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Pregnancy Upregulates Large-Conductance Ca²⁺-Activated K⁺ Channel Activity and Attenuates Myogenic Tone in Uterine Arteries

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Summary

Uterine vascular tone significantly decreases while uterine blood flow dramatically increases during pregnancy. However, the complete molecular mechanisms remain elusive. We hypothesized that increased BK_{Ca} channel activity contributes to the decreased myogenic tone of uterine arteries in pregnancy. Resistance-sized uterine arteries were isolated from nonpregnant and near-term pregnant sheep. Electrophysiological studies revealed a greater whole-cell K⁺ current density in pregnant than nonpregnant uterine arteries. Tetraethylammonium and iberoitoxin inhibited K^+ currents to the same extent in uterine arterial myocytes. The BK_{Ca} channel current density was significantly increased in pregnant uterine arteries. In accordance, tetraethylammonium significantly increased pressure-induced myogenic tone in pregnant uterine arteries and abolished the difference in myogenic responses between pregnant and nonpregnant uterine arteries. Activation of protein kinase C produced a similar effect to tetraethylammonium by inhibiting BK_{Ca} channel activity and increasing myogenic tone in pregnant uterine arteries. Chronic treatment of nonpregnant uterine arteries with physiologically relevant concentrations of 17β -estradiol and progesterone caused a significant increase in the BK_{Ca} channel current density. Western blot analyses demonstrated a significant increase of the $\beta 1$, but not α , subunit of BK_{Ca} channels in pregnant uterine arteries. In accordance, steroid treatment of nonpregnant uterine arteries resulted in an upregulation of the β 1, but not α , subunit expression. The results indicate that the steroid hormone-mediated upregulation of the $\beta 1$ subunit and BK_{Ca} channel activity may play a key role in attenuating myogenic tone of the uterine artery in pregnancy.

Keywords

uterine artery; pregnancy; BK_{Ca} channel; myogenic tone; steroids; protein kinase C

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Introduction

Uterine blood flow increases substantially during pregnancy, which is essential both for the growth and survival of the fetus and for cardiovascular well-being of the mother. Maladaptation of the uteroplacental circulation during pregnancy is associated with high incidence of clinical complications including preeclampsia and fetal intrauterine growth restriction. While the mechanisms underlying the adaptation of uterine circulation to pregnancy are complex and poorly understood, recent studies have demonstrated that pressure-dependent myogenic reactivity plays a pivotal physiological role in the regulation of uterine circulation, and that decreased uterine arterial myogenic tone contributes significantly to the adaptation of uterine vascular hemodynamics in pregnancy.^{1,2}

The molecular mechanisms underlying this attenuated myogenic tone of uterine arteries in pregnancy remain elusive. Previous studies have suggested a possible role of large-conductance Ca²⁺-activated K⁺ (BK_{Ca}) channels in the regulation of uterine circulation during pregnancy.^{3–5} The BK_{Ca} channel in vascular smooth muscle contains pore-forming α subunit (encoded by KCNMA1) and up to four accessory identical β 1 subunits (encoded by KCNMB1).⁶ The β 1 subunit is predominantly expressed in smooth muscle^{7–9} and enhances the Ca²⁺ sensitivity of BK_{Ca} channels.¹⁰ It has been shown that BK_{Ca} channels play an important role in regulating the resting membrane potential of vascular smooth muscle, and channel blockade by iberoitoxin (IBTX) or tetraethylammonium (TEA) depolarizes the membrane resulting in vasoconstriction.¹¹ The importance of BK_{Ca} channel in the regulation of BK_{Ca} channel, and α or β 1 subunit knockout mice displayed increased vascular tone and elevated blood pressure,^{8,12} although uterine vascular function was not examined.

Little is known about the functional role of BK_{Ca} channels in regulating pressure-dependent myogenic tone of uterine arteries and its adaptation to pregnancy. In the present study, we test the hypothesis that upregulation of BK_{Ca} channel expression and activities accounts for attenuated myogenic tone of uterine arteries in pregnancy. This was achieved by investigating the myogenic reactivity and BK_{Ca} channel current density and protein expression in uterine arteries obtained from nonpregnant and near-term pregnant sheep using functional, electrophysiological and biochemical approaches. Given our recent findings that sex steroid hormones played an important role in downregulating pressure-dependent myogenic tone of the uterine artery in pregnancy,² we further test the hypothesis that sex steroid hormones upregulate BK_{Ca} channel expression and activities in uterine arteries.

Materials and Methods

An expanded Materials and Methods section is available in the online data supplement at http://hyper.ahajournals.org.

Tissue Preparation and Treatment

Resistance-sized uterine arteries were isolated from nonpregnant and near-term pregnant sheep.^{2,13,14} For hormonal treatment, arteries were incubated in phenol red-free DMEM with 1% charcoal-stripped FBS for 48 hours at 37°C in a humidified incubator with 5% $CO_2/95\%$ air in the absence or presence of 17β-estradiol and/or progesterone. All procedures and protocols were approved by the Institutional Animal Care and Use Committee and followed the guidelines by the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Measurement of Myogenic Tone

Pressure-dependent myogenic tone of resistance-sized uterine arteries were measured as described previously.^{2,13,14}

Measurement of BK_{Ca} Channel Current

Smooth muscle cells were enzymatically dissociated from resistance-sized uterine arteries, and whole-cell K⁺ currents were recorded and normalized to cell capacitance as picoampere per picofarad (pA/pF).^{15,16} Only relaxed and spindle-shaped myocytes were used for recording. The BK_{Ca} channel current was determined as the difference between the whole-cell K⁺ current in the absence and presence of IBTX or TEA.

Western Immunoblotting Analysis

Protein abundance of BK_{Ca} channel α and β 1 subunits was measured in freshly isolated resistance-sized, endothelium-intact uterine arteries and after the hormonal treatment by Western blot analysis, as described previously.^{2,13,14}.

Data Analysis

Results were expressed as means \pm SEM obtained from the number of experimental animals given. Differences were evaluated for statistical significance (*P*<0.05) by ANOVA or *t* test, where appropriate.

Results

Pregnancy Upregulates BK_{Ca} Channel Function in Uterine Arteries

The whole-cell K^+ current density in uterine arterial myocytes of pregnant sheep (66.3 \pm 5.2 pA/pF) was significantly greater than that in nonpregnant animals (37.5±3.3pA/pF) at +80 mV (P < 0.05) (Figure 1). TEA at concentrations ≤ 1.0 mmol/L selectively blocks BK_{Ca} channels with negligible effects on voltage-gated K⁺ channels.^{17–19} Exposure of the myocytes to TEA (1.0 mmol/L) or IBTX (100.0 nmol/L) significantly depressed the macroscopic K^+ currents (Figure 1), indicating the functional presence of BK_{Ca} channels in uterine arteries from both nonpregnant and pregnant sheep. As shown in Figure 1, TEA and IBTX inhibited the K⁺ currents to the same extent in uterine arterial myocytes. The TEAand IBTX-sensitive components of K^+ currents in the myocytes at +80 mV were 38.5±2.5% and 40.0±3.1% (P>0.05) in nonpregnant sheep, and 53.2±1.8% and 55.7±2.7% (P>0.05) in pregnant animals, respectively. The finding that TEA and IBTX inhibited the K⁺ currents to the same extent in uterine arterial myocytes validated the specificity of TEA action on the BK_{Ca} channel. The BK_{Ca} current density determined by TEA (35.1±2.9 pA/pF in pregnant myocytes versus 14.1±1.3 pA/pF in nonpregnant myocytes, P<0.05) or by IBTX (32.5±2.1 pA/pF in pregnant myocytes versus 16.8±1.9 pA/pF in nonpregnant myocytes, P<0.05) at +80 mV was significantly increased in pregnant uterine arteries (Figure 2).

Inhibition of BK_{Ca} Channels Increases Pressure-Dependent Myogenic Tone in Uterine Arteries

To determine the functional importance of heightened BK_{Ca} channel activity in regulating uterine arterial vascular tone, pressure-dependent myogenic responses of resistance-sized uterine arteries were measured in the absence or presence of the BK_{Ca} channel inhibitor TEA. Consistent with the previous studies,¹ pressure-dependent myogenic tone in uterine arteries of pregnant sheep was significantly lower than that in uterine arteries of nonpregnant sheep over the physiological range of intravascular pressures (Figure 3). TEA significantly increased the myogenic tone in uterine arteries of pregnant sheep (Figure 3). In uterine arteries of nonpregnant animals, TEA showed no significant effect on the myogenic

reactivity when the data were analyzed as groups (Figure 3). However, paired analysis of data in each animal revealed that TEA appeared to increase the myogenic tone in one out of five animals (Animal #5) (Figure 3). Consisting with this finding, it appeared that the uterine arteries from Animal #5 had higher expression level of the β 1 subunit of BK_{Ca} channels (Figure 3). In the presence of TEA, there were no significant differences in pressure-dependent myogenic tone of uterine arteries between nonpregnant and pregnant animals.

Activation of Protein Kinase C (PKC) Inhibits $\mathsf{BK}_{\mathsf{Ca}}$ Channels and Increases Myogenic Tone in Uterine Arteries

As shown in Figure 4, the activation of PKC by phorbol 12,13-dibutyrate (PDBu) significantly inhibited whole-cell K⁺ currents in uterine arterial myocytes. The inhibition of K⁺ currents at +80 mV by PDBu was significantly greater in the myocytes of pregnant sheep (34.0±2.6%) than that in nonpregnant animals (20.9±4.0%) (Figure 4, P < 0.05). However, there were no significant differences among TEA-, PDBu-, and PDBu plus TEA-produced inhibitions of K⁺ currents in either nonpregnant or pregnant uterine arterial myocytes, respectively (Figure 4), indicating that the PDBu-induced reduction of K⁺ currents is predominantly mediated by inhibiting the BK_{Ca} channel. In accordance, PDBu significantly increased pressure-dependent myogenic tone in uterine arteries of pregnant sheep (Figure 5) to the similar extent as that seen with TEA in Figure 3.

Steroid Hormones Increase the BK_{Ca} Channel Activity in Uterine Arteries

Previous studies demonstrated that 17β-estradiol and progesterone treatment for 48 hours resulted in a significant decrease in myogenic responses in nonpregnant uterine arteries.² The effect of steroid hormones on the BK_{Ca} channel activity was thus determined by the treatment of nonpregnant uterine arteries with 17β -estradiol (0.3 nmol/L) and progesterone (100.0 nmol/L) for 48 hours. As shown in Figure 6, the steroid hormone treatment significantly increased macroscopic whole-cell K⁺ currents in uterine arterial myocytes. Compared with the control myocytes, the current density was increased by ~ 2-fold at +80 mV in hormone-treated myocytes (41.5±5.8 pA/pF versus 24.0±3.4 pA/pF, P<0.05). Whole-cell K⁺ currents of myocytes isolated from both control and steroid hormone-treated uterine arteries were subject to the TEA inhibition. However, TEA produced a significantly greater inhibition of K^+ currents in hormone-treated myocytes (58.6±1.7%) than that in the control (41.3±3.5%) (P<0.05). In accordance, the steroid hormone treatment resulted in ~2.5-fold increase in the current density of BK_{Ca} channels (24.5 \pm 3.7 pA/pF versus 10.4 \pm 2.4 pA/pF, P < 0.05) (Figure 6). To determine the specific effect of 17 β -estradiol and progesterone, nonpregnant uterine arteries were treated with 17β-estradiol (0.3 nmol/L) or progesterone (100.0 nmol/L), respectively, for 48 hours. As shown in Figure 7, 17βestradiol, but not progesterone, produced the similar effect seen with the combined steroids and significantly increased the BK_{Ca} channel current density by 2-fold in uterine arterial myocytes.

Pregnancy and Hormonal Treatment Increase $\mathsf{BK}_{\mathsf{Ca}}$ Channel $\beta 1$ Subunit Expression in Uterine Arteries

As shown in Figure 8, protein abundance of the α subunit detected at about 100 kDa was not significantly different between nonpregnant and pregnant uterine arteries. The β 1 subunit was detected at 39 kDa as seen in the previous findings^{5,20} and its expression levels were significantly greater in pregnant uterine arteries than that in nonpregnant uterine arteries. In accordance, 17 β -estradiol (0.3 nmol/L) and progesterone (100.0 nmol/L) treatment for 48 hours significantly upregulated β 1, but not α , subunit expression in nonpregnant uterine arteries (Figure 8).

Discussion

The present study provides direct evidence for the first time that the BK_{Ca} channel activity in uterine arteries is significantly enhanced during pregnancy. Previous studies demonstrated that intra-arterial infusion of TEA into the uterine artery circulation of late gestation sheep caused a decrease of basal uterine blood flow from 50% to 80% in the absence of systemic effects or a change in contralateral uterine blood flow.^{3,4} This is consistent with the present finding that TEA inhibited K^+ currents by 53% in pregnant uterine arteries. The findings that TEA significantly increased pressure-dependent myogenic tone in uterine arteries of pregnant sheep, and that it abrogated the difference in the myogenic response between nonpregnant and pregnant uterine arteries indicate that the pregnancy-induced attenuation in uterine arterial myogenic tone is conferred primarily by enhanced BK_{Ca} channel activity. BK_{Ca} channels have been shown to play an important role in the regulation of resting membrane potential and control of vascular tone.²¹ These findings support the notion that the heightened BK_{Ca} channel activity is a predominant mechanism in maintaining uteroplacental blood flow in pregnancy. The finding that the β 1 subunit was higher in the nonpregnant animal that also showed the effect of TEA is intriguing, and suggests that the animal was in the follicular phase as shown by Kahn et al.²⁰ This finding further supports the notion that elevated estrogen levels increase the $\beta 1/\alpha$ stoichiometry and BK_{Ca} channel activity,^{20,22} which contributes to the downregulation of myogenic tone of uterine arteries. In agreement, previous studies in overiectomized nonpregnant sheep demonstrated that TEA had no significant effects on basal uterine vascular resistance and blood flow, but it inhibited estrogen-induced rise in uterine blood flow.²³ These findings suggest that the regulation of myogenic tone of uterine arteries by BK_{Ca} channels is modulated by sex steroids.

The question arises as to how pregnancy might affect BK_{Ca} channel function and its regulation of uterine arterial myogenic tone. Previous studies have demonstrated that PKC plays an important role in the regulation of pressure-dependent myogenic response of resistance arteries,^{1, 24, 25}, and a decrease in the PKC signaling pathway accounts for the attenuated myogenic tone of the uterine artery in pregnancy.^{1, 2,13,14,26–30} The BK_{Ca} channel is subject to modulation by PKC and activation of PKC has been shown to inhibit BK_{Ca} channels in vascular smooth muscle cells.^{31–34} The present finding that activation of PKC inhibited BK_{Ca} channel activity and increased pressure-dependent myogenic tone in pregnant uterine arteries provides a functional link of the BKCa channel in pregnancymediated downregulation of PKC and myogenic tone of uterine arteries. The work suggests a mechanism of attenuated PKC in the heightened BKCa channel activity in pregnant uterine arteries. Whereas ANOVA and post-hoc analysis indicated no significant differences between PDBu- and PDBu plus TEA-produced inhibition of the total K⁺ currents in uterine arterial myocytes, it appeared that the combination might produce a slightly greater inhibition than the PDBu alone, which would suggest that PDBu might not completely inhibit BK_{Ca} channels under the current conditions studied.

Consisting with the previous finding of a direct genomic effect of sex steroid hormones in attenuating the PKC activity and myogenic tone of the uterine artery,^{2,13,14} the present study demonstrated that the enhanced BK_{Ca} channel activity seen in pregnant uterine arteries was mimicked by the treatment of nonpregnant uterine arteries with 17β-estradiol and progesterone for 48 hours in an *ex vivo* tissue culture system. The acute and nongenomic effects of estrogen in activating BK_{Ca} channels have been previously reported at high nanomolar to micromolar concentrations that are substantially greater than physiological concentrations,^{23,35–37} To our knowledge, the present study is the first to demonstrate that physiologically relevant concentrations of 17β-estradiol and progesterone have direct genomic effects on upregulating the BK_{Ca} channel activity in vascular smooth muscle cells. Whereas both estrogen and progesterone receptors have been identified in uterine artery

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vascular smooth muscle, 13,38,39 the present study demonstrated that 17 β -estradiol alone was sufficient to increase the BK_{Ca} channel activity in uterine arterial myocytes. The conditions of ovarian cycle of luteal or follicular phases and systemic blood steroid levels were not determined in nonpregnant animals in the present study, and thus we cannot exclude the possibility that prior steroid exposure *in vivo* may have primed the uterine arteries to respond to estrogen. In agreement to the present study, similar findings were obtained in a neuronal cell line GT1-7 in which 17 β -estradiol at physiological concentrations augmented BK_{Ca} channel currents in a genomic manner.⁴⁰

The BK_{Ca} channel in vascular smooth muscle consists of pore-forming α subunit and up to four accessory β 1 subunits.⁶ In ovine uterine arteries, both α and β 1 subunits were detected exclusively in vascular smooth muscle cells with no evidence of their existence in the endothelium.^{5,20,22} In the present study, we found that the α subunit was not significantly different in uterine arteries between nonpregnant and pregnant sheep, suggesting no changes in the BK_{Ca} channel density by pregnancy. This is somewhat different from the previous finding showing about 60% increase in protein abundance of the α subunit in pregnant uterine arteries, although the mRNA was not significantly changed.⁵ One possible reason for this apparent difference may be because the previous study used larger diameter first and second generation uterine arteries while the present study used resistance-sized uterine arteries. In agreement with the present finding, it has been shown in sheep that increased endogenous estrogen in the follicular phase or infusion of exogenous estrogen in ovariectomized nonpregnant ewes have no significant effect on α subunit levels in uterine arteries.^{20,22} The present finding of increased β 1 subunit in pregnant uterine arteries as compared with nonpregnant uterine arteries is consistent with the previous study in sheep.⁵ Although the β^2 subunit was present in uterine artery smooth muscle, the expression was minimal and unchanged by pregnancy.⁵ Unlike the effect of pregnancy, both β 1 and β 2 subunits were found elevated in follicular as compared with luteal phase of the ovarian cycle in nonpregnant sheep, probably due to relatively high estrogen and low progesterone that was produced endogenously by the ovaries.²⁰ Given that pregnancy is a state with substantially higher levels of estrogen and progesterone as compared with the nonpregnant state, it is possible that high progesterone inhibits the expression of $\beta 2$ subunit. Similar findings were obtained in the ovariectomized estrogen-treated sheep showing an increase in the β 1 subunit although the β 2 subunit was not examined.²²

The increase in $\beta 1$ subunit expression of the BK_{Ca} channel would have functional consequences. It has been shown that $\beta 1$ subunits of BK_{Ca} channels play a pivotal role in regulating vascular tone and blood pressure.⁸ The association of $\beta 1$ subunits with α subunits are critical in regulating the Ca²⁺ sensitivity of BK_{Ca} channels.^{10,41} Downregulation of the β 1 subunit and altered α : β 1 stoichiometry decrease the Ca²⁺ sensitivity of the channel, resulting in decreased BK_{Ca} channel activity.⁴² In contrast, upregulation of the β 1 subunit increases the Ca²⁺ sensitivity of BK_{Ca} channel and channel activity in vascular smooth muscle.⁴³ In the present study, the increased $\beta_{1:\alpha}$ subunit stoichiometry in the uterine arteries of pregnant animals is likely to augment the Ca²⁺ sensitivity of the BK_{Ca} channel and facilitate the activation of the channel and the consequent reduction in myogenic tone of the uterine arteries in pregnancy. The previous findings of the increased $\beta 1$ subunit mRNA and protein levels in uterine arteries and myometrial smooth muscle after prolonged exposure of ovariectomized sheep or mice to estrogen suggested a possible role of the steroid hormone in modulating BK_{Ca} channel expression. The present study provides clear evidence of the direct effect of steroid hormones on upregulating the β 1 subunit in the uterine artery. Although the molecular mechanisms remain elusive, the estrogen responsive elements have been identified at the promoter of ovine β 1 subunit gene.

PERSPECTIVES

The present study suggests a possible mechanism of 17β -estradiol-mediated upregulation of the β 1 subunit in the heightened BK_{Ca} channel activity in uterine arteries during pregnancy, resulting in the reduced myogenic tone of uterine artery in pregnancy. Given that the BK_{Ca} channel plays a pivotal role in regulating vascular tone and thus blood flow and pressure, dysregulation of the stoichiometric composition of BK_{Ca} channels and the channel activity in the uterine artery is likely to contribute significantly to the maladaptation of uterine vascular hemodynamics in pregnancies complicated by preeclampsia. Indeed, reductions in uteroplacental blood flow and chronic uteroplacental ischemia in a variety of animal models lead to a hypertension state that closely resembles preeclampsia in women.^{44,45} The present findings provide an understanding of the mechanisms of the BK_{Ca} channel in uterine vascular adaptation to pregnancy, and may suggest new insights of therapeutic strategies by enhancing the BK_{Ca} channel activity in vascular smooth muscle that may be beneficial for pregnant women with preeclampsia. Additionally, the present study offers insights into the mechanisms in the hormonal regulation of the BKCa channel activity and myogenic tone of resistance arteries in general, and improves our understanding of vascular benefits of hormone replacement therapy in postmenopausal women, given the well-established finding that premenopausal women are at lower risk of developing hypertension and coronary heart disease than men of the same age and that the cardiovascular risk increases only after the cessation of ovarian function.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Figure 1. Effect of TEA and IBTX on K⁺ currents in uterine arteries

Smooth muscle cells were freshly isolated from nonpregnant (**panel A**) and pregnant (**panel B**) uterine arteries and whole-cell K⁺ currents were recorded in the absence or presence of TEA (1.0 mmol/L) or IBTX (100.0 mmo/L). Data are means \pm SEM of 4–12 cells from 4–7 animals of each group. * *P* < 0.05, versus control.

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Figure 2. Effect of pregnancy on BK_{Ca} current density in uterine arteries Smooth muscle cells were freshly isolated from nonpregnant (NPUA) and pregnant (PUA) uterine arteries. BK_{Ca} current density was determined in the presence of TEA (**panel A**) or IBTX (**panel B**). Data are means \pm SEM of 4–12 cells from 4–7 animals of each group. **P* < 0.05, versus NPUA.

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Figure 3. Effect of TEA on myogenic tone in uterine arteries

Pressure-dependent myogenic tone of nonpregnant (NPUA) and pregnant (PUA) uterine arteries was determined in the absence or presence of TEA (1.0 mmol/L for 20 minutes) in each animal in a pair wise fashion. Protein abundance of BK_{Ca} channel β 1 subunit was determined by Western blot analysis. **P* < 0.05, presence of TEA versus absence of TEA.

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Figure 4. Effect of PDBu on K⁺ currents in uterine arteries

Smooth muscle cells were freshly isolated from nonpregnant (**panel A**) and pregnant (**panel B**) uterine arteries and whole-cell K⁺ currents were recorded in the absence or presence of TEA (1.0 mmol/L), PDBu (1.0 μ mol/L) or PDBu plus TEA, respectively. Bar graphs present the effects of TEA, PDBu or PDBu plus TEA on K⁺ current densities obtained at +80 mV. Data are means ± SEM of 5 cells from 5 animals of each group. * *P* < 0.05, versus control.

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Figure 5. Effect of PDBu on myogenic tone in pregnant uterine arteries

Pressure-dependent myogenic tone of pregnant uterine arteries was measured in the absence or presence of PDBu (100.0 nmol/L for 15 minutes). Data are means \pm SEM of tissues from 5–6 animals of each group. **P* < 0.05, versus –PDBu.

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Figure 6. Effect of steroid hormone treatment on ${\rm BK}_{\rm Ca}$ channel activity in nonpregnant uterine arteries

Nonpregnant uterine arteries were treated with 17 β -estradiol (E2 β , 0.3 nmol/L) plus progesterone (P4, 100.0 nmol/L) for 48 hours. Myocytes were then isolated and whole-cell K⁺ currents were recorded in the absence or presence of TEA (1.0 mmol/L). **Panel A**: K⁺ currents in control myocytes. * *P* < 0.05, –TEA versus +TEA; **Panel B**: K⁺ currents in hormone-treated myocytes. * *P* < 0.05, –TEA versus +TEA; **Panel C**: BK_{Ca} current densities in control and hormone-treated myocytes. * *P* < 0.05, +E2 β /P4 versus –E2 β /P4. Data are means ± SEM of 7 cells from 4 animals of each group.

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Figure 7. Specific effect of 17β -estradiol and progesterone on BK_{Ca} channel activity in nonpregnant uterine arteries

Nonpregnant uterine arteries were treated with 17 β -estradiol (E2 β , 0.3 nmol/L) or progesterone (P4, 100.0 nmol/L) separately for 48 hours. Myocytes were then isolated and BK_{Ca} current densities were determined at +80 mV in the presence of TEA. Data are means \pm SEM of 4 cells from 4 animals of each group. * *P* < 0.05, versus control.

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Figure 8. Effect of pregnancy and steroid hormones on $BK_{\mbox{Ca}}$ channel subunit expression in uterine arteries

Protein abundance of BK_{Ca} channel α and β 1 subunits was determined by Western blot analyses. **Panel A**: Freshly isolated nonpregnant (NPUA) and pregnant (PUA) uterine arteries. * *P* < 0.05, PUA versus NPUA. **Panel B**: Nonpregnant uterine arteries were treated with 17 β -estradiol (E2 β , 0.3 nmol/L) plus progesterone (P4, 100.0 nmol/L) for 48 hours. * *P* < 0.05, E2 β /P4 versus control. Data are means ± SEM of tissues from 5–6 animals of each group.