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Trace amine-associated receptor 1 is a stereoselective binding site for compounds in the amphetamine class

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Abstract

The demonstrated ability of amphetamine to functionally activate the rat trace amine associated receptor 1 (*r*TAAR1) and the subsequent reports of amphetamine activation of TAAR1 in rhesus monkey mouse, human, and human-rat chimeric TAAR1-expressing cell lines has led to speculation as to the role of this receptor in the central nervous system (CNS) responses associated with amphetamine and its analogs. The agonist potencies of ten pairs of enantiomeric amphetamines, including several with known CNS activity, at primate TAAR1 stably expressed in RD-HGA16 cells, robustly indicate the *S*-configuration to be associated with higher potency. Moreover, the rank order of potency to activate TAAR1 parallels the stimulant action reported by humans for the specific amphetamines. Taken together, these data suggest that TAAR1 is a stereoselective binding site for amphetamine and that activation of TAAR1 is involved in the modulation of the stimulant properties of amphetamine and its congeners. In addition, the observed parallel between *h*TAAR1 and *rh*TAAR1 responses supports the rhesus monkey as a highly translational model for developing novel TAAR1-directed compounds as therapeutics for amphetamine-related addictions.

Keywords

Amphetamine; Stereoselective; TAAR1; Binding site; phenylisopropylamine

1. Introduction

Compounds in the amphetamine class have been known for decades to be highly addictive substances¹ producing one or more of at least three distinct effects: an (*S*)-amphetamine-like effect, a DOM-like effect, and a PMMA-like effect.² The DOM-like (hallucinogenic) effect has been associated with activation of the 5-HT₂ family of receptors.^{3–5} In fact, the binding potencies of hallucinogenic amphetamines such as DOM, DOB, DOEt at 5HT_{2A/C} receptors have been shown to correlate with measures of hallucinogenic potencies in rodent and the (*R*)-enantiomer has been found to possess higher potency than the (*S*)-enantiomer,⁶ consistent with binding data at 5-HT_{2A/C} receptors. On the other hand, (*S*)-amphetamine,

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associated with stimulant properties, is bound at NET ($EC_{50} = 7.1$ nM) and DAT ($EC_{50} = 24.8$ nM) in cloned human cells⁷ and with low affinity at SERT ($EC_{50} = 1.77$ μ M)⁷ and at rat adrenergic alpha sites ($K_i \sim 1$ μ M)⁸ but not at 5-HT receptors. Binding sites for (*S*)-[³H]amphetamine have been reported in rat brainstem, hypothalamus, and striatum.⁹ Further studies have characterized two sites for [³H]amphetamine: a low affinity sequestration-site lacking stereospecificity¹⁰ and a high affinity, stereoselective site of (*S*)-[³H]amphetamine incorporation in striatal synaptosomes.¹¹ However, these studies did not implicate either of these two [³H]amphetamine sites with (*S*)-amphetamine-like stimulant effects. The demonstrated ability of amphetamine to functionally activate the rat trace amine associated receptor 1 (*r*TAAR1)¹² and the subsequent reports of amphetamine activation of TAAR1 in rhesus monkey,¹³ rat,¹⁴ mouse,^{14–16} human,^{15, 17} and human-rat chimeric¹⁴ TAAR1-expressing cell lines has led to speculation as to the role of this receptor in the psychostimulant, hallucinogenic and addictive effects associated with amphetamine and its analogs¹⁸ as well as to the suggestion that TAAR1 may contribute a novel mode of action to these hallucinogenic drugs.¹⁹ A recent study of the activation of mouse, rat, and human-rat chimeric TAAR1s by amphetamine, methamphetamine, and *p*-hydroxyamphetamine¹⁴ concluded that this receptor could be a mediator of the effects of these drugs.

We had expressed wild type *h*TAAR1 in CHO cells stably expressing $G_{\alpha 16}$ ^{17, 20} and had developed a high throughput assay for functional TAAR1 agonists²⁰ that we have been using to evaluate a series of amphetamines. Here, we have similarly expressed *rh*TAAR1 and have collected binding data on a spectrum of stereoisomers of amphetamines, including several with known CNS activity, that demonstrate that primate TAAR1 is a stereoselective binding site for compounds in the amphetamine class. The findings suggest an opportunity for the rhesus as a valid model for assessing whether specific TAAR1-active agents may have therapeutic efficacy in humans.

2. Methods

2.1 Cell Lines

2.1.1 Human TAAR1—A cell line expressing *h*TAAR1 was developed as previously described.²⁰ Briefly, *h*TAAR1 cDNA was cloned from Marathon Ready cDNA from human stomach using the Advantage cDNA PCR kit (Clontech, Mountain View, CA). The receptor coding sequence was amplified in 2 parts using primer pairs based on GenBank accession no. AF380185. The 2-part strategy was used because we had difficulty obtaining the full-length cDNA with 1 set of primers. The resultant PCR products were sub-cloned separately into the pcDNA4/HisMax TOPO TA expression vector (Invitrogen, Carlsbad, CA). The cloned PCR products were verified by sequence analysis (Duke University DNA Analysis Facility, Durham, NC). The full-length *h*TAAR1 cDNA was generated by restriction digestion of the 5' and 3' cDNAs and subsequent subcloning into the pcDNA4/HisMax TOPO TA expression vector. Upon reconsideration, we believed that the N-terminal 6 \times his tag in this vector might interfere with intracellular trafficking of the receptor by masking the native N-terminal. We therefore subcloned the coding region into an expression vector without an N-terminal tag. This expression vector contains an EF-1.2 promoter, a neomycin resistance gene, and a C-terminal HA epitope tag (pCEFL). The endogenous stop codon of *h*TAAR1 is included so the HA tag is not incorporated into the expressed *h*TAAR1 protein.

Plasmid DNA from the resulting construct was prepared using Qiagen's Hispeed plasmid purification kit as per manufacturer's instructions. The sequence was verified, and DNA was linearized by *ScaI* digestion prior to transfection of cells. The *h*TAAR1 expression construct was transfected into RD-HGA16 cells (Molecular Devices Corporation, Sunnyvale, CA). RD-HGA16 cells stably express the promiscuous Gq protein, $G_{\alpha 16}$. Expression of $G_{\alpha 16}$

allows coupling of *hTAAR1* to calcium mobilization. These cells were transfected with the *hTAAR1* expression construct using Lipofectamine 2000 reagent as per manufacturer's instructions. Stably transfected cells were selected in 400 $\mu\text{g/ml}$ geneticin (*hTAAR1* selection) and 400 $\mu\text{g/ml}$ hygromycin (*G α 16* selection). Clones were selected from low-density cultures and analyzed for response to β -PEA using the Calcium 3 assay (Molecular Devices) and a FlexStation II³⁸⁴ (Molecular Devices) as per manufacturer's instructions. Three positive clones were chosen for further experiments based on their positive response to β -PEA. All data are from 1 clonal cell line of *hTAAR1*/RD-HGA16 cells.

2.1.2 Rhesus TAAR1—A mammalian expression plasmid (pcDNA3.1 TOPO, Invitrogen) carrying an antibiotic resistance gene and the coding sequence for rhesus monkey TAAR1¹³ was transiently transfected into RD-HGA16 cells using Lipofectamine Plus transfection reagent as per manufacturer's instructions. The cells were transferred to 15 10-cm tissue culture dishes and subjected to antibiotic selection as described above for the *hTAAR1* cells. Surviving clones were expanded and assayed for increased internal calcium concentration in response to treatment by a TAAR1 agonist using a calcium 4 kit and a FlexStation II³⁸⁴ fluorescence plate reader. For *rhTAAR1*, octopamine was the agonist of choice since it had been found that it was a superagonist relative to β -PEA (which we had used for *hTAAR1*).²¹

Screening of 400 clones that were obtained by transfection of the rhesus expression plasmid into RD-HGA16 cells, and that survived antibiotic selection over several transfection events, gave seven "hits." However, none of these showed a reproducible response to 10 μM octopamine on subsequent analysis. Because our successful generation of stable cell lines expressing *hTAAR1* had used the proprietary expression vector pCEFL, which was different from the vector containing the *rhTAAR1*, the *rhTAAR1* coding sequence was subcloned into the pCEFL vector by restriction enzyme digestion, gel extraction, and ligation. Stable cell line generation using this expression plasmid was performed as above. Of 96 clones screened over one transfection event, ten "hits" were identified. Further analysis led to the identification of two cell lines stably expressing the *rhTAAR1*.

3. Results

The potencies of ten pairs of enantiomeric amphetamines (**1–10**) to activate *hTAAR1*^{17, 20} and *rhTAAR1* stably expressed in RD-HGA16 cells, determined as previously described,²⁰ are shown in Table 1. While all the data suggest possible stereoselectivity for the *S*-configuration (see Figure 1), the data for activation of *hTAAR1* are not compelling for five (**2, 3, 5, 8, 10**) of the ten compound pairs. However, the potency data for activation of *rhTAAR1* robustly indicate the *S*-configuration to be preferred for four of these five compounds (**2, 3, 5, 8**). The data for methamphetamine (**10**) are ambiguous.

4. Discussion

While a considerable body of quantitative data relative to potencies to activate *m*- and *r*TAAR1 has been reported,^{22–25} particularly for thyronamine and its analogs, no such data are reported for *rhTAAR1*, and only scant data are available for *hTAAR1*. For the few compounds where cross species comparisons have been possible, differences between human and rodent responses to structural variations have been observed. For example, addition of a *p*-hydroxyl functionality to the aromatic ring of β -PEA (to give tyramine) led to three-fold decreased potency for stimulation of cAMP formation in HEK 293 cells expressing *hTAAR1* modified by replacing the N-terminal amino acids 1–20, the C-terminal amino acids 305–340, and the third intracellular loop corresponding to amino acids 204–258 with the corresponding rat TAAR1 sequences (*h-rTAAR1*), but resulted in five-fold increase in potency in *rTAAR1* cells and in a two-fold reduction in *mTAAR1* cells.^{15, 26} Wainscott et

al. noted that addition of a *p*-hydroxyl functionality to the aromatic ring of β -PEA led to four-fold decreased potency for stimulation of cAMP formation in *hTAAR1*-expressing rGasAV12-664 cells, while potency for stimulation of cAMP formation in *mTAAR1*-expressing rGasAV12-664 cells was increased by an order of magnitude;¹⁵ analogous results were reported for EC₅₀ values for cAMP accumulation in HEK cells stably expressing *rTAAR1* and the chimeric *h-rTAAR1*.¹⁴ Similarly, while the potencies for β -PEA in human and mouse TAAR1 were almost the same (EC₅₀ = 106 nM and 209 nM, respectively), they were two orders of magnitude different for 3-iodothyronamine (EC₅₀ = 1510 nM and 22.4 nM, for human and mouse, respectively). Addition of a *p*-hydroxy functionality to amphetamine virtually abolished activity at *h-rTAAR1* but had minimal effect in *rTAAR1* and a stereoselective effect in *mTAAR1*.¹⁴ These observations may be due to, as has been pointed out,¹³ the significant divergence in sequence between human and rodent TAAR1s (76–78%).²⁷ Since the *rhTAAR1* coding sequence had been found to be 96.9% homologous to the *hTAAR1* coding sequence,²⁸ we undertook to develop a stable expression of *rhTAAR1* in cells stably expressing G _{α 16}, by analogy to our previously developed expression system for *hTAAR1*^{17, 20, 29} for the purpose of making direct comparisons between *hTAAR1* and *rhTAAR1*.

Only scant data relative to the potencies of amphetamine and its analogs to activate TAAR1 are currently available (Table 2). The available data are in reasonable agreement considering that the evaluations have been carried out in different laboratories and using different expression systems and bioassay methodologies. A significant exception are the data reporting (*S*)-amphetamine, with EC₅₀ = 2 nM, to be the most potent agonist identified for *mTAAR1*.¹⁶ Our results (Table 1) for amphetamine (**1**) are similar to the literature data. Specifically, the EC₅₀ (0.6 ± 0.002 μ M) obtained by us for (*S*)-amphetamine ((*S*)-**1**) to stimulate calcium flux by activation of *hTAAR1* transfected into RD-HGA16 cells is in between the EC₅₀ values obtained by evaluating the effectiveness of (*S*)-**1** to promote cAMP accumulation in *hTAAR1*-expressing rGasAV12-664 cells (0.99 ± 0.16 μ M),¹⁵ and by measuring the cAMP signaling associated with activation of *hTAAR1* modified by the addition of a glycosylation site by insertion of the first nine amino acids of the human β 2-adrenergic receptor between the HA-tag and the N-terminus (0.14 ± 0.01 μ M).³⁰ Similarly, for (*R*)-amphetamine ((*R*)-**1**) our EC₅₀ value of 1.3 ± 0.3 μ M, is in the range of 1.7 ± 0.29 μ M and 0.25 ± 0.01 μ M reported by Wainscott¹⁵ and Barak,³⁰ respectively. Our data diverge significantly from those of Barak et al.³⁰ for (*S*)-3,4-methylenedioxymethamphetamine ((*S*)-**4**). In fact, the potency (0.37 ± 0.05 μ M) reported by Barak et al.³⁰ for (*S*)-**4**, is surprisingly high, particularly when considering that effects on potency reported by Barak et al. parallel our previously reported findings for analogs of β -PEA.¹⁷ For example, the EC₅₀ reported for (*S*)-methamphetamine (1.3 ± 0.19 μ M) is in good agreement with the value determined by us (1.5 ± 0.4 μ M), suggesting that N-methylation tends to decrease potency at *hTAAR1*, in agreement with our finding that N-methylation of β -PEA decreased potency at *hTAAR1* by a factor of three.¹⁷ Since we had found that 3,4-methylenedioxy substitution onto the aromatic moiety of β -PEA decreased potency at *hTAAR1* by at least an order of magnitude, an EC₅₀ value of 30 μ M might be expected for (*S*)-MDMA ((*S*)-**4**) in reasonable agreement with the value obtained by us (Table 1) but much higher than the value reported by Barak et al.³⁰ Overall, introduction of oxygenated substituents at the aryl moiety of amphetamine (see **2**, **3**, **4**, **5**, **7**, **8**, **9**) decreased potency at *hTAAR1*, as had been noted for analogous substitution in β -PEA.¹⁷

Based on studies carried out in rat brain synaptosomal preparations it has been suggested that the discrete behavioral effects of high and low doses of (*S*)-amphetamine ((*S*)-**1**) might be attributable to the low affinity amphetamine sequestration site and the high affinity amphetamine transport site, respectively.^{10, 11} Specifically, low doses of (*S*)-**1** would be bound by the high affinity amphetamine transport site thereby leading to increased

monoamine release. Studies of the effects of ephedrine-related structures on biogenic amine uptake and release in stably and transiently transfected cells expressing mainly human cloned transporters demonstrated high activity for (*S*)-**1** at norepinephrine ($EC_{50} = 7.1$ nM) and dopamine ($EC_{50} = 24.8$ nM) transporters,⁷ and it was suggested that DA release by (*S*)-**1** may contribute to its behavioral effects. Co-localization of TAAR1 with DAT in some, but not all, dopamine neurons has been reported, and it has been shown that activation of TAAR1 plays a mediatory role in DAT regulation.^{31, 32} In particular, it has been shown that β -PEA and methamphetamine effects in cells expressing TAAR-DAT significantly exceed those observed in cells expressing DAT only. Consistent with this conclusion is the higher potency of (*S*)-**1** in rat synaptosomes relative to cloned human DAT cells (EC_{50} 60 nM vs 240 nM). These data suggest that the efficacy of different TAAR1 agonists at DAT would be related to their affinity for TAAR1 and their efficiency as substrates at the DAT. It is thus possible that the stimulant properties of (*S*)-amphetamine ((*S*)-**1**), and of analogs that generalize to (*S*)-**1**, may be regulated by TAAR1. Some of the similarities between behaviors elicited by β -PEA and (*S*)-**1**, which were observed decades ago, may also be accounted for by binding at TAAR1. For example, β -PEA was found to be as effective as amphetamine in a place preference paradigm in rats although it was significantly less potent than either amphetamine isomer.³³

It has been long known that the addition of substituents to the aromatic moiety of amphetamine results in cognitive and behavioral outcomes distinct from those associated with (*S*)-amphetamine. Thus, an early study described (*S*)-**1** as a central stimulant with anorectic properties and effects on cardiovascular and thermoregulatory processes.³⁴ The same report described the effects of aryl substituted phenylisopropylamines as retaining amphetamine-like effects while exhibiting LSD-like activity.³⁴ Considering what relevance the *in vitro* potencies of the series of psychoactive amphetamines to activate primate TAAR1 may have to the *in vivo* effects of these compounds in primates is clearly premature. First, because the distribution parameters in primates are not known and, second, no validated studies in primates are available. Keeping these caveats in mind, it is nevertheless interesting to note that our results are consistent with the possibility that activation of TAAR1 contributes to amphetamine-like effects. In particular, our data show (*S*)-**3** to be twice as potent as (*R*)-**3** at both *rh*TAAR1 and *h*TAAR1, and (*R*)-**4** to be inactive at *h*TAAR1 while (*S*)-**4** retained weak activity, consistent with the observation that for MDA (**3**) and MDMA (**4**) the (*S*)- isomers, but not the (*R*)-isomers, substitute for (*S*)-amphetamine in drug discrimination studies in rats^{2, 35} and with the notion that psychomotor stimulant activity is associated with the (*S*)-configuration of these agents.³⁶ In humans doses of 100 mg (or greater) of (*S*)-**3** or (*S*)-**4**, but not of (*R*)-**3** or (*R*)-**4**, have been reported to lead to excitation.³⁷ Similar effects have been reported for 2.6 mg doses of (*S*)-DOM ((*S*)-**8**), but not for the enantiomeric (*R*)-**8**;³⁷ our data show (*S*)-**8** to be equipotent to (*S*)-**1** in *rh*TAAR1, consistent with the lower effective *in vivo* dose of (*S*)-**8** relative to (*S*)-**3** or (*S*)-**4**.³⁷ The lack of observed agonist activity for (*S*)-**8** in *h*TAAR1 is surprising. Our data show (*S*)-*m*-methoxyamphetamine ((*S*)-**5**) to have relatively high potency at both *rh*TAAR1 and *h*TAAR1, suggesting that (*S*)-**5** would have significant amphetamine-like effects. Drug discrimination studies in rats showed racemic *m*-methoxyamphetamine (**5**) to produce amphetamine-like effect and to generalize to (*S*)-**1**³⁸ but not to DOM (**8**); examination of the individual enantiomers of **5** in drug discrimination studies showed significantly reduced response rates and disruption of behavior by both enantiomers.³⁹ Relatively high potency at *h*TAAR1 has also been found for (*S*)-2,5-dimethoxy-4-ethylamphetamine ((*S*)-**9**), again suggesting that it may produce amphetamine-like effects. The literature data for racemic (*S*)-**9** imply some anxiogenic effects in humans at low dose (25 mg);³⁷ but no clear-cut amphetamine-like stimuli of either of the individual enantiomers or of the racemate were detected in a study in which human subjects were given oral doses of 1–4 mg of **9**.⁴⁰ The bromo analog (*S*)-**7** has been reported to have no effect in humans at doses of 0.5–1.0 mg. If,

indeed, psychostimulant effects are associated with activation of TAAR1 this observation may be consistent with the low agonist potency observed for (*S*)-**7** in *h*TAAR1. There are no literature reports concerning the effects of (*S*)-**7** in rhesus monkeys. While we did not collect quantitative data on *N*-norfenfluramine (**6**), the fact that only (*S*)-**6** was found to be active at *h*TAAR1 is consistent with the finding that (*S*)-**6** was more potent than (*R*)-**6** in counteracting amphetamine induced increase in locomotor activity in rats.⁴¹ While this effect is likely to be associated with activation of 5-HT₂ activation, it does not preclude involvement of TAAR1.

While our data suggest a role for TAAR1 in eliciting amphetamine-like stimulant effects, it must be borne in mind that the observed *in vivo* effects are likely to result from interaction with both TAAR1 and monoamine transporters. Thus it has been shown that the selective TAAR1 agonist RO5166017 fully prevented psychostimulant-induced and persistent hyperdopaminergia-related hyperactivity in mice.⁴² This effect was found to be DAT-independent, since suppression of hyperactivity was observed in DAT-KO mice.⁴²

The collected information leads us to conclude that TAAR1 is a stereoselective binding site for amphetamine and that TAAR1 activation by amphetamine and its congeners may contribute to the stimulant properties of this class of compounds. Since the observed *S*-stereoselectivity for activation of TAAR1 is inconsistent with the known *R*-stereoselectivity for hallucinogenic activity of compounds in the amphetamine class, we conclude that TAAR1 is not a mediator of hallucinogenic activity of these agents. Our results show a reasonable parallel between *h*TAAR1 and *rh*TAAR1 responses to a series of substituted amphetamines, supporting the rhesus monkey as a highly translational model for developing novel TAAR1-directed compounds as therapeutics for amphetamine-related addictions.

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Abbreviations

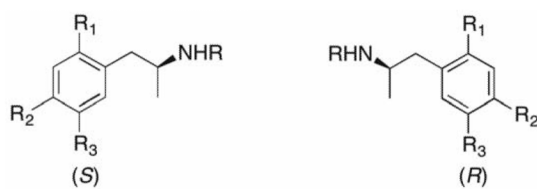
TAAR	trace amine-associated receptor
NET	norepinephrine transporter
DAT	dopamine transporter
SERT	serotonin transporter
5-HT	5-hydroxytryptamine
LSD	lysergic acid diethylamide
CHO	Chinese hamster ovary
DOM	2,5-dimethoxy-4-methylamphetamine
PMMA	p-methoxymethamphetamine
MDA	3,4-methylenedioxyamphetamine
MDMA	3,4-methylenedioxymethamphetamine

CNS	central nervous system
DOB	4-Bromo-2,5-dimethoxyamphetamine
DOEt	2,5-Dimethoxy-4-ethylamphetamine
KO	knock out

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Compound No.	R1	R2	R3	R
1	H	H	H	H
2	H	OH	H	H
3	H	OCH ₂ O		H
4	H	OCH ₂ O		CH ₃
5	H	H	OCH ₃	H
6	H	H	CF ₃	H
7	OCH ₃	Br	OCH ₃	H
8	OCH ₃	CH ₃	OCH ₃	H
9	OCH ₃	CH ₂ CH ₃	OCH ₃	H
10	H	H	H	CH ₃

Figure 1.
Structures of Enantiomeric Amphetamines

Table 1

Potencies to activate TAAR1

No	Compound Name	EC ₅₀ (μM)	
		Human	Rhesus
(S)-1	(S)-amphetamine	0.6 ± 0.002	1.01 ± 0.08
(R)-1	(R)-amphetamine	1.3 ± 0.03	1.86 ± 0.03
(S)-2	(S)- <i>p</i> -hydroxyamphetamine	2.96 ± 0.1	0.101 ± 0.02
(R)-2	(R)- <i>p</i> -hydroxyamphetamine	3.16 ± 1.2	0.424 ± 0.2
(S)-3	(S)-3,4-methylenedioxyamphetamine	6.57 ± 1.2	1.35 ± 0.3
(R)-3	(R)-3,4-methylenedioxyamphetamine	11.73 ± 6.3	2.48 ± 0.5
(S)-4	(S)-3,4-methylenedioxymethamphetamine	73.7 ± 31	16.1 ± 6.8
(R)-4	(R)-3,4-methylenedioxymethamphetamine	inactive	7.4 ± 0.5
(S)-5	(S)- <i>m</i> -methoxyamphetamine	1.9 ± 0.9	3.3 ± 1.8
(R)-5	(R)- <i>m</i> -methoxyamphetamine	6.5 ± 4.5	inactive
(S)-6	(S)-norfenfluramine	active*	ND
(R)-6	(R)-norfenfluramine	inactive	ND
(S)-7	(S)-4-bromo-2,5-dimethoxyamphetamine	15.34 ± 0.1	2.14 ± 0.08
(R)-7	(R)-4-bromo-2,5-dimethoxyamphetamine	31.94 ± 6.6	13.9 ± 5
(S)-8	(S)-4-methyl-2,5-dimethoxyamphetamine	inactive	0.9 ± 0.04
(R)-8	(R)-4-methyl-2,5-dimethoxyamphetamine	inactive	inactive
(S)-9	(S)-4-ethyl-2,5-dimethoxyamphetamine	5.02 ± 1.8	inactive
(R)-9	(R)-4-ethyl-2,5-dimethoxyamphetamine	inactive	inactive
(S)-10	(S)-methamphetamine	1.5 ± 0.4	5.3 ± 0.5
(R)-10	(R)-methamphetamine	3.3 ± 1.7	2.5 ± 1.2

ND= not determined

* 43% of maximum in screen, using 10 μM in hTAAR1/G 16 CHO-K1 cells loaded with Calcium 4 dye and analyzed using a FlexStation 2 microplate reader (Molecular Devices).

Table 2

Literature Data for Potencies to Activate TAAR1

Compound	EC ₅₀ (μM)			
	Rat	Mouse	Human-rat	Human
<i>(S)</i> -1	0.81 ¹⁴	0.21 ¹⁴	1.12 ¹⁴	0.99 ± 0.16 ¹⁵
	0.44 ± 0.01 ¹²	0.002 ± 0.001 ¹⁶		0.14 ± 0.01 ³⁰
	1.2 ± 0.07 ¹⁵			
<i>(R)</i> -1	0.28 ¹⁴	4.96 ¹⁴	3.09 ¹⁴	1.7 ± 0.29 ¹⁵
	0.21 ± 0.04 ¹²	0.065 ± 0.053 ¹⁶		0.25 ± 0.01 ³⁰
	1.4 ± 0.6 ¹⁵			
<i>(rac)</i> -2	0.05 ± 0.01 ¹²			
<i>(S)</i> -2	0.19 ¹⁴	0.28 ¹⁴	>5.42 ¹⁴	
<i>(R)</i> -2	0.06 ¹⁴	5.65 ¹⁴	>8.54 ^{12,14}	
<i>(rac)</i> -4	1.7 ± 1.2 ¹²			
<i>(S)</i> -4				0.37 ± 0.05 ³⁰
<i>(S)</i> -10	0.89 ¹⁴	0.92 ¹⁴	4.44 ¹⁴	1.3 ± 0.19 ³⁰
		0.07 ± 0.08 ¹⁶		
<i>(R)</i> -10	1.19 ¹⁴	2.44 ¹⁴	9.83 ¹⁴	