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Effects of Pulmonary Inhalation on Hyperpolarized Krypton-83 Magnetic Resonance T₁ Relaxation

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Abstract

The ⁸³Kr magnetic resonance (MR) relaxation time T₁ of krypton gas in contact with model surfaces was previously found to be highly sensitive to surface composition, surface to volume ratio, and surface temperature. The current work explored aspects of pulmonary ⁸³Kr T₁ relaxation measurements in excised lungs from healthy rats using hyperpolarized (hp) ⁸³Kr with approximately 4.4 % spin polarization. MR spectroscopy without spatial resolution was applied to the *ex vivo* lungs that actively inhale hp ⁸³Kr through a custom designed ventilation system. Various inhalation schemes were devised to explore the influence of anatomical dead space upon the measured ⁸³Kr T₁ relaxation times. The longitudinal ⁸³Kr relaxation times in the distal airways and the respiratory zones were independent of the lung inhalation volume, with T₁ = 1.3 s and T₁ = 1.0 s, depending only on the applied inhalation scheme. The obtained data was highly reproducible between different specimens. Further, the ⁸³Kr T₁ relaxation times in excised lungs were unaffected by the presence of up to 40% oxygen in the hp gas mixture. The results support the possible importance of ⁸³Kr as a biomarker for evaluating lung function.

1. Introduction

Since the first reports of pulmonary MRI with hyperpolarized (hp) ¹²⁹Xe (Albert et al., 1994) and hp ³He (Middleton et al., 1995) approximately 1 ¹/₂ decades ago, a variety of pulmonary diagnostic MRI techniques have been developed that are based on these two noble gas isotopes with nuclear spin I = $\frac{1}{2}$ (Moller *et al.*, 2002; Cherubini and Bifone, 2003; Lutey et al., 2008; Santyr et al., 2009). The immediate task for advancing hp gas based MR techniques is to better understand lung function and to develop new biomarkers for various lung pathologies. Demonstrated examples are direct gas density imaging that provides insights into ventilation defects caused by chronic obstructive pulmonary disease (COPD) and smoking (de Lange et al., 1999) or restricted ventilation caused by asthma (Altes et al., 2001; Samee et al., 2003; Tustison et al., 2010). The apparent diffusion contrast (ADC) is related to alveolar size and is a biomarker for COPD with some research suggesting that it is a sensitive approach for detecting the early stages of COPD (Yablonskiy et al., 2002; Peces-Barba et al., 2003; Fain et al., 2005; Altes et al., 2006; Woods et al., 2006; van Beek et al., 2009). The large gyromagnetic ratio of 3 He makes the relaxation of this isotope (and to a lesser extent the relaxation of ¹²⁹Xe) sensitive to the presence of paramagnetic substances (Abragam, 1961). Molecular oxygen, O₂, is the only paramagnetic molecule present in sufficient quantities in the inhaled gas to influence 3 He relaxation. The T₁ relaxation of 3 He can therefore provide information about alveolar oxygen partial pressure (Fischer et al., 2005; Cieslar et al., 2007; Yu et al., 2008). Hp ¹²⁹Xe, with high tissue solubility and 300

ppm chemical shift range (Goodson, 2002), has been explored to study gas exchange in lungs (Ruppert *et al.*, 2000; Driehuys *et al.*, 2006; Ruppert *et al.*, 2007; Driehuys *et al.*, 2009; Cleveland *et al.*, 2010). Hp ¹²⁹Xe was also explored for molecular imaging using functionalized xenon biosensors (Hilty *et al.*, 2006; Schroder *et al.*, 2006) and was infused directly into blood for MR imaging applications (Cleveland *et al.*, 2009).

The recent development of hp 83Kr MR (Pavlovskaya et al., 2005; Cleveland et al., 2006; Cleveland et al., 2007; Cleveland et al., 2008a; Cleveland et al., 2008b) provides potentially a new pulmonary biomarker that utilizes the nuclear electric quadrupole moment of ⁸³Kr as a probe for surfaces. Hyperpolarization of the spin I = 9/2 isotope ⁸³Kr is achieved by spin exchange optical pumping (SEOP) in a stopped flow mode for 10 - 15 min (Walker and Happer, 1997; Cleveland et al., 2006). The gas phase relaxation time, T₁, is on the order of hundreds of seconds at atmospheric pressure (Brinkmann and Kuhn, 1980; Cleveland and Meersmann, 2008a, b) and is therefore sufficiently long to separate the hp ⁸³Kr from the rubidium vapor used in the SEOP process without substantial depolarization (Cleveland et al., 2006). The separation of the highly reactive and therefore toxic alkali metal is crucial for any pulmonary application. The ⁸³Kr gas phase relaxation time, T₁, is shortened in the presence of a surface and is inversely dependent on the surface-to-volume ratio (Stupic et al., 2006). In porous media, the T_1 of ⁸³Kr is affected by surface composition (Stupic *et al.*, 2006; Cleveland et al., 2008b), surface hydration (Cleveland et al., 2007), and surface temperature (Cleveland et al., 2006). The sensitivity of ⁸³Kr relaxation to surfaces that are in contact with the hp gas arises from interactions of the nuclear electric quadrupole moment with the electric field gradient (EFG) from the electronic environment surrounding the krypton nucleus. In the case of noble gas atoms, an EFG is typically caused by distortions of the electron cloud of the noble gas atom during surface adsorption. This effect can provide T₁ contrast in MRI (Pavlovskaya et al., 2005; Cleveland et al., 2008b) that may complement information obtained from hp 3 He or hp 129 Xe MRI data.

Previous reports with hp 83 Kr provided the proof of concept of 83 Kr relaxation as a surface sensitive contrast agent in model systems (Pavlovskaya *et al.*, 2005) and demonstrated the feasibility of hp 83 Kr MRI in rat lungs (Cleveland *et al.*, 2008a). This work explores the effects that influence pulmonary hp 83 Kr T₁ relaxation measurements using excised lungs of healthy rats. A spatially non-resolved MR spectroscopy study of hp 83 Kr relaxation as a function of pulmonary inhalation in excised rat lungs using natural abundance krypton (i.e. 11.5 % 83 Kr) is reported here expanding the potential role of this new technique for evaluating and monitoring lung function in health and disease.

2. Materials and Methods

2.1. Spin exchange optical pumping (SEOP) of ⁸³Kr

Hyperpolarized (hp) 83 Kr was produced with two 35 W diode array lasers (Spectra Physics, Santa Clara, Ca) line-narrowed to 0.3 nm at the Rb D1 transition of 794.7 nm. The laser power available for SEOP was approximately 30 W for each of the lasers after accounting for losses in the optical elements used for the experiments. The lasers were oriented such that simultaneous illumination from the front and back of the cylindrical glass SEOP cell (Pyrex glass, no surface treatment, ID = 28 mm, length = 125 mm) was achieved. Stopped flow SEOP was employed as previously described (Cleveland *et al.*, 2008a) using 10 – 20 min laser irradiation at a pump cell pressure of 135 kPa. Each SEOP cell was loaded with 1 g of Rb (99.75%; Alfa Aesar, Ward Hill, MA) and the krypton mixture was produced from research grade gases (Airgas, Radnor, PA) comprised of 25% krypton (99.995% pure), 5% nitrogen (99.9997% pure), and 70% helium (99.9999% pure). The SEOP system was constructed of glass and PTFE tubing using brass valves and unions as necessary. Due to fast quadrupolar relaxation on siliconized or silanized surfaces (Wu et al., 1990; Stupic et

al., 2006) all glassware that was in contact with hp 83 Kr was used without surface treatment of any kind. All other materials in contact with hp 83 Kr were also used as supplied without modification of the surfaces. During operation the SEOP cell containing the gas mixture was kept at a temperature of 438 ± 5 K using a temperature controlled air flow to provide even heating across the cell. All SEOP operation took place in the fringe field of the superconducting magnet at approximately 50 mT. 40 ml of the Rb free hp gas was transferred through equalization to ambient pressure into the injection syringe with variable storage volume V_A (see figure 1). Note that the ambient pressure is $p \approx 83$ kPa at the laboratory location in 1500 m elevation. An air-cooled condenser placed at the outlet of the SEOP cell was used to separate the Rb vapor from the hp gas. The hp gas was then transferred further (section 3.2), without separation of the krypton from the SEOP mixture, for ventilation into the lungs (section (2.3)).

2.2. Animal care and usage

Following the University of Colorado Health Sciences Center approved protocol, 30 healthy, male Sprague-Dawley rats (Charles River Laboratories, Inc., Wilmington, MA) rats (175 – 400 g) at the time of lung excision were anesthetized with ketamine (80 mg/kg) (University of Colorado Hospital pharmacy) and xylazine (16 mg/kg) (MWT Veterinary Supply, Meridian, ID). 100 USP units heparin (American Pharmaceutical Partners, Inc., Schaumburg, IL) injected into the anesthetized animal via the right ventricle of the heart were allowed to circulate for 10 - 15 s before lungs were placed on a ventilator. The lungs were perfused with 50 ml of Belzer-MPS solution (UW Kidney Preservation Solution, Trans-Med Corporation, Elk. River, MN) and the trachea was clamped at the time of inhalation to avoid collapsing the airways while removing the heart and lungs *en bloc* from the chest cavity. The excised lungs were then cannulated with a plastic adapter tube and positioned 5 mm above the bifurcation of the lungs (Cleveland *et al.*, 2008a).

2.3. Ex vivo lung ventilation

Following excision, the lungs, with the heart still attached, were immediately transferred into a Pyrex ventilation chamber (inner diameter = 34 mm and height = 100 mm) with the main trachea pointing downwards as shown in figure 1(a-c). The lung trachea was connected via the adapter tube to the access opening in the bottom of the ventilation chamber that allowed for ventilation after the lungs were immersed in ~60 ml of Belzer-MPS solution. The lungs were then inflated to 5-6 ml of air and transported to the imaging facility at a temperature of 277 K.

Prior to all experiments, the lungs submerged in the ventilation chamber were inflated to 1 ml above the maximum inhalation volume used in the measurements. This procedure was undertaken to check if any gas bubbling was occurring from the sutures connecting the trachea to the adapter tube, or from the lung itself since any leakage could affect the apparent relaxation behavior. All lungs were kept at 290 K for the experiments, a temperature that corresponded to the bore temperature of the superconducting magnet.

Lung inhalation was accomplished through suction above the Belzer-MPS solution using a syringe (i.e. ventilation syringe) attached to the top of the ventilation chamber (Cleveland *et al.*, 2008a) as shown in figure 1. The applied suction allowed for the lung to inhale to the desired inhalation volume V_i (see figure 1(c)). The ventilation syringe was calibrated prior to the experiments by correlating the applied 'suction volume' V_s with the displacement volume of the Belzer-MPS solution and therefore with the inhaled gas volume V_i in the lungs. The displacement volume was determined through the rise of the liquid level in the ventilation chamber. After calibration, this procedure enables a good control of the inhaled gas volume V_i during the experiments inside the MR magnet when the lungs are shielded

from the view. Note that the ventilation chamber enabled the *ex vivo* lungs to actively inhale the gas, rather than being passively inflated by a pressurized gas.

2.4. Longitudinal relaxation measurements

T₁ measurements were obtained with a 43 mm (inner diameter) saddle coil tuned to 15.4 MHz ⁸³Kr resonance frequency at 9.4 T. The lung ventilation chamber of 40 mm diameter allowed the rat lungs to expand without contacting the ventilation chamber walls. A pulse sequence consisting of 32 small (constant 12°) flip angle RF pulses spaced evenly in time at intervals of $\tau = 0.2$ s was used to collect a spectrum at each time interval during inhalation by the lung and subsequent breath hold. Only the spectra at sufficient time after breath hold were used for analysis. Specifically, data recorded at 0.4 s after the observed maximum integrated intensity was used for T₁ fitting. This 'starting point' for the fitting was defined as experimental number n = 0 at time $t = n \cdot \tau = 0$ s. The natural logarithm of integrated signal, S(n), normalized by its value at n = 0, was fitted as a function of the experimental number n using:

$$\ln \frac{S(n)}{S(n=0)} = \left(\ln(\cos(\theta)) - \frac{\tau}{T_1} \right) \cdot n.$$
[1]

Equation (1) takes the polarization destruction occurring from the $\theta = 12^{\circ}$ flip angle RF pulse into account (Gao *et al.*, 1997; Cleveland *et al.*, 2008b). The linear fitting was applied to data obtained between t = 0 s up to t = 2.6 s because of non-linearity occurring after this time in all data sets.

3. Results and Discussion

3.1. ⁸³Kr hyperpolarization

⁸³Kr SEOP with the two 30 W line-narrowed diode array lasers generated a signal enhancement of 9300 times the thermally polarized ⁸³Kr signal at 9.4 T magnetic field strength and 290 K temperature. The spin polarization P of the thermally polarized spin I = 9/2 system was calculated through equation (2) (Stupic *et al.*, 2011).

$$P = \frac{|\gamma| B_0}{3k_B T} (I+1)$$
^[2]

where γ is the gyromagnetic ratio, k_B is the Boltzmann constant, and $\hbar = h/2\pi$ is the Planck constant. Note that for I = $\frac{1}{2}$, equation (2) leads to $P = |\gamma| \frac{\hbar B_0}{2k_B T}$ that is commonly used for ³He and ¹²⁹Xe. Equation (2) is valid for temperatures $T \gg |\gamma| \frac{\hbar B_0}{k_B}$, a condition that is fulfilled for ⁸³Kr at B₀ = 9.4 T at ambient temperature since $|\gamma| \frac{\hbar B_0}{k_B} \approx 10^{-3} K$, and the thermal polarization is:

$$P_{{}^{83}Kr}^{9.47,290K} = 4.70 \times 10^{-6}$$
^[3]

A SEOP signal enhancement of 9300 corresponds therefore to approximately 4.4 % spin polarization, a 2 fold improvement in spin polarization compared to previous work (Cleveland *et al.*, 2008a). This improvement was likely achieved because of a 1.2 fold increase in laser power, better laser alignment, and predominately laser line narrowing (i.e 0.3 nm vs. 2 nm used previously). The enhancement was calculated by comparison of the hp signal to the signal of thermally polarized krypton at 10.430 MPa with 116 scans and 600 s

delay time between scans resulting to a signal with a signal to noise ratio of 130. The same radio frequency coil, probe head, pulse duration, and power were used for both samples.

The 4.4 % spin polarization was obtained from a mixture of 25% krypton, 5% nitrogen, and 70% helium. Similar to ¹²⁹Xe (Driehuys *et al.*, 1996; Mortuza *et al.*, 2003) and ¹³¹Xe (Stupic *et al.*, 2011) higher polarization is expected from SEOP with more dilute krypton mixtures. However, the increase in signal caused by the increase in polarization is offset by the signal decrease from dilution. In the case of ¹²⁹Xe, this concern is solved by separating the xenon from the other gases through a freeze-thaw cycle using liquid nitrogen. Unfortunately, the same method would severely depolarize the ⁸³Kr hyperpolarized state due to fast ⁸³Kr quadrupolar relaxation in the frozen state with $T_1 < 10$ s at 77 K (Cowgill and Norberg, 1976). A 25% krypton mixture has been shown to be a good compromise (Cleveland *et al.*, 2007) as long as practical hp ⁸³Kr concentration techniques are not available. It should be noted that effectively less than 3% of the ⁸³Kr isotope is present in the 'hp-gas' used in this work.

3.2 Testing of ex vivo hp ⁸³Kr lung ventilation

Ventilation of the lungs with hp ⁸³Kr was achieved through a custom build delivery system that is depicted in figure 1. Following polarization buildup in the SEOP cell for 10 – 20 min at 135 kPa pressure, hp ⁸³Kr was delivered to a pre-evacuated (pressure < 10 Pa) 50 ml borosilicate glass syringe (Chemglass, Vineland, NJ) – i.e. injection syringe with variable storage volume V_A - by pressure equalization as shown in figure 1(a). After delivery of approximately 40 ml of hp ⁸³Kr into V_A , the valve between V_A and the SEOP cell (denoted as valve #1) was closed, while the valve (i.e. valve #3)) between the syringe V_A and the transfer line with storage volume V_B , was opened. This allowed for the controlled injection of hp ⁸³Kr from the syringe V_A into storage container V_B (see figure 1(b)) while keeping the gas pressure approximately constant. Using this delivery method, a sudden pressure rise and therefore premature entry of hp ⁸³Kr into the lung is prevented without the need for a valve in the high magnetic field detection region.

Any excess hp ⁸³Kr delivered beyond the capacity of storage container V_B (expandable from 6 ml to 30 ml) was exhausted to open air. The hp ⁸³Kr was further transferred from V_B into the lung using the ventilation syringe V_s as described in section (2.3). The ventilation system enabled the *ex vivo* lungs to actively inhale the gas. Active inhalation was preferred over passive inflation by a pressurized gas because it is a better way to prevent depolarization of the fast relaxing hp ⁸³Kr. However, as an additional advantage, the active inhalation method more closely resembles the *in vivo* breathing process (i.e. through pleural pressure reduction) when compared to a forced inflation method using a pressurized gas.

Figure 2 shows a series of 32 individual MR spectra (without spatial encoding) that were recorded every 0.2 s using a 12° flip angle radiofrequency pulse before and during lung inhalation with hp ⁸³Kr. During the initial 2 s no signal was visible, demonstrating that no amount of MR detectable hp ⁸³Kr was forced into the lung during gas transfer from the injection syringe V_A into storage container V_B . The hp ⁸³Kr is inhaled into the lung when a suction is applied to the ventilation chamber at t = -1.2 s. At approximately t = -0.6 s, the lungs are reaching the desired inhalation volume V_i . The ⁸³Kr T₁ relaxation reduces the observed signal as time progresses. At t = 0 s, the lungs are presumed to be static and lung movement no longer contributes to the temporal changes in the hp ⁸³Kr signal. The behavior of the signal decay at t > 0s is discussed in the following section.

3.3. Determination of longitudinal ⁸³Kr relaxation in lungs

No attempt was made to spatially resolve the relaxation measurements; however, higher ⁸³Kr MR signal intensities may become available as a result of technological developments and/or a reduction in the prohibitively high costs for isotopically enriched ⁸³Kr. Note that in this publication the wording 'different zones of the lung' refers solely to differences between the more proximal airways (the major conducting airways, i.e. trachea and bronchi), the more distal airways (the conducting airways at higher generation of branching, namely bronchioles), and the respiratory zones (i.e. the alveolar ducts and the alveolar sacs). The differentiation of these zones is intentionally kept coarse in order to allow for an initial interpretation of the results.

The hp ⁸³Kr T₁ times in lungs where obtained from a series of 32 spectra, separated each by 0.2 s, each following a 12° flip angle radiofrequency pulse as described in more detail in sections 2.4 and 3.2. In order to best measure the relaxation behavior, the series of MR scans were timed to commence approximately with the start of the inhalation process thus allowing the first few spectra to capture a part of the actual inhalation. The decay of the signal in the spectra during the "breath hold" that followed the inhalation provided the crucial data for determining the T₁ relaxation time. A typical set of spectra, that is used for the relaxation measurements, is shown in figure 3(a) where the inhalation of 20 ml hp ⁸³Kr by an excised rat lung is monitored. Figure 4(a) shows the inhalation of 6 ml hp ⁸³Kr under otherwise identical conditions.

In both experiments, the lung inhalation with hp 83 Kr during the initial phase leads to a rapid increase in signal intensity observed in the spectral array until a maximum is reached. The occurrence of the signal maximum corresponds approximately to the point where the lungs have reached the selected inhalation volume V_i. Note that the maximum signal intensity does not necessarily coincide exactly with the end of the inhalation process because of insufficient (0.2 s) time resolution and also because of longitudinal relaxation during the inhalation phase. Therefore, data points used for the T₁ fittings start two spectra after the maxima (corresponding to 0.4 s after the maximum signal intensity) when the signal intensities start to follow a mono-exponential decay behavior as can be visualized in the semi-logarithmic graphs in figures 3(b) and 4(b). After this experimental time point, labeled as t = 0 s and marked in figures 3(b) and 4(b) by a diamond marker, the changes in the hp 83 Kr signal intensity are assumed to be caused by longitudinal relaxation (and perhaps by diffusion and convection – see arguments below) but not by changes in the lung volume.

Data used in the T_1 fittings extended from t = 0 s to t = 2.6 s without considering spectra collected after 2.6 s. The 2.6 s cutoff was chosen to eliminate deviations from monoexponential relaxation behavior as is apparent in the semi-logarithmic graphs. This deviation is likely caused by vastly different relaxation behavior between the relatively smaller respiratory zones and the larger airways. Depending on the inhalation level, a very high percentage of the signal intensity will initially arise from the smaller respiratory zones, in particular alveolar sacs and alveolar ducts but also by the nearby respiratory bronchioles with similar diameter as the alveoli and perhaps the distal airways such as the terminal bronchioles. Depending on the total ventilation volume, the respiratory zone accounts typically for more than 95 % of the lung volume. A single rate constant appears to be a good description of the hp⁸³Kr relaxation from this region (see fitted line in 3(b) and 4(b)). However, as time progressed, fast longitudinal relaxation in the high surface-to-volume ratio alveoli reduced the relative contribution of the hp ⁸³Kr signal of this distal region compared to that of the proximal larger airways where the $^{\hat{8}3}$ Kr T₁ relaxation is slower. The relative contribution of the larger airways (with vastly different T₁ values) to the signal intensity will therefore rise over time and the decay will no longer have the appearance of single exponential relaxation. This point appears to be reached at t = 2.6 s, a time that is slightly

more than two times the T_1 time of ⁸³Kr in the alveolar space, when the alveolar signal has decayed to about one tenth or less of the original intensity. Further data analysis after t = 2.6 s is difficult because of low signal to noise ratios. (Depending on the inhalation volume, the initial signal to noise ratio, S/N, is between 189 and 75 at t = 0 s. The S/N drops to values between 12 and 4 at t = 2.6 s). Further, the hp ⁸³Kr in the larger airways will contribute substantially to the signal after this time and is therefore expected to decay at various relaxation time constants depending on the diameter of the airway. The situation may be further complicated by the possibility of migration of hp ⁸³Kr from the adapter tube were the ⁸³Kr relaxation is relatively slow with $T_1 \gg 1$ s. Although the adapter tube is outside the MR detection region, diffusion and convection of hp ⁸³Kr into the airways within the detection region may lead to an apparent slower ⁸³Kr relaxation in the lungs. If present, this effect will be pronounced for longer experimental time scales.

3.5. Influence of different inhalation schemes

Previously, ⁸³Kr T₁ was shown to be inversely dependent on the surface-to-volume ratio, S/V, of porous systems (Stupic *et al.*, 2006). The inverse dependence of ⁸³Kr T₁ times on the surface-to-volume should also affect the ⁸³Kr T₁ relaxation in various regions of the lungs. Three different inhalation schemes were developed as a way to, as well as is possible, discriminate hp ⁸³Kr relaxation behavior in the larger airways from the relaxation in the alveoli and distal airways. The goal was to obtain first insights in ⁸³Kr longitudinal relaxation in these regions.

In scheme 1, only hp $^{83}\rm{Kr}$ gas mixture was used for inhalation. This scheme was applied for the experiments in figures 2, 3 and 4. The outer glass tube of storage container V_B had a volume of 6 ml but an additional fitted glass extension tube was available to expand the volume of stored hp $^{83}\rm{Kr}$ gas to V_B = 30 ml. The extended storage container V_B enabled the lungs to inhale up to V_i = 20 ml of hp $^{83}\rm{Kr}$ depending on the suction applied by the ventilation syringe. Inhalation volumes V_i > 20 ml would have likely exceeded the lung capacity and were not attempted.

Scheme 1 does not discriminate between major airways and alveoli because the inhaled hp 83 Kr is present in the respiratory zone and the anatomical dead space (i.e. the airway encompasses all generations of branching down to the terminal bronchioles) as sketched in figure 5(a). The lung regions filled with hp 83 Kr are depicted in light blue color (or light gray in black and white representation) and all regions contribute to the observed signal in this inhalation scheme. However, the airways contribute only to about 5 % of the total lung volume. As a consequence, relaxation initially appears to be mono-exponential but then deviates from this behavior as time passes as discussed in the previous section.

Scheme 2 uses non-hp gas that is not detectable by the MR scanner at these experimental conditions, in combination with hp ⁸³Kr. The non-hp gas is referred to in this publication as 'thermal' or 'dark' gas. In scheme 2, a determined volume V_i^{dark} of thermal gas was inhaled into the lungs prior to ventilation with hp ⁸³Kr. Thermally polarized ⁸³Kr, which does not produce a detectable MR signal under the conditions used in these experiments, can serve as dark gas. Normal air, which does not contribute at all to signals at the ⁸³Kr frequency, can also be used (see section (3.8) on the effects of oxygen). Specifically, volumes of $V_i^{dark}=6$ ml and $V_i^{dark}=12$ ml of thermal gas were introduced into the lungs prior to the transfer of hp ⁸³Kr into storage container V_B. Subsequently, hp ⁸³Kr was transferred into V_B and a selected volume V_i^{hp} of ⁸³Kr was inhaled. The total inhalation volume was therefore $V_i=V_i^{dark}+V_i^{hp}$.

The rationale for inhalation scheme 2 is to keep hp ⁸³Kr as much as possible in the proximal airways and not in the alveoli as sketched to different degrees in figures 5(b) and 5(c). In these sketches, dark gas (i.e. thermal gas with no appreciable MR signal) is symbolized by red color (dark gray in 'black and white' representation) while hp 83 Kr is represented by light blue (light gray). The underlying assumption is that gas inhaled during the initial phase of the breath take is more likely to be found in the distal airway and alveoli region than gas inhaled during the later phase of the breath take. Admittedly, this assumption constitutes a simplified model of the breathing process that ignores that different lobes of the lung expand at different times in the breathing cycle and/or other complexities such as mixing that occur during inhalation. Furthermore, the presence of many surface-to-volume ratios at different generations of branching of the airways can not be quantified. Even in the simplified model, the largest contribution to the hp⁸³Kr signal will originate from the alveolar region of the lung due to the small total overall volume of the conducting airways, which typically contribute 5% or less to the overall lung volume (Fowler, 1948; Heller et al., 1999). Nevertheless, scheme 2 should direct hp 83Kr more towards the proximal airways than scheme 1 and scheme 2 is therefore expected to provide the slowest longitudinal relaxation.

As an example for data obtained from one lung specimen (N = 1), figure 6 shows a resulting set of decay curves for the three different inhalation schemes using a total inhalation volume of $V_i = 20$ ml. Two different volumes of thermal gas, $V_i^{dark} = 6$ ml and $V_i^{dark} = 12$ ml have been used with scheme 2. Figure 6 shows a tendency that is further confirmed when the averages of T₁ values of lung specimens from 5 individual rats (N = 5) with at least two repeat measurements per specimen are taken. As reported in table 1, the slowest T₁ time of 1.48 ± 0.07 s was indeed observed with scheme 2 using $V_i^{dark} = 12$ ml whereas T₁ of 1.31 ± 0.05 s was observed for $V_i^{dark} = 6$ ml. By comparison, the data for V_i = 20 ml using scheme 1 with $V_i^{dark} = 0$ was T₁ = 1.25 ± 0.07 s. The error values shown are one standard deviation of the calculated averaged value obtained from at least 10 T₁ measurements. The data demonstrates the increase in the relaxation time with increasing 'dark' inhaled gas volume.

Scheme 3 used an initial inhalation of V_i^{hp} =6 ml hp ⁸³Kr gas immediately followed by inhalation of thermal gas until the selected total inhalation volume V_i was reached. In this case, the transfer of dark gas had to occur rapidly after the hp ⁸³Kr inhalation in order to minimize polarization losses through fast ⁸³Kr relaxation in the lungs. This was accomplished by transferring hp ⁸³Kr into the minimum possible storage volume of V_B = 6 ml (i.e. with glass extension tubes removed from storage container V_B). The lungs were then inflated using suction with the ventilation syringe to achieve the final inhalation volume

 $V_i = V_i^{hp} + V_i^{dark} = 6ml + V_i^{dark}$. Assuming relatively little gas mixing in the narrow tube of storage container V_B during the approximately 1 s inhalation period, any gas inhaled beyond

 V_i^{hp} =6 ml would be largely thermal gas that enters storage container V_B through its exhaust opening to the ambient atmosphere (figure 1).

Using analogous arguments to those presented for scheme 2, the procedure of scheme 3 should direct hp 83 Kr towards the respiratory zones and the distal airways as opposed to the larger proximal airways (figure 5(d)). As a consequence, the already weak hp 83 Kr signal from the relatively small volume of the proximal airways would be further suppressed at moderate inhalation volumes. Even the relative signal contribution from the distal airways is probably reduced compared to that from the alveoli. This assumption is supported by the observation that the steepest slope occurs for scheme 3, as shown in figure 6. Scheme 3 leads to an averaged relaxation time of $T_1 = 1.00 \pm 0.08$ s for $V_i = 20$ ml (see table 1).

3.6. Longitudinal relaxation times in lungs as a function of lung inhalation

The inhalation schemes discussed in the previous section were applied to a series of inhalation volumes ranging from $V_i = 3$ to $V_i = 20$ ml in rat lungs. The resulting T_1 times are presented as a function of V_i in figure 7 and numerically in table 1. Each relaxation time is the average of measurements from lung specimen of five individual rats and of at least two T_1 fittings per inhalation volume and specimen. The relatively small deviation in the relaxation data obtained from lungs of different individual rats (similar age and weight, 3 months old and 350-425 g, respectively) substantiates the generality of the results. One trend discussed in context with figure 6 is observed for all inhalation volumes: Scheme 2 provides the longest T_1 values for a chosen total inhalation volume V_i , followed by scheme 1, while the shortest T_1 times are found for scheme 3.

As the inhalation volume increases for all inhalation schemes, T_1 times become either shorter or remain constant. A statistically significant slowing of the relaxation with increasing V_i is never observed. The T_1 times found with inhalations in scheme 2 decrease monotonically with increasing V_i over the measured range. The relaxation time decrease is more pronounced for V_i^{dark} =12 ml than for V_i^{dark} =6 ml This indicates that the contribution from the slow relaxing signal of the larger airways to the overall signal cannot be neglected at any inhalation volume V_i for scheme 2.

Experiments using scheme 3 were designed to suppress the hp ⁸³Kr signals from the larger airway and therefore to obtain the T₁ times of ⁸³Kr in respiratory zones. Scheme 3 was performed with total inhalation volumes V_i ranging from 9 to 20 ml. As with scheme 1, the T₁ times with scheme 3 decrease and eventually stabilize at about 12 ml total inhalation volume V_i. While some mixing may have occurred in the inhalation process, the hp ⁸³Kr gas at high V_i is expected to be more confined to the respiratory zones and the distal airways such as bronchioles and smaller bronchia. This assumption is supported by the measurements with scheme 3 that resulted to T₁ \approx 1.0 s for V_i \geq 12ml. This value is significantly shorter than the corresponding value of T₁ \approx 1.3 s obtained with scheme 1 for V_i \geq 12ml.

3.7. Consequences for models of alveolar recruitment

The complete absence of an increase in ⁸³Kr T₁ times with increasing lung inhalation in the distal part of the lung warrants consideration. This observation is reminiscent to the finding that inhalation volume independent ADC is observed in diffusion measurement with 3 He in lungs (see for instance (Gierada et al., 2009) However, ADC is determined on much shorter timescales, typically 1 - 3 ms, and the hp gas probes therefore a much smaller region (Yablonskiy et al., 2002) than the relaxation measurements of 2.6 s duration reported here. Note that a free diffusing gas of similar composition would display centimeter-sized mean displacements during the 2.6 s time period, as estimated from D = 0.63 cm²s⁻¹ for diffusion of krypton in helium and $D = 0.15 \text{ cm}^2\text{s}^{-1}$ for diffusion of krypton in nitrogen at 293 K (Lide, 2001). The inhalation volume independent 83 Kr T₁ times in the more distal part of the airways indicate constant surface to volume ratio in the area probed by the krypton atoms (Stupic et al., 2006). This finding is somewhat unexpected since the surface, S, to volume, V, ratio (S/V) of the lung should decrease with increasing inhalation volume, assuming the S/V of the alevoli behave similar to an expanding balloon or sphere where $V \propto r^3$ and $S \propto r^3$ r^2 . However an apparently constant surface to volume ratio observed through hp ⁸³Kr MR could be explained by rapid recruitment of alveolar regions into the inflated state of constant surface to volume ratio without further changes during continuing inhalation. Although the surface to volume ratio of the lung should decrease with increasing inhalation (based again on the underlying assumption of an expanding sphere model), the hp ⁸³Kr MR signal originates only from inflated alveoli that are already expanded to their final surface to

volume ratio. Alveoli that are not yet inflated are still void of hp ⁸³Kr and cannot contribute to the MR signal.

Following this assumption, the gradual process of inhalation might be a consequence of many different alveolar regions expanding rapidly but at different times rather than of a gradual, continuously ongoing alveolar expansion. The size of these regions could range from macroscopic areas of the lung lobes down to small groups of individual alveoli. Arguments based on surface tension would also support the uniform expansion of neighboring alveoli with common water / surfactant surface layers (Escolar and Escolar, 2004). Alternatively, the underlying assumption that the S/V of expanding alveoli behave similar to that of expanding (micro) spheres maybe wrong. Whatever cause will ultimately be identified to be responsible for the inhalation volume independence of the ⁸³Kr T₁ relaxation, it leads to easily reproducible ⁸³Kr relaxation measurements that may facilitate future diagnosis of damaged lungs for which analysis of T₁ times may provide a biomarker.

3.8. ⁸³Kr T₁ times in rat lungs using breathable hp ⁸³Kr mixtures with oxygen

Molecular oxygen in breathable noble gas mixtures causes paramagnetic relaxation of the noble gas nuclei. For instance, the longitudinal relaxation time of ¹²⁹Xe is reduced from hours (or even days in the case of ³He) to 10 - 20 s in the presence of 20% oxygen (Jameson *et al.*, 1988; Saam *et al.*, 1995; Moller *et al.*, 2002). However, the T₁ time of a nuclear spin *I* caused by intermolecular paramagnetic relaxation is dependent on the square of the gyromagnetic ratio γ of this nuclear spin (Abragam, 1961).

$$\frac{1}{T_1^{para}} \propto \gamma_1^2 \tag{4}$$

A 50 fold reduced paramagnetic relaxation rate, T_1^{-1} , for ⁸³Kr compared to ¹²⁹Xe (Cleveland *et al.*, 2008b) is therefore suggested by the different gyromagnetic ratios: $(\gamma_{129_{Xe}}/\gamma_{83_{Kr}})^2 \approx 50$. However, the relaxation also follows a $T_1^{-1} \propto r^{-6}$ dependence, with r as the distance of the nuclei from the paramagnetic spins during the noble gas – oxygen interaction. The distance, r, can be estimated by the van der Waals radii of the noble gas atoms, resulting to the ratio $r_{129_{Xe}}^{-6}/r_{83_{Kr}}^{-6} \approx 0.55$. Further factors are the different rates for binary gas phase collision and, if

 $r_{129\chi_e}$, r_{83K_r} voids. Further factors are the different factors for only gas phase conston and, if present, the different lifetimes for noble gas – oxygen van der Waals complexes. However, it is unlikely that these factors will offset the dominating influence originating from the large

difference in gyromagnetic ratios (note that $(\gamma_{3_{He}}/\gamma_{83_{Kr}})^2 \approx 390$). Therefore, the low gyromagnetic ratio of 83 Kr (4% of that of 1 H) provides a distinct advantage in preserving hyperpolarization in the presence of oxygen. This advantage can of course only be utilized if the inherently low signal intensity with such low γ value is sufficiently boosted through hyperpolarization.

The effect of oxygen upon 83 Kr relaxation in rat lungs was studied by inhaling 18 ml of a mixture of hp 83 Kr with molecular oxygen using inhalation scheme 1. After oxygen delivery using the injection syringe V_A, hp 83 Kr was added and a 10 s delay allowed for the mixing of the two gases within the 27 mm inner diameter syringe. The gas mixture was then transferred into storage container V_B where it was available for inhalation by the lung. The relaxation time of 83 Kr in a mixture with 20% oxygen was 1.25 ± 0.01 s in three replicate measurements with a single lung specimen. Increasing the oxygen concentration to 40%, the T₁ value was 1.22 ± 0.01 s from two replicate measurements with the same specimen. The T₁ times obtained with both oxygen mixtures are within the error of T₁ = 1.25 ± 0.07 s

recorded in the absence of oxygen under otherwise identical conditions. Thus, the presence of oxygen in a breathable gas mixture has no appreciable effect on ⁸³Kr T₁ in the rat lungs. A 20% oxygen mixture leads, however, to an approximately 25% reduction in the MR signal because of the reduced hp ⁸³Kr concentration and the relaxation during the 10 s mixing phase before inhalation.

4. Conclusions

Alkali metal vapor free hp 83 Kr with 4.4 % spin polarization was generated in a mixture of 25 % krypton in helium and nitrogen. The resulting hp 83 Kr MR signal intensity was sufficient despite the natural abundance of krypton gas with only 11.5 % 83 Kr and the experimental 4-fold dilution in the gas mixture. A custom built hp-gas delivery system and a ventilation chamber allowed for hp 83 Kr to be actively inhaled *ex vivo* by isolated rat lungs. The 83 Kr T₁ times in the rat lungs ranged from 1.0 to 2.2 s depending on the lung inhalation volume and the applied inhalation scheme.

The 83 Kr T₁ relaxation accelerated with increasing lung inhalation volume when the inhalation scheme did not suppress the hp 83 Kr signal from the proximal airway region. This effect was attributed to the change in the relative contribution of the slower relaxing hp 83 Kr in proximal airways compared to the faster relaxing hp 83 Kr in distal airways. With higher inhalation volume, this ratio shifted towards the hp 83 Kr signal in the alveolar region and thus the apparent relaxation time was shorter.

The hp ⁸³Kr relaxation times were independent of the inhalation volume if the signal from the proximal airways was largely suppressed through usage of an inhalation schemes that directed hp ⁸³Kr towards the distal airways. Depending on the degree of hp ⁸³Kr direction towards the distal airways, the inhalation volume independent relaxation times was $T_1 = 1.3$ s (scheme 1) and $T_1 = 1.0$ s (scheme 3).

At least three aspects of this observation merit further consideration: (1) The observed T_1 times in rat lungs were long enough for *in vivo* usage of hp ⁸³Kr MRI since rats typically breathe at a rate of 1 to 5 times per second. (2) The inhalation volume independent hp ⁸³Kr T_1 relaxation in the distal airways allowed for reproducible measurements. This may be useful for diagnostic studies if pulmonary diseases affect the ⁸³Kr T_1 times. This possibility is further strengthened by the observation of very little deviation in the ⁸³Kr T_1 times between different lung specimens. (3) The inhalation volume independent hp ⁸³Kr T_1 relaxation suggests a constant, inhalation level independent surface to volume ratio in the alveolar region. This preliminary finding supports alveolar recruitment models that postulate a rapid inflation of individual alveolar regions into an expanded state rather than a slow expansion throughout the breathing process.

Finally, the quadrupolar driven T_1 relaxation of ⁸³Kr in the rat lungs was not affected by the presence of up to 40% oxygen. This finding may further facilitate future use of hp ⁸³Kr for pulmonary studies *in vivo*.

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References

- Abragam, A. The Principles of Nuclear Magnetism. Oxford, UK: Oxford University Press; 1961.
- Albert MS, Cates GD, Driehuys B, Happer W, Saam B, Springer CS, Wishnia A. Biological Magnetic-Resonance-Imaging Using Laser Polarized Xe-129. Nature. 1994; 370:199–201. [PubMed: 8028666]
- Altes TA, Mata J, de Lange EE, Brookeman JR, Mugler JP. Assessment of lung development using hyperpolarized helium-3 diffusion MR imaging. J Magn Reson Imaging. 2006; 24:1277–83. [PubMed: 17096396]
- Altes TA, Powers PL, Knight-Scott J, Rakes G, Platts-Mills TAE, de Lange EE, Alford BA, Mugler JP, Brookeman JR. Hyperpolarized He-3 MR lung ventilation imaging in asthmatics: Preliminary findings. J Magn Reson Imaging. 2001; 13:378–84. [PubMed: 11241810]
- Brinkmann D, Kuhn D. Nuclear Magnetic-Relaxation of Kr-83 in Krypton Gas. Physical Review A. 1980; 21:163–7.
- Cherubini A, Bifone A. Hyperpolarised xenon in biology. Progress in Nuclear Magnetic Resonance Spectroscopy. 2003; 42:1–30.
- Cieslar K, Stupar V, Canet-Soulas E, Gaillard S, Cremillieux Y. Alveolar oxygen partial pressure and oxygen depletion rate mapping in rats using He-3 ventilation imaging. Magnetic Resonance in Medicine. 2007; 57:423–30. [PubMed: 17152086]
- Cleveland ZI, Cofer GP, Metz G, Beaver D, Nouls J, Kaushik SS, Kraft M, Wolber J, Kelly KT, McAdams HP, Driehuys B. Hyperpolarized Xe-129 MR Imaging of Alveolar Gas Uptake in Humans. Plos One. 2010; 5
- Cleveland ZI, Meersmann T. Binary-collision-induced longitudinal relaxation in gas-phase Kr-83. Journal of Chemical Physics. 2008a; 129:244304. [PubMed: 19123504]
- Cleveland ZI, Meersmann T. Density-independent contributions to longitudinal relaxation in Kr-83. Chemphyschem. 2008b; 9:1375–9. [PubMed: 18574827]
- Cleveland ZI, Moller HE, Hedlund LW, Driehuys B. Continuously Infusing Hyperpolarized Xe-129 into Flowing Aqueous Solutions Using Hydrophobic Gas Exchange Membranes. Journal of Physical Chemistry B. 2009; 113:12489–99.
- Cleveland ZI, Pavlovskaya GE, Elkins ND, Stupic KF, Repine JE, Meersmann T. Hyperpolarized Kr-83 MRI of lungs. Journal of Magnetic Resonance. 2008a; 195:232–7. [PubMed: 18948043]
- Cleveland ZI, Pavlovskaya GE, Stupic KF, LeNoir CF, Meersmann T. Exploring hyperpolarized 83Kr by remotely detected NMR relaxometry. Journal of Chemical Physics. 2006; 124:044312. [PubMed: 16460167]
- Cleveland ZI, Pavlovskaya GE, Stupic KF, Wooten JB, Repine JE, Meersmann T. Detection of Tobacco Smoke Deposition by Hyperpolarized Krypton-83 MRI. Magnetic Resonance Imaging. 2008b; 26:270–8. [PubMed: 17826938]
- Cleveland ZI, Stupic KF, Pavlovskaya GE, Repine JE, Wooten JB, Meersmann T. Hyperpolarized Kr-83 and Xe-129 NMR relaxation measurements of hydrated surfaces: Implications for materials science and pulmonary diagnostics. Journal of the American Chemical Society. 2007; 129:1784– 92. [PubMed: 17243679]
- Cowgill DF, Norberg RE. Pulsed Nmr-Studies of Self-Diffusion and Defect Structure in Liquid and Solid Krypton. Physical Review B. 1976; 13:2773–81.
- de Lange EE, Mugler JP, Brookeman JR, Knight-Scott J, Truwit JD, Teates CD, Daniel TM, Bogorad PL, Cates GD. Lung air spaces: MR imaging evaluation with hyperpolarized He-3 gas. Radiology. 1999; 210:851–7. [PubMed: 10207491]
- Driehuys B, Cates GD, Miron E, Sauer K, Walter DK, Happer W. High-volume production of laserpolarized Xe-129. Applied Physics Letters. 1996; 69:1668–70.
- Driehuys B, Cofer GP, Pollaro J, Mackel JB, Hedlund LW, Johnson GA. Imaging alveolar-capillary gas transfer using hyperpolarized Xe-129 MRI. Proceedings of the National Academy of Sciences of the United States of America. 2006; 103:18278–83. [PubMed: 17101964]
- Driehuys B, Moller HE, Cleveland ZI, Pollaro J, Hedlund LW. Pulmonary Perfusion and Xenon Gas Exchange in Rats: MR Imaging with Intravenous Injection of Hyperpolarized Xe-129. Radiology. 2009; 252:386–93. [PubMed: 19703880]

- Escolar JD, Escolar A. Lung histeresis: a morphological view. Histol Histopath. 2004; 19:159–66.
- Fain SB, Altes TA, Panth SR, Evans MD, Waters B, Mugler JP, Korosec FR, Grist TM, Silverman M, Salerno M, Owers-Bradley J. Detection of age-dependent changes in healthy adult lungs with diffusion-weighted He-3 MRI. Academic Radiology. 2005; 12:1385–93. [PubMed: 16253850]
- Fischer MC, Kadlecek S, Yu JS, Ishii M, Emami K, Vahdat V, Lipson DA, Rizi RR. Measurements of regional alveolar oxygen pressure using hyperpolarized He-3 MRI. Academic Radiology. 2005; 12:1430–9. [PubMed: 16253855]
- Fowler WS. Lung Function Studies .2. the Respiratory Dead Space. Americam Journal of Physiology. 1948; 154:405–16.
- Gao JH, Lemen L, Xiong JH, Patyal B, Fox PT. Magnetization and diffusion effects in NMR imaging of hyperpolarized substances. Magnetic Resonance in Medicine. 1997; 37:153–8. [PubMed: 8978645]
- Gierada DS, Woods JC, Bierhals AJ, Bartel ST, Ritter JH, Choong CK, Das NA, Hong C, Pilgram TK, Chang YV, Jacob RE, Hogg JC, Battafarano RJ, Cooper JD, Meyers BF, Patterson GA, Yablonskiy DA, Conradi MS. Effects of Diffusion Time on Short-Range Hyperpolarized He-3 Diffusivity Measurements in Emphysema. J Magn Reson Imaging. 2009; 30:801–8. [PubMed: 19787725]
- Goodson BM. Nuclear magnetic resonance of laser-polarized noble gases in molecules, materials, and organisms. Journal of Magnetic Resonance. 2002; 155:157–216. [PubMed: 12036331]
- Heller H, Konen-Bergmann M, Schuster KD. An algebraic solution to dead space determination according to Fowler's graphical method. Computers and Biomedical Research. 1999; 32:161–7. [PubMed: 10337497]
- Hilty C, Lowery TJ, Wemmer DE, Pines A. Spectrally resolved magnetic resonance imaging of a xenon biosensor. Angewandte Chemie-International Edition. 2006; 45:70–3.
- Jameson CJ, Jameson AK, Hwang JK. NUCLEAR-SPIN RELAXATION BY INTERMOLECULAR MAGNETIC DIPOLE COUPLING IN THE GAS-PHASE - XE-129 IN OXYGEN. Journal of Chemical Physics. 1988; 89:4074–81.
- Lide, DR., editor. CRC Handbook of Chemistry and Physics. Boca Raton, FL: CRC Press; 2001.
- Lutey BA, Lefrak SS, Woods JC, Tanoli T, Quirk JD, Bashir A, Yablonskiy DA, Conradi MS, Bartel ST, Pilgram TK, Cooper JD, Gierada DS. Hyperpolarized He-3 MR imaging: Physiologic monitoring observations and safety considerations in 100 consecutive subjects. Radiology. 2008; 248:655–61. [PubMed: 18641256]
- Middleton H, Black RD, Saam B, Cates GD, Cofer GP, Guenther R, Happer W, Hedlund LW, Johnson GA, Juvan K, Swartz J. Mr-Imaging with Hyperpolarized He-3 Gas. Magnetic Resonance in Medicine. 1995; 33:271–5. [PubMed: 7707920]
- Moller HE, Chen XJ, Saam B, Hagspiel KD, Johnson GA, Altes TA, de Lange EE, Kauczor HU. MRI of the lungs using hyperpolarized noble gases. Magnetic Resonance in Medicine. 2002; 47:1029–51. [PubMed: 12111949]
- Mortuza MG, Anala S, Pavlovskaya GE, Dieken TJ, Meersmann T. Spin-exchange optical pumping of high-density xenon-129. Journal of Chemical Physics. 2003; 118:1581–4.
- Pavlovskaya GE, Cleveland ZI, Stupic KF, Meersmann T. Hyperpolarized Krypton-83 as a New Contrast Agent for Magnetic Resonance Imaging. Proceedings of the National Academy of Sciences of the United States of America. 2005; 102:18275–9. [PubMed: 16344474]
- Peces-Barba G, Ruiz-Cabello J, Cremillieux Y, Rodriguez I, Dupuich D, Callot V, Ortega M, Arbo MLR, Cortijo M, Gonzalez-Mangado N. Helium-3 MRI diffusion coefficient: correlation to morphometry in a model of mild emphysema. European Respiratory Journal. 2003; 22:14–9. [PubMed: 12882445]
- Ruppert K, Brookeman JR, Hagspiel KD, Mugler JP. Probing lung physiology with xenon polarization transfer contrast (XTC). Magnetic Resonance in Medicine. 2000; 44:349–57. [PubMed: 10975884]
- Ruppert K, Mata JF, Wang HTJ, Tobias WA, Cates GD, Brookeman JR, Hagspiel KD, Mugler JP. XTC MRI: Sensitivity improvement through parameter optimization. Magnetic Resonance in Medicine. 2007; 57:1099–109. [PubMed: 17534927]

- Saam B, Happer W, Middleton H. Nuclear-Relaxation of He-3 in the Presence of O-2. Physical Review A. 1995; 52:862–5. [PubMed: 9912313]
- Samee S, Altes T, Powers P, de Lange EE, Knight-Scott J, Rakes G, Mugler JP, Ciambotti JM, Alford BA, Brookeman JR, Platts-Mills TAE. Imaging the lungs in asthmatic patients by using hyperpolarized helium-3 magnetic resonance: Assessment of response to methacholine and exercise challenge. Journal of Allergy and Clinical Immunology. 2003; 111:1205–11. [PubMed: 12789218]
- Santyr GE, Lam WW, Parra-Robles JM, Taves TM, Ouriadov AV. Hyperpolarized noble gas magnetic resonance imaging of the animal lung: Approaches and applications. Journal of Applied Physics. 2009; 105
- Schroder L, Lowery TJ, Hilty C, Wemmer DE, Pines A. Molecular imaging using a targeted magnetic resonance hyperpolarized biosensor. Science. 2006; 314:446–9. [PubMed: 17053143]
- Stupic KF, Cleveland ZI, Pavlovskaya GE, Meersmann T. Quadrupolar Relaxation of Hyperpolarized Krypton-83 as a Probe for Surfaces. Solid State Nuclear Magnetic Resonance. 2006; 29:79–84. [PubMed: 16202568]
- Stupic KF, Cleveland ZI, Pavlovskaya GE, Meersmann T. Hyperpolarized Xe-131 NMR spectroscopy. Journal of Magnetic Resonance. 2011; 208:58–69. [PubMed: 21051249]
- Tustison NJ, Altes TA, Song G, de Lange EE, Mugler JP, Gee JC. Feature Analysis of Hyperpolarized Helium-3 Pulmonary MRI: A Study of Asthmatics Versus Nonasthmatics. Magnetic Resonance in Medicine. 2010; 63:1448–55. [PubMed: 20512846]
- van Beek EJR, Dahmen AM, Stavngaard T, Gast KK, Heussel CP, Krummenauer F, Schmiedeskamp J, Wild JM, Sogaard LV, Morbach AE, Schreiber LM, Kauczor HU. Hyperpolarised He-3 MRI versus HRCT in COPD and normal volunteers: PHIL trial. European Respiratory Journal. 2009; 34:1311–21. [PubMed: 19541712]
- Walker TG, Happer W. Spin-exchange optical pumping of noble-gas nuclei. Review of Modern Physics. 1997; 69:629–42.
- Weibel ER. What makes a good lung? The morphometric basis of lung function. Swiss Med Wkly. 2009; 139:375–86. [PubMed: 19629765]
- Woods JC, Choong CK, Yablonskiy DA, Bentley J, Wong J, Pierce JA, Cooper JD, Macklem PT, Conradi MS, Hogg JC. Hyperpolarized He-3 diffusion MRI and histology in pulmonary emphysema. Magnetic Resonance in Medicine. 2006; 56:1293–300. [PubMed: 17058206]
- Wu Z, Happer W, Kitano M, Daniels J. Experimental Studies of Wall Interactions of Adsorbed Spin-Polarized Xe-131 Nuclei. Physical Review A. 1990; 42:2774–84. [PubMed: 9904348]
- Yablonskiy DA, Sukstanskii AL, Leawoods JC, Gierada DS, Bretthorst GL, Lefrak SS, Cooper JD, Conradi MS. Quantitative in vivo assessment of lung microstructure at the alveolar level with hyperpolarized He-3 diffusion MRI. Proceeding of the National Academy of Sciences of the United States of America. 2002; 99
- Yu J, Ishii M, Law M, Woodburn JM, Emami K, Kadlecek S, Vahdat V, Guyer RA, Rizi RR. Optimization of scan parameters in pulmonary partial pressure oxygen measurement by hyperpolarized He-3 MRI. Magnetic Resonance in Medicine. 2008; 59:124–31. [PubMed: 18050348]



Figure 1.

Overview of experimental setup and gas delivery system for hp ⁸³Kr MR experiments. (a) Experimental setup for the hp⁸³Kr delivery to the lung.⁸³Kr is hyperpolarized through SEOP in the pumping cell at 135 kPa pressure, while valves #1 and #3 are closed and valve #2 is opened to evacuate the syringe. Valve #2 is then closed and valve #1 is opened to deliver hp ⁸³Kr through pressure equalization into storage container V_A. Valve 1 is closed again and valve 3 is opened to allow for hp 83 Kr injection into the storage container with volume V_B. The lungs are fully immersed in Belzer-MPS solution (UW Kidney Preservation Solution) with the trachea pointing downward and connected to the external opening below the ventilation chamber. (b) Expanded view of gas delivery to storage container V_B with valve # 3 opened. The hp 83 Kr passes over the lung orifice as it is slowly injected into V_B while the pressure of the system stays approximately constant, thus preventing any premature entry of hp 83Kr into the lung. Excess hp 83Kr delivered beyond the capacity of storage container V_B (expandable from 6 ml to 30 ml) is exhausted to open air. (c) Delivery of hp⁸³Kr to the lungs (valve #3 closed). A syringe connected to the top of the ex vivo lung ventilation chamber creates a suction, thus causing inhalation by the lungs and therefore active inhalation of the hp 83 Kr gas into the lung. The suction volume V_s of the syringe was correlated to the inhalation volume Vi prior to the experiments by monitoring the level of the liquid in the ventilation chamber as a function of V_s.



Figure 2.

Time resolved series of hp 83 Kr MR spectra for an excised rat lung with V_i = 15 ml inhalation of 25% krypton in a mixture with helium/nitrogen (inhalation scheme 1). The MR signals were acquired from a series of 12° RF pulses evenly spaced every 0.2 s. No signal is initially visible, demonstrating that gas transfer into storage container V_B does not lead to MR detectable amounts of hp 83 Kr in the lung. As suction is applied to the outside of the lung and the lung inhales hp 83 Kr, the MR signal becomes observable and starts to increase over time. The exact timing of the manually applied suction is not known but is also not needed for the analysis. Approximately coinciding with the end of the inhalation period, T₁ relaxation leads to an observable decay in the T₁ the signal over time. At t = 0 s, labeled with a diamond marker, lung movement no longer contributes to the change in signal intensity.



Figure 3.

Experimental data and fitting for a $V_i = 20$ ml lung inhalation of an excised rat lung with hp ⁸³Kr using inhalation scheme 1 (25% krypton in helium/nitrogen; natural abundance ⁸³Kr). (a) 0.2 s time resolved hp ⁸³Kr MR signal intensity behavior found in *ex vivo* rat lung analogue to figure 2 but extending further out in time. The diamond marker, also marked at t = 0 s, is the time point where only T₁ decay signal is observed during 'breath hold' without interference from lung movement. (b) Plot of the natural logarithm of the integrated intensity from the data shown in 3(a) versus time. Three time periods are labeled as follows: Data acquired prior to t = 0 s (cosses). Only data recorded between t = 0 to 2.6 s (solid circles) is fitted to a linear function with a correction factor for the polarization destroyed by the RF pulse to calculate the T₁ value using equation (1).



Figure 4.

Experimental and fitting data for a $V_i = 6$ ml lung inhalation of an *ex vivo* rat lung (inhalation scheme 1) at conditions otherwise identical to those described for the experiment in figure 3 (all symbols and markings used as in figure 3).



Figure 5.

Sketch of gas distribution for hp ⁸³Kr colored in blue (light gray in black and white representation) and thermal gas (non-hp gas) colored in red (dark gray) in a lung for the three different inhalation schemes. (a) Inhalation scheme 1: only hp ⁸³Kr is inhaled, i.e.

 $V_i^{hp} = V_i$; (b) <u>inhalation scheme 2</u>: $V_i^{dark} = 6ml$ of thermal gas inhaled followed by inhalation of

hp ⁸³Kr to reach the desired inhalation volume V_i; (c) also <u>inhalation scheme 2</u>: $V_i^{dark} = 12ml$ of thermal gas inhaled followed by inhalation of hp ⁸³Kr to reach the desired inhalation

volume V_i; (d) <u>inhalation scheme 3</u>: $V_i^{hp} = 6ml$ of hp ⁸³Kr inhaled followed by inhalation of thermal gas to reach the desired V_i. The sketch of the lung loosely follows the architecture of the lung described by Weibel (Weibel, 2009).



Figure 6.

Natural log of the integrated hp ⁸³Kr signal intensities from an excised rat lung (from N = 1 animal) as a function of time and normalized to their values at t = 0 s (i.e. experimental number n = 0). Presented here are the results from four different inhalation schemes, each with a total inhalation volume of V_i = 20 ml. <u>Inhalation scheme 1</u>: only hp ⁸³Kr is inhaled, i.e. $V_i^{hp} = V_i = 20ml$ (closed circles). <u>Inhalation scheme 2</u>: $V_i^{dark} = 6ml$ of thermal gas inhaled prior to $V_i^{hp} = 14ml$ of hp ⁸³Kr (open triangles) and $V_i^{dark} = 12ml$ thermal gas inhaled prior to $V_i^{hp} = 8ml$ hp ⁸³Kr (filled triangles). <u>Inhalation scheme 3</u>: $V_i^{hp} = 6ml$ of hp ⁸³Kr inhaled prior to inhalation of $V_i^{dark} = 14ml$ of MR non-contributing gas (open circles). Each line is the result of a linear fitting and includes a compensation term for spin destruction by the applied 12° pulses. T₁ values obtained from this fitting are presented in figure 7 and table 1.



Figure 7.

Average longitudinal relaxation times as a function of inhalation volume for different inhalation schemes using the corresponding symbols from figure 6 (i.e. inhalation scheme 1 (closed circles); inhalation scheme 2 with $V_i^{dark}=6ml$ prior to V_i^{hp} inhalatiom (open triangles); inhalation scheme 2 with $V_i^{dark}=12ml$ prior to V_i^{hp} inhalation (filled triangles); and inhalation scheme 3 with $V_i^{dark}=6ml$ prior to inhalation of V_i^{dark} (open circles)). The dashed line used for scheme 3 was extended to the lower inhalation volumes of scheme 1 as a reference. The T₁ values are presented in table 1 and are averages from T₁ data of lung specimens from 5 individual rats (N = 5) with at least two repeat measurements per specimen. The error values are one standard deviation from the calculated averaged value.

Table 1

Longitudinal relaxation times for *ex vivo* experiments using rat lungs at different inhalation volumes V_i and different inhalation schemes. T_1 values are averages of T_1 data of lung specimens from 5 individual rats with at least two repeat measurements per specimen. The error values shown are one standard deviation from the calculated averaged value obtained from the (at least) ten T_1 measurements. (Note that the fittings were not error weighted by the signal to noise ratio of the spectra and the fitting accuracy was not considered for the error calculation. However, the fitting error of the individual T_1 measurements is about one order of magnitude smaller than the standard deviation value between the individual T_1 measurements. Examples of relaxation curves are shown in figure 6).

Inhalation volume $V_i = V_i^{dark} + V_i^{hp}$	Inhalation scheme 1 Only hp 83 Kr gas $V_i = V_i^{hp}$	Inhalation scheme 2 V_i^{dark} =6 ml thermal gas at start of inhalation followed by hp ⁸³ Kr	Inhalation scheme 2 $V_i^{dark} = 12 \text{ ml}_{thermal}$ gas at start of inhalation followed by hp ⁸³ Kr	Inhalation scheme 3 $V_i^{hp} = 6 \text{ ml}_{hp} s_{^3Kr}$ gas at start of inhalation followed by thermal (non-hp) gas
	T ₁ [s]	T ₁ [s]	T ₁ [s]	T ₁ [s]
3 ml	2.11 ± 0.75			
6 ml	1.57 ± 0.16			
9 ml	1.43 ± 0.10	2.00 ± 0.16		1.20 ± 0.13
12 ml	1.31 ± 0.08	1.57 ± 0.07		1.07 ± 0.08
15 ml	1.29 ± 0.12	1.42 ± 0.09	2.19 ± 0.29	1.01 ± 0.09
18 ml	1.28 ± 0.08	1.36 ± 0.05	1.58 ± 0.08	1.01 ± 0.07
20 ml	1.25 ± 0.07	1.31 ± 0.05	1.48 ± 0.07	1.00 ± 0.08
20 ml (20% O_2) ^{<i>a</i>}	1.25 ± 0.01			
20 ml (40% O ₂) ^b	1.22 ± 0.01			

a,b The measurements with 20% and 40% oxygen were limited to 1 rat lung specimen (total) with three and two repeat measurements respectively.