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# Selection on codon usage and base composition in Drosophila americana

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We have used a polymorphism dataset on introns and coding sequences of X-linked loci in Drosophila americana to estimate the strength of selection on codon usage and/or biased gene conversion (BGC), taking into account a recent population expansion detected by a maximumlikelihood method. Drosophila americana was previously thought to have a stable demographic history, so that this evidence for a recent population expansion means that previous estimates of selection need revision. There was evidence for natural selection or BGC favouring GC over AT variants in introns, which is stronger for GC-rich than GC-poor introns. By comparing introns and coding sequences, we found evidence for selection on codon usage bias, which is much stronger than the forces acting on GC versus AT basepairs in introns.

Keywords: Drosophila americana; codon usage; biased gene conversion; population expansion

# 1. INTRODUCTION

In bacteria, yeast, Drosophila and plants, there is evidence for selection on codon usage at synonymous coding sites, probably because of selection on translational efficiency and/or accuracy [\[1\]](#page-2-0). Several population genetic studies of Drosophila have used polymorphism data to estimate the intensity of selection on codon usage  $[2-7]$  $[2-7]$  $[2-7]$  $[2-7]$ . In addition, genome evolution is affected by the process of biased gene conversion (BGC), which tends to favour GC over AT basepairs in the meiotic products of GC/AT heterozygotes, and acts in a similar way to directional selection [\[8\]](#page-2-0). Its effects and strength can be inferred from polymorphism data on non-coding sequences [[9,10\]](#page-2-0).

Here, we present results on the nature and intensity of selection and/or BGC on non-coding and synonymous sites, using polymorphism data on X-linked loci of Drosophila americana, a close relative of Drosophila virilis. The virilis group diverged from the Drosophila melanogaster group about 62 Ma [[11](#page-2-0)] and has somewhat different patterns of codon usage and base composition [[12,13\]](#page-2-0) making it of special interest for studies of these genomic features. Drosophila americana has been used in evolutionary genetic studies for several decades [[14](#page-2-0)–[18\]](#page-3-0). It has a well-defined ecology, independent of

Electronic supplementary material is available at [http:](http://dx.doi.org/10.1098/rsbl.2011.0601)//[dx.doi.org](http://dx.doi.org/10.1098/rsbl.2011.0601)/ 10.1098/[rsbl.2011.0601](http://dx.doi.org/10.1098/rsbl.2011.0601) or via [http:](http://rsbl.royalsocietypublishing.org)//[rsbl.royalsocietypublishing.org](http://rsbl.royalsocietypublishing.org). human activity [\[14](#page-2-0)], and might thus be expected to have a relatively stable demographic history, which is advantageous for estimating the parameters of natural selection from polymorphism data [[3](#page-2-0)].

This paper presents, to our knowledge, the first analysis of a species in the *virilis* group to detect both selection on codon usage and BGC from polymorphism data, using a population genetic method that allows for a recent population size change [[19](#page-3-0)], whereas a previous study of selection on codon usage assumed demographic equilibrium [\[3\]](#page-2-0). We provide evidence for a recent population expansion, and for selection on codon usage at synonymous sites, as well as selection or BGC favouring GC over AT in GC-rich introns.

### 2. MATERIAL AND METHODS

For DNA extractions, we used males from 14 D. americana isofemale lines from the HI99 population on the south bank of the Missouri River [\(http:](http://www.biology.uiowa.edu/mcallister/HI.html)//[www.biology.uiowa.edu](http://www.biology.uiowa.edu/mcallister/HI.html)/mcallister/HI.html), provided by Bryant McAllister. About 85 per cent of genomes from this population have a fusion between the X and chromosome 4 [\[15](#page-2-0),[16](#page-2-0)]. Because genes located near the fusion region or in inversions may suffer from hitchhiking effects of the rearrangements, regions affected by the  $X/4$  fusion or known segregating inversions were excluded.

Details of DNA extraction, amplification, sequencing and alignment of sequences are provided in the electronic supplementary material. The resulting dataset contains sequences for 32 introns sampled from 18 loci, including 12 short introns and 20 long introns (electronic supplementary material, figure S1). We also obtained the coding sequences of 15 X-linked genes, and retrieved four additional X-linked coding sequences from Maside & Charlesworth [\[17\]](#page-3-0), in order to compare synonymous sites and introns. Sequences were deposited in GenBank (accession numbers JN246676-JN246926).

Using the codon preference table for *D. virilis* from Betancourt  $et al. [20]$  $et al. [20]$  $et al. [20]$ , we assigned preferred (P) and unpreferred (U) alternatives to each synonymous site in both species, and then used parsimony to determine whether the synonymous site change within D. americana was  $P > P$ ,  $U > U$ ,  $P > U$  or  $U > P$ . Similarly, we obtained the counts and frequencies of  $AT > TA$ ,  $GC > CG$ ,  $GC > AT$  and  $AT > GC$  polymorphic changes for each intron in the *D. americana* intron dataset to test for selection or BGC favouring GC over AT basepairs [\[9,10\]](#page-2-0).

We used the maximum-likelihood (ML) method of Zeng & Charlesworth [[4](#page-2-0)], as modified by Haddrill et al. [\[6\]](#page-2-0), for fitting the observed frequencies of variants to models of selection and demography, to estimate the strength of selection/BGC on  $U > P$  synonymous polymorphisms or  $GC > AT$  basepairs and the extent of mutational bias in favour of  $GC > AT$  versus  $GC > AT$  changes, allowing for the possibility of a recent population size change in D. americana. Details are given in the electronic supplementary material.

# 3. RESULTS

Our major findings are presented below; other results are described in the electronic supplementary material. The mean values of various summary statistics are shown in [table 1](#page-1-0). The mean diversity and divergence values are broadly consistent with those reported previously, even after excluding the four coding sequences in common with Maside & Charlesworth [[17\]](#page-3-0). There are no significant differences in mean Tajima's D values between the different classes of sites, or in variation and divergence values among intronic versus synonymous sites. The consistently negative Tajima's D values suggest a recent population expansion [[21\]](#page-3-0), as confirmed by the analysis below.

We first examined selection on variants affecting codon usage, using data on 19 X-linked coding sequences. There are four classes of mutations:  $P > P$ and  $U > U$  (expected to be selectively nearly neutral),  $P > U$  (potentially deleterious), and  $U > P$  mutations (potentially advantageous) [[2](#page-2-0)[,22](#page-3-0)]. Selection favouring

<span id="page-1-0"></span>Table 1. Summary statistics of polymorphism and divergence for the different classes of sites. (S is the number of segregating sites;  $\pi$  and  $\theta_{\rm w}$  are the standard measures of the nucleotide diversity based on the mean pairwise divergence per nucleotide site between alleles and the number of segregating sites, respectively;  $D$  is the number of fixed differences between  $D$ . virilis and D. americana;  $K_{\text{JC}}$  is the mean Jukes-Cantor-corrected divergence from D. virilis; and  $D_T$  is Tajima's D statistic.)

category	S	$\pi$ (%) (s.e.)	$\theta_{\rm W}$ (%) (s.e.)		$K_{\rm IC}$ (%) (s.e.)	$D_{\rm T}$ (s.e.)
introns	803	2.17(0.23)	2.32(0.22)	609	9.98(0.92)	$-0.75(0.09)$
synonymous	173	1.96 (0.56)	1.77(0.32)	245	9.84(1.01)	$-0.73(0.13)$
non-synonymous	29	0.09(0.03)	0.11(0.03)	68	0.78(0.22)	$-0.93(0.20)$

P versus U variants is usually expected to yield an excess of  $P > U$  over  $U > P$  variants [[2](#page-2-0)–[4\]](#page-2-0). Consistent with this, we found nearly three times as many  $P > U$  variants as  $U > P$  variants (162 versus 56). In addition,  $P > U$ variants are disproportionately present at low frequencies compared with  $U > P$  variants (figure 1); the mean frequency of  $U > P$  mutations over the segregating sites in the sample was significantly higher than that of both  $P > U$  changes (Wilcoxon's  $W = 916.5$ ,  $p = 0.022$ ) and the pooled  $P > P$  and  $U > U$  changes  $(W = 856, p = 0.030).$ 

We also explored the possible effect of BGC on intronic base composition, which is expected to favour GC over AT variants [\[8\]](#page-2-0). The total numbers of  $GC > AT$  and  $AT > GC$  variants over the set of 32 introns are similar (248 versus 242), whereas the mean frequency of  $AT > GC$  variants is higher than that of  $GC > AT$  variants (0.28 versus 0.19) ( $W =$ 197.5,  $p = 0.002$ ).

We also analysed these datasets by the method of Zeng & Charlesworth [\[4,6](#page-2-0)]. The ML estimates of mutational bias under all models examined indicate higher rates of mutations towards  $P > U$  and  $GC >$ AT variants compared with the reverse mutations, as found in previous Drosophila studies [\[4\]](#page-2-0). The contrasts between the model with no expansion, but with all other parameters fitted  $(L_0)$ , and the other models  $(L<sub>1</sub>)$  indicate a recent 4.2-fold increase in population size ([table 2](#page-2-0)), with an ML estimate of the time since the event of  $\tau = 0.11$ , where  $\tau$  is the number of generations since the expansion divided by twice the current effective population size.

To test for selection on codon usage, we compared the full  $L_1$  model with the reduced version with  $\gamma_{\text{cod}} = 0$ , where  $\gamma_{\text{cod}}$  is the estimate of the strength of selection/BGC at a synonymous site, scaled by four times the effective population size before the expansion. The full model has strong statistical support  $(\chi_1^2 = 29.9, p < 0.0001)$ , with  $\gamma_{\text{cod}} = 1.6$ , implying selection in favour of preferred codons, consistent with the patterns of  $P > U$  versus  $U > P$ variants described above. To test for selection/BGC on intronic variants, we compared the full  $L_1$  model with  $\gamma_{\text{int}} = 0 \ (\chi_1^2 = 8.27, \ p = 0.004)$ . Selection or BGC in favour of GC intronic basepairs is thus implied, with  $\gamma_{int} = 0.36$ . We tested whether  $\gamma_{cod}$  is significantly larger than  $\gamma_{\text{int}}$ , by comparing a model with a single  $\gamma$  for both categories: the full  $L_1$ model is significantly more likely than that with  $\gamma_{\text{cod}} = \gamma_{\text{int}} \quad (\chi_1^2 = 14.6, \, p < 0.0001).$  We similarly found that the  $\gamma_{int}$  estimates are significantly different for introns with high and low GC content  $(\chi_1^2 = 18.9,$  $p < 0.0001$ ).



Figure 1. The frequency classes of polymorphisms for different types of synonymous site changes (see text for explanation of P and U). The numbers above each type of synonymous site change indicate the percentages of the total number of synonymous polymorphisms contributed by each type. Black bars, less than 0.2; grey bars, greater than or equal to 0.2 and less than or equal to 0.8; unfilled bars, greater than 0.8.

#### 4. DISCUSSION

Our analysis provides evidence for a fairly large, recent increase in population size in *D. americana*, within a time-span of approximately  $0.11 \times 2N_e$  generations. This is consistent with the results for another widespread North American species, Drosophila pseudoobscura [[6\]](#page-2-0). Given the mean silent site diversity values of about 2 per cent (table 1), using the standard formula for equilibrium neutral diversity  $(4N_e\mu)$  together with the D. melanogaster mutation rate estimate of  $3.5 \times 10^{-9}$ [\[23](#page-3-0)], we estimate that the current  $N_e$  of D. americana is about 1.4 million, implying that the expansion took place about 308 000 generations ago. Assuming five generations per year for this slowly breeding species [\[14](#page-2-0)], this corresponds to 61 600 years, although there is considerable uncertainty about the exact value.

The results in [table 2](#page-2-0) show that both synonymous sites and intron sequences in *D. americana* are influenced by selection and/or BGC, even after the recent population expansion was taken into account. The  $\gamma$ estimate of about 1.6 for selection favouring preferred over unpreferred codons is in line with values for other Drosophila species [[6](#page-2-0)[,19](#page-3-0)], but is lower than the value of 2.6 found previously in D. americana [[3](#page-2-0)], suggesting that population expansion caused the strength of selection to be overestimated, as expected theoretically [\[4\]](#page-2-0).

<span id="page-2-0"></span>



Consistent with other evidence from Drosophila for selection or BGC favouring GC over AT base pairs in non-coding sequences [10,[19\]](#page-3-0), we found evidence for natural selection or BGC favouring GC over AT basepairs. As in Haddrill & Charlesworth [10], selection/BGC appears to be significantly stronger in GC-rich compared with GC-poor introns, consistent with the idea that the intensity of BGC shapes the GC content of genomes [8].

As preferred codons are mostly GC-ending, selection for codon usage largely works in the same direction as BGC. The difference in  $\gamma$  between the synonymous sites and introns almost certainly reflects the action of selection on codon usage bias at synonymous sites, possibly in addition to the effects of BGC, whereas the apparent selection on intron sites may result from BGC alone [8]. This difference could also be owing to a higher rate of recombination in exons than in introns, resulting in a higher rate of BGC in exons [3], although we did not find any evidence for this (see electronic supplementary material).

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- 1 Hershberg, R. & Petrov, D. A. 2008 Selection on codon bias. Annu. Rev. Genet. 42, 287-299. ([doi:10.1146](http://dx.doi.org/10.1146/Annurev.Genet.42.110807.091442)/ [Annurev.Genet.42.110807.091442\)](http://dx.doi.org/10.1146/Annurev.Genet.42.110807.091442)
- 2 Akashi, H. 1995 Inferring weak selection from patterns of polymorphism and divergence at silent sites in Drosophila DNA. Genetics 139, 1067–1076.
- 3 Maside, X. L., Lee, A. W. S. & Charlesworth, B. 2004 Selection on codon usage in Drosophila americana. Curr. Biol. 14, 150-154. (doi:10.1016/[J.Cub.2003.](http://dx.doi.org/10.1016/J.Cub.2003.12.055) [12.055](http://dx.doi.org/10.1016/J.Cub.2003.12.055))
- 4 Zeng, K. & Charlesworth, B. 2009 Estimating selection intensity on synonymous codon usage in a nonequili-brium population. Genetics 183, 651-662. [\(doi:10.](http://dx.doi.org/10.1534/Genetics.109.101782) 1534/[Genetics.109.101782](http://dx.doi.org/10.1534/Genetics.109.101782))
- 5 Comeron, J. M. & Guthrie, T. B. 2005 Intragenic Hill–Robertson interference influences selection intensity on synonymous mutations in Drosophila. Mol. Biol. Evol. 22, 2519–2530. [\(doi:10.1093](http://dx.doi.org/10.1093/molbev/msi246)/molbev/msi246)
- 6 Haddrill, P. R., Zeng, K. & Charlesworth, B. 2011 Determinants of synonymous and nonsynonymous variability in three species of Drosophila. Mol. Biol. Evol. 28, 1731–1743. ([doi:10.1093](http://dx.doi.org/10.1093/molbev/msq354)/molbev/msq354)
- 7 dos Reis, M. & Wernisch, L. 2009 Estimating translational selection in eukaryotic genomes. Mol. Biol. Evol. 26, 451 –461. ([doi:10.1093](http://dx.doi.org/10.1093/Molbev/Msn272)/Molbev/Msn272)
- 8 Marais, G. 2003 Biased gene conversion: implications for genome and sex evolution. Trends Genet. 19, 330-338. (doi:10.1016/[S0168-9525\(03\)00116-1](http://dx.doi.org/10.1016/S0168-9525(03)00116-1))
- 9 Galtier, N., Bazin, E. & Bierne, N. 2006 GC-biased segregation of noncoding polymorphisms in *Drosophila*. Genetics 172, 221-228. (doi:10.1534/[genetics.105.](http://dx.doi.org/10.1534/genetics.105.046524) [046524](http://dx.doi.org/10.1534/genetics.105.046524))
- 10 Haddrill, P. R. & Charlesworth, B. 2008 Non-neutral processes drive the nucleotide composition of noncoding sequences in *Drosophila*. Biol. Lett. 4, 438-441. (doi:10.1098/[Rsbl.2008.0174](http://dx.doi.org/10.1098/Rsbl.2008.0174))
- 11 Tamura, K., Subramanian, S. & Kumar, S. 2004 Temporal patterns of fruit fly (Drosophila) evolution revealed by mutation clocks. Mol. Biol. Evol. 21, 36–44. [\(doi:10.](http://dx.doi.org/10.1093/molbev/msg236) 1093/molbev/[msg236](http://dx.doi.org/10.1093/molbev/msg236))
- 12 McVean, G. A. & Vieira, J. 2001 Inferring parameters of mutation, selection and demography from patterns of synonymous site evolution in Drosophila. Genetics 157,  $245 - 257$ .
- 13 Heger, A. & Ponting, C. P. 2007 Variable strength of translational selection among 12 Drosophila species. Genetics 177, 1337–1348. (doi:10.1534/[genetics.107.070466\)](http://dx.doi.org/10.1534/genetics.107.070466)
- 14 Throckmorton, L. H. 1982 The virilis species group. In The genetics and biology of Drosophila (eds M. Ahsburner, H. L. Carson & J. N. Thompson), pp. 227 –296. New York, NY: Academic Press.
- 15 Vieira, J., McAllister, B. F. & Charlesworth, B. 2001 Evidence for selection at the fused1 locus of Drosophila americana. Genetics 158, 279 –290.
- 16 McAllister, B. F. 2002 Chromosomal and allelic variation in Drosophila americana: selective maintenance of a

<span id="page-3-0"></span>chromosomal cline. Genome 45, 13–21. ([doi:10.1139](http://dx.doi.org/10.1139/g01-112)/ [g01-112\)](http://dx.doi.org/10.1139/g01-112)

- 17 Maside, X. & Charlesworth, B. 2007 Patterns of molecular variation and evolution in Drosophila americana and its relatives. Genetics 176, 2293–2305. ([doi:10.1534](http://dx.doi.org/10.1534/Genetics.107.071191)/ [Genetics.107.071191](http://dx.doi.org/10.1534/Genetics.107.071191))
- 18 Vieira, C. P., Almeida, A., Dias, J. D. & Vieira, J. 2006 On the location of the gene(s) harbouring the advantageous variant that maintains the X4 fusion of Drosophila americana. Genet. Res. 87, 163-174. ([doi:10.](http://dx.doi.org/10.1017/S0016672306008147) 1017/[S0016672306008147](http://dx.doi.org/10.1017/S0016672306008147))
- 19 Zeng, K. & Charlesworth, B. 2010 Studying patterns of recent evolution at synonymous sites and intronic sites in Drosophila melanogaster. J. Mol. Evol. 70, 116-128. (doi:10.1007/[S00239-009-9314-6](http://dx.doi.org/10.1007/S00239-009-9314-6))
- 20 Betancourt, A. J., Welch, J. J. & Charlesworth, B. 2009 Reduced effectiveness of selection caused by a lack of recombination. Curr. Biol. 19, 655 –660. ([doi:10.1016](http://dx.doi.org/10.1016/J.Cub.2009.02.039)/ [J.Cub.2009.02.039](http://dx.doi.org/10.1016/J.Cub.2009.02.039))
- 21 Tajima, F. 1989 The effect of change in population size on DNA polymorphism. Genetics 123, 597-601.
- 22 Haddrill, P. R., Bachtrog, D. & Andolfatto, P. 2008 Positive and negative selection on noncoding DNA in Drosophila simulans. Mol. Biol. Evol. 25, 1825–1834. [\(doi:10.1093](http://dx.doi.org/10.1093/Molbev/Msn125)/Molbev/Msn125)
- 23 Keightley, P. D., Trivedi, U., Thomson, M., Oliver, F., Kumar, S. & Blaxter, M. L. 2009 Analysis of the genome sequences of three Drosophila melanogaster spontaneous mutation accumulation lines. Genome Res. 19, 1195–1201. (doi:10.1101/[Gr.091231.109\)](http://dx.doi.org/10.1101/Gr.091231.109)