Regulation of stomatal density by the GTL1 transcription factor for improving water use efficiency

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> stoma (pl. stomata) is the pore I formed by two guard cells found predominantly in the leaf epidermis. Plants control stomatal aperture (opening and closing) and/or number (density) to regulate carbon dioxide (CO₂) uptake and water loss (transpiration), which is necessary to optimize plant growth, development, and fitness in response to various environmental conditions. Recently, we identified Arabidopsis GT2-LIKE 1 (GTL1) as a transcriptional repressor of STOMATAL DENSITY AND DISTRIBUTION 1 (SDD1), a negative regulator of stomatal density. GTL1 directly interacts with the SDD1 promoter regulating stomatal density, transpiration and water use efficiency (WUE). Here we discuss potential GTL1 orthologs in other plant species.

The plant epidermal surface consists of pavement cells and stomata, and regulates gas exchange of carbon dioxide (CO_2) and water (H₂O) vapor between the leaf and air.1 Plants have physiological and developmental mechanisms controlled by genetic pathways to regulate stomatal aperture and/or number (density) to optimize fitness by controlling WUE in various environmental conditions, particularly under water deficit.1 Stomatal density in leaves is controlled by two genetic pathways: the stomatal cell lineage pathway (basal pathway) that results in the development of guard cells from post-protodermal cells and the negative regulatory pathway that inhibits cells adjacent to stomata from developing into stomata.^{2,3} The interaction and competition between stomatal cell lineage and

negative regulatory pathways is finely controlled to determine stomatal density for environmental adaptation.^{4,5}

The basal pathway is regulated by several basic-helix-loop-helix (bHLH) transcription factors such as *SPEECHLESS* (*SPCH*), *MUTE*, and *FAMA*, which specifically control the three developmentally distinct steps: asymmetric cell division that produces a meristemoid and a larger sister cell, transition of a meristemoid into a guard mother cell, and guard cell differentiation.^{2,3} Each step is also regulated by the interaction of INDUCER OF CBF EXPRESSION 1 (ICE1)/SCREAM (SCRM 1) or ICE2/ SCRM2.⁶

STOMATAL DENSITY AND DISTRIBUTION I (SDDI) that encodes a subtilisin-like serine protease is the furthest upstream genetic determinant in the negative regulatory pathway.^{7,8} SDDI processes propeptides to ligands that activate receptors such as TOO MANY MOUTHS (TMM) and ERECTA (ER), which transduces the signal to the mitogen-activated protein kinase (MAPK) pathway and suppresses stomatal development.

We recently identified the GTL1 transcription factor as a determinant that regulates *SDD1* expression and stomatal density⁹ (Fig. 1). Loss-of GTL1 function results in reduction of transpiration with similar biomass accumulation and net CO₂ assimilation, which improves integrated and instantaneous WUE, and drought tolerance.⁹ Similar level of net CO₂ assimilation in *gtl1* plants, despite their lower stomatal conductance, may indicate that net CO₂ assimilation is still at a maximum

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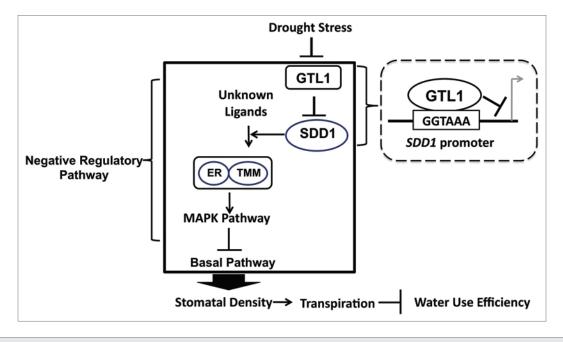


Figure 1. Model of how the GTL1 transcription factor regulates stomatal development in Arabidopsis. GTL1 binds to the GT3 box (GGTAAA) in the *SDD1* promoter and transrepresses *SDD1* expression. SDD1 presumably processes unknown ligands for the TMM-ER receptor complexes to activate the MAPK pathway, thereby repressing the basal pathway. Repression or induction of *SDD1* expression increases or decreases stomatal density, respectively. GTL1 fine-tunes *SDD1* expression to optimize water use efficiency.

rate in gtl1 plants under our conditions, or that gtl1 plants have a higher carboxylation efficiency of ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco).¹ The determination of any effects of GTL1 on CO₂ assimilation requires further investigation. Reduced transpiration in gtl1 plants is attributable to the reduction of stomatal density in abaxial leaves.⁹ GTL1 protein directly interacts with the *SDD1* promoter to repress its expression.⁹ Taken together, our results indicate that GTL1 is a transcriptional repressor of *SDD1* that negatively regulates stomatal density, which is necessary for fine-tuning of stomatal density to regulate WUE (Fig. 1). We postulate that GTL1 receives an environmental signal and transduces this signal to the regulation of stomatal development for an adaptive response that prevents excessive water loss.

GTL1 Orthologs in Other Plant Species

GTL1 is a member of the GT2 transcription factor family containing two DNA-binding domains.¹⁰⁻¹² The Basic

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Species	Locus name
Poplar	Pt01s31660, Pt01s45870, Pt02s06900, Pt05s21410, Pt5s21420, Pt19s02650, Pt11s15050
Peach	Pp001704, Pp002848, Pp003808, Pp021097
Grape	Vv01019722001, Vv01026588001
Cucumber	Cs193210, Cs204000, Cs250070
Tomato	S14g071360, Sl11g005380, Sl12g056510
Рарауа	Cp21.194, Cp21.195, Cp2611.1
Soybean	Gm2g09050, Gm4g39400, Gm6g15500, Gm10g36950, Gm10g36960, Gm16g28240, Gm20g30640, Gm20g30650
Rice	Os2g43300, Os3g02240.1, Os4g45750, Os10g37240
Maize	Zm016649, Zm063203, Zm113585
Sorgum	Sb1g017120, Sb1g049740, Sb4g033390, Sb6g023980

Local Alignment Search Tool (BLAST) in Phytozome (www.phytozome.net) was used to identify GTL1 orthologs in the plant species Arabidopsis thaliana (At, Arabidopsis), Carica papaya (Cp, papaya), Cucumis sativus (Cs, Cucumber), Populus trichocarpa (Pt, Poplar), Prunus persica (Pp, Peach), Solanum lycopersicum (Sl, Tomato), Sorghum bicolor (Sb, Sorghum) and Vitis vinifera (Vv, Grape). We identified GTL1 orthologs similar to AtGTL1 based on the amino acid sequence identity and the existence of two DNA-binding domains (Table 1). The most similar AtGTL1 orthologs identified were in poplar (Pt01s45870), peach (Pp001704), grape (Vv01019722001), and cucumber (Cs193210). Interestingly, monocots such as rice, maize and sorghum also have GTL1 orthologs that are more similar to AtGTL1 than AtGT2 or AtGTL2. Considering the evidence indicating that stomatal development pathways in monocot and dicot may be similar despite different stomatal morphology,¹³ we hypothesize that GTL1 orthologs in monocots positively regulate stomatal density and transpiration, and therefore may play a role in regulating WUE. Suppression of GTL1 orthologs in other plant species by tools such as RNAi suppression may provide a means to improve WUE in important economic plants.¹⁴

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