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C9ORF72, the new gene on the block, causes C9FTD/ALS: new insights provided by neuropathology

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New avenues of investigation into sporadic neurodegenerative disease are often revealed by genetic discoveries in familial disease. In frontotemporal dementia, it seems that genetic and molecular discoveries are reported in pairs. *MAPT* mutations in FTDP-17 with tauopathy (June 1998), PGRN mutations in familial FTLD-U (July 2006), TDP-43 as the major protein component of inclusions in FTLD-U (October 2006)—each of these were reported by different groups in two papers in the same month [2, 3, 6, 11, 18, 20]. Just 2 months ago, DeJesus et al. [8] and Renton et al. [21] in back-to-back publications in Neuron reported that mutations in a noncoding region of *C9ORF72*, coding for a protein of unknown function that is highly conserved across species, resulted in expansion of a GGGGCC hexanucleotide repeat, and was the cause of chromosome 9p-linked FTD and ALS. The recommended terminology for FTD and ALS associated with *C9ORF72* mutations is C9FTD/ALS [8].

This issue of *Acta Neuropathologica* continues this pattern, with two articles describing the neuropathology in cases of FTLD-TDP, FTLD-MND, and ALS with *C9ORF72* (C9)-expanded repeats [1, 17]. In the first paper, the Mayo Jacksonville group report clinical and pathologic heterogeneity within a group of 20 cases of C9FTD/ALS [17]. In the second, the Kings College London group compares the neuropathology among 14 C9FTD/ALS cases and 18 FTD/ALS cases without the expanded repeat [1]. The pathology has been previously reported, both in a subset of cases not known to be linked to chromosome 9p, and in cases with known 9p linkage, but the two articles in this issue of *Acta Neuropathologica* are the first to link specific pathologic findings to cases of C9FTD/ALS [1, 5, 12, 13, 17, 19].

The C9FTD/ALS cases in both papers include cases of FTLD-TDP, FTLD-MND, and ALS. There are fewer FTLD-MND cases in the London group's paper—1 of 14 cases compared to 8 of 20 cases in the Mayo Jacksonville report, but that may simply reflect the difference in how this group determines the final neuropathologic diagnosis, because four of their "ALS/MND" cases have TDP-43 positive inclusions in frontal cortex and dentate gyrus. The Mayo paper sub-typed the FTLD-TDP pathology and interestingly found that some of the C9FTD/ALS cases had Mackenzie type 1 pathology. Because the C9-linked cases reported thus far have been only Mackenzie type 3, the pathology of C9FTD/ALS, therefore, seems to be more heterogeneous than previously thought.

Some of the pathology appears to be unique to C9FTD/ALS. All 20 of the Mayo Jacksonville and all 14 of the Kings College London cases have ubiquitin + (Mayo) or p62+ (Kings)/TDP-43-inclusions in cerebellar granular neurons that are not found in cases without

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The London group also screened hippocampus and neocortex with p62 and TDP-43. Similar to their previous report, all C9 cases also had abundant unusual "star"-shaped inclusions in the hippocampal pyramidal layer, and to a lesser extent in frontal and temporal cortex, along with occasional NIIs in both regions, that were p62 positive and mostly negative for TDP-43 [13]. The Mayo group used ubiquitin on cerebellum, but not on hippocampus or neo-cortex, and therefore did not report these inclusions. Presumably, the star-shaped inclusions, like the cerebellar inclusions, are also positive with ubiquitin, although it has been reported that p62 is incorporated into inclusions before ubiquitin [15]. In C9 cases, this remains undetermined, since it appears that in their previous report the London group used both p62 and ubiquitin in the cerebellum but only p62 in the hippocampus and cortex [13]. Importantly, the Kings College group previously reported that the star-shaped and cerebellar granular neuron inclusions were negative for neurofilament, alpha-internexin, and FUS, and their current paper adds optineurin and the C9ORF72 protein to this list [1, 13].

On the other hand, the Mayo group found fine granular TDP-43 positivity in hippocampal CA2–CA4 regions, which they call "synaptic" labeling, and fine TDP-43 positive neurites predominantly in CA1. Although the Kings College group described very occasional p62 positive dystrophic neurites in the hippocampus in 9 of 14 cases, 5 of which had occasional TDP positivity, it appears that they did not find the same fine neurites in CA1 and synaptic labeling in CA2–4 with TDP-43 immunohistochemistry. This may be due to differences in immunostaining protocols, as TDP-43 results vary greatly, depending mostly on the antigen retrieval methodology.

The p62+/TDP-inclusions in the cerebellar granular layer, hippocampal pyramidal neurons, and neocortex signify that there is an as yet undiscovered protein involved in c9FTD/ALS. Likely suspects, neurofilament, alpha-internexin, and FUS, were previously excluded, and now optineurin and C9ORF72 are also excluded [1, 13]. At the rapid rate new knowledge is currently learned in this field, it will likely soon be identified. What will the identity of this new protein reveal about the molecular process underlying FTLD and ALS? The proteins involved in both ALS and FTLD now include ubiquitin, p62, TDP-43, FUS, ubiquilin2, and VCP. Ubiquitin, ubiquilin2, p62, and VCP are involved in ubiquitin-mediated protein degradation and TDP-43 and FUS bind to DNA, RNA, and protein and are involved in exon skipping, gene and splicing regulation, and transcription repression [7, 10, 14, 16, 18, 22]. Mutations in p62, also known as sequestosome 1, were recently reported in familial and sporadic ALS [9]. While the function of the C9ORF72 protein is unknown, the fact that the mutation is in an intron adds C9FTD/ALS to the class of noncoding repeat expansion disorders that include myotonic dystrophies DM1 and DM2, fragile-X-associated tremor/ ataxia syndrome FXTAS, and several spinocerebellar ataxias (SCA8, SCA10, SCA31, SCA36) [8]. Abnormal RNA metabolism has been suggested in both ALS and FTD and the presence of RNA nuclear foci in C9FTD/ALS brains suggests that alternative mRNA splicing is dysregulated. RNA misprocessing may be the underlying mechanism in the pathogenesis of C9FTD/ALS, and understanding this mechanism can lead to new therapeutic targets [4, 8]. New findings, such as the clinical and pathologic details of C9FTD/ALS presented in the two papers featured in this issue of Acta Neuropathologica, reveal the increasing complexity of the overlap between FTD and ALS [1, 17].

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