

Structure of a bovine gene for P-450c21 (steroid 21-hydroxylase) defines a novel cytochrome P-450 gene family

(adrenal gland/steroid hormone/molecular evolution/DNA sequence)

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ABSTRACT P-450c21, a cytochrome P-450 enzyme [steroid 21-monooxygenase (steroid 21-hydroxylase), EC 1.14.99.10], mediates the 21-hydroxylation of glucocorticoid and mineralocorticoid hormones in the adrenal gland. The complete sequence of a bovine P-450c21 gene shows it is 3447 base pairs long and contains 10 exons. The intron/exon organization and encoded amino acid sequence indicate that P-450c21 represents a unique family of genes in the P-450 gene superfamily. Primer extension and S1 nuclease protection experiments identified several cap sites for initiation of transcription; the principal cap site produces mRNA with a 5' untranslated region only 11 bases long. S1 nuclease protection experiments confirm that P-450c21 is actively expressed in the adrenal and the testis, an organ not known to secrete 21-hydroxylated steroids.

21-Hydroxylation is a key enzymatic step in the synthesis of glucocorticoid and mineralocorticoid hormones. The 21-hydroxylation of progesterone to desoxycorticosterone or of 17-hydroxyprogesterone to 11-desoxycortisol is mediated by P-450c21, a cytochrome P-450 enzyme [steroid 21-monooxygenase (steroid 21-hydroxylase), EC 1.14.99.10] bound to endoplasmic reticulum. 21-Hydroxylation was the first enzymatic activity ascribed to any cytochrome P-450 (1). Cytochrome P-450s metabolize xenobiotic agents and endogenous steroid substrates. Although cytochrome P-450s are related in size, sequence, and spectral characteristics, they are encoded by several widely divergent families of genes belonging to a large gene superfamily (2). P-450c21 is of great medical interest as about 1 in 5000 persons has an autosomal recessive disorder of P-450c21 causing congenital adrenal hyperplasia, variously manifested by genital ambiguities, virilization, cardiovascular collapse, and death (3). We recently identified a bovine genomic DNA clone containing a P-450c21 gene and showed that cattle have two copies of this gene and synthesize two sizes of P-450c21 mRNA in the adrenal cortex (4). We now report the sequence of >6.5 kilobases (kb) of bovine genomic DNA containing this gene. Nuclease S1 protection experiments indicate this gene is active in the bovine adrenal cortex and, surprisingly, in the testis, which is not known to secrete 21-hydroxylated steroids. Analysis of the structure of this gene indicates that this P-450c21 gene is only distantly related to other P-450 gene families and hence represents a novel member of the P-450 gene superfamily.

MATERIALS AND METHODS

DNA sequencing was done by the dideoxy method using ³⁵S-labeled dNTPs and gradient gels (5).

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The 27-base primer 3' GACGATCGACCGCGAGTG-GACGATACC 5' complementary to bases 45-71 was mechanically synthesized, purified, and labeled (4). For primer extension, 0.1 pmol of ³²P-labeled primer and 10 μg of bovine adrenocortical poly(A) RNA were hybridized for 2 hr at 65°C in 10 μl of 0.3 M NaCl/5 mM EDTA/10 mM Tris·HCl, pH 8.3, and then made to 50 μl, having final concentrations of 1 mM (each) dNTP, 10 mM Tris (pH 8.3), 10 mM dithiothreitol, 10 mM MgCl₂, 640 units of human placental ribonuclease inhibitor (Amersham) per ml, and 500 units of reverse transcriptase (Life Sciences, St. Petersburg, FL) per ml. After 30 min at 42°C, the reaction was stopped with 4 μl of 0.25 M EDTA.

S1 nuclease mapping was modified from Sudhof *et al.* (6). One picomole of ³²P-labeled 27-mer was used to initiate DNA synthesis on an M13 clone of a 412-base *HincII/Pst I* fragment of the P-450c21 gene corresponding to nucleotides -275 to +137 employing unlabeled dNTPs and the Klenow fragment of DNA polymerase I. The reaction proceeded 30 min at 25°C and then was heated to 65°C for 10 min. After *Bam*HI digestion, the labeled, 359-base oligonucleotide-initiated probe was gel-purified (4). About 10⁴ cpm of 359-base probe and 50 μg of total RNA in 20 μl of 80% deionized formamide/0.4 M NaCl/1 mM EDTA/40 mM Pipes, pH 6.4, were denatured 10 min at 85°C and then hybridized 3 hr at 48°C. One hundred eighty microliters of 0.2 M NaCl, 30 mM NaOAc (pH 5.5), 1 mM ZnSO₄, 20 mg of denatured salmon sperm DNA per ml, and 100-300 units of S1 nuclease (Bethesda Research Laboratories) was added and incubated 1 hr at 37°C.

RESULTS AND DISCUSSION

Structure of the P-450c21 Gene. We manually synthesized 23- and 30-base oligonucleotides (4) corresponding to different regions of a 486-base bovine P-450c21 cDNA fragment (7) and used these to screen a bovine genomic DNA library. Clone λE11 was shown to contain P-450c21 sequences by initiating dideoxy sequencing directly on full-length double-stranded phage without subcloning into M13 (4). The bovine genomic DNA in λE11 was mapped and subcloned into pUC13 as a series of overlapping clones. These pUC13 clones were mapped further and appropriate fragments were subcloned into M13 for sequencing (Fig. 1). As we had no cDNA probe, coding regions and intron/exon boundaries were identified by maintenance of an open reading frame and by comparison to canonical splice-junction site sequences (8), since no exceptions to the canonical splicing rules have been found in any P-450 gene (9-15). Comparison of our sequence with the complete amino acid sequence of porcine

Abbreviations: kb, kilobase(s); bp, base pair(s).

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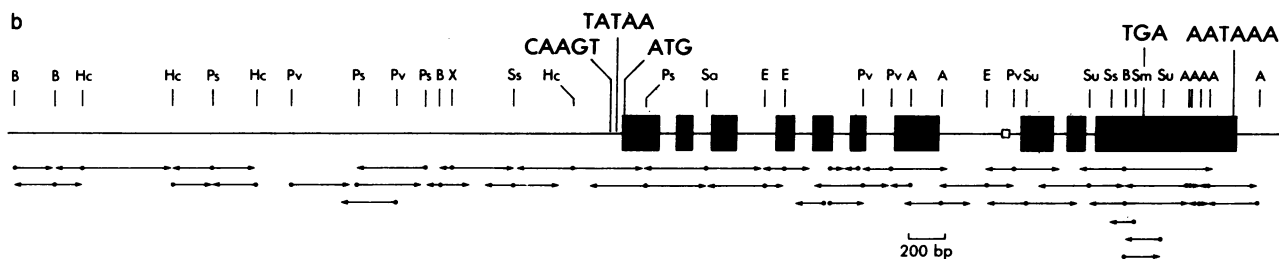


FIG. 1. (a) Sequence of the bovine P-450c21 gene. Base numbering (indicated with plus and minus signs) begins with the guanosine residue (asterisk) representing the principal RNA transcriptional initiation site. Two minor transcriptional initiation sites at -35 and -72 are also indicated by asterisks (see Fig. 2). Where two numbers appear in the left-hand margin, the upper number refers to the amino acid sequence. Spaces between introns and exons and between codons are for clarity only and do not represent absent bases. The TATAA and CAAGTA regions upstream from the principal transcriptional start site and the AATAAAA sequence 22 bp from the poly(A) site are underlined. (b) Restriction map of the bovine P-450c21 gene and sequencing strategy. The solid boxes represent exons, and the small open box represents a region of repeated DNA (see text). Only the restriction sites used in sequencing are shown: A, *Alu* I; B, *Bam*HI; E, *Eco*RI; Hc, *Hinc*II; K, *Kpn* I; Ps, *Pst* I; Pv, *Pvu* II; Sa, *Sal* I; Sm, *Sma* I; Ss, *Sst* I; Su, *Sau*3A; X, *Xba* I. Arrows indicate the direction and extent of dideoxy sequencing. Arrows not originating at restriction sites represent sequencing initiated by the two unique oligonucleotides (4) used to identify λ E11 or by the 27-mer used for primer extension. Each sequencing reaction was performed at least twice, generally on different clones of a given template. Rare areas of ambiguity were resolved by Maxam-Gilbert sequencing. bp, Base pairs.

P-450c21 (John Shively, personal communication) confirms our sequence.

Excluding the flanking DNA, the gene contains 3447 bp divided into 10 exons; other sequenced P-450 genes are all 6-23 kb long and contain 7 or 9 exons (9-15). Unlike other P-450 genes, exon 1 encodes a 5' untranslated segment of mRNA only 11 bases long; 5' untranslated regions of this size are unusual but have been reported as short as 7 bases (17). The nine introns, designated A through I, vary from 48 to 507 bp. Introns A and B contain perfect 12-bp inverted repeats at loci 253-264 and 487-498. Intron I is very short (48 bp), maintains an open reading frame, and lacks a consensus lariat junction sequence (18) and hence resembles exonic DNA. However, it has perfect splice sites, and no P-450 sequenced to date, including porcine P-450c21 (John Shively, personal communication), has amino acids corresponding to this region; furthermore, a bovine P-450c21 cDNA fragment lacks this region (Michael Waterman, personal communication): hence, the sequence designated intron I appears to be an intron. Intron G contains a 48-bp region of alternating pyrimidines and purines, including the sequence (CA)₂₀; this region may be able to form Z-DNA (19) and might thus be involved in the regulated expression of P-450c21. Hybridization of labeled total bovine genomic DNA to restriction fragments of the gene show that this intron is the only one containing highly repetitive DNA (ref. 4 and other data not shown). The rat P-450b and P-450e genes induced by phenobarbital have similar (CA)_n sequences in their 5' flanking DNA but not in introns (13, 15), whereas rat methylcholanthrene-inducible P-450c (9) and mouse dioxin-inducible P₁-450 (11) have similar structures in introns.

Analysis of the 5' flanking region showed no glucocorticoid-regulatory elements, indicating it is unlikely that the P-450c21 gene is regulated by feedback of the glucocorticoid hormones to whose synthesis it contributes. Similarly, by comparison to the sequence of the murine cDNA (20), no evidence was found for the 3' end of an adjacent C'4 gene encoding the fourth component of complement, even though the two human and mouse P-450c21 genes lie <4 kb 3' to the C'4 genes (21, 22). Bases -444 to -415 are closely homologous (18 of 29 bases) to the putative cAMP regulatory site of several eukaryotic genes (23), but the significance of this is not clear, as cAMP regulation of P-450c21 is probable (24) but not established.

The gene sequence predicts synthesis of a 496 amino acid protein containing a leucine-rich hydrophobic amino-terminal region reminiscent of leader peptides of secreted proteins. As P-450c21 is a membrane-bound enzyme (25), this region is

probably involved in affixing P-450c21 to the smooth endoplasmic reticulum. Amino acids 1-16 and 30-47 are hydrophobic regions lacking charged amino acids, each

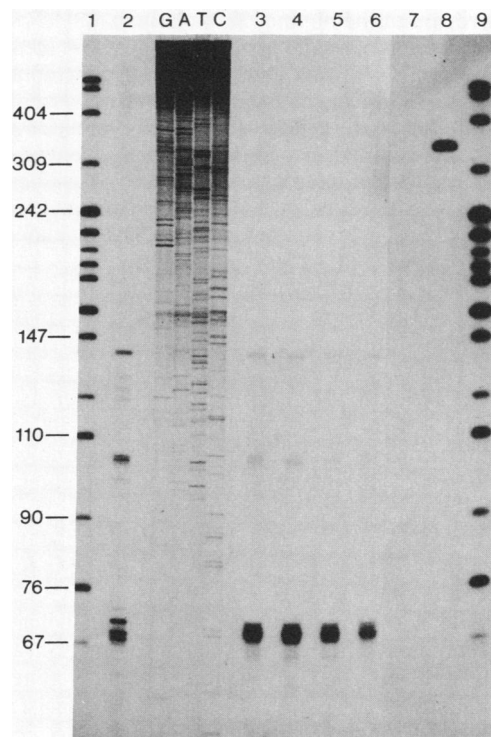


FIG. 2. Location of the transcriptional start site(s) by primer extension and S1 nuclease protection. A 5% acrylamide/7 M urea gel contains the following. Lanes 1 and 9, ³²P-labeled *Hpa* II-cut pBR322 size markers (indicated in bp). Lane 2, primer-extended cDNA initiated from the 27-mer on a bovine adrenal RNA template. Lanes G, A, T, and C, dideoxy sequencing ladder of the 5' end of the gene initiated by the 27-mer on an M13 clone of the 412-base *Hinc*II/*Pst* I fragment. Lanes 3-8, S1 nuclease protection of the ³²P-labeled transcript of this same M13 clone hybridized to bovine RNA. Lanes 3-5, 50 μ g of total bovine adrenal RNA and 500, 750, and 1500 units of S1 nuclease per ml, respectively. Lanes 6 and 7, 50 μ g of total bovine testis and brain RNA, respectively, with 750 units of S1 nuclease. Lane 8, the 359-base probe [corresponding to bases 71 (3' end of 27-mer) to -275, plus 13 bp of the M13 polylinker site] mixed with 50 μ g of tRNA and no S1 nuclease. All samples were electrophoresed on a single gel; the photograph of lanes 7-9 is from a longer autoradiographic exposure of the gel.

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bP450c21
rP450b
rP450c
bP450scc      mlsrglplrsalvkacppilstvgegwhrvrtggegagistktprryseipspgdngwnlyhfwreksqrihfrhienfqkygpiy
mpsvyq

c21      MVLAGLLLLLTLTLAGAHLWG----RWKLRNLHLPLVPGFLH-----LQPNLPHLLSLT----QKLGPVYRLRLGLQ*EAVVNLNKR
b      Mepsillllallvgfllllvrvy---hpksRgmf-PPgprlpllgm----LLQldrggllnSfmg*1reKyGdVftvhlGpr-pVWnlCgtd
c      fpaftAtellLlavttfcigfwwrvtrtwvpgkLksPPgpgWGlPfighvtlglknPhLSlTKLS-----QqyGdVlqiRiGet-pVWVLSqln
scc      rekLgmlesvyiIhpedaVHLf---kfgesyperydIPPWlayhry-----yqkPigvlfkKsgTvw-----kk-drVVLNtev

c21      TIEEAMIRKWDVAFGRQPQPS*---YKLSVQRQDISLGDYSLLWKAHRKLTFRSALLLGTFRS-----SMEPWVDQLTQEFCE*RMVQA-GA
b      TtkEAlvggaeDFsGRgtLav---iepi fkey* -gvi fangerWkAlrrfslatmrdfgmg-----krSvEerVqeeaqclve-eLRksq*GA
c      TtkqAlvkqgdDFkGRPdlyS---ftLiangqsmtdfnDsgplWArRrrLagnAlksfsiasdptlasscylEehVskeaeYlis-kfkqkln-ae
scc      mapEAlknfipllnpsvQdfvslLhKrikQqsgskfvGDikedl fhfafesitnvmfGeRl-----gmLEtVnpeaQkF

c21      PVTIQKEFSL--LTCSLICYLTFGNK*EDTLVIAHHCVDLIMKRW-DHMSIQILDMVPFLR*FFP-NPGLWRLKQAENRDMVEKQLTRHK*E
b      PldptfLFgc--iTanIICtIVGer-fdytdrgfLrllelfyrtfslsSfseq*vfeFfsgFlkyfPGahRqislnlqeilidyighivekhra
c      vghfdpFkyLvsvanvICAicFgrr-yDhdqge llsiVmlsnefgevgtGsyPaDfiPiLR-yLP-NssLdaFkdlnkkfysfmkKlikeHy-r
scc      --iDavykmFHtsVpLLnvppeLyrLfrktwrhdvaawd-----

c21      SMAV-GQWRMDID-YMLQGVGRQVEEGPQLLEGHVMSVVDLFIQ---GTETASTLSWAVAFLLHHPD---*IQRRLEEL-DRELGPASCS
b      tldp-napRDfiDtYLLmek* -eksnhhte fihenlmiSllsLfa---GTETsstTLrygflmlLkyPhva---ekvQkEi-DqviG---Shr
c      tfeK*GhIRDiTdsLiehqdRrldEnamQLsddkVitiVFDLfga*gfdtITTAIsWslmylvtmnr-----IQRkiQEEL*dsvtlG---rdr
scc      ifnk-aekyteifvqdlrrktefRmypGilyclLksekMlleDvkan---iTEmlAgvnttsmtLqwhlyemarslnvQmLreevLnarrqae

c21      RVTYKDRARPLLNATIAEVLRLRPVPLALPHR-TTRPSS*IFGYDIPE-GMVIIPNLQGAHLDE-TVWEGPHEFRP*DRFLFEPGAN-----P
b      lptLDRskmPytdAvhEiqRfsdIVPigVPHR-vtkdtn- frGylLPk*nteVtYPrssAllyp-cyfdhPdsFnP-ehFLdangaLkk---se
c      qprlsDRpqlPyLeAfIletFRhssFVPtiPhs*TiRdts-LnGfyIPK-GhcVfvNqvmhDq*elWgdPnEFRP-eRFLtssgtldkhlse
scc      qdisKmlqmvPLLAskEtLRhLhSivtqlr---yPsdvlvlgdYlTPa-ktlVqvaiyamgrDp-affssPdkFqP-trWlskdkdl ihf---

c21      SALAFGC-GARVCLGESLARLELFVLLRLQLAFTLPPPPVG---ALPSLQDPYCYGNLKVQFP-QVRLQPRGVEAGAWESASAQ
b      afmpFst*GkRiCLGEGjARnELfffttiLQnFsvsshlap---kdldLpke-sGikkippty-QicfsaR
c      kvilPGL-GkRiCIGetigREvFLfLailLQqmfsvsPge---kvdmtpayglLtkharcehf-QVgmrsGqghlqa
scc      mlqFGw-GvRqCvGrriaEelmtlFLihlLenFkvenqhiGdvdti fnLiltPdkpiflvvrPFPQppQa
    
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FIG. 3. Relationship of P-450c21 to other P-450 sequences. Amino acid sequences of bovine P-450c21, rat P-450b (19), rat P-450c (9), and bovine P-450scc (39) are aligned to show regions of homology. The alignments and insertions of gaps were done manually as described (40). Asterisks in the sequences indicate the locations of intervening sequences; dashes indicate gaps.

sufficiently long to anchor the protein to the membrane (26). Amino acids 35–52 are 50% identical to the membrane-spanning region of the bovine (27) or human (28) low density lipoprotein receptor; amino acids 35–91 are 32% identical to amino acids 1324–1381 of human complement component C3a (29); the significance of these similarities is unknown. The overall sequence is 85% identical to that for porcine P-450c21, determined by amino acid sequencing (John Shively, personal communication). Of the 76 residues differing between the two species, 13 are at the carboxyl terminus, which is poorly conserved among other closely related forms of cytochrome P-450 (9–15).

Expression of P-450c21. Cattle (4), mice (22, 30), and human beings (21, 31) have two P-450c21 genes. The human and murine genes lie in the major histocompatibility loci in tight linkage to genes for the fourth component of complement. In man the array is C4A–21A–C4B–21B (21), whereas in mice the array is C4(Slp)–21A–C4–21B (30, 32), with all four genes having a left-to-right transcriptional orientation in both species. Deletion of the human P-450c21 B gene eliminates all 21-hydroxylase activity, suggesting the A gene is normally nonfunctional (31); by contrast, gene-specific oligonucleotide hybridizations indicate only the mouse A gene is functional (32). As different members of the pair of P-450c21 genes appear to be inactivated in mouse and man, such inactivation must have occurred after mammalian speciation about 85 million years ago. Therefore, both P-450c21 genes might remain functional in cattle; this might be consistent with finding two distinct sizes of P-450c21 mRNA in bovine adrenal cortex (4) but only a single size of P-450c21 mRNA in mouse (22) and human (unpublished) adrenal cortex.

To determine if our bovine P-450c21 gene was transcrip-

tionally active and to locate the site(s) of transcriptional initiation (cap site), we performed primer extension and nuclease S1 protection experiments. The two experiments are entirely consistent in identifying cap sites (Fig. 2). Most P-450c21 mRNA molecules begin at one of three guanosine residues 10–12 bases 5' to the ATG initiation codon; some of the mRNA begins at 46 or 83 bases 5' to the ATG, and rare mRNA molecules may be initiated at other loci (Fig. 2 and other gels at higher resolution). The sequence TATAA is found 30 bases from the principal cap site, typical of most eukaryotic genes. The sequence GCTCAAGTA is found 65 bp from the cap site and closely resembles the consensus sequence 5' GG_TCAATCT 3' found 65–90 bp 5' to the cap site of many eukaryotic genes (33). The sequence GG-GCAAGGA found 108 bp from the cap site resembles the consensus "CAAT box" more distantly. These "CAAT" sequences may function as weak promoters initiating transcription of the minor P-450c21 mRNA species originating at bases –35 and –72. Bovine adrenal RNA did not protect the *Sst* I/*Hinc*II fragment from bases –275 to –623 from S1 nuclease digestion, demonstrating that the CAAT and TATAA regions of –562 and –532 do not function as a promoter.

21-Hydroxylase activity is found in a wide variety of extraadrenal tissues (34), but the adult testis is not known to synthesize 21-hydroxylated steroids (35). The nature of extraadrenal 21-hydroxylase is controversial; its activity appears to remain when adrenal 21-hydroxylase is impaired by congenital adrenal hyperplasia (36) and, unlike adrenal 21-hydroxylase, it appears to be regulated by estrogen (34). Furthermore, rabbit hepatic P-450 isozyme 1 readily 21-hydroxylates progesterone (37) but is structurally no more

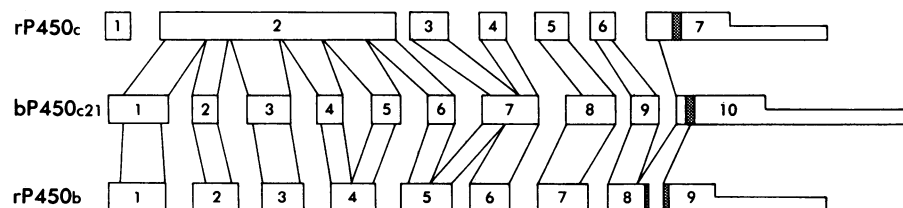


FIG. 4. Exon structures of selected P-450 genes. Exons of rat phenobarbital-inducible P-450b (15), rat 3-methylcholanthrene-inducible P-450c (9), and bovine P-450c21 are drawn to scale, but the spaces between exons do not correlate with the lengths of the corresponding introns. Lines connecting exons of different P-450 genes show homologous regions of exons. The stippled area represents the heme-binding site. The narrow regions of the 3' terminal exons indicate the portion of that exon encoding the 3' untranslated regions of the corresponding mRNAs.

related to P-450c21 than are other hepatic P-450s (38). To determine if the P-450c21 gene is expressed in nonadrenal tissues, we performed S1 nuclease protection experiments with several bovine RNAs. The result with testicular RNA (Fig. 2, lane 6) is indistinguishable from that obtained with bovine adrenal RNA, indicating that this P-450c21 gene is expressed in adrenal cortex and testis. By contrast, bovine brain RNA (Fig. 2, lane 7) and liver, kidney, and ovary RNA (not shown) did not protect the labeled P-450c21 gene fragment from S1 nuclease digestion.

Evolution of the P-450c21 Gene. The P-450c21 gene has 10 exons spanning 3.5 kb, distinguishing it from rabbit and rat P-450 genes induced by phenobarbital, which have 9 exons spanning 14–23 kb (13–15), and the rat and mouse P-450 genes induced by 3-methylcholanthrene or dioxin, which have 7 exons spanning 6–7 kb (9–12). To examine potential relationships among various P-450 sequences, we aligned (Fig. 3) the amino acid sequences of bovine P-450c21 with rat hepatic P-450b (induced by phenobarbital) (15), rat hepatic P-450c (induced by 3-methylcholanthrene) (9), and bovine adrenal P-450scc (39), a mitochondrial P-450 also engaged in steroid hormone synthesis (16). These amino acid alignments in turn indicate homologies among the various exons of the three genes (Fig. 4). Inspection of these exon domains suggests that a precursor to these genes was divided into at least 12 exons. Based upon similar analysis of dioxin-inducible mouse P-450 genes and the rat P-450b and P-450e genes, Gonzales *et al.* (11) concluded that an ancestral P-450 gene had a minimum of 14 exons.

Of the 496 amino acids in P-450c21, 129 are identical in P-450b and 139 are identical in P-450c, further emphasizing that the families of P-450 genes induced by 3-methylcholanthrene and phenobarbital are about equally distant in evolution from the P-450c21 gene. The sequence of P-450scc was much less related, having only 88 identical amino acids, including none in its first 115 residues. As P-450scc is a mitochondrial enzyme, this great difference may reflect on ancient evolutionary divergence between nuclear genes encoding mitochondrial and cytoplasmic P-450s as well as the presence of a 39 amino acid mitochondrial leader sequence in P-450scc. Thus, gene structure and amino acid sequence indicate that P-450c21 represents a separate family of P-450 genes, only distantly related to other members of the P-450 gene superfamily.

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