

Daunorubicin-TiO₂ nanocomposites as a “smart” pH-responsive drug delivery system

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Abstract: Daunorubicin (DNR) has a broad spectrum of anticancer activity, but is limited in clinical application due to its serious side effects. The aim of this study was to explore a novel “smart” pH-responsive drug delivery system (DDS) based on titanium dioxide (TiO₂) nanoparticles for its potential in enabling more intelligent controlled release and enhancing chemotherapeutic efficiency of DNR. DNR was loaded onto TiO₂ nanoparticles by forming complexes with transition metal titanium to construct DNR-TiO₂ nanocomposites as a DDS. DNR was released from the DDS much more rapidly at pH 5.0 and 6.0 than at pH 7.4, which is a desirable characteristic for tumor-targeted drug delivery. DNR-TiO₂ nanocomposites induced remarkable improvement in anticancer activity, as demonstrated by flow cytometry, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay, and nuclear 4',6-diamidino-2-phenylindole staining. Furthermore, the possible signaling pathway was explored by western blot. For instance, in human leukemia K562 cells, it was demonstrated that DNR-TiO₂ nanocomposites increase intracellular concentration of DNR and enhance its anticancer efficiency by inducing apoptosis in a caspase-dependent manner, indicating that DNR-TiO₂ nanocomposites could act as an efficient DDS importing DNR into target cancer cells. These findings suggest that “smart” DNR delivery strategy is a promising approach to cancer therapy.

Keywords: drug delivery system, daunorubicin, pH-responsive, TiO₂ nanoparticles, cancer, apoptosis

Introduction

With a broad spectrum of anticancer activity, daunorubicin (DNR) remains the first-line treatment for hematological malignancies and solid tumors. Unfortunately, its therapeutic effect is limited due to its serious side effects, including congestive heart failure,¹ which presents a challenge for DNR in clinical application. Therefore, efforts should be made to develop new delivery techniques that reduce its side effects by altering its biodistribution, enhancing its deposition in tumor sites, and improving its therapeutic efficacy.

Recently, a so-called “smart” stimuli-sensitive drug delivery system (DDS) that responds to certain internal or external stimuli, such as pH, temperature, and magnetism, represents a promising approach to improving the efficacy of cancer chemotherapy.^{2,3} It is well known that the extracellular pH of tumors is slightly more acidic than that of blood and normal tissue. Moreover, numerous pH gradients exist in the intracellular endocytic compartment (pH = 4.5–6.5) where cells uptake a DDS via an endocytosis process.⁴ Therefore, an ideal anticancer DDS should retain the drug at pH 7.4 but be able to quickly release it at a relatively lower pH (eg, pH 6.0 or 5.0). An important

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breakthrough in this field is the preparation of a new class of nanosized DDS which can be internalized by tumor cells through the endocytosis pathway to avoid undesired drug release during drug transportation in blood circulation, thus improving effective release of the anticancer drug in tumor tissue or within tumor cells.⁵

Furthermore, there has been growing interest in metal nanomaterials as anticancer drug carriers recently, which represents a promising approach to tumors, although it is still under investigation.^{6,7} Barick et al reported a method for entrapping drugs at sites capable of complexing with transition metal ions and suggested that drug release is dependent on the pH of the medium and nature of the materials which encapsulate the drug.⁷ It is also suggested that this general strategy could be readily extended to other inorganic carriers.⁷

In view of these developments, the aim of this paper was to explore a novel “smart” pH-responsive DDS based on titanium dioxide (TiO₂) nanoparticles that is chemically stable, environmentally friendly, and noncytotoxic.^{8,9} This DDS could provide a simple and easy way to enable intelligently controlled release of DNR and enhance its chemotherapeutic efficiency. To the best of the authors’ knowledge, no study has been carried out on TiO₂ nanoparticles for “smart” stimuli-sensitive DDS and is investigated here for the first time. In this study, DNR was loaded on TiO₂ nanoparticles to construct DNR-TiO₂ nanocomposites as a DDS. Loading efficiency and influence of pH value on drug release behavior of the DDS were investigated by high-performance liquid chromatography. Additionally, tumor cellular uptake and cytotoxicity of the DDS were evaluated in K562 leukemia cells.

Materials and methods

Chemicals and apparatus

DNR was purchased from Pharmacia Italia SpA (Nerviano, Italy), and its stock solution was freshly prepared and stored in the dark at 4°C. Phosphate buffer solution (PBS) with different pH value (pH 7.4, 6.0, 5.0) was prepared with double distilled water. TiO₂ nanoparticles (P25; Evonik Industries AG, Essen, Germany) were suspended in sterilized PBS. Caspase 3 antibody, β-actin antibody, and horseradish peroxidase-conjugated immunoglobulin G antibody were obtained from Nanjing KeyGen Biotech Co, Ltd, (Nanjing, China). 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) and 4,6-Diamidino-2-phenylindole (DAPI) were obtained from Sigma-Aldrich Corporation (St Louis, MO) and stored in the dark. All other reagents were of analytical grades. The images of transmission electron microscope, electron diffraction, and high-resolution transmission electron

microscope were obtained by a JEM-2100 transmission electron microscope (JEOL Ltd, Tokyo, Japan). Fluorescent microscopic images were shot on an Olympus IX51 inverted microscope (Olympus Corporation, Tokyo, Japan). Optical density at 492 nm was recorded by multiwell spectrophotometer reader (MK3; Thermo Labsystems, Vantaa, Finland).

Construction of DNR-TiO₂ nanocomposites

For a typical drug loading experiment, 2 mL aqueous solution of DNR (2 mg/mL) was added to 1 mL aqueous suspension of TiO₂ nanoparticles (10 mg/mL). The above reaction mixture was continuously agitated with a stirrer at 50 g and kept overnight in the dark to construct nanocomposites of DNR-loaded TiO₂ nanoparticles which acted as the DDS. DNR-loaded TiO₂ nanoparticles were separated from the free-standing drug molecules through centrifugation at 5000 g for 20 minutes and the supernatant was determined by high-performance liquid chromatography, allowing estimation of drug encapsulation efficiency and loading efficiency. Encapsulation efficiency and loading efficiency were calculated by the following equations:

$$\text{Encapsulation efficiency (\%)} = \left(\frac{\text{Amount of drug in DNR-TiO}_2 \text{ nanocomposites}}{\text{initial amount of drug}} \right) \times 100$$

$$\text{Loading efficiency (\%)} = \left(\frac{\text{Amount of drug in DNR-TiO}_2 \text{ nanocomposites}}{\text{amount of DNR-TiO}_2 \text{ nanocomposites}} \right) \times 100$$

Drug release

Release behavior of DNR from DNR-TiO₂ nanocomposites was investigated at pH 5.0 (approximate pH in endosomes or lysosomes), pH 6.0 (pH of the environment around the tumor), and pH 7.4 (pH of physiological blood). DNR-loaded TiO₂ nanoparticles (20 mg) were dispersed in PBS (pH 7.4, 5 mL) and transferred into a dialysis bag (Spectra/Por®; Spectrum Laboratories, Inc, Rancho Dominguez, CA). The dialysis bag was then immersed in 95 mL PBS at pH 5.0, 6.0, or 7.4. The release medium was continuously agitated with a stirrer at 50 g and 37°C. At predetermined time intervals, 2 mL of the external medium was collected and replaced with the same fresh PBS. The amount of released DNR in the medium was then determined by high-performance liquid chromatography (LC-310; Skyray Instrument, Nanjing, China).

Cell culture

K562 leukemia cells were obtained from Institute of Hematology, Chinese Academy of Medical Sciences (Beijing, China)

and cultured in Gibco® Roswell Park Memorial Institute 1640 medium (Life Technologies, Grand Island, NY) supplemented with 10% heat-inactivated fetal bovine serum (Sigma-Aldrich), 100 U/mL penicillin (Sigma-Aldrich), and 100 µg/mL streptomycin (Sigma-Aldrich) at 37.0°C in humidified air with 5% carbon dioxide. Cells were in log phase prior to the following experiments to minimize experimental variations which occur with proliferative status.

Cell internalization studies

Cells (1×10^5 /mL) were seeded in six-well culture plates and grown overnight. They were then treated with either DNR or DNR-TiO₂ nanocomposites (DNR concentration 0.5 µg/mL) for 6 hours. To visualize the cellular uptake of DNR with different treatments, the cells were examined by fluorescent microscopy (λ_{ex} 488 nm, λ_{em} 515 nm) after incubation for 6 hours. Furthermore, the cells were resuspended in PBS after being washed three times. DNR uptake was analyzed by FACSCalibur™ flow cytometer (BD Biosciences, San Jose, CA); the gate was arbitrarily set for the detection of green fluorescent DNR.

Assay of anticancer activity

Cytotoxicity of TiO₂ nanoparticles, DNR, and DNR-TiO₂ nanocomposites was studied in leukemia K562 cells with MTT assays. Cells (1×10^5 /mL) were seeded in 96-well plates and incubated for 24 hours. Growth medium was then replaced with 200 µL of the preprepared medium containing either free DNR or DNR-TiO₂ nanocomposites (DNR concentration 0, 0.125, 0.25, 0.5, and 1 µg/mL). Cells were treated by TiO₂ nanoparticles alone as well to evaluate its cytotoxicity. Cells without any treatment were used as the control group. Cells were further incubated for 48 hours, and relative anticancer activity was assessed with MTT assays. After 48 hours, MTT assay was carried out. In brief, MTT solutions were added after treatment and incubated for an additional 4 hours. Dimethyl sulfoxide (Sinopharm Chemical Reagent Co Ltd, Shanghai, China) was added to solubilize the formazan crystal, and optical density 492 nm was recorded. Cell viability (%) was calculated as follows:

$$\text{Optical density}_{492 \text{ nm in test cells}} / \text{Optical density}_{492 \text{ nm in control cells}} \times 100$$

DAPI staining

Cells were treated with the above methods for 48 hours, and were then fixed with 4% polyoxymethylene prior to washing with PBS. The washed cells were then stained with 1 mg/mL DAPI for 15 minutes in the dark. Staining images were recorded with fluorescent microscope.

Western blot analysis

After the different treatments, caspase 3 expression was detected by western blot. In brief, total protein was isolated, subjected to sodium dodecyl sulfate polyacrylamide gel electrophoresis, and transferred to a polyvinylidene fluoride membrane. After being blocked, the membrane was incubated with primary polyclonal antibodies either anti-caspase 3 or anti-β-actin overnight at 4°C, and subsequently incubated with horseradish peroxidase-conjugated immunoglobulin G antibody as the secondary antibody for 1 hour at room temperature. The protein bands were detected by an enhanced electrochemiluminescence detection system (ECL system, Amersham, UK). After normalization by corresponding β-actin expression, protein expression levels of caspase 3 were determined by densitometry scans.

Statistical analysis

All data are presented as mean ± standard deviation. F-test was used for significance testing, and $P < 0.05$ was considered to be statistically significant. All tests were performed using IBM SPSS software (v 13.0; SPSS Inc, Chicago, IL).

Results and discussion

Characterization of TiO₂ nanoparticles

Transmission electron microscopic images of TiO₂ nanoparticles are shown in Figure 1. Observed TiO₂ nanoparticles had a spherical shape with a diameter of about 25 nm (Figure 1A). The particles had dimensions suitable for escaping rapid renal excretion, as well as avoiding components of the reticular endothelial system. Thus, potentially passive targeting of drugs to tumors was facilitated via enhanced permeation and retention effect, and drug accumulation in tumor cells after endocytosis was increased.⁴ Also, the selected-area electron diffraction (Figure 1B) patterns confirmed that TiO₂ nanoparticles were single crystalline with a

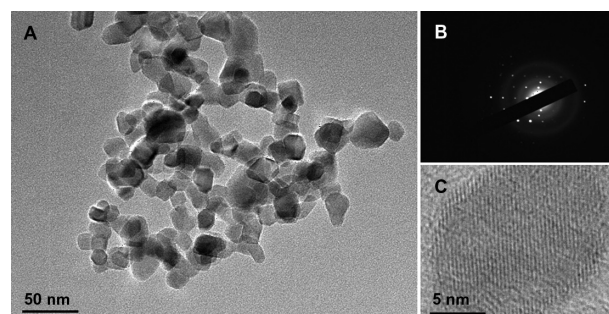


Figure 1 Transmission electron microscopic image (A), electron diffraction patterns (B), and high-resolution transmission electron microscopic image (C) of titanium dioxide nanoparticles.

hexagonal-wurtzite structure. The fine microstructures of TiO_2 nanoparticles were further characterized by high-resolution transmission electron microscope (Figure 1C), which showed that TiO_2 nanoparticles were highly crystalline.

DNR- TiO_2 nanocomposites as a DDS

A previous study showed that the analog of DNR, doxorubicin, has three potential metal binding sites.⁷ One is the nitrogen atom in the sugar moiety and the other two are the chelating sites of quinone and phenolic oxygens on both sides of the anthracycline aromatic moiety.⁷ In the present study, the capacity of TiO_2 nanoparticles as a drug carrier for loading DNR was explored. Color change of pure DNR from red to orange was observed during the loading process, indicating that DNR molecules could form complexes with TiO_2 nanoparticles. The more likely metal binding sites of DNR were deduced to be the two keto-phenolate functions: $\text{C}=\text{O}$ and $\text{C}-\text{O}^-$ (six-membered chelate), due to the low affinity of metal ions for the nitrogen atom donor. In addition, DNR, which is positively charged, can be easily self-assembled onto the surface of TiO_2 nanoparticles through electrostatic interaction as the relative surface of TiO_2 nanoparticles is negatively charged in pH 7.4 aqueous solutions.^{8,10} Thus, the

authors believe that DNR was absorbed onto the TiO_2 nanoparticles by forming complexes with transition metal titanium and by electrostatic interaction. Figure 2 shows a proposed schematic representation of DNR loading onto TiO_2 nanoparticles through the formation of DNR- TiO_2 nanocomposites as a DDS. Encapsulation efficiency and loading efficiency of DNR-loaded TiO_2 nanoparticles were assessed and calculated as $65.46\% \pm 6.82\%$ and $20.63\% \pm 3.55\%$, respectively. Results show that DNR-loaded TiO_2 nanoparticles may efficiently act as anticancer drug delivery carrier.

pH-triggered drug release behavior

As shown in Figure 3, release of drug molecules depended on pH of the medium and release time. Drug release at pH 7.4 was slow and sustained, with release ratio at about 18% within 48 hours. However, at lower pH, DNR release rate was much faster, with approximately 67% (pH 6.0) and 86% (pH 5.0) of the drug released within 48 hours. Protonation of the drug (dissociation of DNR- TiO_2 nanocomposites) occurred at lower pH, which released chemisorbed drug molecules into the medium. Furthermore, surface charges of TiO_2 nanoparticles turned positive at lower pH, which blunted the electrostatic interaction of DNR and TiO_2 nanoparticles

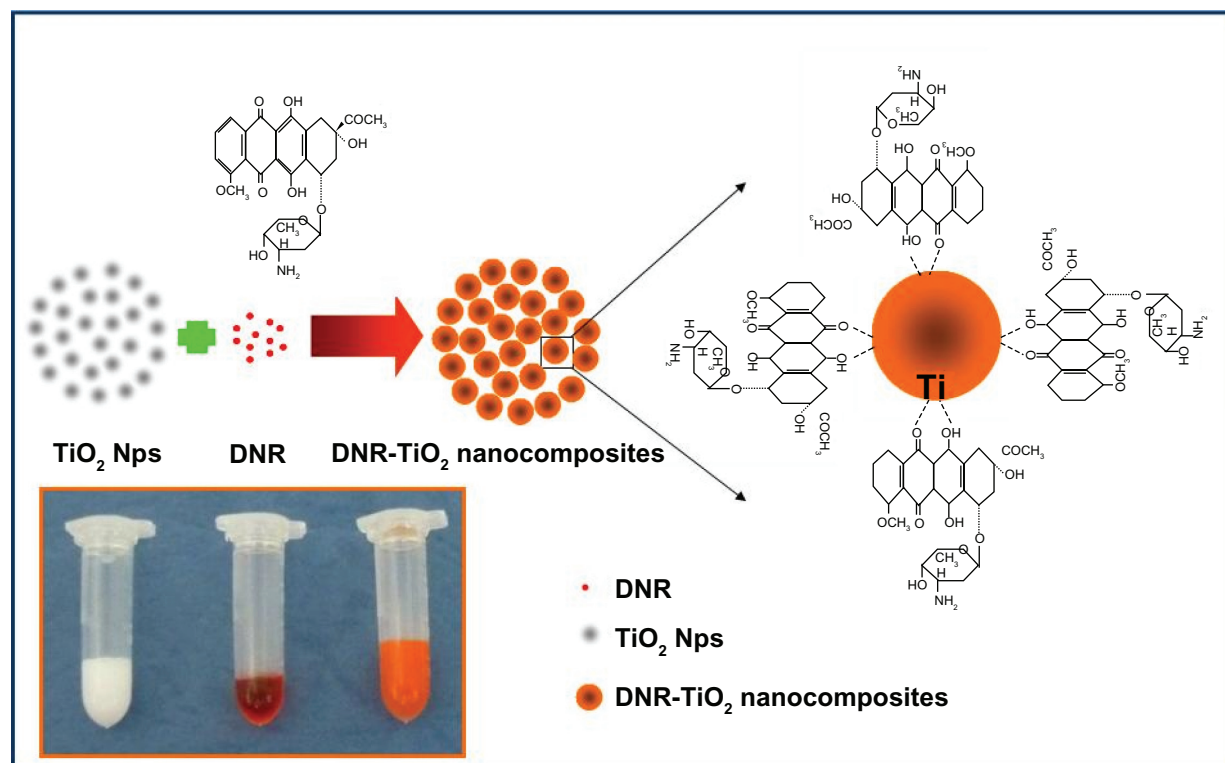


Figure 2 Schematic representation and photographic image (inset) of daunorubicin loading onto titanium dioxide nanoparticles through the formation of daunorubicin-titanium dioxide nanocomposites.

Abbreviations: DNR, daunorubicin; Nps, nanoparticles; TiO_2 , titanium dioxide.

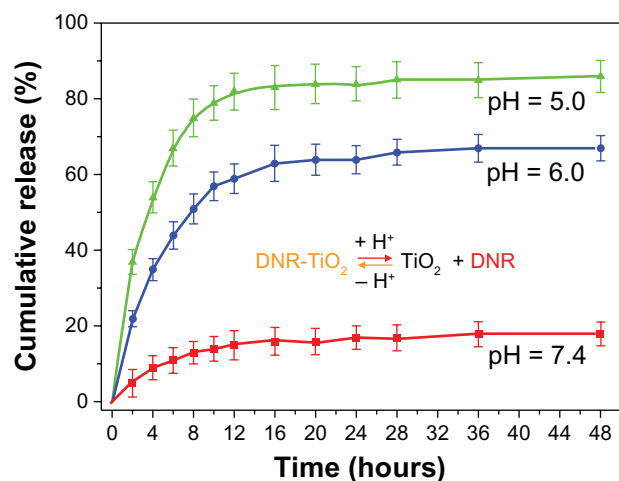


Figure 3 In vitro daunorubicin release behavior at pH 7.4, 6.0, and 5.0. **Abbreviations:** DNR, daunorubicin; DNR-TiO₂, daunorubicin-titanium dioxide nanocomposites; TiO₂, titanium dioxide.

and facilitated the drug release process. These results demonstrate pH-triggered drug release behavior from the novel DDS, which is of particular interest. It may be hypothesized that most DNR will remain in the carrier for a considerable time period at normal physiological conditions (pH 7.4), indicating the potential for prolonged DNR retention time in blood circulation and thereby greatly reducing the side effects to normal tissues. On the other hand, once DNR-loaded TiO₂ nanoparticles are taken up by tumor cells via endocytotic process, a faster release may occur at lower local pH, ie, surrounding the tumor site or inside the endosome and lysosome of tumor cells, leading to significant improvement in cancer treatment efficacy.

Cellular uptake

Fluorescent DNR allows facile observation of the cellular uptake of free DNR and DNR-loaded TiO₂ nanoparticles by K562 cells, quantitatively detected by flow cytometry without additional markers. Figure 4A illustrates the typical results of fluorescent microscopy of intracellular DNR. The internalization of DNR loaded onto TiO₂ nanoparticles (Figure 4A, right panel) was much higher after 6 hours of incubation compared with the free DNR (Figure 4A, left panel), resulting in stronger fluorescence intensity in K562 cells. The relative intracellular fluorescence intensity of DNR analyzed by flow cytometry allows quantitative comparison of the cases mentioned above (Figure 4B). Intracellular fluorescence intensity for the group of DNR-TiO₂ nanocomposites produced 65% ± 3.7% enhancements compared to the group of free DNR. These results clearly show that DNR-TiO₂ nanocomposites as a DDS increased

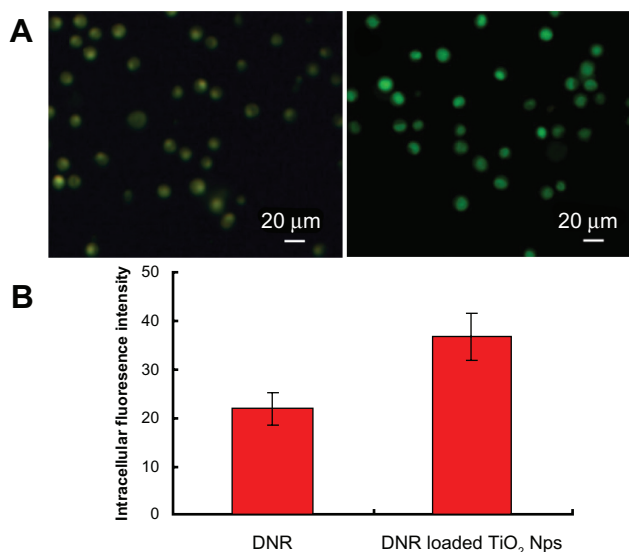


Figure 4 Fluorescent microscopic images of K562 cells (A) treated with free daunorubicin (left panel) and titanium dioxide nanoparticles loading daunorubicin as a drug delivery system (right panel). Comparison of the respective average intracellular fluorescence intensity is also shown (B). Concentrations of daunorubicin and titanium dioxide nanoparticles are 0.5 µg/mL and 10 µg/mL, respectively. **Note:** Data expressed as mean ± standard deviation (n = 3). **Abbreviations:** DNR, daunorubicin; Nps, nanoparticles; TiO₂, titanium dioxide.

the cellular uptake of DNR substantially. TiO₂ nanoparticles demonstrated significant properties of a drug carrier, which represents a promising approach to cancer therapy.

Anticancer activity in vitro

A common characteristic of a nanoparticle-based DDS is the rapid internalization and intracellular localization into acidic endosomes and/or lysosomes of cells.^{11,12} Due to this common pathway of cellular uptake, there has been significant interest in methods to engineer acid-triggered responses from DDS for rapid intracellular release of therapeutics.¹³ To further explore the anticancer efficiency of the novel pH-responsive DDS based on TiO₂ nanoparticles for DNR, K562 cells were cultured with free DNR at different concentrations (0, 0.125, 0.25, 0.5, and 1 µg/mL) and DNR-TiO₂ nanocomposites with equivalent DNR concentration for 48 hours. Cytotoxicity results were estimated by MTT assay (Figure 5).

A cytotoxicity test of the nanomaterial is the first-level evaluation before its biomedical applications. When treated by TiO₂ nanoparticles, about 95% of the cells survived (Figure 5, blue line), which is consistent with a previous report.¹⁰ The lack of cytotoxicity of TiO₂ nanoparticles thus ensures a wide potential range of applications in the field of biomedical science and cancer therapy. Compared with DNR alone (Figure 5, green line), viability of K562 cells treated by DNR-TiO₂ nanocomposites obviously decreases (Figure 5, black line). Results also indicate that with increasing concentrations

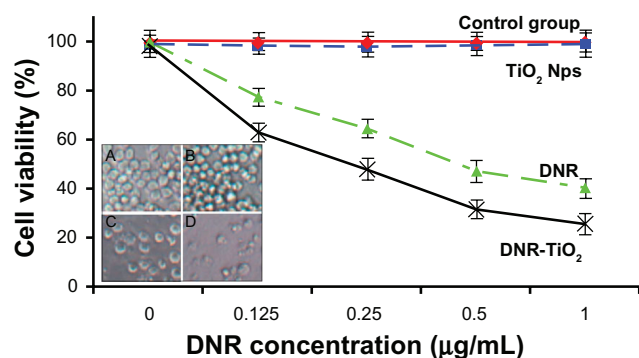


Figure 5 Cytotoxic effect of daunorubicin or daunorubicin-titanium dioxide nanocomposites on K562 leukemia cells. Microscopic images of K562 cells after different treatments for 48 hours are shown inset: (A) untreated cells as control, (B) titanium dioxide nanoparticles, (C) daunorubicin alone, and (D) daunorubicin-titanium dioxide nanocomposites. Concentrations of daunorubicin and titanium dioxide nanoparticles are 1 µmol/L and 10 µg/mL, respectively.

Note: Data expressed as mean ± standard deviation (n = 3).

Abbreviations: DNR, daunorubicin; DNR-TiO₂, daunorubicin-titanium dioxide nanocomposites; Nps, nanoparticles; TiO₂, titanium dioxide.

of DNR, lethality increased, suggesting a dose-dependent effect in vitro. Increased cytotoxicity may be due to improved DNR cellular uptake by DNR-TiO₂ nanocomposites (as illustrated above) and to the induced release of DNR from TiO₂.

The process of controlled release is of particular interest in achieving DNR delivery with TiO₂. At normal physiological conditions (pH 7.4), it is expected that most DNR conjugated with TiO₂ will remain in the carrier for a considerable time period. As well documented, a nanoparticle-based DDS could be internalized through the endocytosis pathway.¹⁴ In the endocytic compartment, pH values range 4.5–6.5;¹⁵ and at this low pH, protonation of the drug occurs.

Protonation will then trigger the release of chemisorbed drug molecules and turn surface charges of TiO₂ nanoparticles to positive, blunting the electrostatic interaction of DNR and TiO₂ nanoparticles which facilitates the drug release process. Consequently, a faster release of DNR from DNR-TiO₂ nanocomposites occurs once the carrier is taken up by the tumor cells via endocytic process. Therefore, a sufficiently high concentration of DNR can be generated within a reasonably short period of time, thereby greatly promoting the cell-killing effects efficiently. Considering that a high dose of DNR may lead to serious side effects, especially congestive heart failure,¹ DNR-TiO₂ nanocomposites as a DDS improves therapeutic efficacy without a high usage of DNR, thus potentially minimizing its toxic side effects.

As shown in the inset of Figure 5, optical microscopic observations confirm MTT assay results. K562 cells without any treatment attached to the plate with normal spherical shape (Figure 5A). Notably, cells treated with TiO₂ nanoparticles had no morphological changes (Figure 5B). It is obvious that DNR (Figure 5C) and the even more effective DNR-TiO₂ nanocomposites (Figure 5D) could kill more cancer cells and cause more significant morphological changes, indicating an increased probability in cell death.

Morphologic characterization of apoptosis

To explore the mechanism of distinct improvement in anticancer activity induced by the novel pH-responsive DDS, nuclear DAPI staining was performed. To confirm the existence of

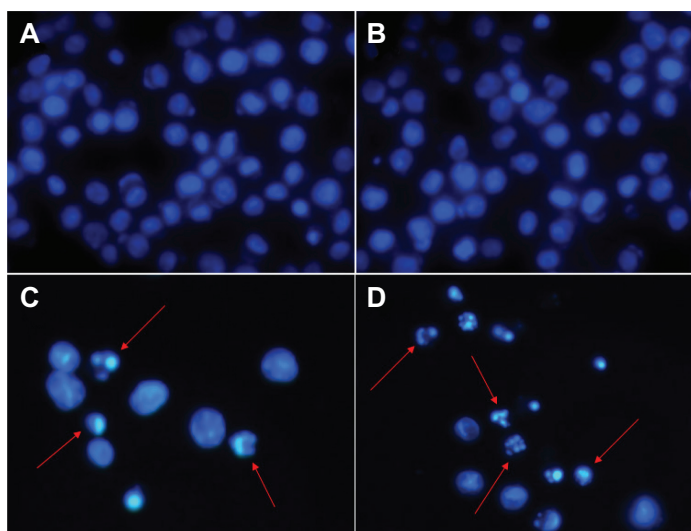


Figure 6 Nuclear morphologic changes of K562 leukemia cells after different treatments for 48 hours: (A) untreated cells as control, (B) titanium dioxide nanoparticles, (C) daunorubicin alone, and (D) daunorubicin-titanium dioxide nanocomposites. Concentrations of daunorubicin and titanium dioxide nanoparticles are 0.5 µg/mL and 10 µg/mL, respectively.

Notes: Magnification: ×400. Arrows indicate cells with apoptotic nuclear condensation and fragmentation.

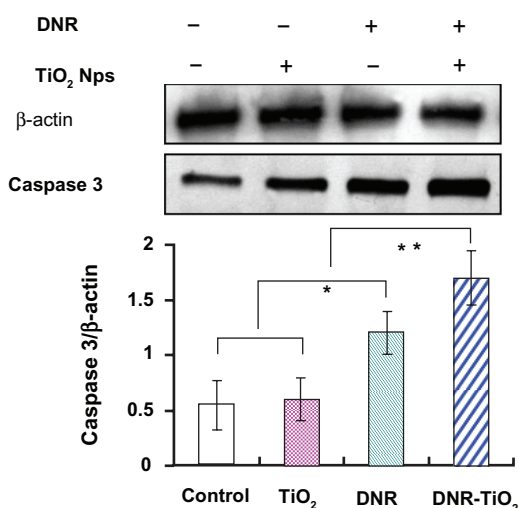


Figure 7 Expression of caspase 3 in K562 cells by western blotting analysis. After normalization by corresponding β -actin expression, protein expression levels of caspase 3 were determined by densitometry scans to obtain quantitative data.

Note: Data expressed as mean \pm standard deviation ($n = 3$).

Abbreviations: DNR, daunorubicin; DNR-TiO₂, daunorubicin-titanium dioxide nanocomposites; Nps, nanoparticles; TiO₂, titanium dioxide.

apoptosis, morphological changes of the cells were studied. Evaluation of normal or apoptotic cells depends on morphological characterization. Normal nuclei (smooth nuclear) and apoptotic nuclei (condensed or fragmented chromatin) were easily distinguished. As shown in Figure 6, nuclear morphology analysis showed characteristic apoptotic changes, such as chromatin condensation, convoluted nuclei with cavitations, fragmentation of the nucleus, and apoptotic bodies in K562 cells

after treatment with DNR and DNR-TiO₂ nanocomposites. There was almost no apoptotic evidence in the control group (Figure 6A) and TiO₂ group (Figure 6B). When cells were treated with DNR-TiO₂ nanocomposites (Figure 6D), typical apoptotic morphology was more apparent than that of cells treated with DNR alone (Figure 6C). These findings strongly indicate that DNR-TiO₂ nanocomposites killed the cancer cells by inducing apoptosis rather than necrosis.

Caspase 3 activity during induced apoptosis

Nuclear DNA is the main subcellular site of DNR action for its anticancer activity. DNR can efficiently accumulate in nucleus, intercalate DNA, and act as a cytostatic and apoptotic agent against tumor cells. To explore the possible signaling pathways through which DNR-TiO₂ nanocomposites induced distinct improvement in anticancer activity, the changes in expression levels of the apoptosis-regulating protein, caspase 3, were examined by western blot. As shown in Figure 7, when K562 cells were treated with DNR and DNR-TiO₂ nanocomposites for 48 hours, levels of caspase 3 protein were significantly upregulated compared to control group. Meanwhile, the upregulated levels of caspase 3 protein in DNR-TiO₂ nanocomposites group were slightly higher than those in DNR group ($P < 0.05$). However, the caspase 3 protein was not obviously altered when the cells were treated with TiO₂ nanoparticles ($P > 0.05$). Caspase activation is generally considered

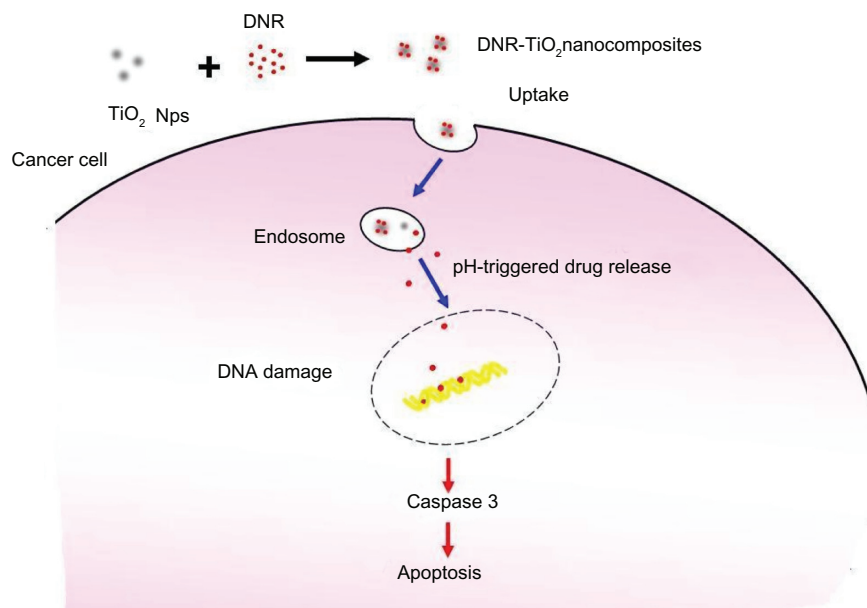


Figure 8 Schematic illustration of the possible process of distinct improvement in anticancer activity by the novel pH-responsive drug delivery system based on titanium dioxide nanoparticles for daunorubicin.

Abbreviations: DNR, daunorubicin; Nps, nanoparticles; DNR-TiO₂, daunorubicin-titanium dioxide nanocomposites; TiO₂, titanium dioxide.

to be a key hallmark of apoptosis, and caspase 3 is the main effector caspase that is involved in apoptosis.¹⁶ These results indicate that the studied DDS induces distinct improvement in anticancer activity in a caspase-dependent manner.

Based on the above studies, Figure 8 schematically illustrates the possible processes that lead to the distinct improvement in anticancer activity. DNR can be easily self-assembled onto the surface of TiO₂ nanoparticles by forming complexes with transition metal titanium to construct DNR-TiO₂ nanocomposites as a DDS. Drug release shows strong dependence on pH value, which is higher at lower pH. Thereby, TiO₂ nanoparticles load an amount of DNR molecules and carry them into cells, increasing intracellular concentration of DNR dramatically, and thus enhancing the suppression of cancer cell proliferation. The therapeutic concentration inside cancer cells is increased, which enhances the efficiency of drugs per dosage and, accordingly, solves some of the limitations of anticancer drugs, thereby exhibiting tremendous application potential in cancer therapy. Finally, apoptosis, a preferred mode of killing cancer cells in cancer therapy, is induced synergistically, resulting in a distinct improvement in anticancer activity.

Conclusion

In the present study, a pH-responsive nanocomposite composed of self-assembled DNR and TiO₂ nanoparticles was developed for a “smart” stimuli-sensitive DDS. Drug release from the DDS was significantly accelerated by decreasing pH from 7.4 to 5.0, which is of particular interest to cancer therapy due to the acidic extracellular tumor environment and intracellular endosomal and/or lysosomal compartments. DNR-TiO₂ nanocomposites increased the accumulation of DNR in tumor cells and induced caspase-dependent apoptosis, enhancing anticancer activity and potentially minimizing the toxic side effects of DNR. All these characteristics demonstrate that the novel pH-responsive DDS based on TiO₂ nanoparticles for DNR is a promising strategy for clinical practice.

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Disclosure

The authors report no conflicts of interest in this work.

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