

REVIEW

Bone marrow and the control of immunity

Ende Zhao^{1,2,7}, Huanbin Xu^{3,7}, Lin Wang², Ilona Kryczek¹, Ke Wu², Yu Hu², Guobin Wang² and Weiping Zou^{1,4,5,6}

Bone marrow is thought to be a primary hematopoietic organ. However, accumulated evidences demonstrate that active function and trafficking of immune cells, including regulatory T cells, conventional T cells, B cells, dendritic cells, natural killer T (NKT) cells, neutrophils, myeloid-derived suppressor cells and mesenchymal stem cells, are observed in the bone marrow. Furthermore, bone marrow is a predetermined metastatic location for multiple human tumors. In this review, we discuss the immune network in the bone marrow. We suggest that bone marrow is an immune regulatory organ capable of fine tuning immunity and may be a potential therapeutic target for immunotherapy and immune vaccination.

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INTRODUCTION

Bone marrow is the tissue comprising the center and the epiphysis of bones, which is the place where new blood cells are produced. Bone marrow has been long thought to be a hematopoietic organ. However, it is well known that B cells are produced and matured in the bone marrow. Antigen-specific antibody producing, long-term lived plasma cells are largely found in the bone marrow. Thus, bone marrow contributes to humoral immune responses. Although normal bone marrow lacks the organized T- and B-cell areas, bone marrow is a nest for function, migration and selective retainment of innate and adaptive immune cells. In this review, we discuss the immune networks in the bone marrow. We suggest that bone marrow is an immune regulatory organ capable of fine tuning immunity and may be a potential therapeutic target for immunotherapy and immune vaccination.

BONE MARROW STRUCTURE

Bone is an organ composed of cortical and trabecular bone, cartilage, hemopoetic and connective tissues. Spongy or trabecular bone is composed of a lattice of fine bone plates filled with hematopoietic marrow, fat containing marrow or blood vessels. Arterial vessels enter the marrow through foramina nutricia and then divide into several arterioles. Small arterioles and capillaries from these vessels span throughout the bone marrow and supply sinusoids, which are interconnected by intersinusoidal capillaries (Figures 1 and 2).¹ The bone marrow cavity in trabecular bone is subdivided into four regions: endosteal, subendosteal, central and perisinusoidal. Bone marrow consists of a hematopoietic component (parenchyma) and a vascular component (stroma) (Figure 1). The parenchyma includes hematopoietic stem cells (HSCs) and hematopoietic progenitor cells (Figure 1), which are not randomly distributed in the bone marrow but rather are localized close

to the endosteum of the bone and more around blood vessels (Figure 2).

Bone marrow stroma contains multipotential non-hematopoietic progenitor cells (Figure 1c) capable of differentiating into various tissues of mesenchymal origin, including osteoblasts, endothelial cells, reticular cells, fibroblasts and adipocytes. The bone marrow's microvasculature with a single layer endothelium forms sinusoid (Figure 1b), which radially distributes around the draining central sinus. The vasculature provides the barrier between the bone marrow compartment as a functional and spatial entity from extralymphoid organ and the peripheral circulation.¹

The stromal cells including endothelial cells provide signals for migration of individual leukocytes into and out of the bone marrow, involving in rolling/extravasations along the vascular endothelium. Compared with other organ-specific endothelial cells, bone marrow-derived endothelial cells constitutively express certain cytokines and adhesion molecules like vascular cell adhesion molecule 1 (VCAM-1 or CD106) and E-selectin (Figure 3). Additionally, organ regeneration relies on the presence of endothelial precursors among the grafted cell population, which improves vascularization of damaged tissues or by secretion of pro-angiopoietic factors by the infused cells.² Therefore, it not only implies a role of bone marrow microvasculature system in stem cell mobilization and development, but also indicates that they play key roles in migration and maintenance of leukocyte function in bone marrow environment.

IMMUNE CELLS IN THE BONE MARROW

Immune system is functionally compartmentalized into primary lymphoid organs and secondary lymphoid tissues where immune responses are initiated and maintained. T-cell areas of secondary

¹Department of Surgery, University of Michigan School of Medicine, Ann Arbor, MI, USA; ²Union Hospital, Huazhong University of Science and Technology School of Medicine, Wuhan, China; ³Tulane National Primate Research Center, Tulane University School of Medicine, Covington, LA, USA; ⁴Graduate Program in Immunology, University of Michigan School of Medicine, Ann Arbor, MI, USA; ⁵Graduate Program in Biology, University of Michigan School of Medicine, Ann Arbor, MI, USA and ⁶University of Michigan Comprehensive Cancer Center, University of Michigan School of Medicine, Ann Arbor, MI, USA

⁷These two authors contributed equally to this work.

Correspondence: Dr Weiping Zou, University of Michigan School of Medicine, C560B MSRB II, 1150 W. Medical Center Dr., Ann Arbor, MI 48109-0669, USA.

E-mail: wzou@med.umich.edu

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Bone marrow structure

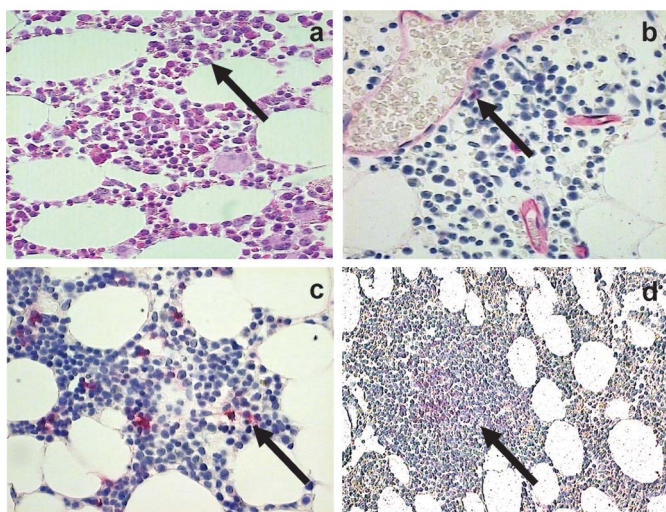


Figure 1 Bone marrow morphology and key cellular components: bone marrow core biopsy sections were subjected to HE staining (a), anti-CD34 staining (b), anti-CD38 staining (c) and anti-CD20 staining (d). The stained sections were revealed with fast red, analyzed by conventional microscope and images were shown with $\times 40$ (a–c) and $\times 10$ (d) magnification. The positive cells were judged by positive staining (red, b–d) as well as the morphology. The control mAb staining reveals no positive cells (not shown). HE, hematoxylin and eosin; mAb, monoclonal antibody.

lymphoid organs have a unique architecture and cellular composition which is thought to be a prerequisite for primary T-cell responses.³ Bone marrow displays structural and functional features resembling a secondary lymphoid organ, and contains follicle-like structures similar to lymph nodes or spleen, although it lacks the organized T- and B-cell areas (Figure 1d). Bone marrow microenvironment provides

Bone marrow structure and potential T cell including Treg anatomical localization in the bone marrow

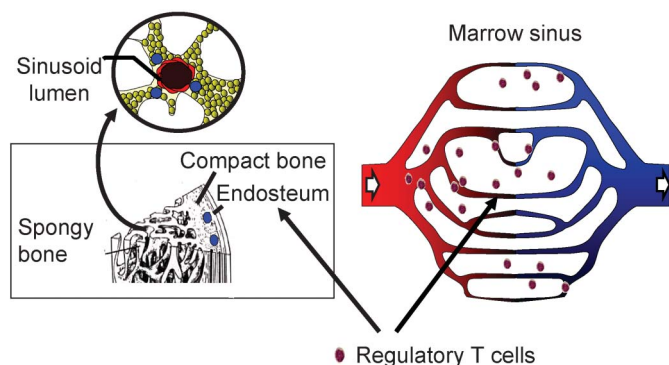


Figure 2 Bone marrow structure. The bone marrow is encased by cortical bone and traversed by trabecular bone. Bone marrow consists of a highly organized meshwork of thin-walled capillary-venous with extracellular matrix that fills the space between the bony trabeculae. The artery and the periosteal capillary network are the two sources of arterial blood for the bone marrow. By successive bifurcations, small branches of the artery ultimately form the capillary-venous sinus network. Murine and human bone marrow harbor immune cells including Treg cells. T cells including Treg cells may reside in the marrow sinus. Treg, regulatory T.

Potential molecular signals for T cell including Treg, OB, OC and DC interaction in the bone marrow

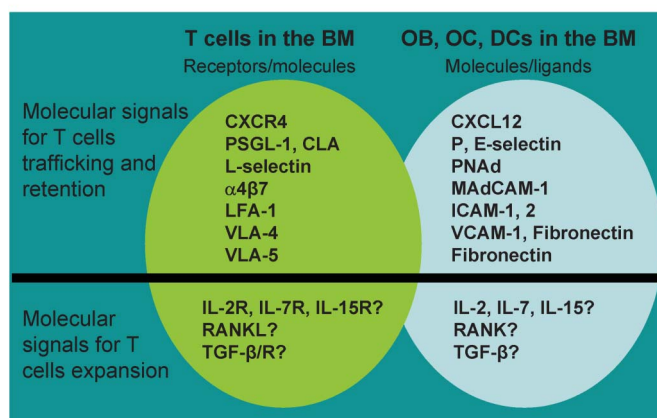


Figure 3 Molecular and cellular interaction between T cells and marrow sinus: T cells including Treg cells might be in the sinus areas or/and endosteum areas. The organization of molecular and cellular niches is known to have a key role in regulating T-cell immunity. Bone marrow contains a large amount of chemokines, adhesion and integrin molecules. These molecular signals may be important for immune cell bone marrow trafficking, retention and expansion. These structural and molecular niches may play a role in bone pathology in cancer patients with bone metastasis including prostate cancer and breast cancer. BM, bone marrow; CLA, conjugated linoleic acid; DC, dendritic cell; ICAM, intercellular adhesion molecule; LFA, leucocyte function antigen; OB, osteoblast; OC, osteoclast; PSGL, P-selectin glycoprotein ligand; RANKL, receptor activator of NF- κ B ligand; TGF, transforming growth factor; Treg, regulatory T; VCAM, vascular cell adhesion molecule; VLA, very late antigen.

appropriate support for T cells to develop in the absence of the thymus.⁴ Lymphoid follicles in the bone marrow are increased during infections, inflammation and autoimmunity.

Bone marrow is vascularized by blood (Figure 1b), not by lymphatic vessels, and could represent a major part of the lymphocyte recirculation network, with billions of lymphocytes recirculating through it per day. It has been shown that bone marrow contains various immune cells (Table 1). Approximately 8%–20% of bone marrow mononuclear cells are lymphocytes, with a T cell/B cell ratio of 5 : 1.^{5,6} Bone marrow lymphocytes are distributed throughout stroma and parenchyma, and condensed in lymphoid follicles. Approximately 1% of the bone marrow mononuclear population represents plasma cells, which can produce antibodies. Mouse bone marrow contains 1%–5% CD3⁺ T cells in mononuclear cells.^{6,7} Among T cells, there are about 1.5% CD4⁺ T cells and 2.0%–2.5% CD8⁺ T cells.^{8–11} Interestingly, approximately one-third of CD4⁺ T cells are CD4⁺CD25⁺ regulatory T (Treg) cells,¹² and the CD4/CD8 ratio in the bone marrow is 1 : 2, which is inverted as compared to both peripheral lymph nodes and the

Table 1 Immune cells in the bone marrow

Immune cells	Percent	Reference
CD4 ⁺ T cells	~1.5%	10, 11
CD8 ⁺ T cells	2–2.5%	8, 9
Regulatory T cells (Treg)	~0.5%	10–12
CD11c ⁺ dendritic cells (DCs)	1–2%	6, 7, 13
B cells	~1%	63
Plasma cell	~0.5%	5, 66
Natural killer T (NKT) cells	0.4–4%	10, 14–16
Mesenchymal stem cells (MSCs)	0.01–0.1%	109, 117
Myeloid-derived suppressor cells (MDSCs)	20–30%	97, 100

blood.^{8,10} Two-thirds of bone marrow T cells express surface markers indicative of antigen experience, such as CD44^{hi} and CD122⁺, whereas most T cells in spleen and peripheral lymph nodes exhibit naive phenotypes.⁸ In addition to T cells, there are 1%–2% CD11c⁺ dendritic cells^{6,7,13} and 0.4%–4% natural killer T (NKT) cells in bone marrow.^{14–16} Therefore, bone marrow contains substantial amount of immune cells. Altogether, the evidences suggest that bone marrow is a lymphoid organ which may play a key role in immunity.

CD4⁺ T cell

Bone marrow contains a high proportion of CD4⁺ T cells displaying a memory phenotype, which express high levels of CD44 in mice and low levels of CD45RA in humans (Figures 2 and 3).^{8,11} Similar to the CD8⁺ T cells, basal homeostatic proliferation and survival of CD4⁺ memory T cells are regulated by IL-7, identified as the dominant cytokine, and IL-15, an accessory cytokine.¹⁷ Although human CD4⁺ memory T cells proliferate *in vitro* in response to IL-7 and IL-15,¹⁸ studies in mice showed that acute homeostatic proliferation of 'memory-phenotype' CD4⁺ T cells is independent of IL-7 and IL-15.¹⁹ Both cytokines were also ruled out to participate in CD4⁺ memory T-cell survival, because CD4⁺ memory T cells deficient for CD132 (γ -chain, jointly used by IL-2, IL-4, IL-7, IL-9, IL-15, and IL-21 receptors) are effectively maintained *in vivo*, but other work demonstrated that IL-7 is actually required for survival of both memory phenotype and T-cell receptor transgenic CD4⁺ memory T cells.²⁰

Treg cell

Naturally occurring CD4⁺CD25⁺ Treg cells represent 5%–10% of the CD4⁺ T cells. However, our group has reported that 30% of CD4⁺ T cells are functional Treg cells in bone marrow.¹² In patients with prostate cancer bone marrow metastasis, bone marrow Treg cells are further increased (Zou *et al.*, unpubl. data). It suggests that bone marrow is a preferential site for migration, or selective retainment and function of Treg cells.^{12,21,22} Furthermore, we have demonstrated that CXCR4/CXCL12 (CXC chemokine ligand 12, stromal cell-derived factor-1) signaling mediates Treg cell trafficking to bone marrow (Figures 2 and 3).¹² Mouse Treg cells are known to reduce the severity of graft-versus-host disease (GVHD).^{6,23} Granulocyte colony-stimulating factor (G-CSF) decreases bone marrow CXCL12, and in turn mobilizes bone marrow Treg cells. These findings may help explain why G-CSF administration reduces the severity and mortality in acute GVHD.^{13,24}

Given that high levels of Treg cells are found in the bone marrow, bone marrow transplantation may result in increased numbers of Treg cells and in turn lead to a reduction of autoantibody production and protection from lethality caused by severe GVHD.²⁵ Additionally, antigen presentation by bone marrow dendritic cells can induce the expansion of CD4⁺CD25⁺ T cells while simultaneously activating their ability to suppress cytokine secretion by effector T cells. Therefore, agents that mobilize Treg cells from bone marrow would be therapeutically beneficial in some clinical settings.

It is well known that Treg cells are implicated in the pathogenesis of autoimmune diseases, tumors and organ transplantation.^{26–29} Our unpublished data indicate that Treg cells may regulate bone biology. For example, Treg cells can suppress osteoclastogenesis and ameliorate osteolytic bone resorption and destruction. The levels of bone marrow Treg cells are much higher in patients with prostate cancer. Patients with prostate cancer often have bone metastases with bone precipitation as a pathological characteristic. It is assumed that bone marrow Treg cells may contribute to bone pathology in patients with prostate

cancer. The studies in our laboratory will address this possibility. Nonetheless, bone marrow Treg cells might be a novel therapeutic strategy for clinical diseases and transplantation.^{30,31}

IL-17⁺CD4⁺ T (Th17) cell

IL-17 (originally termed CTLA8, also known as IL-17A) belongs to a family of six members and has been of great interest recently owing to the discovery that the production of IL-17 characterizes a subset of CD4⁺ helper T cells (Th17 cells).³² The development of Th17 cells is coupled to signal transducer and activator of transcription 3 and the transcription factor ROR γ t and depends on IL-6 and transforming growth factor (TGF)- β , which is shared by regulatory T cells for development and function.³³ Th17 cells are characterized by the production of a distinct profile of effector cytokines, including IL-17 (or IL-17A), IL-17F, IL-21 and IL-22.^{34–36} Th17 cells play an important role in inflammation, autoimmune disease and tumor.^{37–39}

Th17 cells are induced and elevated in multiple myeloma by the elevated cytokine production of IL-6, IL-1 β and TGF- β in bone marrow microenvironment.³³ IL-17 in turn promotes myeloma cell growth and suppresses immune function in multiple myeloma.³³ It is also reported that elevated Th17 cells mediate bone resorption and destruction by inducing osteoclast formation in multiple myeloma patients.⁴⁰ Bone marrow mesenchymal stem cells (MSCs) inhibit the differentiation and function of Th17 cells and ameliorate multiple sclerosis in an experimental autoimmune encephalomyelitis model.^{41,42} Administration of bone marrow stromal cells ameliorates experimental autoimmune myasthenia gravis by reducing Th17 cells.⁴³ Several groups have shown that Th17 cells contribute to GVHD after bone marrow transplantation.^{44,45} There is evidence showing increased Th17 cells in low risk myelodysplastic syndrome.⁴⁶ Thus, Th17 cells play an important role in bone marrow-mediated immunity and may serve as a therapeutic target for bone marrow-related diseases.

CD8⁺ T cell

Bone marrow is the preferred site for proliferation of memory CD8⁺ T cells.⁴⁷ Antigen specific memory CD8⁺ T cells receive proliferative signals by IL-7 and/or IL-15 in the bone marrow. It suggests that the bone marrow is a 'niche' for the antigen-independent proliferation of memory CD8⁺ T cells. Memory CD8⁺ T lymphocyte populations in bone marrow display a CD44-positive phenotype with a higher percentage of HLA-DR molecules, which suggests that CD8⁺ T cells in bone marrow are in an activated state. High numbers of tumor-associated antigen-specific CD8⁺ T cells were shown to persist in the bone marrow for several months after acute infection or tumor development.^{48,49} Adoptive transfer of bone marrow cells from lymphochoriomeningitis virus-immunized mice to immunodeficient recipients provides antiviral protection.⁹ Thus, memory CD8⁺ T cells in the bone marrow are able to mount an effective secondary response. A long time after priming, memory CD8⁺ T cells proliferate more extensively in the bone marrow than they do in either secondary lymphoid or extra-lymphoid organs and undergo basal proliferation in the bone marrow.^{47,50} Indeed, bone marrow-resident T cells are functionally distinct from those in other compartments.¹⁰ Compared to their blood counterparts, bone marrow-derived CD8⁺ T cells induce milder GVHD and possess higher anti-tumor activity.⁶

Bone marrow also functions as a site of recruitment and retention for central memory T cells by providing specific recruitment signals that mediate the recruitment of central memory T cells from the blood.⁸ Central memory T cells constitute the largest endogenous

subset of CD8⁺ T cells in murine bone marrow and are also prominent in human bone marrow. There are also naive CD8⁺ T cells and effector memory CD8⁺ T cells in bone marrow, but these populations are smaller (at least in mice) than naive T cells in secondary lymphoid organs or effector memory T cells in spleen, liver and lung.⁵¹ Accordingly, adoptively transferred central memory T cells from immunized mice accumulated and colonized in recipient bone marrow more effectively than effector T cells and naive T cells. Overall, the bone marrow may be an important organ for CD8⁺ T cell-mediated immunity.

NKT cell

NKT cells are defined as T cells bearing the common natural killer cell marker such as NK1.1 (NKR-P1C) in mice or CD161 (NKR-P1A) in humans, expressing IL-2R β (CD122), Ly49 family receptors and T-cell receptor.^{52,53} In normal adult mice, NKT cells are found in thymus, bone marrow, liver and spleen at a level of 0.5–1.5 million cells per organ. NKT cells are rare in lymph nodes and virtually absent from gut intraepithelial lymphocytes. Besides thymus and liver, NKT cells can be generated in bone marrow of nude mice.^{14,54} Reconstitution of adult thymectomized irradiated mice with syngeneic bone marrow cells gives rise to NKT cells in the recipient organs,^{55,56} suggesting that NKT cells can develop extrathymically from the bone marrow. Peripheral NKT cells are rapidly deleted upon activation and replaced by NKT cells that have been generated by *de novo* proliferation in bone marrow. Thus, bone marrow plays a major role in restoring NKT cell homeostasis.

Bone marrow NKT cells may play immune stimulatory and inhibitory roles in the regulation of bone marrow transplantation immunity^{15,57,58} and tumor immunity.^{59,60} Adoptive transfer of donor NKT cells significantly ameliorates GVHD in a murine model of bone marrow transplantation.⁵⁸ The relative protection against GVHD was contributed to, to some extent, the population of NKT cells in bone marrow.^{58,61} NKT cells are also believed to be among the most important anti-cancer cell populations in the mouse, causing rejection of malignant cells *in vivo*.^{59,60} Thus, bone marrow NKT cells could differentially regulate immune responses in different settings.

B cell

Bone marrow is major organ for the development and maturation of B cells. B cells are generated from HSCs and developed in bone marrow before they egress into peripheral blood to reach peripheral lymphoid organs. Specific cellular niches for B-cell development include CXCL12-expressing cells and IL-7-expressing cells.⁶² Immature B cells, like pro-B and pre-B stage cells, are regulated by extrinsic signals from the bone marrow during their development.⁶³ B-cell precursors and plasma cells reside in the specific niches and move between the niches as development proceeds.⁶²

The majority of serum antibody is produced by terminally differentiated plasma cells. These non-dividing cells differ from memory B cells in typical B-cell markers, including major histocompatibility (MHC) class II and surface immunoglobulin. The main function of plasma cells is to continuously secrete large quantities of specific antibody. In contrast, memory B cells do not spontaneously secrete antibody. These cells proliferate and differentiate into antibody-secreting cells following appropriate stimulation. Interestingly, some plasma cells are long-lived. Most importantly, the long-lived plasma cells are found in the bone marrow. The bone marrow is a reservoir for long-lived plasma cells and is involved in the maintenance of long immunity.^{64,65} Antigen-specific bone marrow plasma cells have been

detected for more than 300 days post-viral infection.⁶⁴ The persistence of plasma cells within the bone marrow might be supported by soluble factors and/or cell–cell contact in the bone marrow microenvironment, in which one critical element is the bone marrow reticular stromal cells. Stromal cells provide growth factors as well as cell contact-dependent signals, such as IL-6 and cell contact-mediated signals (e.g., very late antigen-4) (Figure 2).⁶⁶ In addition to stromal elements, plasma cells could interact with various other bone marrow resident cells, including developing lymphoid and myeloid lineage cells. Therefore, bone marrow plasma cells are not intrinsically long-lived. Bone marrow stromal cells provide survival factors to them. The plasmablasts can migrate to the bone marrow^{67–69} and differentiate into memory plasma cells after docking with CXCL12-expressing stromal cells.^{62,69} The overall impact of these interactions supports the longevity of the plasma cells in bone marrow. Therefore, bone marrow contributes to humoral immune responses.

Neutrophil

Neutrophils are an essential component of innate immune system and may represent a critical link between the innate and adaptive immune system.⁷⁰ They are differentiated from stem cells in the bone marrow by a process termed granulopoiesis.⁷¹ Neutrophils are generated at a rate of 1 to 2 $\times 10^{11}$ cells per day in a normal adult human under normal condition.⁷² Several myeloid transcription factors are essential for granulopoiesis, including CCAAT-enhanced binding protein α PU.1 and growth factor independent-1 (GFI-1).⁷³ PU.1 is an ETS family transcription factor encoded by spleen focus forming virus proviral integration oncogene in human⁷⁴ and absolutely required for myeloid lineage commitment.^{75,76} The balance between PU.1 and CCAAT-enhanced binding protein α determines the commitment of granulocytes and monocytes.^{77–79} High expression of PU.1 drives monocytic differentiation and CCAAT-enhanced binding protein α promotes granulocytogenesis.^{72,80} GFI-1 is also necessary for neutrophil differentiation and it is upregulated during granulocytic lineage commitment.^{81,82} G-CSF is critical in regulating granulopoiesis at several stages. G-CSF directs the commitment of multipotent progenitor cells down to the myeloid lineage, stimulates proliferation of granulocytic precursors and reduces the transit time of neutrophils through granulocytic compartment.^{83,84}

Bone marrow is a large pool for the mature neutrophils.⁸⁵ There are 1%–2% of mature neutrophils in the circulation in mice.⁸⁶ The majority of neutrophils are reserved in the bone marrow. A large amount of neutrophils can be mobilized rapidly in response to infection and stress, which suggests that the bone marrow reserve is critical for host defense. CXCR4/CXCL12 signaling pathway plays a crucial role for maintaining neutrophils in bone marrow (Figure 3).^{87,88} It is reported that administration of G-CSF reduces the expression of CXCR4 on bone marrow neutrophils and Treg cells, and the levels of bone marrow CXCL12.¹² This explains why G-CSF administration mobilizes neutrophils from bone marrow to peripheral circulation.^{89,90}

Once released from the bone marrow, neutrophils circulate in the peripheral blood and have a relatively short half-life (about 6–8 h).^{71,91} Bone marrow serves as an important site for neutrophil clearance under homeostatic conditions. Senescent neutrophils home back to bone marrow depending on G α i subunit of the heterotrimeric G-protein.⁹² Senescent neutrophils highly express CXCR4⁹³ and may home back to bone marrow *via* the CXCR4/CXCL12 chemokine axis.⁸⁸ Once senescent neutrophils return to bone marrow, they are phagocytosed and destroyed by resident stromal macrophages in bone marrow, which in turn stimulates the production of G-CSF by bone

marrow macrophages after uptake of apoptotic neutrophils and subsequently G-CSF acts as a positive feedback to promote granulopoiesis and regulate neutrophils release.^{91,92} Thus, bone marrow plays an important role in the homeostasis of neutrophils.

Dendritic cell

Dendritic cells (DCs) play a key role in linking innate and adaptive immune responses.^{13,94} Circulating DCs migrate to the bone marrow where they are retained better than in spleen, liver and lung tissues.⁹⁵ Homing of DCs to the bone marrow depends on constitutively expressed VCAM-1 and endothelial selectins in bone marrow microvessels (Figure 3). The migration of DCs to non-lymphoid organs might be advantageous for 'boosting' memory responses to previously antigen-experienced T cells.⁹⁶ Bone marrow DCs are able to trigger central memory T cells-mediated responses with antigen-dependent contacts. Bone marrow-derived dendritic cells are distinct from counterpart DCs in extralymphoid tissue, and are essential for innate immunity to intracellular infection. Bone marrow-derived dendritic cells are able to uptake the blood-derived cell-associated tumor-associated antigen, process them and induce antigen-specific systemic protective T cells-mediated immunity.⁵ Thus, bone marrow DCs are functionally important in adaptive immunity.

Myeloid-derived cell

Myeloid-derived cells are a heterogeneous population of cells consisting of myeloid progenitors, immature myeloid cells and macrophages.⁹⁷ Immature myeloid cells that generated in bone marrow differentiate into mature myeloid cells under normal condition. But under pathological conditions (e.g., tumor), immature myeloid cells are expanded and activated, and become myeloid-derived suppressor cells (MDSCs), and acquire immunosuppressive activity by producing immune suppressive factors such as arginase I or inducible nitric oxidase synthase or TGF- β .^{97,98} MDSCs express Gr-1 and CD11b, the surface marker for myeloid cell lineage in mice.⁹⁹ In healthy mice, 20%–30% of whole bone marrow cells express Gr-1⁺CD11b⁺ phenotype.¹⁰⁰ Based on the two different epitopes of Gr-1 antibodies, Ly6G and Ly6C, mouse MDSCs are subdivided into granulocytic MDSCs (CD11b⁺Ly6G⁺Ly6C^{low}) and monocytic MDSCs (CD11b⁺Ly6G⁺Ly6C^{hi}).^{101,102} MDSCs may be defined as cells with CD14⁻CD11b⁺HLA-DR^{dim} phenotype in human.^{103,104} MDSCs are shown to have the ability to induce the development and differentiation of Treg cells through cytokine production or cell–cell interaction,^{105,106} which indicates that MDSCs and Treg cells might cooperate to regulate immune response.⁹⁷ Adoptive transfer of MDSCs inhibits T-cell alloresponses, prevents GVHD and prolongs the survival of mice.^{107,108} MDSC–T cell interactions play an important role in protective anti-tumor immunity, infection, inflammation and GVHD.¹⁰⁷ Bone marrow MDSCs may be an important regulator of immune response and may serve as a potential therapeutic target for several clinical diseases.

MSC

In adult life, stem cells of mesenchymal lineage (MSCs), which comprise 0.01%–0.1% of total adult bone marrow cells, are mainly confined to the bone marrow,¹⁰⁹ where they are multipotential non-hematopoietic progenitor cells capable of differentiating into various tissues of mesenchymal origin. Human MSCs express human leukocyte antigen (HLA) class I but not HLA class II¹¹⁰ or costimulatory molecules CD80 (B7.1), CD86 (B7.2) or CD40.¹¹¹ MSCs produce cytokines and growth factors for hematopoiesis and may attract

infused HSCs to the bone marrow by inducing expression of homing receptors.^{112,113}

In the bone marrow, MSCs display a great potential role in immunoregulatory activity, which comes from the observation that MSCs from various species can exert profound immunosuppression by inhibiting T-cell proliferation in responses to polyclonal stimuli and to their cognate peptide in a MHC-independent manner.^{24,114–116} Although the conflicting results about MSC-mediated immunosuppression have been produced with different methods and species, overall data suggest that some soluble factors and mechanisms of cell–cell interaction might contribute to the inhibition of MSCs.^{24,115,116} It is reported that MSCs could inhibit naive and memory T-cell responses to their cognate antigens, but it does not appear to be antigen-specific.²⁴ MSCs also induce T-cell anergy¹¹⁷ or T-cell apoptosis,¹¹⁸ suppress T-cell IFN- γ production and increase IL-4 secretion, and enhance Treg cell compartment.¹¹⁹ MSCs selectively targets antigen-experienced T-cell responses in the contact with antigen-presenting cells in a non-cognate fashion, sparing those that have not been activated by T-cell receptor engagement.¹¹⁶ Moreover, MSCs appear to discriminate between cellular responses to alloantigens and recall antigens.¹²⁰ The expression of early activation markers such as CD25 and CD69 on T cell is unaffected in the presence of MSCs, but IFN- γ production is reduced. The inhibitory effect of MSCs is directed mainly at the level of cell proliferation without interfering with early T-cell activation. MSCs do not preferentially target any T-cell subset. MSCs have a role in inhibiting proliferation and affecting differentiation, antibody production and chemotactic behavior of B cells.^{117,121} MSCs may inhibit differentiation of hematopoietic progenitors into DCs and promote anti-inflammatory cytokine production of monocyte-derived DCs.^{122,123} MSCs suppress cytokine-induced proliferation and prevent cytokine production and cytotoxic activity of natural killer cells.^{124,125} In the context of limited understanding about MHC expression on MSCs, it is difficult to ascribe specific T-cell receptor/MHC/peptide interactions to their mechanism of immunoregulation. Notably, the inhibitory effect of MSCs does not require the presence of antigen-presenting cells and is not mediated through Treg cells.¹¹⁶ MSCs have been reported to be used in clinical trials for autologous or allogeneic engraftment of bone marrow transplants,¹²⁶ osteogenesis imperfect,¹²⁷ stroke,¹²⁸ myocardial infarction¹²⁹ and GVHD.¹³⁰ Therefore, bone marrow MSCs have an important role in immune regulation and are potential candidates for clinical treatment.

Cytokines and chemokines

Bone marrow-derived cells including leukocytes, HSCs and stromal cells could secrete lots of cytokines. Stroma cells and cells of hemopoietic lineage in the bone marrow produce both IL-7 and IL-15. IL-7 is a 'stromal cytokine' produced by a variety of stromal tissues including those in bone marrow. The production of IL-7 by bone marrow stromal cells is thought to be essential for early B lymphocyte development in mouse (but not in human),^{131–133} and secreted IL-7 in bone marrow is postulated to play a critical role in post-thymic T-cell homeostasis.^{54,134} IL-15 promotes basal homeostatic proliferation and survival of memory T cells in different experimental systems, whereas IL-7 performs an overlapping and complementary role during acute homeostatic proliferation in lymphopenic environments.^{135,136} Recently, exogenous IL-15 was shown to replace exogenous IL-2 therapy during the treatment of established, non-manipulated poorly immunogenic tumors.¹³⁷ IL-21 plays a role in the proliferation and maturation of natural killer cell populations from bone marrow, and

contributes to the proliferation of T/B cell populations and anti-tumor effect.^{138,139}

Chemokines could also be involved in the generation and regulation of bone marrow immune cells. Mobilization and interactions of stromal cell/hematopoietic precursors are thought to be controlled by cytokines, particularly, chemokines. Among chemokines, CXCL12 is particularly intriguing. CXCL12-expressing cells are a small population of bone marrow stromal cells scattered throughout bone marrow, such as osteoblasts, marrow fibroblasts and endothelial cells.¹⁴⁰ CXCL12-mediated interaction of progenitors with the bone marrow vascular niche allows the progenitors to relocate to a microenvironment that is permissive and instructive for megakaryocyte maturation and thrombopoiesis.²³ It has been recently determined that chemokine stimulation of HSCs and bone marrow-derived endothelial cells by CXCL12 leads to an enhancement in transendothelial and stromal migration *via* activation of adhesion molecules, in addition to its well-known ability to stimulate motility.^{141,142} Signals for the translocation of HSCs from the fetal liver to bone marrow are provided by CXCL12. In response to CXCL12, HSCs or lymphocytes that express its specific seven transmembrane-span G protein-coupled CXCR4 receptor leave the fetal liver and colonize in the bone marrow, where they finally establish hematopoiesis.^{143–145} As we discussed above, CXCR4/CXCL12 signaling also regulates the bone marrow trafficking of memory T cells, Treg cells and neutrophils.^{12,67,68,87,88,93}

In addition to the role of chemokines in bone marrow, the adhesion molecules also regulate leukocytes migration to the bone marrow (Figure 3). Normal bone marrow sinusoids express VCAM-1 and E-selectin. The migration of B cells, CD4⁺ and CD8⁺ T cells to bone marrow is impaired in conditional VCAM-1-deficient mice, resulting in reduced B cells and T cells in bone marrow and mild leukocytosis in peripheral blood.¹⁴⁶ E-selectin and VCAM-1 are necessary for recruitment of hematopoietic progenitor cells to bone marrow.¹⁴⁷ Neutralizing antibody to E-selectin attenuates CD8⁺ central memory T-cell rolling in bone marrow.⁸ Thus, bone marrow plays an important role in immune cells homeostasis *via* the expression of cytokines, chemokines and adhesion molecules.

CONCLUDING REMARKS

Bone marrow is well-known as a primary hematopoietic organ. However, bone marrow contains high levels of multiple immune cell subsets with important and unique functionalities. It is evident that bone marrow can supplant the secondary lymphoid tissue either as a site of primary immune response or memory response. Immune regulation occurs in the bone marrow microenvironment in cell–cell contact manners or/and through soluble factors including cytokines. Thus, bone marrow is an immune regulatory organ, which may importantly affect systemic immunity and therapeutic efficacy of conventional and immune therapy/vaccination. Given that multiple human cancers including breast and prostate cancer preferentially metastasize to bone marrow, specific cellular and molecular niches in the bone marrow including high levels of Treg cells and MDSCs may impact tumor bone metastasis and contribute to bone pathology in cancer patients with bone marrow metastasis. Therefore, understanding the immune regulatory mechanisms in the bone marrow microenvironment will generate significant insight into human bone biology and immunology. Furthermore, given the unique functionalities of bone marrow memory T cells, one may expect that bone marrow serves as an excellent source of immune cells for adoptive immunotherapy for both malignancies and infectious diseases.

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- Kopp HG, Avecilla ST, Hooper AT, Rafii S. The bone marrow vascular niche: home of HSC differentiation and mobilization. *Physiology (Bethesda)* 2005; **20**: 349–356.
- Rafii S, Lyden D. Therapeutic stem and progenitor cell transplantation for organ vascularization and regeneration. *Nat Med* 2003; **9**: 702–712.
- Tripp RA, Topham DJ, Watson SR, Doherty PC. Bone marrow can function as a lymphoid organ during a primary immune response under conditions of disrupted lymphocyte trafficking. *J Immunol* 1997; **158**: 3716–3720.
- Dejbakhsh-Jones S, Jerabek L, Weissman IL, Strober S. Extrathymic maturation of alpha beta T cells from hemopoietic stem cells. *J Immunol* 1995; **155**: 3338–3344.
- Schirmacher V, Feuerer M, Fournier P, Ahlert T, Umansky V, Beckhove P. T-cell priming in bone marrow: the potential for long-lasting protective anti-tumor immunity. *Trends Mol Med* 2003; **9**: 526–534.
- Feuerer M, Beckhove P, Mahnke Y, Hommel M, Kyewski B, Hamann A *et al*. Bone marrow microenvironment facilitating dendritic cell: CD4 T cell interactions and maintenance of CD4 memory. *Int J Oncol* 2004; **25**: 867–876.
- Feuerer M, Beckhove P, Garbi N, Mahnke Y, Limmer A, Hommel M *et al*. Bone marrow as a priming site for T-cell responses to blood-borne antigen. *Nat Med* 2003; **9**: 1151–1157.
- Mazo IB, Honczarenko M, Leung H, Cavanagh LL, Bonasio R, Weninger W *et al*. Bone marrow is a major reservoir and site of recruitment for central memory CD8⁺ T cells. *Immunity* 2005; **22**: 259–270.
- Slifka MK, Whitmire JK, Ahmed R. Bone marrow contains virus-specific cytotoxic T lymphocytes. *Blood* 1997; **90**: 2103–2108.
- Zeng D, Hoffmann P, Lan F, Huie P, Higgins J, Strober S. Unique patterns of surface receptors, cytokine secretion, and immune functions distinguish T cells in the bone marrow from those in the periphery: impact on allogeneic bone marrow transplantation. *Blood* 2002; **99**: 1449–1457.
- Price PW, Cerny J. Characterization of CD4⁺ T cells in mouse bone marrow. I. Increased activated/memory phenotype and altered TCR Vbeta repertoire. *Eur J Immunol* 1999; **29**: 1051–1056.
- Zou L, Barnett B, Safah H, Larussa VF, Evdemon-Hogan M, Mottram P *et al*. Bone marrow is a reservoir for CD4⁺CD25⁺ regulatory T cells that traffic through CXCL12/CXCR4 signals. *Cancer Res* 2004; **64**: 8451–8455.
- Banchereau J, Briere F, Caux C, Davoust J, Lebecque S, Liu YJ *et al*. Immunobiology of dendritic cells. *Annu Rev Immunol* 2000; **18**: 767–811.
- Sykes M. Unusual T cell populations in adult murine bone marrow. Prevalence of CD3⁺CD4⁺CD8⁺ and alpha beta TCR⁺NK1.1⁺ cells. *J Immunol* 1990; **145**: 3209–3215.
- Higuchi M, Zeng D, Shizuru J, Gworek J, Dejbakhsh-Jones S, Taniguchi M *et al*. Immune tolerance to combined organ and bone marrow transplants after fractionated lymphoid irradiation involves regulatory NK T cells and clonal deletion. *J Immunol* 2002; **169**: 5564–5570.
- Zeng D, Gazit G, Dejbakhsh-Jones S, Balk SP, Snapper S, Taniguchi M *et al*. Heterogeneity of NK1.1⁺ T cells in the bone marrow: divergence from the thymus. *J Immunol* 1999; **163**: 5338–5345.
- Fry TJ, Mackall CL. The many faces of IL-7: from lymphopoiesis to peripheral T cell maintenance. *J Immunol* 2005; **174**: 6571–6576.
- Geginat J, Sallusto F, Lanzavecchia A. Cytokine-driven proliferation and differentiation of human naive, central memory, and effector memory CD4⁺ T cells. *J Exp Med* 2001; **194**: 1711–1719.
- Tan JT, Ernst B, Kieper WC, LeRoy E, Sprent J, Surh CD. Interleukin (IL)-15 and IL-7 jointly regulate homeostatic proliferation of memory phenotype CD8⁺ cells but are not required for memory phenotype CD4⁺ cells. *J Exp Med* 2002; **195**: 1523–1532.
- Lantz O, Grandjean I, Matzinger P, Di Santo JP. Gamma chain required for naive CD4⁺ T cell survival but not for antigen proliferation. *Nat Immunol* 2000; **1**: 54–58.
- Wei S, Kryczek I, Edwards RP, Zou L, Szeliga W, Banerjee M *et al*. Interleukin-2 administration alters the CD4⁺FOXP3⁺ T-cell pool and tumor trafficking in patients with ovarian carcinoma. *Cancer Res* 2007; **67**: 7487–7494.
- Wei S, Kryczek I, Zou W. Regulatory T-cell compartmentalization and trafficking. *Blood* 2006; **108**: 426–431.
- Avecilla ST, Hattori K, Heissig B, Tejada R, Liao F, Shido K *et al*. Chemokine-mediated interaction of hematopoietic progenitors with the bone marrow vascular niche is required for thrombopoiesis. *Nat Med* 2004; **10**: 64–71.
- Bartholomew A, Sturgeon C, Siatskas M, Ferrer K, McIntosh K, Patil S *et al*. Mesenchymal stem cells suppress lymphocyte proliferation in vitro and prolong skin graft survival in vivo. *Exp Hematol* 2002; **30**: 42–48.
- Herrmann MM, Gaertner S, Stadelmann C, van den Brandt J, Bosche R, Budach W *et al*. Tolerance induction by bone marrow transplantation in a multiple sclerosis model. *Blood* 2005; **106**: 1875–1883.
- Kryczek I, Liu R, Wang G, Wu K, Shu X, Szeliga W *et al*. FOXP3 defines regulatory T cells in human tumor and autoimmune disease. *Cancer Res* 2009; **69**: 3995–4000.
- Wilke CM, Wu K, Zhao E, Wang G, Zou W. Prognostic significance of regulatory T cells in tumor. *Int J Cancer* 2010; **127**: 748–758.

- 28 Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P *et al*. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 2004; **10**: 942–949.
- 29 Issa F, Wood KJ. CD4⁺ regulatory T cells in solid organ transplantation. *Curr Opin Organ Transplant* 2010; **15**: 757–764.
- 30 Zou W. Immunosuppressive networks in the tumour environment and their therapeutic relevance. *Nat Rev Cancer* 2005; **5**: 263–274.
- 31 Zou W. Regulatory T cells, tumour immunity and immunotherapy. *Nat Rev Immunol* 2006; **6**: 295–307.
- 32 Zou W, Restifo NP. T(H)17 cells in tumour immunity and immunotherapy. *Nat Rev Immunol* 2011; **10**: 248–256.
- 33 Prabhala RH, Pelluru D, Fulciniti M, Prabhala HK, Nanjappa P, Song W *et al*. Elevated IL-17 produced by Th17 cells promotes myeloma cell growth and inhibits immune function in multiple myeloma. *Blood* 2010; **115**: 5385–5392.
- 34 Kryczek I, Banerjee M, Cheng P, Vatan L, Szeliga W, Wei S *et al*. Phenotype, distribution, generation, and functional and clinical relevance of Th17 cells in the human tumor environments. *Blood* 2009; **114**: 1141–1149.
- 35 Kryczek I, Wei S, Vatan L, Escara-Wilke J, Szeliga W, Keller ET *et al*. Cutting edge: opposite effects of IL-1 and IL-2 on the regulation of IL-17⁺ T cell pool IL-1 subverts IL-2-mediated suppression. *J Immunol* 2007; **179**: 1423–1426.
- 36 Liang SC, Tan XY, Luxenberg DP, Karim R, Dunussi-Joannopoulos K, Collins M *et al*. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. *J Exp Med* 2006; **203**: 2271–2279.
- 37 Kryczek I, Wei S, Szeliga W, Vatan L, Zou W. Endogenous IL-17 contributes to reduced tumor growth and metastasis. *Blood* 2009; **114**: 357–359.
- 38 Kryczek I, Bruce AT, Gudjonsson JE, Johnston A, Aphale A, Vatan L *et al*. Induction of IL-17⁺ T cell trafficking and development by IFN- γ : mechanism and pathological relevance in psoriasis. *J Immunol* 2008; **181**: 4733–4741.
- 39 Kryczek I, Wei S, Zou L, Altuwajri S, Szeliga W, Kolls J *et al*. Cutting edge: Th17 and regulatory T cell dynamics and the regulation by IL-2 in the tumor microenvironment. *J Immunol* 2007; **178**: 6730–6733.
- 40 Noonan K, Marchionni L, Anderson J, Pardoll D, Roodman GD, Borrello I. A novel role of IL-17-producing lymphocytes in mediating lytic bone disease in multiple myeloma. *Blood* 2010; **116**: 3554–3563.
- 41 Ghannam S, Pene J, Torcy-Moquet G, Jorgensen C, Yssel H. Mesenchymal stem cells inhibit human Th17 cell differentiation and function and induce a T regulatory cell phenotype. *J Immunol* 2010; **185**: 302–312.
- 42 Bai L, Lennon DP, Eaton V, Maier K, Caplan AI, Miller SD *et al*. Human bone marrow-derived mesenchymal stem cells induce Th2-polarized immune response and promote endogenous repair in animal models of multiple sclerosis. *Glia* 2009; **57**: 1192–1203.
- 43 Kong QF, Sun B, Wang GY, Zhai DX, Mu LL, Wang DD *et al*. BM stromal cells ameliorate experimental autoimmune myasthenia gravis by altering the balance of Th cells through the secretion of IDO. *Eur J Immunol* 2009; **39**: 800–809.
- 44 Kappel LW, Goldberg GL, King CG, Suh DY, Smith OM, Ligh C *et al*. IL-17 contributes to CD4-mediated graft-versus-host disease. *Blood* 2009; **113**: 945–52.
- 45 Chen X, Vodanovic-Jankovic S, Johnson B, Keller M, Komorowski R, Drobyski WR. Absence of regulatory T-cell control of TH1 and TH17 cells is responsible for the autoimmune-mediated pathology in chronic graft-versus-host disease. *Blood* 2007; **110**: 3804–3813.
- 46 Kordasti SY, Afzali B, Lim Z, Ingram W, Hayden J, Barber L *et al*. IL-17-producing CD4⁺ T cells, pro-inflammatory cytokines and apoptosis are increased in low risk myelodysplastic syndrome. *Br J Haematol* 2009; **145**: 64–72.
- 47 Becker TC, Coley SM, Wherry EJ, Ahmed R. Bone marrow is a preferred site for homeostatic proliferation of memory CD8 T cells. *J Immunol* 2005; **174**: 1269–1273.
- 48 Marshall DR, Turner SJ, Belz GT, Wingo S, Andreansky S, Sangster MY *et al*. Measuring the diaspora for virus-specific CD8⁺ T cells. *Proc Natl Acad Sci USA* 2001; **98**: 6313–6318.
- 49 Masopust D, Vezy V, Marzo AL, Lefrancois L. Preferential localization of effector memory cells in nonlymphoid tissue. *Science* 2001; **291**: 2413–2417.
- 50 Parretta E, Cassese G, Barba P, Santoni A, Guardiola J, Di Rosa F. CD8 cell division maintaining cytotoxic memory occurs predominantly in the bone marrow. *J Immunol* 2005; **174**: 7654–7664.
- 51 Weninger W, Crowley MA, Manjunath N, von Andrian UH. Migratory properties of naive, effector, and memory CD8⁺ T cells. *J Exp Med* 2001; **194**: 953–966.
- 52 Kronenberg M. Toward an understanding of NKT cell biology: progress and paradoxes. *Annu Rev Immunol* 2005; **23**: 877–900.
- 53 MacDonald HR. NK1.1⁺ T cell receptor- α/β ⁺ cells: new clues to their origin, specificity, and function. *J Exp Med* 1995; **182**: 633–638.
- 54 Tan JT, Dudl E, LeRoy E, Murray R, Sprent J, Weinberg KI *et al*. IL-7 is critical for homeostatic proliferation and survival of naive T cells. *Proc Natl Acad Sci USA* 2001; **98**: 8732–8737.
- 55 Kikly K, Dennert G. Evidence for extrathymic development of TNK cells. NK1⁺ CD3⁺ cells responsible for acute marrow graft rejection are present in thymus-deficient mice. *J Immunol* 1992; **149**: 403–412.
- 56 Sato K, Ohtsuka K, Hasegawa K, Yamagawa S, Watanabe H, Asakura H *et al*. Evidence for extrathymic generation of intermediate T cell receptor cells in the liver revealed in thymectomized, irradiated mice subjected to bone marrow transplantation. *J Exp Med* 1995; **182**: 759–767.
- 57 Haraguchi K, Takahashi T, Hiruma K, Kanda Y, Tanaka Y, Ogawa S *et al*. Recovery of V α 24⁺ NKT cells after hematopoietic stem cell transplantation. *Bone Marrow Transplant* 2004; **34**: 595–602.
- 58 Margalit M, Ilan Y, Ohana M, Safadi R, Alper R, Sherman Y *et al*. Adoptive transfer of small numbers of DX5⁺ cells alleviates graft-versus-host disease in a murine model of semi-allogeneic bone marrow transplantation: a potential role for NKT lymphocytes. *Bone Marrow Transplant* 2005; **35**: 191–197.
- 59 Smyth MJ, Crowe NY, Hayakawa Y, Takeda K, Yagita H, Godfrey DI. NKT cells - conductors of tumor immunity? *Curr Opin Immunol* 2002; **14**: 165–171.
- 60 Dean J, McCarthy D, Lawler M, Doherty DG, O'Farrelly C, Golden-Mason L. Characterization of NKR⁺ T-cell subsets in human bone marrow: implications for immunosurveillance of neoplasia. *Clin Immunol* 2005; **114**: 42–51.
- 61 Zeng D, Lewis D, Dejbakhsh-Jones S, Lan F, Garcia-Ojeda M, Sibley R *et al*. Bone marrow NK1.1⁺ and NK1.1⁺ T cells reciprocally regulate acute graft versus host disease. *J Exp Med* 1999; **189**: 1073–1081.
- 62 Tokoyoda K, Egawa T, Sugiyama T, Choi BI, Nagasawa T. Cellular niches controlling B lymphocyte behavior within bone marrow during development. *Immunity* 2004; **20**: 707–718.
- 63 Sandel PC, Gendelman M, Kelsoe G, Monroe JG. Definition of a novel cellular constituent of the bone marrow that regulates the response of immature B cells to B cell antigen receptor engagement. *J Immunol* 2001; **166**: 5935–5944.
- 64 Slifka MK, Antia R, Whitmire JK, Ahmed R. Humoral immunity due to long-lived plasma cells. *Immunity* 1998; **8**: 363–372.
- 65 Manz RA, Thiel A, Radbruch A. Lifetime of plasma cells in the bone marrow. *Nature* 1997; **388**: 133–134.
- 66 Minges Wols HA, Underhill GH, Kansas GS, Witte PL. The role of bone marrow-derived stromal cells in the maintenance of plasma cell longevity. *J Immunol* 2002; **169**: 4213–4221.
- 67 Hargreaves DC, Hyman PL, Lu TT, Ngo VN, Bidgol A, Suzuki G *et al*. A coordinated change in chemokine responsiveness guides plasma cell movements. *J Exp Med* 2001; **194**: 45–56.
- 68 Hauser AE, Debes GF, Arce S, Cassese G, Hamann A, Radbruch A *et al*. Chemotactic responsiveness toward ligands for CXCR3 and CXCR4 is regulated on plasma blasts during the time course of a memory immune response. *J Immunol* 2002; **169**: 1277–1282.
- 69 Tokoyoda K, Hauser AE, Nakayama T, Radbruch A. Organization of immunological memory by bone marrow stroma. *Nat Rev Immunol* 2010; **10**: 193–200.
- 70 Nathan C. Neutrophils and immunity: challenges and opportunities. *Nat Rev Immunol* 2006; **6**: 173–182.
- 71 Christopher MJ, Link DC. Regulation of neutrophil homeostasis. *Curr Opin Hematol* 2007; **14**: 3–8.
- 72 Borregaard N. Neutrophils, from marrow to microbes. *Immunity* 2010; **33**: 657–670.
- 73 Rosmarin AG, Yang Z, Resendes KK. Transcriptional regulation in myelopoiesis: hematopoietic fate choice, myeloid differentiation, and leukemogenesis. *Exp Hematol* 2005; **33**: 131–143.
- 74 Ray D, Culine S, Tavittain A, Moreau-Gachelin F. The human homologue of the putative proto-oncogene Spi-1: characterization and expression in tumors. *Oncogene* 1990; **5**: 663–668.
- 75 Nerlov C, Graf T. PU.1 induces myeloid lineage commitment in multipotent hematopoietic progenitors. *Genes Dev* 1998; **12**: 2403–2412.
- 76 Iwasaki H, Somoza C, Shigematsu H, Duprez EA, Iwasaki-Arai J, Mizuno S *et al*. Distinctive and indispensable roles of PU.1 in maintenance of hematopoietic stem cells and their differentiation. *Blood* 2005; **106**: 1590–1600.
- 77 Reddy VA, Iwama A, Iotzova G, Schulz M, Elsasser A, Vangala RK *et al*. Granulocyte inducer C/EBP α inactivates the myeloid master regulator PU.1: possible role in lineage commitment decisions. *Blood* 2002; **100**: 483–490.
- 78 Dahl R, Walsh JC, Lancki D, Laslo P, Iyer SR, Singh H *et al*. Regulation of macrophage and neutrophil cell fates by the PU.1:C/EBP α ratio and granulocyte colony-stimulating factor. *Nat Immunol* 2003; **4**: 1029–1036.
- 79 Laslo P, Spooner CJ, Warmflash A, Lancki DW, Lee HJ, Sciammas R *et al*. Multilineage transcriptional priming and determination of alternate hematopoietic cell fates. *Cell* 2001; **126**: 755–766.
- 80 Zhang DE, Zhang P, Wang ND, Hetherington CJ, Darlington GJ, Tenen DG. Absence of granulocyte colony-stimulating factor signaling and neutrophil development in CCAAT enhancer binding protein α -deficient mice. *Proc Natl Acad Sci USA* 1997; **94**: 569–574.
- 81 Karsunky H, Zeng H, Schmidt T, Zevnik B, Kluge R, Schmid KW *et al*. Inflammatory reactions and severe neutropenia in mice lacking the transcriptional repressor Gfi1. *Nat Genet* 2002; **30**: 295–300.
- 82 Hock H, Hamblen MJ, Rooke HM, Traver D, Bronson RT, Cameron S *et al*. Intrinsic requirement for zinc finger transcription factor Gfi-1 in neutrophil differentiation. *Immunity* 2003; **18**: 109–120.
- 83 Lord BI, Bronchud MH, Owens S, Chang J, Howell A, Souza L *et al*. The kinetics of human granulopoiesis following treatment with granulocyte colony-stimulating factor in vivo. *Proc Natl Acad Sci USA* 1989; **86**: 9499–9503.
- 84 Richards MK, Liu F, Iwasaki H, Akashi K, Link DC. Pivotal role of granulocyte colony-stimulating factor in the development of progenitors in the common myeloid pathway. *Blood* 2003; **102**: 3562–3568.
- 85 Cartwright GE, Athens JW, Wintrobe MM. The kinetics of granulopoiesis in normal man. *Blood* 1964; **24**: 780–803.
- 86 Semerad CL, Liu F, Gregory AD, Stumpf K, Link DC. G-CSF is an essential regulator of neutrophil trafficking from the bone marrow to the blood. *Immunity* 2002; **17**: 413–423.
- 87 Eash KJ, Means JM, White DW, Link DC. CXCR4 is a key regulator of neutrophil release from the bone marrow under basal and stress granulopoiesis conditions. *Blood* 2009; **113**: 4711–4719.

- 88 Martin C, Burdon PC, Bridger G, Gutierrez-Ramos JC, Williams TJ, Rankin SM. Chemokines acting via CXCR2 and CXCR4 control the release of neutrophils from the bone marrow and their return following senescence. *Immunity* 2003; **19**: 583–593.
- 89 Semerad CL, Christopher MJ, Liu F, Short B, Simmons PJ, Winkler I *et al*. G-CSF potently inhibits osteoblast activity and CXCL12 mRNA expression in the bone marrow. *Blood* 2005; **106**: 3020–3027.
- 90 Christopher MJ, Liu F, Hilton MJ, Long F, Link DC. Suppression of CXCL12 production by bone marrow osteoblasts is a common and critical pathway for cytokine-induced mobilization. *Blood* 2009; **114**: 1331–1339.
- 91 Rankin SM. The bone marrow: a site of neutrophil clearance. *J Leukoc Biol* 2010; **88**: 241–251.
- 92 Furze RC, Rankin SM. The role of the bone marrow in neutrophil clearance under homeostatic conditions in the mouse. *FASEB J* 2008; **22**: 3111–3119.
- 93 Nagase H, Miyamasu M, Yamaguchi M, Imanishi M, Tsuno NH, Matsushima K *et al*. Cytokine-mediated regulation of CXCR4 expression in human neutrophils. *J Leukoc Biol* 2002; **71**: 711–717.
- 94 Guernonprez P, Valladeau J, Zitvogel L, Thery C, Amigorena S. Antigen presentation and T cell stimulation by dendritic cells. *Annu Rev Immunol* 2002; **20**: 621–667.
- 95 Cavanagh LL, Bonasio R, Mazo JB, Halin C, Cheng G, van der Velden AW *et al*. Activation of bone marrow-resident memory T cells by circulating, antigen-bearing dendritic cells. *Nat Immunol* 2005; **6**: 1029–1037.
- 96 Sallusto F, Geginat J, Lanzavecchia A. Central memory and effector memory T cell subsets: function, generation, and maintenance. *Annu Rev Immunol* 2004; **22**: 745–763.
- 97 Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol* 2009; **9**: 162–174.
- 98 Gabrilovich D. Mechanisms and functional significance of tumour-induced dendritic-cell defects. *Nat Rev Immunol* 2004; **4**: 941–952.
- 99 Kusmartsev S, Nefedova Y, Yoder D, Gabrilovich DI. Antigen-specific inhibition of CD8⁺ T cell response by immature myeloid cells in cancer is mediated by reactive oxygen species. *J Immunol* 2004; **172**: 989–999.
- 100 Kusmartsev S, Gabrilovich DI. Role of immature myeloid cells in mechanisms of immune evasion in cancer. *Cancer Immunol Immunother* 2006; **55**: 237–245.
- 101 Movahedi K, Guillemins M, van den Bossche J, van den Bergh R, Gysemans C, Beschin A *et al*. Identification of discrete tumor-induced myeloid-derived suppressor cell subpopulations with distinct T cell-suppressive activity. *Blood* 2008; **111**: 4233–4244.
- 102 Youn JI, Nagaraj S, Collazo M, Gabrilovich DI. Subsets of myeloid-derived suppressor cells in tumor-bearing mice. *J Immunol* 2008; **181**: 5791–5802.
- 103 Ochoa AC, Zea AH, Hernandez C, Rodriguez PC. Arginase, prostaglandins, and myeloid-derived suppressor cells in renal cell carcinoma. *Clin Cancer Res* 2007; **13**: 721s726s.
- 104 Almand B, Clark JI, Nikitina E, van Beynen J, English NR, Knight SC *et al*. Increased production of immature myeloid cells in cancer patients: a mechanism of immunosuppression in cancer. *J Immunol* 2001; **166**: 678–689.
- 105 Yang R, Cai Z, Zhang Y, Yutzy WH 4th, Roby KF, Roden RB. CD80 in immune suppression by mouse ovarian carcinoma-associated Gr-1⁺CD11b⁺ myeloid cells. *Cancer Res* 2006; **66**: 6807–6815.
- 106 Huang B, Pan PY, Li Q, Sato AI, Levy DE, Bromberg J *et al*. Gr-1⁺CD115⁺ immature myeloid suppressor cells mediate the development of tumor-induced T regulatory cells and T-cell anergy in tumor-bearing host. *Cancer Res* 2006; **66**: 1123–1131.
- 107 Ribechini E, Greifenberg V, Sandwick S, Lutz MB. Subsets, expansion and activation of myeloid-derived suppressor cells. *Med Microbiol Immunol* 2010; **199**: 273–281.
- 108 Highfill SL, Rodriguez PC, Zhou Q, Goetz CA, Koehn BH, Veenstra R *et al*. Bone marrow myeloid-derived suppressor cells (MDSCs) inhibit graft-versus-host disease (GVHD) via an arginase-1-dependent mechanism that is up-regulated by interleukin-13. *Blood* 2010; **116**: 5738–5747.
- 109 Caplan AI. Mesenchymal stem cells. *J Orthop Res* 1991; **9**: 641–650.
- 110 Le Blanc K, Tammik C, Rosendahl K, Zetterberg E, Ringden O. HLA expression and immunologic properties of differentiated and undifferentiated mesenchymal stem cells. *Exp Hematol* 2003; **31**: 890–896.
- 111 Klyushnenkova E, Mosca JD, Zernitkina V, Majumdar MK, Beggs KJ, Simonetti DW *et al*. T cell responses to allogeneic human mesenchymal stem cells: immunogenicity, tolerance, and suppression. *J Biomed Sci* 2005; **12**: 47–57.
- 112 Majumdar MK, Thiede MA, Haynesworth SE, Bruder SP, Gerson SL. Human marrow-derived mesenchymal stem cells (MSCs) express hematopoietic cytokines and support long-term hematopoiesis when differentiated toward stromal and osteogenic lineages. *J Hematother Stem Cell Res* 2000; **9**: 841–848.
- 113 Cheng L, Hammond H, Ye Z, Zhan X, David G. Human adult marrow cells support prolonged expansion of human embryonic stem cells in culture. *Stem Cells* 2003; **21**: 131–142.
- 114 Le Blanc K, Tammik L, Sundberg B, Haynesworth SE, Ringden O. Mesenchymal stem cells inhibit and stimulate mixed lymphocyte cultures and mitogenic responses independently of the major histocompatibility complex. *Scand J Immunol* 2003; **57**: 11–20.
- 115 Di Nicola M, Carlo-Stella C, Magni M, Milanese M, Longoni PD, Matteucci P *et al*. Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. *Blood* 2002; **99**: 3838–3843.
- 116 Krampera M, Glennie S, Dyson J, Scott D, Laylor R, Simpson E *et al*. Bone marrow mesenchymal stem cells inhibit the response of naive and memory antigen-specific T cells to their cognate peptide. *Blood* 2003; **101**: 3722–3729.
- 117 Glennie S, Soeiro I, Dyson PJ, Lam EW, Dazzi F. Bone marrow mesenchymal stem cells induce division arrest anergy of activated T cells. *Blood* 2005; **105**: 2821–2827.
- 118 Plumas J, Chaperot L, Richard MJ, Molens JP, Bensa JC, Favrot MC. Mesenchymal stem cells induce apoptosis of activated T cells. *Leukemia* 2005; **19**: 1597–1604.
- 119 Aggarwal S, Pittenger MF. Human mesenchymal stem cells modulate allogeneic immune cell responses. *Blood* 2005; **105**: 1815–1822.
- 120 Potian JA, Aviv H, Ponzio NM, Harrison JS, Rameshwar P. Veto-like activity of mesenchymal stem cells: functional discrimination between cellular responses to alloantigens and recall antigens. *J Immunol* 2003; **171**: 3426–3434.
- 121 Corcione A, Benvenuto F, Ferretti E, Giunti D, Cappiello V, Cazzanti F *et al*. Human mesenchymal stem cells modulate B-cell functions. *Blood* 2006; **107**: 367–372.
- 122 Jiang XX, Zhang Y, Liu B, Zhang SX, Wu Y, Yu XD *et al*. Human mesenchymal stem cells inhibit differentiation and function of monocyte-derived dendritic cells. *Blood* 2005; **105**: 4120–4126.
- 123 Nauta AJ, Kruisselbrink AB, Lurvink E, Willemze R, Fibbe WE. Mesenchymal stem cells inhibit generation and function of both CD34⁺-derived and monocyte-derived dendritic cells. *J Immunol* 2006; **177**: 2080–2087.
- 124 Sotiropoulou PA, Perez SA, Gritzapis AD, Baxevas CN, Papamichail M. Interactions between human mesenchymal stem cells and natural killer cells. *Stem Cells* 2006; **24**: 74–85.
- 125 Spaggiari GM, Capobianco A, Abdelrazik H, Becchetti F, Mingari MC, Moretta L. Mesenchymal stem cells inhibit natural killer-cell proliferation, cytotoxicity, and cytokine production: role of indoleamine 2,3-dioxygenase and prostaglandin E2. *Blood* 2008; **111**: 1327–1333.
- 126 Lazarus HM, Haynesworth SE, Gerson SL, Rosenthal NS, Caplan AI. *Ex vivo* expansion and subsequent infusion of human bone marrow-derived stromal progenitor cells (mesenchymal progenitor cells): implications for therapeutic use. *Bone Marrow Transplant* 1995; **16**: 557–564.
- 127 Horwitz EM, Prockop DJ, Fitzpatrick LA, Koo WW, Gordon PL, Neel M *et al*. Transplantation and therapeutic effects of bone marrow-derived mesenchymal cells in children with osteogenesis imperfecta. *Nat Med* 1999; **5**: 309–313.
- 128 Bang OY, Lee JS, Lee PH, Lee G. Autologous mesenchymal stem cell transplantation in stroke patients. *Ann Neurol* 2005; **57**: 874–882.
- 129 Chen SL, Fang WW, Ye F, Liu YH, Qian J, Shan SJ *et al*. Effect on left ventricular function of intracoronary transplantation of autologous bone marrow mesenchymal stem cell in patients with acute myocardial infarction. *Am J Cardiol* 2004; **94**: 92–95.
- 130 Le Blanc K, Frassonni F, Ball L, Locatelli F, Roelofs H, Lewis I *et al*. Mesenchymal stem cells for treatment of steroid-resistant, severe, acute graft-versus-host disease: a phase II study. *Lancet* 2008; **371**: 1579–1586.
- 131 Miller JP, Izon D, DeMuth W, Gerstein R, Bhandoola A, Allman D. The earliest step in B lineage differentiation from common lymphoid progenitors is critically dependent upon interleukin 7. *J Exp Med* 2002; **196**: 705–711.
- 132 Peschon JJ, Morrissey PJ, Grabstein KH, Ramsdell FJ, Maraskovsky E, Gliniak BC *et al*. Early lymphocyte expansion is severely impaired in interleukin 7 receptor-deficient mice. *J Exp Med* 1994; **180**: 1955–1960.
- 133 Roifman CM, Zhang J, Chitayat D, Sharfe N. A partial deficiency of interleukin-7R alpha is sufficient to abrogate T-cell development and cause severe combined immunodeficiency. *Blood* 2000; **96**: 2803–2807.
- 134 Schluns KS, Kieper WC, Jameson SC, Lanchaud L. Interleukin-7 mediates the homeostasis of naive and memory CD8 T cells *in vivo*. *Nat Immunol* 2000; **1**: 426–432.
- 135 Becker TC, Wherry EJ, Boone D, Murali-Krishna K, Antia R, Ma A *et al*. Interleukin 15 is required for proliferative renewal of virus-specific memory CD8 T cells. *J Exp Med* 2002; **195**: 1541–1548.
- 136 Kieper WC, Tan JT, Bondi-Boyd B, Gapin L, Sprent J, Ceredig R *et al*. Overexpression of interleukin (IL)-7 leads to IL-15-independent generation of memory phenotype CD8⁺ T cells. *J Exp Med* 2002; **195**: 1533–1539.
- 137 Klebanoff CA, Finkelstein SE, Surman DR, Lichtman MK, Gattinoni L, Theoret MR *et al*. IL-15 enhances the *in vivo* antitumor activity of tumor-reactive CD8⁺ T cells. *Proc Natl Acad Sci USA* 2004; **101**: 1969–1974.
- 138 Di Carlo E, Comes A, Orengo AM, Rosso O, Meazza R, Musiani P *et al*. IL-21 induces tumor rejection by specific CTL and IFN-gamma-dependent CXC chemokines in syngeneic mice. *J Immunol* 2004; **172**: 1540–1547.
- 139 Parrish-Novak J, Dillon SR, Nelson A, Hammond A, Sprecher C, Gross JA *et al*. Interleukin 21 and its receptor are involved in NK cell expansion and regulation of lymphocyte function. *Nature* 2000; **408**: 57–63.
- 140 Nagasawa T, Hirota S, Tachibana K, Takakura N, Nishikawa S, Kitamura Y *et al*. Defects of B-cell lymphopoiesis and bone-marrow myelopoiesis in mice lacking the CXC chemokine PBSF/SDF-1. *Nature* 1996; **382**: 635–638.
- 141 Peled A, Grabovsky V, Habler L, Sandbank J, Arenzana-Seisdedos F, Petit I *et al*. The chemokine SDF-1 stimulates integrin-mediated arrest of CD34⁺ cells on vascular endothelium under shear flow. *J Clin Invest* 1999; **104**: 1199–1211.
- 142 Peled A, Kollet O, Ponomarev T, Petit I, Franitza S, Grabovsky V *et al*. The chemokine SDF-1 activates the integrins LFA-1, VLA-4, and VLA-5 on immature human CD34⁺ cells: role in transendothelial/stromal migration and engraftment of NOD/SCID mice. *Blood* 2000; **95**: 3289–3296.
- 143 Lapidot T, Kollet O. The essential roles of the chemokine SDF-1 and its receptor CXCR4 in human stem cell homing and repopulation of transplanted immune-deficient NOD/SCID and NOD/SCID/B2m(null) mice. *Leukemia* 2002; **16**: 1992–2003.

- 144 Ma Q, Jones D, Springer TA. The chemokine receptor CXCR4 is required for the retention of B lineage and granulocytic precursors within the bone marrow microenvironment. *Immunity* 1999; **10**: 463–471.
- 145 Avigdor A, Goichberg P, Shivtiel S, Dar A, Peled A, Samira S *et al*. CD44 and hyaluronic acid cooperate with SDF-1 in the trafficking of human CD34⁺ stem/progenitor cells to bone marrow. *Blood* 2004; **103**: 2981–2989.
- 146 Koni PA, Joshi SK, Temann UA, Olson D, Burkly L, Flavell RA. Conditional vascular cell adhesion molecule 1 deletion in mice: impaired lymphocyte migration to bone marrow. *J Exp Med* 2001; **193**: 741–754.
- 147 Frenette PS, Subbarao S, Mazo IB, von Andrian UH, Wagner DD. Endothelial selectins and vascular cell adhesion molecule-1 promote hematopoietic progenitor homing to bone marrow. *Proc Natl Acad Sci USA* 1998; **95**: 14423–14428.