Pharmacokinetics and pharmacodynamics following maintenance doses of prasugrel and clopidogrel in Chinese carriers of CYP2C19 variants

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WHAT IS ALREADY KNOWN ABOUT THIS SUBJECT

Observations from clinical studies have demonstrated that:

- CYP2C19 genotype does not have a clinically relevant effect on active metabolite concentrations or platelet inhibition in prasugrel-treated subjects.
- Variability of response to clopidogrel is related to the presence of reduced function CYP2C19 alleles.
- Lower concentrations of clopidogrel active metabolite and reduced platelet inhibition lead to increased adverse cardiovascular events in patients with acute coronary syndromes.

WHAT THIS STUDY ADDS

- Observations from previous studies related to the effect of CYP2C19 genotype on plasma drug exposure and platelet response to prasugrel and clopidogrel are confirmed in this prospectively-stratified, randomized, crossover, genetic study in healthy Chinese subjects.
- In this study, healthy subjects receiving prasugrel had higher exposure to its active metabolite and greater pharmacodynamic responses (inhibition of platelet aggregation) compared with clopidogrel treated subjects, regardless of CYP2C19-predicted phenotype group.
- The exposure to the active metabolite of clopidogrel was lower in CYP2C19 IM compared with RM, and in CYP2C19 PM compared with IM. The inhibition of platelet aggregation was lowest in clopidogrel treated CYP2C19 PM subjects.

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AIMS

This open-label, two-period, randomized, crossover study was designed to determine the effect of CYP2C19 reduced function variants on exposure to active metabolites of, and platelet response to, prasugrel and clopidogrel.

METHODS

Ninety healthy Chinese subjects, stratified by CYP2C19 phenotype, were randomly assigned to treatment with prasugrel 10 mg or clopidogrel 75 mg for 10 days followed by 14 day washout and 10 day treatment with the other drug. Eighty-three subjects completed both treatment periods. Blood samples were collected at specified time points for measurement of each drug's active metabolite (Pras-AM and Clop-AM) concentrations and determination of inhibition of platelet aggregation (IPA) by light transmittance aggregometry. CYP2C19 genotypes were classified into three predicted phenotype groups: rapid metabolizers [RMs (*1/*1)], heterozygous or intermediate metabolizers [IMs (*1/*2, *1/*3)] and poor metabolizers [PMs (*2/*2, *2/*3)].

RESULTS

Pras-AM exposure was similar in IMs and RMs (90% CI 0.85, 1.03) and slightly lower in PMs than IMs (90% CI 0.74, 0.99), whereas Clop-AM exposure was significantly lower in IMs compared with RMs (90% CI 0.62, 0.83), and in PMs compared with IMs (90% CI 0.53, 0.82). IPA was more consistent among RMs, IMs and PMs in prasugrel treated subjects (80.2%, 84.2% and 80.2%, respectively) than in clopidogrel treated subjects (59.7%, 56.2% and 36.8%, respectively; $P < 0.001$).

CONCLUSIONS

Prasugrel demonstrated higher active metabolite exposure and more consistent pharmacodynamic response across all three predicted phenotype groups compared with clopidogrel, confirming observations from previous research that CYP2C19 phenotype plays an important role in variability of response to clopidogrel, but has no impact on response to prasugrel.

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Introduction

Prasugrel, a thienopyridine prodrug, is metabolized *in vivo* to an adenosine diphosphate (ADP)-receptor antagonist that potently inhibits ADP-induced platelet aggregation. Prasugrel, co-administered with aspirin, has been approved in the European Union, United States and several other countries for the prevention of atherothrombotic events in patients with acute coronary syndromes (ACS) who undergo percutaneous coronary intervention (PCI). Clopidogrel is also a thienopyridine prodrug indicated for the reduction of atherosclerotic events in patients with recent stroke, recent myocardial infarction or established peripheral arterial disease. Although prasugrel and clopidogrel are both thienopyridine prodrugs that require *in vivo* conversion to their active metabolites, Pras-AM and Clop-AM, the pathways leading to active metabolite formation significantly differ between the two drugs (Figure 1) [1].

Multiple enzymes appear to be involved in the absorption and metabolism of prasugrel and clopidogrel [2–4]. The conversion of prasugrel and clopidogrel to Pras-AM and Clop-AM depends in part on the activity of cytochrome P450 (CYP) 2C19, which plays a greater role in the metabolism of clopidogrel than in that of prasugrel (Figure 1) [2, 3]. Prasugrel is not detected in plasma since it is rapidly hydrolyzed during absorption, forming the inactive thiolactone metabolite R-95913. R-95913 is metabolized to Pras-AM primarily by CYP3A4 and CYP2B6, and to a lesser extent by CYP2C9 and CYP2C19 [2, 5, 6]. Pras-AM is subsequently metabolized to inactive compounds by S-methylation or conjugation with cysteine [5]. Clopidogrel is converted to its active metabolite in the liver through two sequential CYP-mediated steps, being first converted to 2-oxoclopidogrel (via CYP2C19, CYP1A2 and CYP2B6) and then to Clop-AM (via CYP3A4, CYP2C19, CYP2C9 and CYP2B6) [3]. CYP2C19 contributes significantly to both oxidative steps in clopidogrel metabolism [3].

Pras-AM and Clop-AM produce their antiplatelet effects by irreversibly binding to the $P2Y_{12}$ ADP receptor. Once exposed, platelets are inactivated for the remainder of their life span. *In vitro* studies have shown that, on a molar basis, Pras-AM and Clop-AM are equipotent in terms of inhibition of the platelet receptor $P2Y_{12}$; however, compared with clopidogrel, prasugrel is metabolized to its active metabolite more efficiently, resulting in higher inhibition of platelet aggregation (IPA) and a more rapid onset of action [1, 7, 8].

Clinical studies have shown that the pharmacodynamic (PD) response to clopidogrel varies, with 20% to 30% of patients classified as poor responders, nonresponders or resistant to clopidogrel [9, 10]. Studies in healthy subjects have shown that IPA is decreased in subjects receiving

Figure 1

Prasugrel and clopidogrel metabolic pathways to their active metabolites

clopidogrel who have at least one CYP2C19 reducedfunction allele [11–15]; however, the PD response to prasugrel is not affected by polymorphisms associated with carrying CYP2C19 variants with reduced function [12, 16].

The presence of reduced function CYP2C19 alleles has also been associated with reduced responsiveness to, and thus reduced therapeutic benefit of, clopidogrel in cardiovascular patients [1, 16–19]. The major CYP2C19 alleles include three single-nucleotide polymorphisms (*2, *3, *17) [20]. The presence of CYP2C19 loss of function alleles *2 and *3 may result in lower exposure to Clop-AM and thus decreased platelet inhibition in clopidogrel treated patients [12], whereas carriers of *17 may have increased platelet response with an increased risk of bleeding, but not greater efficacy [21]. However, in a study designed to find variant CYP2C19 alleles associated with ethnic background, only 4% of Chinese subjects, compared with 18% of Swedes and Ethiopians, were CYP2C19*17 carriers [22].

Of interest, recent studies have presented evidence of poor clinical outcomes, including an increased risk of major adverse cardiovascular events (MACE) for clopidogrel treated patients with ACS who are carriers of reduced function CYP2C19 alleles, particularly for those who undergo PCI [16, 19, 23]. In contrast, an analysis of two studies evaluating the effects of CYP2C19 genotype on clinical outcomes found that the efficacy of clopidogrel compared with placebo was not affected by CYP2C19 genotype in some patients with ACS [20]. However, one explanation for this different finding may be that only 18% of the patients underwent PCI and 14.5% underwent PCI with stent placement. Because of the demonstrated negative effect of reduced function CYP2C19 on the generation of Clop-AM and the reduced PD response to clopidogrel, a black box warning was recently added to the clopidogrel label indicating the key role of CYP2C19 in clopidogrel metabolism and efficacy (http://products.sanofi-aventis.us/PLAVIX/PLAVIX. html) [24].

The genetic variants resulting in reduced CYP2C19 function are more frequent in East Asians than in Caucasians [25, 26]. In Chinese, the major CYP2C19 genotypes are *1/*1 and *1/*2, each of which comprise about 43% of the population [25]. In Caucasians these genotypes comprise approximately 40% and 19% of the population, respectively. Since the reduced-function *1/*2 allele is 'enriched' in Asians compared with Caucasians, studies that evaluate the effect of this allele can be sufficiently powered with fewer Asian subjects than Caucasian subjects. The current study was therefore conducted in a healthy Chinese population to evaluate the effects of this variation using a prospectively stratified design. Specifically, the objective of this study was to determine the effect of predicted CYP2C19 phenotype on plasma exposure to the active metabolites and platelet response to Pras-AM and Clop-AM.

METHODS

Study design

This single-centre, open-label, two-period, randomized, crossover study was conducted at the Lilly-NUS Centre for Clinical Pharmacology Pte Ltd, National University of Singapore, between 7 December 2006 and 2 April 2008. The study was conducted in accordance with the Declaration of Helsinki and consistent with applicable guidelines for good clinical practice.The study was approved by the local ethics committee (National Healthcare Group – Domain Specific Review Board, reference number C/06/361) and written informed consent was obtained from each subject.

Subjects

Subjects were healthy men and women of Chinese ancestry (having all four grandparents of Chinese origin) 21 to 60 years of age, and had a body mass index (BMI) of 18.5 to 29.9 kg m^{-2} with no clinically important physical findings or abnormalities in laboratory results, including platelet function tests at screening. Genetic testing for determination of CYP2C19 genotype was also performed, unless such data for a subject were already available. Exclusion criteria included a history of bleeding disorders or reasonable suspicion of vascular malformations at screening.Aspirin, nonsteroidal anti-inflammatory drugs and medications other than study drugs were not permitted within 14 days prior to or at any time during the study unless deemed necessary by the clinical investigator to treat adverse events.

Treatment administered

Study participants were randomly assigned to daily treatment with prasugrel 10 mg or clopidogrel 75 mg for 10 days during period 1, followed by a minimum 14 day washout before beginning period 2 daily treatment with the other drug for 10 days (Figure 2). Study drug was administered in the clinical research unit (CRU) on days 1, 2 and 10 of each period, with other dosing days conducted on an outpatient basis.

Pharmacokinetic measurements

Plasma concentrations of Pras-AM and Clop-AM were measured to enable estimation of PK parameters for each metabolite. Subjects were admitted to the CRU the day before PK sampling days and remained in the CRU until discharge on the mornings of days 2 and 11.Blood samples (3 ml each) were collected via a venous cannula or direct venipuncture into ethylenediamine tetraacetic acid (EDTA) tubes at specified time points on days 1 and 10 in each of the two study periods. The blood samples were reacted with 25μ of 500μ m $3'$ -methoxyphenacyl bromide in acetonitrile within 30 s of collection to derivatize and stabilize the active metabolites [27]. Plasma samples harvested from blood samples were stored at approximately -70°C in labelled polypropylene tubes until shipment on dry ice to the analytical laboratory.

Figure 2

Study design and patient disposition. Ninety subjects received \geq 1 dose of study drug. In period 1, 45 subjects completed 10 days of prasugrel and 41 subjects completed 10 days of clopidogrel. In period 2, 43 subjects completed 10 days of clopidogrel and 40 subjects completed 10 days of prasugrel. Clop, clopidogrel; *n*, number of subjects; Pras, prasugrel

Plasma concentrations were determined using validated liquid chromatography methods with tandem mass spectrometric detection as previously described [27, 28]. The lower limit of quantitation was 0.5 ng ml^{-1} for both Pras-AM and Clop-AM.

Pharmacodynamic measurements

Blood samples for PD measurements were collected at various time points on days 1, 2, 3, 5, 9 and 10 of each period; venous blood samples of approximately 9 ml (two tubes of 4.5 ml each) were obtained via direct venipuncture. Platelet aggregation using light transmittance aggregometry (LTA) was measured at the CRU on a platelet aggregation profiler-4 optical aggregometer, with temperature maintained at 37°C. Platelet aggregation in platelet-rich plasma was measured with 20μ M ADP as agonist. Platelet counts in platelet-rich plasma were adjusted to approximately 250 000 μ l⁻¹ using autologous platelet-poor plasma. In addition, citrated blood samples (2 ml each) were collected to perform an exploratory analysis assessing the correlation of LTA platelet aggregation results with those using the Accumetrics VerifyNow™ (VN) P2Y12 assay system (Accumetrics, San Diego, California, United States) as well as the flow cytometric assessment of vasodilator-stimulated phosphoprotein (VASP) phosphorylation using a commercially available kit (BioCytex, Marseille, France). In addition, subjects were characterized as PD nonresponders if they had \leq 20% IPA to 20 μ m ADP as assessed by LTA [29], $>$ 240 P2Y₁₂ reactivity units (PRU) as assessed by the VN P2Y12 assay [30] or >50% platelet reactivity index (PRI) as assessed by VASP phosphorylation [31].

Genotype and phenotype groups

A 6 ml blood sample was collected at screening to determine CYP2C19 genotype by polymerase chain reaction. During the study, DNA was extracted from peripheral blood anticoagulated with EDTA using standard methods to genotype for CYP2C19. The Drug Metabolizing Enzyme and Transporter gene assay system [32] was used to gen-

Table 1

Summary of demographic and CYP2C19-predicted phenotype results

BMI, body mass index; RM, rapid metabolizers; IM, intermediate metabolizers; *n*, number of subjects; PM, poor metabolizers; SD, standard deviation.

erate genotype data. Using validated computer systems, the genetic variants measured were converted to the common consensus allele nomenclature [33] for CYP2C19. Genotype assignments were also reviewed manually.

Subjects were classified by CYP2C19 genotype into metabolic phenotypes based on functional predictions established by the literature [33, 34]. The three classifications consisted of rapid metabolizers [RMs; wild-type homozygotes having two alleles with normal enzyme activity (*1/*1)], heterozygous or intermediate metabolizers [IMs; heterozygotes with one allele having normal enzyme activity and one allele having little or no activity (1*/2* and 1*/3*)] and poor metabolizers [PMs; two alleles with little or no activity $(2*/2*)$ and $2*/3*)$] (Table 1) [35, 36].

Safety analyses

Safety data were collected in the form of vital signs, clinical examinations, laboratory tests and adverse events. A follow-up visit for each subject was conducted within 14 days of the subject's last dose of study drug.

Pharmacokinetic analysis

Pharmacokinetic parameter estimates of Pras-AM and Clop-AM were calculated by standard noncompartmental methods of analysis using WinNonlin® Enterprise Version 5.0.1 and were summarized by treatment and CYP2C19-predicted phenotype. Following the dose of prasugrel or clopidogrel on days 1 and 10, the estimated PK parameters included the area under the plasma concentration–time curve (AUC) from the time of dosing through the sampling time of the last measurable concentration [AUC(0,*t*last)], the maximum observed plasma concentration (C_{max}), and the time of observed *C*max (*t*max). Log transformed *C*max and AUC estimates were evaluated to estimate ratios of least squares (LS) geometric means and the corresponding 90% confidence intervals (CIs). An analysis of covariance model was used to detect differences in PK parameter estimates between groups.

Statistical analyses accounted for differences in body weight among the three predicted phenotype groups.The inclusion of body weight in the statistical model as an intrinsic factor was based on an integrated analysis of 16 clinical pharmacology studies of prasugrel in which the effect of body weight on AUC(0,t_{last}) and C_{max} was statistically significant; higher Pras-AM exposure was associated with lower body weight [37].

Pharmacodynamic analysis

The PD effects of prasugrel and clopidogrel were compared with each CYP2C19-predicted phenotype group. The PD parameter was IPA measured in response to 20 µm ADP using LTA. Estimates of IPA were calculated for all subjects at each time point using the following formula:

$$
IPA_t = \frac{(MPA_0 - MPA_t)}{MPA_0} \times 100\%
$$

where $MPA₀$ is the maximum platelet aggregation (MPA) at baseline (pre-dose), MPA*^t* is the MPA at time *t* and IPA*^t* is the IPA at time *t*. If baseline MPA in the two periods did not differ at a 0.05 significance level using a linear mixed model, then the average of these values was used; otherwise, the corresponding pre-dose MPA for each period was used in the calculation of IPA. A linear mixed effects model was used to test for prasugrel or clopidogrel effect on IPA between predicted phenotype groups with baseline (pre-dose) IPA as a continuous covariate. The fixed categorical effects included phenotype, time and phenotype by time. Subject and subject by time were fitted as random effects. For each phenotype, the LS geometric mean and two-sided 90% CI were estimated at each time point. The mean differences between phenotypes and the corresponding two-sided 90% CIs were also estimated at each time point.

Results

Subjects

Ninety-five healthy Chinese subjects were enrolled, five of whom were not dosed due to physician or subject decision (Figure 2). Ninety subjects received at least one dose of study drug. Seven were withdrawn before completing the study (six during period 1 and one during period 2) as a result of adverse events considered to be unrelated to study drug. Table 1 summarizes the demographic characteristics and CYP2C19-predicted phenotypes of the study population. Subjects were tested for the *17 allele; however, none were found to be carriers.

Pharmacokinetic results

Figure 3 presents mean plasma concentration–time profiles on day 10 in subjects categorized by CYP2C19 predicted phenotype. Mean plasma concentration–time curves for Pras-AM and Clop-AM were similar for days 1 and 10. Following prasugrel dosing, IMs and RMs had similar Pras-AM concentration–time profiles, but PMs had lower Pras-AM concentrations than did IMs or RMs. Following clopidogrel dosing, both PMs and IMs had lower concentrations of active metabolite than did RMs and PMs had lower concentrations than did IMs. Of note, Pras-AM concentrations in PMs were higher than those of Clop-AM in RMs. A comparison of PK parameter estimates of Pras-AM and Clop-AM by predicted CYP2C19 phenotype on day 10 is summarized in Table 2. The data in Table 2 are adjusted for body weight. The molecular weights of Pras-AM $(349.4 \text{ g mol}^{-1})$ and Clop-AM $(355.9 \text{ g mol}^{-1})$ differ by less

Figure 3

Mean plasma concentration of Pras-AM and Clop-AM (day 10) by CYP2C19-predicted phenotype. Clop-AM, clopidogrel active metabolite, CYP, cytochrome P450; RM, rapid metabolizer; IM, intermediate metabolizer; PM, poor metabolizer; Pras-AM, prasugrel active metabolit. Prasugrel RM (\odot); Clopidogrel RM (\odot); Prasugrel IM (∇); Clopidogrel IM (∇); Prasugrel PM (□); Clopidogrel PM (□)

Table 2

Comparison of PK parameter estimates of prasugrel and clopidogrel active metabolites by CYP2C19-predicted phenotype following daily doses of prasugrel 10 mg or clopidogrel 75 mg for 10 days Comparison of PK parameter estimates of prasugrel and clopidogrel active metabolites by CYP2C19-predicted phenotype following daily doses of prasugrel 10 mg or clopidogrel 75 mg for 10 days

concentration-time curve from the time of dosing throgh the sampling time of the last measurable concentration; CI, confidence interval; C_{max}, maximum observed plasma concentration; CYP, cytochrome P450; RM, rapid metabo concentration-time curve from the time of dosing through the sampling time of the last measurable concentation; CI, confidence interval; maximum observed plasma concentration; CYP, cytochrome P450; RM, rapid metabolizer (phenotype *1/*1); IM, intermediate metabolizer (phenotype *1/*2 & *1/*3); LS, least squares; PM, poor metabolizer (phenotype *2/*2 and *2/*3).

than 2%, and therefore comparison of the relative exposures shown in Table 2 is valid.

Data are also presented from a sensitivity analysis, excluding subject 105. This particular subject from the PM phenotype group had atypically low active metabolite exposure for both prasugrel and clopidogrel on both days 1 and 10, despite having a relatively low body weight (59.9 kg) and BMI (19.0 kg m⁻²). The retrospective analysis was conducted to account for any disproportionate influence this anomalous subject may have had on comparisons in the multivariate analysis. For Pras-AM, the LS geometric means ratios (90% CI) of AUC(0,*t*last) with and without subject 105 data were 0.85 (0.74, 0.99) and 0.92 (0.80, 1.06), respectively, for the comparison of PMs to IMs, and 0.80 (0.69, 0.93) and 0.87 (0.75, 1.00), respectively, for the comparison of PMs with RMs.Therefore, after excluding subject 105 data, the exposure to Pras-AM was not significantly different between PMs and RMs or IMs. For Clop-AM, excluding subject 105 data slightly changed the LS geometric means ratios and 90% CIs between phenotype groups, but did not change conclusions. Data presented in Table 2 also illustrate that, although Pras-AM and Clop-AM AUCs were not compared statistically, exposures to Pras-AM were numerically several fold higher than those of Clop-AM for all predicted phenotype groups.

Pharmacodynamic results

A summary of IPA to 20 µM ADP (LTA) for day 10 is presented in Table 3 and Figure 4 shows mean PD results for all sampling times for days 1 through 10. Mean IPA was higher (*P* < 0.05) for prasugrel than for clopidogrel for all three phenotype groups across all time points. Although statistical analyses were not performed, platelet aggregation results obtained for PRU using the VN P2Y12 device, and for PRI as assessed by VASP phosphorylation, showed similar trends to those calculated for IPA to 20 µM ADP using LTA following 10 days of prasugrel 10 mg and clopidogrel 75 mg.

For prasugrel treated subjects,CYP2C19-predicted phenotype group was not associated with platelet response. For clopidogrel treated subjects, CYP2C19-predicted phenotype group was consistently associated with a dampened PD effect for IMs and for PMs across all platelet aggregation measures. Figure 5 shows individual PK and

PD responses to prasugrel and clopidogrel post dose on day 10 stratified by CYP2C19-predicted phenotype group and indicates the threshold for nonresponse for each PD assay. For clopidogrel, the number of nonresponders was similar for IPA and PRU but comparatively larger for PRI.

Among prasugrel-treated subjects, only subject 169 (*1/*1 genotype) met the criteria for nonresponse based on PRU. This subject showed a lower IPA at 24 h after the day 10 prasugrel dose compared with day 10 clopidogrel dose. This subject had no measurable Pras-AM concentrations on day 10, which indicated noncompliance on that day, at a minimum. However, in the absence of direct proof of sustained noncompliance, the subject was included in the PD analyses with the exception of PRI, as data assessed by VASP phosphorylation were not available for this subject.

Adverse events

Daily 10 mg doses of prasugrel were safe and well tolerated by healthy Chinese subjects when administered daily for 10 days. Most adverse events were mild in severity and no serious or drug-related severe adverse events were reported. The most frequently reported adverse events were mild bleeding-related events, specifically post procedural bleeding and contusion (i.e. bruising), with a higher incidence following prasugrel (72%) compared with clopidogrel (58%). For both treatments, the majority of bleeding-related adverse events were mild in severity.

Following administration of prasugrel, there appeared to be a higher incidence of drug-related contusion in subjects with lower body weights; contusion occurred in 17 of 34 subjects (50%) weighing <60 kg compared with 11 of 51 subjects (22%) with a body weight >60 kg. There was no apparent correlation between the incidence of bleedingrelated events and phenotype for prasugrel treatment or for clopidogrel treatment. There was, however, a higher incidence of contusion in RMs compared with IMs and PMs following clopidogrel dosing; contusion was reported by 24% of RMs, 12% of IMs and 9% of PMs. There were no safety concerns in terms of clinical laboratory evaluations, vital signs, fundoscopy, and petechiae examinations following daily 10 mg doses of prasugrel or 75 mg doses of clopidogrel.

Table 3

Summary of day 10 IPA to 20 µM ADP (LTA) for 10 mg prasugrel and 75 mg clopidogrel by CYP2C19-predicted phenotype

ADP, adenosine diphosphate; CI, confidence interval; CYP, cytochrome P450; RM, rapid metabolizer (phenotype *1/*1); IM, intermediate metabolizer (phenotype *1/*2 and *1/*3); IPA, inhibition of platelet aggregation; LS, least squares; LTA, light transmittance aggregometry; PM, poor metabolizer (phenotype *2/*2 and *2/*3).

Discussion

Recent studies have indicated that the interpatient variability of response to clopidogrel is related to the presence

Figure 4

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Mean platelet aggregation as assessed by LTA IPA to 20 μ M ADP (A), VN P2Y12 assay (B) and VASP phosphorylation (C) following daily doses of 10 mg prasugrel and 75 mg clopidogrel according to CYP2C19-predicted phenotype. Differences in IPA (A) between prasugrel and clopidogrel were statistically significant (*P* < 0.05) at all time points. ADP, adenosine diphosphate; CYP, cytochrome P450; RM, rapid metabolizer; IM, intermediate metabolizer; IPA, inhibition of platelet aggregation; LTA, light transmittance aggregometry; PM, poor metabolizer; PRI, platelet reactivity $index; PRU, P2Y_{12}$ reactivity units; VASP, vasodilator-stimulated phosphoprotein; VN, Accumetrics VerifyNow™. Prasugrel RM (○); Clopidogrel RM (○); Prasugrel IM (∇); Clopidogrel IM (∇); Prasugrel PM (\square); Clopidogrel PM (\square)

of reduced function CYP2C19 alleles in poor responders and nonresponders, with a corresponding increased risk of adverse cardiovascular events and stent thrombosis [16, 19, 23, 38]. Further, the relative risk of having the CYP2C19*2 loss-of-function gene for patients who are heterozygous (IMs) *vs.* homozygous (PMs) has been a matter of some controversy. However, a meta-analysis of nine studies demonstrated that, compared with RMs, both groups are at increased risk of MACE when on clopidogrel treatment; the relative risk of MACE was 1.50 for IMs and 1.61 for PMs with a 2.51- and 4.78-fold increased risk of stent thrombosis, respectively [39]. These results suggest that clopidogrel-treated IMs and PMs are at greater risk for experiencing adverse cardiovascular events relative to clopidogrel-treated RMs.

It has also been proposed that patients who are nonresponsive to the standard maintenance dose of clopidogrel may have higher IPA if treated with prasugrel as maintenance therapy [17].Two trials have investigated the effects of switching from clopidogrel to prasugrel. In one, healthy subjects received clopidogrel 75 mg maintenance dose for 10 days and switched directly to either a prasugrel 60 mg loading dose or 10 mg maintenance dose for 10 days [40]. In another study, patients with ACS who had received a clopidogrel 900 mg loading dose were randomized to receive a prasugrel 10 mg or clopidogrel 150 mg maintenance dose for 14 days and then switched to the alternative treatment for 14 days [41]. Results of both studies demonstrated that switching from clopidogrel to prasugrel was well tolerated and resulted in improved platelet response [40, 41]. We undertook a prospectively designed, randomized, crossover study in which healthy Chinese subjects were stratified based on CYP2C19 genotype to specifically evaluate the effect of CYP2C19 genetic variation on the PK and PD of prasugrel and clopidogrel.

In the current study, PMs had lower exposure to both Pras-AM and Clop-AM when compared with their respective RMs. After adjusting for body weight as an intrinsic factor that is known to affect Pras-AM exposure, the AUC(0,*t*last) for Pras-AM did not differ between IMs and RMs, but was 20% (90% CI 7%, 31%) lower in PMs than in RMs. In the sensitivity analysis that excluded data from one

Figure 5

Pharmacokinetic and pharmacodynamic responses to clopidogrel and prasugrel in individual subjects on day 10 following daily doses of 10 mg prasugrel and 75 mg clopidogrel stratified by predicted CYP2C19 phenotype. Panel A shows active metabolite exposure, which is consistently higher for prasugrel than for clopidogrel across all 3 groups. Inhibition of platelet aggregation as measured by LTA (B), VN P2Y12 assay (C), and VASP phosphorylation (D) is also shown. The asterisked RM subject in panels B and C is subject 169, who was very likely noncompliant during prasugrel treatment. See text (Pharmacodynamic results) for details. AUC, area under the plasma concentration–time curve; ADP, adenosine diphosphate; Clop-AM, clopidogrel's active metabolite; CYP, cytochrome P450; RM, rapid metabolizer; IM, intermediate metabolizer; IPA, inhibition of platelet aggregation; LTA, light transmittance aggregometry; PM, poor metabolizer; Pras-AM, prasugrel's active metabolite; PRI, platelet reactivity index; PRU, P2Y₁₂ reactivity units; VASP, vasodilator-stimulated phosphoprotein; VN, Accumetrics VerifyNow™

Figure 5 Continued

subject with atypically low exposure for both prasugrel and clopidogrel, the magnitude of the Pras-AM exposure difference between PMs and RMs was reduced to 13% (90% CI 0%, 25%). As with previous comparisons of exposure between Pras-AM and Clop-AM following 10 mg and 75 mg doses, respectively, AUC and *C*max estimates for Pras-AM were substantially higher overall than those for Clop-AM across all predicted phenotype groups (Table 2).

The PD response to prasugrel treatment was unaffected by CYP2C19 genetic variation based on IPA to 20 μ M ADP as measured by LTA, the VN P2Y12 assay and VASP phosphorylation. Consistent with the higher exposure of Pras-AM, IPA was substantially higher during prasugrel treatment than during clopidogrel treatment for all three CYP2C19-predicted phenotype groups. Compared with prasugrel treated RMs, clopidogrel treated RMs had a 72 percentage point lower active metabolite exposure, translating to a 25 percentage point lower IPA. Previous studies showed that Pras-AM and Clop-AM are equipotent and that the PD response to these compounds correlates with the respective exposure to the active metabolite [1, 7–9, 12]. Since the exposure to Pras-AM is significantly greater than that for Clop-AM, the increase in IPA would be correspondingly greater after a prasugrel dose, and actually approaches the asymptote of the exposure–response curve [1]. Accordingly, while the exposure to Pras-AM was lower in CYP2C19 PMs, the impact on the resultant IPA, if any, was negligible, as small changes in exposure to Pras-AM did not affect the PD response to prasugrel.

One limitation of the current study is that the small number of PMs, which comprised only 11 (13%) of the 83 subjects who completed the study, may limit interpretation of results. For example, the atypically low AUC(0,*t*last) for both Pras-AM and Clop-AM in one PM subject 105, had a disproportionately large influence on comparisons between CYP2C19-predicted phenotype groups, particularly for Pras-AM analyses.The low exposure for this subject was not associated with demographic characteristics, bioanalytical assay variability or improper sample handling. Thus, PK analyses are presented both with and without this subject's data for full disclosure (Table 2). It is noteworthy, however, that the 90% CI for the Clop-AM AUC(0,*t*last) in the PM group did not include 1.0 for either analysis, suggesting a real difference in Clop-AM exposure for PMs relative to RMs and IMs.

The results of this study demonstrate that because of differences in the respective metabolic pathways leading to the formation of Pras-AM and Clop-AM and the resultant differences in exposure between the two drugs, the presence of reduced function CYP2C19 alleles does not affect the PD response to prasugrel, but significantly reduces the anti-platelet response to clopidogrel.This difference would also explain the greater IPA and fewer ischaemic events seen with prasugrel, as compared with clopidogrel, in the TRITON-TIMI 38 study [42]. These results are consistent with the findings of a study in which CYP3A was completely inhibited with ketoconazole in subjects treated with prasugrel or clopidogrel [43]. CYP3A is the predominant pathway for Pras-AM generation, yet potent CYP3A inhibition did not affect the extent of active metabolite generation as indicated by AUC. Given that ketoconazole is one of the most potent inhibitors of CYP3A, inhibitors or reduced function of other, less important CYPs would not be expected to affect the overall exposure or pharmacodynamic response to prasugrel maintenance or loading doses.

The results also demonstrate that, compared with clopidogrel, prasugrel has greater exposure to the active metabolite and more consistent PD responses across all three CYP2C19-predicted phenotypes. This genetics study confirms observations from previous research that CYP2C19 phenotype plays an important role in the variability of response to clopidogrel, but has no impact on response to prasugrel.

Competing Interests

RPK, KJW, LS, FN, JAJ, MH and DSS are all employees and shareholders of Eli Lilly and Company. SLC is a former employee of Eli Lilly and Company and received fees for consulting from Eli Lilly and Daichii Sankyo. NAF is a retired employee and shareholder of Eli Lilly and Company. JRW is an employee of Daiichi Sankyo, Inc. and a shareholder of Daiichi Sankyo Company, Ltd.

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