the fourth class ("unclassified") in the EU classification system [11].

The results of this study assessing the acute oral toxicity of CrGly using both male and female Wistar rats, due to lack of lethal effects and no significant changes in gross and microscopic pictures of internal organs, as well as no sign of pathological changes blood biochemical and morphological indices, suggest that the tested compound should be classified (similarly to CrProp) to the fifth category in the United Nations Globally Harmonized System [49] or the fourth class ("unclassified," practically non-toxic) in the European Union classification system [50], which means that CrGly has low toxicity potential and relatively high safety margins for rat.



In conclusion, the acute toxicity of many Cr(III) compounds had been determined, and most of them should be considered acutely non-toxic, unless the ligands are known to be toxic or the compounds are complexes of imines or related ligands that greatly shift the redox potential of Cr(III), such as Cr(bpy)3 or Cr(phen)3 that are known toxins and mutagens [51, 52].

**Acknowledgments** The research described in this paper was funded by the Ministry of Education and Science (Grant No. 2 P06T 03830) in the years 2006–2009.

Conflict of Interest The authors declare that there are no conflicts of interest.

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