



Published in final edited form as:

Mol Genet Metab. 2011 December ; 104(4): 677–681. doi:10.1016/j.ymgme.2011.09.001.

Hepatorenal Findings in Obligate Heterozygotes for Autosomal Recessive Polycystic Kidney Disease

Meral Gunay-Aygun^{1,2}, Ismail Baris Turkbey³, Joy Bryant¹, Kailash T. Daryanani⁴, Maya Tuchman Gerstein¹, Katie Piwnica-Worms¹, Peter Choyke³, Theo Heller⁵, and William A. Gahl^{1,2}

¹Medical Genetics Branch, National Human Genome Research Institute, Bethesda, MD

²The Intramural Program of the Office of Rare Diseases, Bethesda, MD

³Molecular Imaging Program, National Cancer Institute, Bethesda, MD

⁴National Institutes of Health Clinical Center, Bethesda, MD

⁵National Institute of Diabetes and Digestive and Kidney Diseases

1.0 INTRODUCTION

Autosomal recessive polycystic kidney disease (ARPKD) is characterized by progressive cystic degeneration of the kidneys resulting in kidney failure and congenital hepatic fibrosis (CHF) often complicated with portal hypertension^{1–3}. Renal cysts in ARPKD originate from dilated collecting ducts. Most ARPKD patients present perinatally or in childhood; a minority present in adulthood.

ARPKD is caused by mutations in the *PKHD1* gene that encodes fibrocystin/polyductin, a protein that functions on primary non-motile cilia^{4,5}. ARPKD is the most common childhood-onset ciliopathy with an estimated frequency of 1 in 20,000. The carrier frequency for pathogenic *PKHD1* mutations is estimated to be approximately 1 in 70⁶. Given the recessive pattern of inheritance, parents of ARPKD patients are not expected to develop clinical features. However, this has not been systematically studied.

We performed ultrasound (USG) evaluations on 110 parents and a maternal grandmother from 64 independent ARPKD families. We identified various abnormalities, including multiple liver cysts and increased renal medullary echogenicity. Here we present, the imaging, clinical and molecular findings in this group of ARPKD family members.

2.0 METHODS

2.1 Patients

All patients and their parents were enrolled in the protocol, “Clinical Investigations into the Kidney and Liver Disease in Autosomal Recessive Polycystic Kidney Disease/Congenital Hepatic Fibrosis and other Ciliopathies” (www.clinicaltrials.gov, trial NCT00068224),

Address correspondence to: Meral Gunay-Aygun, M.D., NHGRI, NIH, 10 Center Dr. Bldg 10, Rm 10C103, Bethesda, MD 20892, 301 594 4181, mgaygun@mail.nih.gov.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

approved by the NHGRI Institutional Review Board and gave written informed consent. Patients who carried a clinical diagnosis of ARPKD made by a nephrologist were qualified to enroll. Evaluations of the index cases at the NIH Clinical Center included complete abdominal standard probe and high-resolution ultrasound (HR-USG) and magnetic resonance imaging (MRI), comprehensive biochemical testing of blood and urine and sequencing of the *PKHD1* gene. The clinical diagnosis of ARPKD for each index case at NIH was based upon established criteria^{2,7} including typical kidney and liver involvement on imaging and/or biopsy and autosomal recessive inheritance. All parents were evaluated by standard and HR-USG; abdominal MRI and magnetic resonance cholangiopancreatography (MRCP) were performed on parents with abnormalities on standard probe USG.

2.2 Sequencing and Analysis

Complete sequencing of *PKHD1* was performed in all patients. When DNA samples were available, mutation analysis in parents was performed to determine the parental origin of the mutations identified in the proband. Complete sequencing of the *PKHD1* gene was performed in selected parents with imaging abnormalities.

For the longest open reading frame of *PKHD1*, coding exons (2–67) and their intronic boundaries were sequenced in two directions using a Beckman CEQ 8000 system (Beckman Coulter, Inc., Fullerton, CA) and a contract with Agencourt (Beverly, MA). Sequencher Software (GeneCodes, Ann Arbor, MI) was used to analyze DNA variants. The pathogenicity of missense variants was evaluated as described⁸ using segregation analysis, general population frequencies, 3 computational prediction tools (Align GVGD [http://agvgd.iarc.fr/agvgd_input.php]; PolyPhen [<http://coot.embl.de/PolyPhen/>]; and SNAP [<http://cubic.bioc.columbia.edu/services/SNAP/>]) and the splice variant interpretation software NetGene2 Server (<http://www.cbs.dtu.dk/services/NetGene2/>).

2.3 Imaging Studies

Complete abdominal USG evaluations were performed using standard (4 MHz) and high resolution (7 MHz) ultrasonographic probes by a single technologist (K.T.D.) (AVI Sequoia Inc, Mountain View, CA). Magnetic resonance imaging (MRI) was performed on a 1.5 or 3 Tesla machine (Philips Medical Systems, NA, Bothell, Washington; General Electric Healthcare, Waukesha, WI, USA) without intravenous contrast media. USG, MRI and MRCP images were evaluated by a team of 2 radiologists (B.I.T and P.C).

3.0 RESULTS

3.1 Patient Characteristics

In 93 patients the clinical diagnosis of ARPKD was confirmed after NIH evaluations. One family contributed 4 affected siblings, 7 families contributed 2 siblings, and one family contributed an aunt and niece pair leaving 82 independent families. In 73 patients from 63 families the diagnosis was confirmed molecularly; clinical and mutational data from these 73 patients were previously published^{8,9}.

A total of 110 parents (62 mothers and 48 fathers) and a maternal grandmother from 63 independent ARPKD families were available for evaluation. Eighty five parents were from molecularly confirmed families and 25 were from ARPKD families in which the diagnosis was made based on clinical criteria. Mothers' ages ranged from 27 to 66 years (39 ± 7.2), fathers' ages ranged from 29 to 58 years (40 ± 6.3) and ages of the total group ranged from 27 to 66 years (39.5 ± 6.8).

3.2 Imaging Findings

Kidney USG showed increased medullary echogenicity in 6 mothers (5.5 %) whose ages ranged from 30 to 62 years (38.8 ± 11.8) (Table 1, Figure 1A–G). Multiple small liver cysts were identified in 10 parents (9 %) including 5 mothers and 5 fathers (Figure 2A–L). The ages of the parents with multiple liver cysts ranged from 36 to 49 years (44.2 ± 4.1). These liver cysts were mostly small and peripherally located. The findings of increased medullary echogenicity and liver cysts in all patients were detected by both standard and high resolution ultrasound probes. MRCP images were inconclusive in terms of the connectivity of the cysts to the biliary system (Figure 2A–F).

A 44 years old father with multiple liver cysts (Table 1, parent no: 16, Figure 2A) also had unilateral multiple parapelvic kidney cysts (Figure 1H). Multiple unilateral kidney cysts were detected in another parent (#17). A moderately increased liver echo pattern associated with coarse texture suggestive of CHF, was detected in 4 parents (table 1, parents 12, 14, 18 and 19, Figure 2L) including 2 with multiple liver cysts (parents 12 and 14) and two with splenomegaly (parents 14 and 19)¹⁰. A total of 9 parents had mild splenomegaly (spleen lengths 15–16 cm); in 5, this was in the absence of fatty changes of the liver. Incidentally, fatty changes in the liver suggested by increased echogenicity in association with smooth echo texture were detected in 20 parents¹⁰.

Two other parents with multiple liver cysts (Table 1, parent no 11 and 15) at ages 41 and 46, also had 2 kidney cysts each. Five other fathers ages 38 to 46 had single liver cysts. Two mothers at ages 41 and 46 and a father at age 36 had 2 renal cysts and single renal cysts were detected in 7 parents ages 36 to 49.

Other findings, which may be purely coincidental included gall bladder polyps in 3 parents, liver hemangioma in 3 mothers and a father, left duplicated collecting system in one mother and gall stones in one mother. The remainder of the abdominal USG examinations, including pancreas, were normal in all parents.

3.3 Case of Maternal Grandmother

A 76 year-old maternal grandmother was evaluated because she was documented to have CHF on liver biopsy performed at age 75 prompted by the incidental CT finding of abnormal liver parenchyma. USG imaging at NIH (Figure 2M) showed markedly increased liver echogenicity with moderately coarse echotexture; spleen volume was normal at 100 ml and kidneys were unremarkable.

3.4 Biochemical Findings

Liver and kidney-related serum and urine chemistries including serum cystatin C, PTH, and measurements of calcium and creatinine in 24 hour urines were performed in 6 patients including 3 mothers with increased medullary echogenicity (Table 1, parent no: 1, 3 and 5), 2 mothers with multiple liver cysts (Table 1, parents #10 and 11), and the maternal grandmother. All were within normal limits except for elevated alanine aminotransferase (44 U/L, normal 6–41), aspartate aminotransferase (43 U/L, normal 9–34) and gamma glutamyl transferase (151 U/L, normal 7–38) in the grandmother. Specifically, there was no evidence for hypercalciuria, hyperparathyroidism, or renal tubular acidosis.

3.5 *PKHD1* Sequencing Findings

Complete sequencing of the coding exons of *PKHD1* was performed on 16 parents with abnormal imaging findings and the maternal grandmother in an effort to identify additional mutations. These sequences confirmed the mutation transmitted to the proband but did not

reveal any other pathogenic variants. The maternal grandmother was heterozygous for the p.Thr36Met mutation in *PKHD1*.

4.0 DISCUSSION

Our findings on 110 ARPKD parents and a maternal grandmother suggest that carrier status for *PKHD1* mutations creates a predisposition to multiple liver cysts and renal involvement associated with increased medullary echogenicity on USG.

The ultrasound finding of increased medullary echogenicity is commonly reported as “nephrocalcinosis”, a term that describes the deposition of radiologically-demonstrable calcium salts in renal parenchyma^{11,12}. The differential diagnosis of “nephrocalcinosis” diagnosed radiologically includes true histopathological nephrocalcinosis and medullary sponge kidney (MSK), which itself can be associated with calcium depositions (Table 2)^{11,13}. Most cases of MSK are sporadic; the presence of rare dominantly inherited forms of MSK, and its association with different malformation syndromes, suggest that MSK is an inherited developmental disorder^{14–16}. The molecular basis of MSK remains largely unknown, two sequence variants of the *GDNF* gene were recently reported in a few MSK patients¹⁴. While intravenous pyelography is the gold standard for the diagnosis of MSK, computerized tomography has become the preferred diagnostic modality. MSK is clinically symptomatic in only a subset of patients and, therefore, is probably under-diagnosed¹⁵. The prevalence of MSK is estimated at 5 in 10 000 – 100 000^{14,15}. Based upon our findings, we speculate that carrier (heterozygous) status for *PKHD1* mutations may be one underlying molecular cause of MSK. Supporting this hypothesis, the microscopic pathology of MSK resembles that of ARPKD; both are characterized by dilated collecting ducts, although MSK involves only the precalyceal collecting ducts^{14,15}. Furthermore, MSK has been reported in patients with autosomal dominant polycystic kidney disease¹⁷ and congenital hepatic fibrosis¹⁸, most commonly caused by homozygous mutations in *PKHD1*. Given that the estimated carrier frequency for *PKHD1* mutations is 1 in 70 and 5 % of ARPKD parents have increased medullary echogenicity, our data suggest that the frequency of *PKHD1*-related MSK would be approximately 7 in 10 000.

Polycystic liver disease (PLD) occurs most commonly as a part of ADPKD. PLD without renal cysts, often referred to as isolated PLD, is genetically distinct from ADPKD^{19,20}. Clinical diagnostic criteria for PLD are available only for at-risk individuals; ²¹ in patients younger than 40 years, the presence of any liver cysts is considered to be diagnostic of PLD, while in those older than 40, 4 or more cysts are required to differentiate PLD from simple liver cysts²⁰. These criteria, however, need to be further vetted in a larger cohort of PLD patients. Isolated PLD is genetically heterogeneous¹⁹; approximately 20 to 30% of the isolated cases are due to mutations in either *PRKCSH*²² or *SEC63*; the molecular cause of the remainder of isolated PLD is unknown. *PRKCSH* encodes the beta-subunit of glucosidase II, an ER glucosidase that is involved in quality control of newly synthesized glycoproteins. *SEC63* encodes Sec63, an integral ER membrane protein that is a part of the multi-protein translocon, the translocation machinery for integral membrane and secreted proteins. It was recently shown that glucosidase II β and Sec63p are required in mice for adequate expression of a functional complex of the polycystic kidney disease gene products, polycystin-1, polycystin-2 and fibrocystin/polyductin encoded by *Pkhd1*^{23, 24}. This is supported by the fact that the histopathological features of isolated PLD are very similar to those of the PLD in ADPKD; in both disorders cysts originate from biliary microhamartomas, also termed von Meyenburg’s complexes^{25,26}. However, the reason why defects in glucosidase II β and Sec63 do not cause renal cystic disease is unknown.

The majority of PLD patients are clinically asymptomatic, with normal liver enzymes and intact synthetic function^{21,27}. The incidence of isolated PLD in autopsy series^{26,28,29} (2–3 in 10 000) is higher than its estimated clinical incidence (< 1 in 10 000), suggesting that most cases remain undetected. Our data suggest that heterozygous carriers of *PKHD1* mutations are predisposed to PLD. In all ARPKD parents with multiple liver cysts, these cysts were initially detected by standard probe ultrasound; in every case the number of cysts on initial evaluation exceeded 4. Based on the estimated carrier frequency for *PKHD1* mutations of 1 in 70, and our finding that 9 % of ARPKD parents have PLD, the frequency of *PKHD1*-related PLD would be approximately 1 in 1000. Given the small size of the liver cysts and normal liver-related blood chemistries in ARPKD parents, this condition is more likely to remain undiagnosed, explaining the discrepancy between disease frequencies.

In addition to CHF, ARPKD patients might have liver cysts that are in continuity with the biliary tree, differing from the isolated cysts in PLD. MRCP images of the ARPKD parents were not conclusive in terms of the connectivity of the cysts to biliary ducts. However, the small size and peripheral location of these cysts suggest that they are isolated from the biliary tree, similar to the PLD cysts. This difference in the characteristics of liver cysts between children with ARPKD and their parents is surprising. The increased liver echogenicity associated with coarse echotexture identified in some parents and the biopsy proven CHF in the maternal grandmother presented suggest some *PKHD1* carriers are also prone to CHF.

To exclude the possibility that these parents had two *PKHD1* mutations, we performed complete sequencing of the gene. However, we found no additional *PKHD1* mutations in these individuals. Other genes, including those encoding ciliary proteins, may contribute to cyst formation through synergistic heterozygosity^{30,31}; this could explain why only a subset of *PKHD1* mutation carriers develop these liver and or kidney findings.

In summary, our data suggest that carrier status for ARPKD is a predisposition to renal involvement associated with increased medullary echogenicity on USG (possibly MSK), and liver involvement in the form of asymptomatic PLD and CHF in some cases. All these *PKHD1* mutation carriers with abnormal kidney and/or liver imaging were clinically asymptomatic. It remains to be determined if some of these individuals might become symptomatic as they age.

Acknowledgments

We thank the ARPKD Alliance for their extensive support and the patients and their families who generously participated in this investigation. Supported by the Intramural Research Programs of the National Human Genome Research Institute, National Cancer Institute and the NIH Clinical Center.

6.0 References

1. Zerres K, Rudnik-Schoneborn S, Deget F, et al. Autosomal recessive polycystic kidney disease in 115 children: clinical presentation, course and influence of gender. Arbeitsgemeinschaft für Padiatrische, Nephrologie. Acta Paediatr. Apr; 1996 85(4):437–445. [PubMed: 8740301]
2. Guay-Woodford LM, Desmond RA. Autosomal recessive polycystic kidney disease: the clinical experience in North America. Pediatrics. May; 2003 111(5 Pt 1):1072–1080. [PubMed: 12728091]
3. Gunay-Aygun M, Avner ED, Bacallao RL, et al. Autosomal recessive polycystic kidney disease and congenital hepatic fibrosis: summary statement of a first National Institutes of Health/Office of Rare Diseases conference. J Pediatr. Aug; 2006 149(2):159–164. [PubMed: 16887426]
4. Onuchic LF, Furu L, Nagasawa Y, et al. PKHD1, the polycystic kidney and hepatic disease 1 gene, encodes a novel large protein containing multiple immunoglobulin-like plexin-transcription-factor

- domains and parallel beta-helix 1 repeats. *Am J Hum Genet.* May; 2002 70(5):1305–1317. [PubMed: 11898128]
5. Ward CJ, Hogan MC, Rossetti S, et al. The gene mutated in autosomal recessive polycystic kidney disease encodes a large, receptor-like protein. *Nat Genet.* Mar; 2002 30(3):259–269. [PubMed: 11919560]
 6. Zerres K, Rudnik-Schoneborn S, Steinkamm C, Becker J, Mucher G. Autosomal recessive polycystic kidney disease. *J Mol Med.* Apr; 1998 76(5):303–309. [PubMed: 9587064]
 7. Zerres K, Volpel MC, Weiss H. Cystic kidneys. Genetics, pathologic anatomy, clinical picture, and prenatal diagnosis. *Hum Genet.* 1984; 68(2):104–135. [PubMed: 6500563]
 8. Gunay-Aygun M, Tuchman M, Font-Montgomery E, Lukose L, Edwards H, Garcia A, Ausawarat S, Ziegler S, Piwnica-Worms K, Bryant J, Bernardini I, Fischer R, Huizing M, Guay-Woodford L, Gahl W. *PKHD1* Sequence Variations in 78 Children and Adults with Autosomal Recessive Polycystic Kidney Disease and Congenital Hepatic Fibrosis. *Molecular Genetics and Metabolism.* 2010 in press.
 9. Gunay-Aygun M, Font-Montgomery E, Lukose L, et al. Correlation of kidney function, volume and imaging findings, and *PKHD1* mutations in 73 patients with autosomal recessive polycystic kidney disease. *Clin J Am Soc Nephrol.* Jun; 2010 5(6):972–984. [PubMed: 20413436]
 10. Tchelepi H, Ralls PW, Radin R, Grant E. Sonography of diffuse liver disease. *J Ultrasound Med.* Sep; 2002 21(9):1023–1032. quiz 1033–1024. [PubMed: 12216750]
 11. Manz F, Jaschke W, van Kaick G, Waldherr R, Willich E. Nephrocalcinosis in radiographs, computed tomography, sonography and histology. *Pediatr Radiol.* 1980; 9(1):19–26. [PubMed: 7352106]
 12. Stout HA, Akin RH, Morton E. Nephrocalcinosis in routine necropsies; its relationship to stone formation. *J Urol.* Jul; 1955 74(1):8–22. [PubMed: 13243496]
 13. Hoppe B, Kemper MJ. Diagnostic examination of the child with urolithiasis or nephrocalcinosis. *Pediatr Nephrol.* Mar; 2010 25(3):403–413. [PubMed: 19104842]
 14. Torregrossa R, Anglani F, Fabris A, et al. Identification of *GDNF* gene sequence variations in patients with medullary sponge kidney disease. *Clin J Am Soc Nephrol.* Jul; 2010 5(7):1205–1210. [PubMed: 20448065]
 15. Gambaro G, Feltrin GP, Lupo A, Bonfante L, D'Angelo A, Antonello A. Medullary sponge kidney (Lenarduzzi-Cacchi-Ricci disease): a Padua Medical School discovery in the 1930s. *Kidney Int.* Feb; 2006 69(4):663–670. [PubMed: 16395272]
 16. Kuiper JJ. Medullary sponge kidney in three generations. *N Y State J Med.* Nov; 1971 71(22):2665–2669. [PubMed: 5286604]
 17. Torres VE, Erickson SB, Smith LH, Wilson DM, Hattery RR, Segura JW. The association of nephrolithiasis and autosomal dominant polycystic kidney disease. *Am J Kidney Dis.* Apr; 1988 11(4):318–325. [PubMed: 3354568]
 18. Kerr DN, Warrick CK, Hart-Mercer J. A lesion resembling medullary sponge kidney in patients with congenital hepatic fibrosis. *Clin Radiol.* Apr. 1962 13:85–91. [PubMed: 14455328]
 19. Tahvanainen P, Tahvanainen E, Reijonen H, Halme L, Kaariainen H, Hockerstedt K. Polycystic liver disease is genetically heterogeneous: clinical and linkage studies in eight Finnish families. *J Hepatol.* Jan; 2003 38(1):39–43. [PubMed: 12480558]
 20. Qian Q. Isolated polycystic liver disease. *Adv Chronic Kidney Dis.* Mar; 2010 17(2):181–189. [PubMed: 20219621]
 21. Qian Q, Li A, King BF, et al. Clinical profile of autosomal dominant polycystic liver disease. *Hepatology.* Jan; 2003 37(1):164–171. [PubMed: 12500201]
 22. Li A, Davila S, Furu L, et al. Mutations in *PRKCSH* cause isolated autosomal dominant polycystic liver disease. *Am J Hum Genet.* Mar; 2003 72(3):691–703. [PubMed: 12529853]
 23. Fedeles SV, Tian X, Gallagher AR, et al. A genetic interaction network of five genes for human polycystic kidney and liver diseases defines polycystin-1 as the central determinant of cyst formation. *Nat Genet.* 2011; 43(7):639–647. [PubMed: 21685914]
 24. Drenth JP, Martina JA, van de Kerkhof R, Bonifacino JS, Jansen JB. Polycystic liver disease is a disorder of cotranslational protein processing. *Trends Mol Med.* Jan; 2005 11(1):37–42. [PubMed: 15649821]

25. Desmet VJ. Congenital diseases of intrahepatic bile ducts: variations on the theme “ductal plate malformation”. *Hepatology*. Oct; 1992 16(4):1069–1083. [PubMed: 1398487]
26. Karhunen PJ, Tenhu M. Adult polycystic liver and kidney diseases are separate entities. *Clin Genet*. Jul; 1986 30(1):29–37. [PubMed: 3757294]
27. Everson GT, Scherzinger A, Berger-Leff N, et al. Polycystic liver disease: quantitation of parenchymal and cyst volumes from computed tomography images and clinical correlates of hepatic cysts. *Hepatology*. Nov-Dec; 1988 8(6):1627–1634. [PubMed: 3192176]
28. Melnick PJ. Polycystic liver; analysis of seventy cases. *AMA Arch Pathol*. Feb; 1955 59(2):162–172. [PubMed: 13227714]
29. Feldman M. Polycystic disease of the liver. *Am J Gastroenterol*. Jan; 1958 29(1):83–86. [PubMed: 13497992]
30. Dipple KM, McCabe ER. Phenotypes of patients with “simple” Mendelian disorders are complex traits: thresholds, modifiers, and systems dynamics. *Am J Hum Genet*. Jun; 2000 66(6):1729–1735. [PubMed: 10793008]
31. Vockley J, Rinaldo P, Bennett MJ, Matern D, Vladutiu GD. Synergistic heterozygosity: disease resulting from multiple partial defects in one or more metabolic pathways. *Mol Genet Metab*. Sep-Oct; 2000 71(1–2):10–18. [PubMed: 11001791]

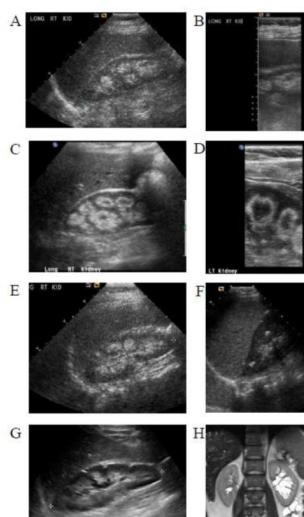


Figure 1.

Kidney USG images of parents of patients with ARPKD. Standard probe (A) and high resolution (B) renal USG images of 62 years old mother (Table 1, parent no 1), showing increased medullary echogenicity consistent with nephrocalcinosis. Standard probe (C) and high resolution (D) renal USG images of 33 years old mother (Table 1, parent no 6) showing medullary nephrocalcinosis occupying the outer rim of the medulla creating a donut like appearance involving the medullary pyramids. E. Renal USG of 33 year old mother (Table 1, parent no 2) demonstrating echogenic medulla consistent with nephrocalcinosis. F. Multiple echogenic foci within renal medulla of 39 year old mother (Table 1, parent no 4). G. Increased medullary echogenicity of 30 year old mother (Table 1, parent no 5). H. Multiple parapelvic cysts of left kidney of 44 year old father (Table 1, parent no 16), who also had innumerable small liver cysts (see Figure 2A).

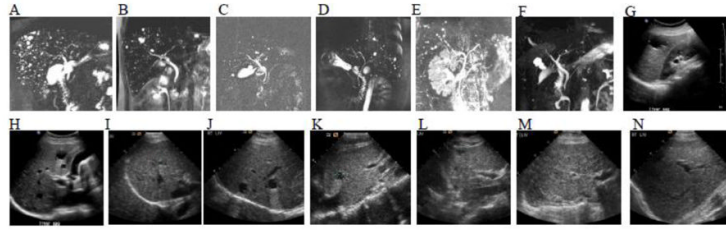


Figure 2.

MRCP and USG images of parents of patients with ARPKD showing multiple liver cysts. **A.** Innumerable small liver cysts ranging from a few mm to 1 cm are visible throughout the liver of 44 year old father (Table 1, parent no 16), who also had multiple parapelvic cysts of the left kidney (see Figure 1H); his right kidney appeared normal. **B.** MRCP of 41 year old mother (Table 1, parent no 11) showing innumerable small liver cysts; the majority are less than a half cm, a few are about a cm, the largest is about 2 cm. **C.** Multiple liver cysts, the largest about 1.3 cm adjacent to the gallbladder, others less than a centimeter in a 42 year old father (Table 1, parent no: 13). **D.** Innumerable small cysts present in both lobes of the liver of a 36 years old father (Table 1, parent no: 12). **E.** Multiple subcentimeter cysts throughout the liver of 46 years old mother (Table 1, Parent no: 15). MRCP (**F**) and USG images (**G** and **H**) of a 42 years old mother (Table 1, parent no: 10) demonstrating multiple liver cysts, the largest measuring 3 cm. **I.** Multiple liver cysts on USG of 49 years old father (Table 1, Parent no: 8). **J.** Forty nine year old father (Table 1, parent no: 9) with multiple liver cysts. **K.** Forty five years old mother (Table 1, parent no: 7) with multiple liver cysts on USG. **L.** USG of a 48 years old mother (Table 1, parent no: 14) demonstrating moderately echogenic liver with multiple cysts. **M.** Liver USG of the 76 year old grandmother showing heterogeneous appearance of the liver parenchyma with increased echogenicity in comparison to normal (**N**).

Table 1

Kidney and Liver imaging findings in parents of children with ARPKD

Parent No	Diagnosis in Offspring	PKHD1 mutation	Sex	Age	Affected status
1	ARPKD-mutation confirmed	p.Arg1624Trp	F	62	Bilateral nephrocalcinosis
2	ARPKD-clinical diagnosis	-	F	33	Bilateral nephrocalcinosis
3	ARPKD-mutation confirmed	p.Thr3035fs or p.Ala293Val**	F	36	Bilateral nephrocalcinosis
4	ARPKD-mutation confirmed	p.Thr36Met or p.Ile222Val**	F	39	Bilateral nephrocalcinosis
5	ARPKD-mutation confirmed	p.Thr36Met	F	30	Bilateral nephrocalcinosis
6	ARPKD-clinical diagnosis	-	F	33	Bilateral nephrocalcinosis
7	ARPKD-mutation confirmed	p.Thr36Met	F	45	Multiple liver cysts
8	ARPKD-mutation confirmed	p.Thr36Met	M	49	Multiple liver cysts
9	ARPKD-clinical diagnosis	-	M	49	Multiple liver cysts
10	ARPKD-mutation confirmed*	-	F	42	Multiple liver cysts
11	ARPKD-mutation confirmed	p.Thr36Met	F	41	Multiple liver cysts, 2 kidney cysts
12	ARPKD-mutation confirmed	p.Phe2374fs	M	36	Multiple liver cysts, moderately hyperechoic liver
13	ARPKD-mutation confirmed	p.Thr36Met	M	42	Multiple liver cysts, gall bladder polyp
14	ARPKD-mutation confirmed	p.Val1741Met	F	48	Multiple liver cysts, moderately hyperechoic liver, splenomegaly
15	ARPKD-mutation confirmed	p.Ile246Thr	F	46	Multiple liver cysts, splenomegaly, 2 kidney cysts
16	ARPKD-mutation confirmed	p.Trp2690Arg or IVS55+1G>A**	M	44	Multiple liver cysts, unilateral multiple kidney cysts
17	ARPKD-mutation confirmed	p.Arg2573Cys or unknown**	F	40	Unilateral multiple kidney cysts
18	ARPKD-mutation confirmed	p.Tyr486X	M	46	Moderately hyperechoic liver, single liver cyst, single kidney cyst
19	ARPKD-mutation confirmed	p.Thr3035fs or p.Ala293Val**	M	36	Moderately hyperechoic liver, splenomegaly
20	ARPKD-mutation confirmed	p_Ala1254fs	M	41	Gall bladder polyp
21	ARPKD-mutation confirmed	p.Thr36Met	M	44	Gall bladder polyps
22	ARPKD-clinical diagnosis	-	M	30	Splenomegaly

F: female, M: male,

* In this patient only one *PKHD1* mutation that was inherited from the father was identified,

** In these families parental origin of the mutations could not be determined since parental DNA was not available.

Table 2

Conditions associated with the radiological diagnosis of nephrocalcinosis

Primary hyperparathyroidism (OMIM #145000 and #145001)
Distal (type 1) renal tubular acidosis (OMIM #179800)
Medullary sponge kidney
Neonatal furosemide therapy
X-linked hypophosphatemic rickets (Phosphate and vitamin D) (OMIM #307800)
Cystinosis (OMIM #219800)
Hyperphosphatasia
Hypomagnesemia
Idiopathic hypercalciuria
Vitamin D intoxication
Hyperoxaluria (OMIM #259900)
Hypothyroidism
Cushing's syndrome
Prolonged immobilization
Bartter's syndrome (OMIM #607364)
Glycogen storage disease Type 1 (OMIM #232200)
Lowe's oculocerebrorenal syndrome (OMIM #309000)
Dent's Disease (OMIM #300009)
Acetazolamide
Tuberculosis
Sarcoidosis
Carrier status for autosomal recessive polycystic kidney disease