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# Dihydrochalcone glycosides from *Oxytropis myriophylla*

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## Abstract

Chemical investigations of the 70% alcohol extract of *Oxytropis myriophylla* (Pall.) DC. (Leguminosae) have afforded the new natural product neohesperidin dihydrochalcone (**1**) and the known phloretin-4'-O- $\beta$ -D-glucopyranoside (**2**), which was the first reported from the genus *Oxytropis*. This paper reports the isolation and full spectroscopic characterization of compounds **1** and **2** by NMR, UV, IR and MS data.

## Background

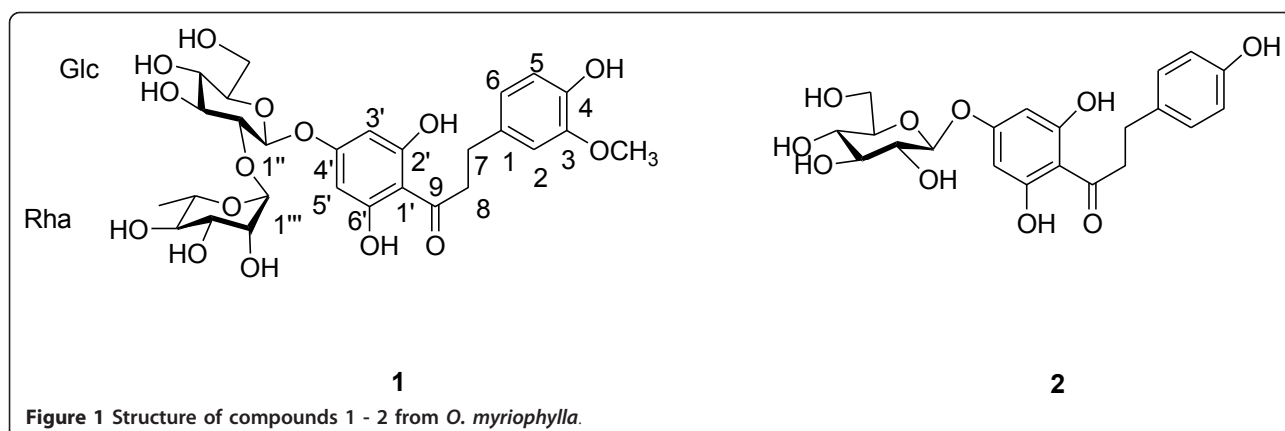
*Oxytropis* is an important genus of the family Leguminosae, also known as "Locoweed". The whole plant of *Oxytropis myriophylla* (Leguminosae) is an important Mongolian medicine, widely distributed in the southeast and northwest areas of China. It has traditionally been used to treat various diseases of rubella and influenza, applied to swelling and throat pain, together with different types of bleeding. Pharmacological research on this plant showed anti-oxidant and anti-inflammatory properties [1,2]. Primary investigation on this plant has led to the isolation of several flavonoids, triterpene glycosides and ligands [3,4], and in a continuation of studies on its minor constituents, the new natural product neohesperidin dihydrochalcone (**1**) and the known phloridzin (**2**), the first reported from the genus *Oxytropis* were obtained. This paper reports the isolation and full spectroscopic characterization of compound **1** by NMR, UV, IR and MS data.

## Results and Discussion

The phytochemical study of 70% ethanol extract obtained from the whole plant of *O. myriophylla* afforded two compounds, including a new natural product **1** neohesperidin dihydrochalcone and a known metabolite phloretin-4'-O- $\beta$ -D-glucopyranoside (**2**) (Figure 1). The structures of **1** and **2** were elucidated by detailed spectroscopic analysis and comparison of their spectroscopic data with those reported in the literatures.

Compound **1** was obtained as a white amorphous powder. Its molecular formula was assigned as  $C_{28}H_{36}O_{15}$  on the basis of the  $^{13}C$  NMR data and negative HRESIMS ( $m/z$  635.19586 [ $M + Na$ ] $^-$ ), calcd. for 635.19519. The UV spectrum exhibited absorption maxima at 284 nm (band II), which were suggestive of a dihydrochalcone skeleton. The IR spectrum of **1** illustrated a broad band at  $3374\text{ cm}^{-1}$  and  $1630\text{ cm}^{-1}$ , consistent with the presence of hydroxy group and carbonyl functions. The occurrence of a dihydrochalcone skeleton in the molecule could be easily deduced from the  $^1H$  NMR spectrum (Table 1), in which compound **1** showed the signals for an exchangeable proton at  $\delta$  12.23 (1H, s, H-6'), ABX-type aromatic protons at  $\delta$  {6.65 d,  $J = 2.0\text{ Hz}$ , H-2}, 6.60 [d,  $J = 8.0\text{ Hz}$ , H-5], 6.80 [dd,  $J = 8.0, 2.0\text{ Hz}$ , H-6]} on A-ring, one singlet at high field in the aromatic region  $\delta$  6.00 [s, H-3', 5'] on B-ring, together with two mutually coupled methylene groups [ $\delta$  3.27 (2H, t,  $J = 8.2\text{ Hz}$ , H-8) and 2.75 (2H, t,  $J = 8.2\text{ Hz}$ , H-7)]. These above data suggested the occurrence of a dihydrochalcone skeleton with the A-ring tri-substituted and B-ring penta-substituted in the molecule. In addition, the  $^1H$  NMR also exhibited signals due to one  $\beta$ -glucopyranosyl unit [ $\delta$  5.08 (d,  $J = 7.0\text{ Hz}$ , H-1'')]. The  $J$  value (7.0 xBHz) of the anomeric proton concluded the  $\beta$ -configuration of the glucose moiety. These NMR features were resembled to those of compound **2**, phloretin-4'-O- $\beta$ -D-glucopyranoside [5], except for the existence of an additional set of signals arising from one  $\alpha$ -L-rhamnopyranosyl [ $\delta$  5.04, (1H, br s, H-1''') and 1.25 (3H, d,  $J = 6.2\text{ Hz}$ , H-6''')], a methoxyl groups [ $\delta$  3.70 (3H, s)] reduced from  $^1H$  NMR,  $^{13}C$  NMR and HMQC-TOCSY in **1**. In addition, 1, 3, 4-trisubstituted phenyls in A-ring

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compound **1**, instead of 1, 4-disubstituted aromatic A-rings in the above-mentioned glycoside. The attachment of the glucopyranosyl moiety was deduced to be at C-4' according to glycosylation rule. It is observed that the chemical shifts of glucosyl C-2'' was downfield shifted to  $\delta$  77.1 in **1**, from  $\delta$  74.7 in **2**, suggesting that the  $\alpha$ -L-

rhamnopyranosyl group was located at the glucosyl C-2'' in **1**. The conclusion was further confirmed by the HMBC spectrum in which the anomeric proton of the glucopyranosyl moiety at  $\delta$  5.08 (d, H-1'') showed long range correlation with C(4') ( $\delta$  163.17). Moreover, the other correlations of H-(OCH<sub>3</sub>) ( $\delta$  3.70) with C-3, H-1'' ( $\delta$  5.04) with C-2'', respectively, in the HMBC spectrum confirmed the structure of compound **1**. Therefore, the structure of **1** was determined to be neohesperidin dihydrochalcone.

**Table 1** <sup>13</sup>C (125 MHz) and <sup>1</sup>H (500 MHz) NMR spectroscopic data for **1** in DMSO-d<sub>6</sub>.

| position         | $\delta_{\text{H}}$             | $\delta_{\text{C}}$ |
|------------------|---------------------------------|---------------------|
| 1                |                                 | 130.4               |
| 2                | 6.65 (1H, d, <i>J</i> = 2.0 Hz) | 115.7               |
| 3                |                                 | 146.3               |
| 4                |                                 | 145.8               |
| 5                | 6.60 (1H, d, <i>J</i> = 8.0 Hz) | 112.4               |
| 6                | 6.80 (1H, d, <i>J</i> = 8.0 Hz) | 118.7               |
| 7                | 2.75 (2H, t, <i>J</i> = 8.2 Hz) | 29.4                |
| 8                | 3.27 (2H, t, <i>J</i> = 8.2 Hz) | 45.4                |
| 9                |                                 | 204.9               |
| 1'               |                                 | 105.3               |
| 2'               |                                 | 162.9               |
| 3'               | 6.00 (1 H, s)                   | 94.8                |
| 4'               |                                 | 163.7               |
| 5'               | 6.00 (1 H, s)                   | 94.8                |
| 6'               |                                 | 162.9               |
| OCH <sub>3</sub> | 3.70 (3H, s)                    | 55.7                |
| 1''              | 5.08 (1H, d, <i>J</i> = 7.0 Hz) | 97.1                |
| 2''              | 3.50                            | 77.1                |
| 3''              | 3.34                            | 76.5                |
| 4''              | 3.50                            | 69.4                |
| 5''              | 3.37                            | 76.8                |
| 6''              | 3.50~3.65                       | 60.3                |
| 1'''             | 5.04 (1H, br s)                 | 100.6               |
| 2'''             | 3.82                            | 70.4                |
| 3'''             | 3.02                            | 70.5                |
| 4'''             | 3.02                            | 71.9                |
| 5'''             | 3.82                            | 68.3                |
| 6'''             | 1.25 (3H, d, <i>J</i> = 7.0 Hz) | 18.1                |

In a previous study, the structure of **1** as we presented in this paper is an artificial sweetener derived from citrus, which had been synthesized by neohesperidin [6]. The known compound, phloretin-4'-O- $\beta$ -D-glucopyranoside (**2**) was the first reported from the *Malus pumila* and this compound was found to show insect-antifeedant activity [7]. Particularly worth mentioning is phloretin-4'-O- $\beta$ -D-glucopyranoside (**2**) is bitter, as opposed to the sweetness of **1** [6]. The spectral data of **2** are in full agreement with those reported previously. To the best of our knowledge, the full NMR assignments of neohesperidin dihydrochalcone (**1**) are reported for the first time, on the basis of detailed spectroscopic analysis. And phloretin-4'-O- $\beta$ -D-glucopyranoside (**2**) has not previously been reported from any plant of *Oxytropis*.

## Experimental

### General

UV spectra were measured with a UV 210A Shimadzu spectrometer (Shimadzu, Kyoto, Japan). IR spectra were recorded on an IR-450 spectrometer (Shimadzu, Kyoto, Japan). <sup>1</sup>H and <sup>13</sup>C NMR, HMQC, HMBC, and HMQC-TOCSY spectra were performed in pyridine on a Bruker DRX-500 spectrometer (500 MHz for <sup>1</sup>H, and 125 for <sup>13</sup>C). ESIMS and HRESIMS were taken on an AutoSpe 3000 spectrometer (VG, Manchester, UK). The macroporous resin AB-8 (pore radius 130 ~ 140 Å, Tianjin Haiguang Chemical Factory, Tianjin, China), MCI-gel CHP20P (Mitsubishi Chemical Co., Kyoto, Japan), ODS-

A (YMC Co. Ltd., Kyoto, Japan), and silica gel (Qingdao Haiyang Chemical Co., Qingdao, China) were used for chromatography.

#### Plant material

The whole plant of *O. myriophylla* was collected from Inner Mongolia Province, China, in June 2006, and identified by Prof. Dong Yu in Inner Mongolia Medical College. The voucher specimen (No. 0648209) was deposited with the KUN Herbarium of Kunming Institute of Botany, Chinese Academy of Sciences.

#### Extraction and Isolation

The whole plant (3.0 kg) of *O. myriophylla* was cut into small pieces and extracted with 70% EtOH at refluxed temperature, and then concentrated in *vavco* to yield a crude extract (256 g). The EtOH crude extract (200 g) was chromatographed on a AB-8 (10 × 100 cm) and eluted with H<sub>2</sub>O:EtOH (1:0 - 0:1) and to give six fractions (0%, 30%, 50%, 70%, 90% and 100% EtOH fractions). The 70% EtOH fraction (12 g) was purified by MCI-gel CHP20P and ODS-A eluted with a step gradient of H<sub>2</sub>O:MeOH (1:0 - 0:1), and silica gel (CHCl<sub>3</sub>:MeOH:H<sub>2</sub>O, 9:1:0.1 - 7:3:0.5), respectively, to give **1** (12 mg) and **2** (27 mg).

#### Authors' contributions

GS carried out the chemical analysis-structure elucidation and drafted the Manuscript; SW carried out the chemical and biological studies; BL conceived of the study and its design and coordination of the scientific teams. All authors have read and approved the final manuscript.

#### Competing interests

The authors declare that they have no competing interests.

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