

First Detection of *bla*_{IMI-2} Gene in a Clinical *Escherichia coli* Strain

The dissemination of carbapenem-resistant *Enterobacteriaceae* is increasing worldwide in the last decade, mainly due to the acquisition of beta-lactamase genes encoding carbapenemases. Among class A carbapenemases, KPC is widespread (14); however, IMI enzymes have been described so far only in *Enterobacter* genus (1, 10, 15). The detected *bla*_{IMI} genes are linked to a gene encoding a LysR-type transcriptional regulator, and as previously reported, the *bla*_{IMI-1} gene is located in the chromosome of *Enterobacter cloacae* (10), whereas *bla*_{IMI-2} is related to plasmids (1, 15). We report here the first description of a *bla*_{IMI-2}-positive *Escherichia coli* strain.

(Part of this study was presented at the 21st European Congress of Clinical Microbiology and Infectious Diseases Congress, Milan, Italy, 7 to 10 May 2011.)

A carbapenem-resistant *E. coli* strain, W635, was recovered from a blood sample of an elderly oncologic patient (without any history of travel) who was admitted with sepsis and treated with piperacillin-tazobactam at a Spanish hospital in 2010. *E. coli* W635 was ascribed to a new sequence type registered as ST1998 (<http://mlst.ucc.ie/mlst/dbs/Ecoli>) and showed resistance to imipenem (IPM), meropenem (MEM), ertapenem (ETP), doripenem, ampicillin (AMP), ticarcillin, amoxicillin-clavulanic acid (AMC), cephalothin, streptomycin, nalidixic acid, ciprofloxacin, norfloxacin, sulfonamides, and trimethoprim and intermediate resistance to aztreonam and chloramphenicol by the CLSI disk diffusion method (3). A class A carbapenemase phenotype was demonstrated in this strain by double-disk synergy test (5, 9).

After multiplex PCR (8) and subsequent sequencing, a partial sequence of the *bla*_{IMI-2} gene was detected in *E. coli* W635. To gain insight into the *bla*_{IMI-2} gene and its surrounding structure, the flanking regions were amplified by PCR using specific primers designed in this work (according to GenBank accession number [AY780889](http://www.ncbi.nlm.nih.gov/GenBank/AY780889)). The LysR-type regulator gene (*bla*_{IMI-2R}) was found upstream of the *bla*_{IMI-2} gene, and their genetic environment was studied by inverse PCR using PvuII and BglII restriction enzymes. Sequence analysis revealed a total fragment of 6,184 bp that was deposited in GenBank database with the accession number [JN412066](http://www.ncbi.nlm.nih.gov/GenBank/JN412066). A new insertion sequence of 1,321 bp, designed ISEc36 by ISFinder (<http://www-is.biotoul.fr/>), was detected upstream of the *bla*_{IMI-2R} gene. This IS belongs to the IS3 family and IS2 group, and it exhibits an identity of 92% with respect to the ISEc27 sequence (GenBank accession number [AY857617](http://www.ncbi.nlm.nih.gov/GenBank/AY857617)).

Testing for the presence of genes implicated in other antimicrobial resistances (beta-lactams, aminoglycosides, quinolones, trimethoprim, and sulfonamides) and the study of mutations in *gyrA* and *parC* genes, as well as the characterization of integrons and *sul2* gene environment, were performed by PCR and sequencing (4, 11, 13). Table 1 shows the MICs and genotypic results.

Two different transconjugants from W635 were obtained by mating experiments, using *E. coli* CSH26 as recipient and plates supplemented with either MEM (8 µg/ml) or AMP (50 µg/ml) and rifampin (100 µg/ml). Detection and typing of plasmids of *E. coli* W635 and transconjugant strains were carried out by PCR-based replicon typing (2, 6). Plasmids of strain W635 and its transconjugants belonging to incompatibility group I1 (IncI1) and F (IncF) were subtyped by plasmid multilocus sequence typing (7, 12). *E. coli* W635 contained the following typeable plasmids: IncI1 (ST26, CC-26), IncF (F43:A⁻:B⁻; Y2 variant:A⁻:B⁻), and ColE_{TP}. The location of *bla*_{IMI-2} in strain W635 and its transconjugants was studied by pulsed-field gel electrophoresis (PFGE)-S1 nuclease and PFGE-XbaI Southern blotting and hybridization (4) with *bla*_{IMI-2}, IncI1, IncF, and ColE_{TP} probes. The *bla*_{IMI-2} gene in *E. coli* W635 was detected in an IncF plasmid of approximately 48.5 kb. The size of this plasmid is smaller than found in previously reported results that described the *bla*_{IMI-2} gene located on transferable plasmids with sizes of 66 kb or 80 kb (1, 15).

This is the first report of a carbapenem-resistant *E. coli* strain carrying the class A carbapenemase IMI-2. The *bla*_{IMI-2} gene located in a conjugative plasmid and linked to mobile elements might significantly spread between different members of the *Enterobacteriaceae*, and therefore this emerging resistance mechanism should be tracked in the future.

Nucleotide sequence accession number. The nucleotide sequence of the novel genetic environment of *bla*_{IMI-2} gene determined in this study was included in the GenBank database with the accession number [JN412066](http://www.ncbi.nlm.nih.gov/GenBank/JN412066).

Published ahead of print 21 November 2011

Address correspondence to Yolanda Sáenz, ysaenz@riojasalud.es.

Copyright © 2012, American Society for Microbiology. All Rights Reserved.

doi:10.1128/AAC.05478-11

TABLE 1 Resistance phenotype, genotype, and genetic elements of *E. coli* strain W635, transconjugants (TC), and recipient strain^a

| Strain | MIC (µg/ml) of ^b : | | | | | | Resistance gene(s) | Amino acid change(s) detected | | Class 1 integron | Incompatibility group detected (size of plasmid) | | |
|-----------------|-------------------------------|-------|------|-------|------|--------|----------------------------------------------------------|-------------------------------|------------------|------------------|--------------------------------------------------|--------------------|---------------------------------------------------------------|
| | AMP | ATM | IPM | MEM | STR | TMP | | GyrA | ParC | | | | |
| W635 donor | >128 | 8 | 128 | 32 | 128 | >128 | <i>bla</i> _{TEM-1b} <i>bla</i> _{IMI-2} | <i>strA-strB</i> ^c | <i>sul1 sul2</i> | S83L, D87N | S80I | <i>dfrA1 aadA1</i> | IncI1 (97 kb), IncF (48.5 kb), ColE _{TP} (<10 kb) |
| TC1 | >128 | 8 | 128 | 16 | >128 | >128 | <i>bla</i> _{TEM-1b} <i>bla</i> _{IMI-2} | <i>strA-strB</i> | <i>sul1 sul2</i> | ND | ND | <i>dfrA1 aadA1</i> | IncI1/IncF (160 kb), ^d ColE _{TP} (<10 kb) |
| TC15 | >128 | 0.125 | 0.25 | 0.032 | >128 | >128 | <i>bla</i> _{TEM-1b} – | <i>strA-strB</i> | <i>sul1 sul2</i> | ND | ND | <i>dfrA1 aadA1</i> | IncI1 (97 kb) |
| CSH26 recipient | 16 | 0.125 | 0.25 | 0.032 | 4 | <0.125 | – | – | – | ND | ND | ND | ND |

^a ND, not determined; –, not detected.

^b AMP, ampicillin; ATM, aztreonam; IPM, imipenem; MEM, meropenem; STR, streptomycin; TMP, trimethoprim.

^c *strA-strB* genes were linked to the *sul2* gene (*repC-sul2-strA-strB-ISCR2*).

^d IncI1 and IncF were detected by hybridization in a plasmid of approximately 160 kb in the *bla*_{IMI-2}-positive TC1 transconjugant.

ACKNOWLEDGMENTS

We thank Patricia Siguier for analyzing the new ISEc36 (<http://www-is.biotoul.fr/>).

The study did not receive financial support from third parties. M. López has a fellowship from the Gobierno de La Rioja.

REFERENCES

1. Aubron C, Poirel L, Ash RJ, Nordmann P. 2005. Carbapenemase-producing *Enterobacteriaceae*, U.S. rivers. *Emerg. Infect. Dis.* 11:260–264.
2. Carattoli A, et al. 2005. Identification of plasmids by PCR-based replicon typing. *J. Microbiol. Methods* 63:219–228.
3. Clinical Laboratory Standards Institute. 2011. Performance standards for antimicrobial susceptibility testing. Twenty-First International Supplement. M100–S21. Clinical and Laboratory Standards Institute, Wayne, PA.
4. de Toro M, et al. 2010. In vivo selection of *aac(6′)-Ib-cr* and mutations in the *gyrA* gene in a clinical *qnrS1*-positive *Salmonella enterica* serovar Typhimurium DT104B strain recovered after fluoroquinolone treatment. *J. Antimicrob. Chemother.* 65:1945–1949.
5. Doi Y, et al. 2008. Simple disk-based method for detection of *Klebsiella pneumoniae* carbapenemase-type beta-lactamase by use of a boronic acid compound. *J. Clin. Microbiol.* 46:4083–4086.
6. García-Fernández A, Fortini D, Veldman K, Mevius D, Carattoli A. 2009. Characterization of plasmids harbouring *qnrS1*, *qnrB2* and *qnrB19* genes in *Salmonella*. *J. Antimicrob. Chemother.* 63:274–281.
7. García-Fernández A, et al. 2008. Multilocus sequence typing of IncII plasmids carrying extended-spectrum beta-lactamases in *Escherichia coli* and *Salmonella* of human and animal origin. *J. Antimicrob. Chemother.* 61:1229–1233.
8. Hong SG, Hong SK, Huh JY, Kang MS. 2010. Multiplex PCR for rapid detection of genes encoding class A carbapenemases. *Clin. Microbiol. Infect.* 16(suppl. s2):S552. doi:10.1111/j.1469-0691.2010.03239.x.
9. Nordmann P, Cuzon G, Naas T. 2009. The real threat of *Klebsiella pneumoniae* carbapenemase-producing bacteria. *Lancet Infect. Dis.* 9:228–236.
10. Rasmussen BA, et al. 1996. Characterization of IMI-1 beta-lactamase, a class A carbapenem-hydrolyzing enzyme from *Enterobacter cloacae*. *Antimicrob. Agents Chemother.* 40:2080–2086.
11. Sáenz Y, et al. 2004. Mechanisms of resistance in multiple-antibiotic-resistant *Escherichia coli* strains of human, animal, and food origins. *Antimicrob. Agents Chemother.* 48:3996–4001.
12. Villa L, García-Fernández A, Fortini D, Carattoli A. 2010. Replicon sequence typing of IncF plasmids carrying virulence and resistance determinants. *J. Antimicrob. Chemother.* 65:2518–2529.
13. Vinué L, et al. 2010. Genetic environment of *sul* genes and characterisation of integrons in *Escherichia coli* isolates of blood origin in a Spanish hospital. *Int. J. Antimicrob. Agents.* 35:492–496.
14. Walther-Rasmussen J, Høiby N. 2007. Class A carbapenemases. *J. Antimicrob. Chemother.* 60:470–482.
15. Yu YS, Du XX, Zhou ZH, Chen YG, Li LJ. 2006. First isolation of *bla*_{IMI-2} in an *Enterobacter cloacae* clinical isolate from China. *Antimicrob. Agents Chemother.* 50:1610–1611.

Beatriz Rojo-Bezares

Área de Microbiología Molecular
Centro de Investigación Biomédica de La Rioja (CIBIR)
Logroño, Spain

Carmen Martín

Laboratorio de Microbiología
Hospital San Pedro
Logroño, Spain

María López

Área de Bioquímica y Biología Molecular
Universidad de La Rioja
Logroño, Spain

Carmen Torres

Área de Bioquímica y Biología Molecular
Universidad de La Rioja
Logroño, Spain

Yolanda Sáenz

Área de Microbiología Molecular
Centro de Investigación Biomédica de La Rioja (CIBIR)
Logroño, Spain