

The structure of rat 28S ribosomal ribonucleic acid inferred from the sequence of nucleotides in a gene

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ABSTRACT

The nucleotide sequence of a rat 28S rRNA gene was determined. The 28S rRNA encoded in the gene contains 4718 nucleotides and the molecular weight estimated from the sequence is 1.53×10^6 . The guanine and cytosine content is 67%. The sequence of rat 28S rRNA diverges appreciably from that of *Saccharomyces carlsbergensis* 26S rRNA (about 50% identity), but more closely approximates that of *Xenopus laevis* 28S rRNA (about 75% identity). Rat 28S rRNA is larger than the analogous nucleic acids from yeast (3393 nucleotides) and *X. laevis* (4110 nucleotides) ribosomes. The additional bases are inserted in specific regions and tend to be rich in guanine and cytosine. 5.8S rRNA can interact with 28S rRNA by extensive hydrogen bonding at two sites near the 5' end of the latter.

INTRODUCTION

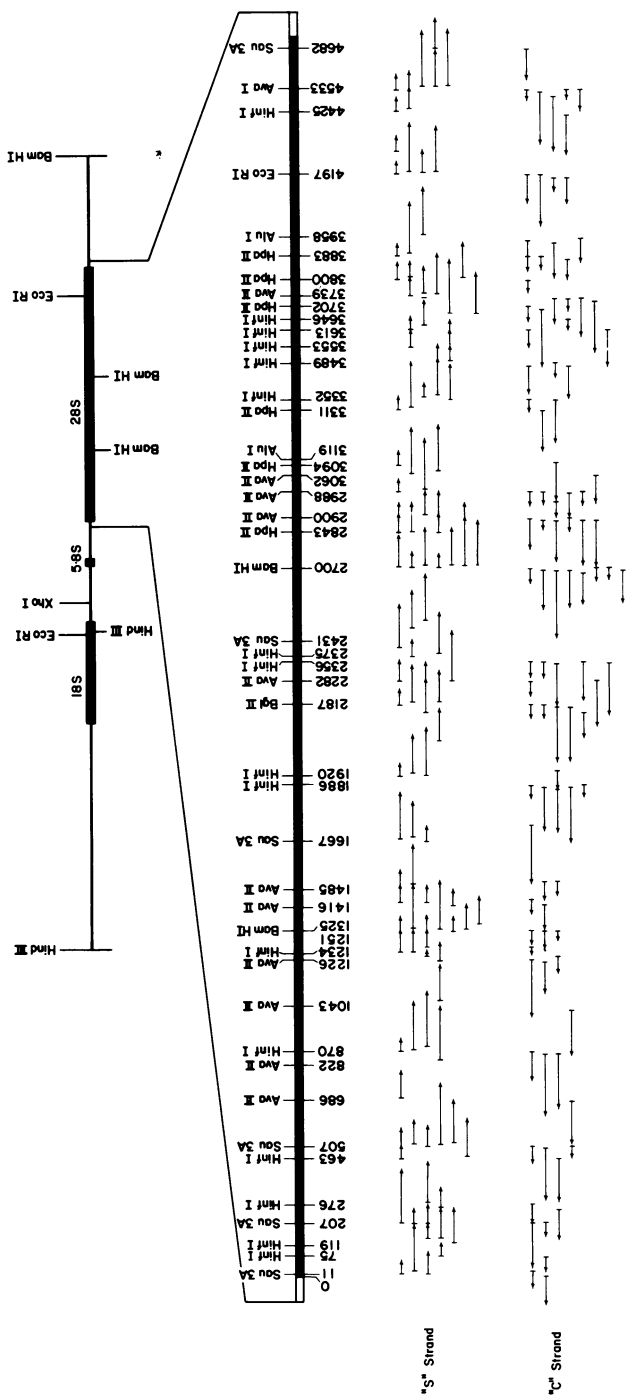
Solution of the structure of ribosomes requires information on the chemistry of the constituents. Eukaryotic ribosomes contain four molecules of RNA: 18S rRNA in the 40S subparticle, and 5S, 5.8S, and 28S RNAs in the 60S subparticle. The covalent structure of 5S (1), 5.8S (2,3), and 18S (4) RNAs from rat liver ribosomes have been determined. We have now established the structure of rat 28S rRNA from the sequence of a gene. For that purpose, we cloned a rat 28S rRNA gene carried on a phage (Charon 4A) in plasmid vectors and determined the sequence of the DNA.

EXPERIMENTAL PROCEDURES

DNA from the λ phage Charon 4A in which rat ribosomal RNA genes had been cloned (5) was kindly provided by T.D. Sargent and J. Bonner of California Institute of Technology. The DNA from the recombinant plasmid p2.7 was provided by L.I. Rothblum of Baylor College of Medicine. The procedures used for the preparation of restriction endonuclease fragments and for the determination of the sequence of nucleotides in DNA have been described or cited before (4).

RESULTS AND DISCUSSION

The Determination of the Sequence of 28S rDNA. A restriction map of the region of the Charon 4A recombinant λ phage containing the rDNA genes had been constructed (4). In



order to amplify the 28S rDNA and to facilitate purification, the gene was subcloned in pACYC184 (6) at the EcoRI sites to form pRRR228 (4). The EcoRI fragment of 6.4 kb contains most of the 28S rRNA gene (it lacks only the 3' end) and two internal transcribed spacers. The 3' end of a 28S rRNA gene was contained in an EcoRI-BamHI fragment of 2.7 kb (7) which was subcloned in pBR322 to form the recombinant plasmid pBR 288 (originally designated p2.7) (Fig. 1).

A restriction map of pRRR228 revealed that a combination of the enzymes BamHI, EcoRI, and XhoI would release six fragments, three of which contained portions of the 28S rRNA gene. The three fragments had an approximate length in kbs of 2.9 (XhoI-BamHI), 1.4 (BamHI-BamHI), and 1.5 (BamHI-EcoRI) (Fig. 1). An analysis of the cleavage pattern generated from the 1.4 and 1.5 kb fragments with restriction endonucleases identified enzymes suitable for production of subfragments convenient for a determination of the nucleotide sequence and for formation of overlapping oligonucleotides (Fig. 1). The 2.9 kb XhoI-BamHI fragment which contains the 5' end of the 28S rRNA gene, the 5.8S rRNA gene, and most of the internal transcribed spacer was digested with Sau3A. Four of the subfragments were shown by Southern hybridization (8) to contain portions of the 28S rRNA gene. The 3' end of the 28S rRNA gene was isolated from pBR288 using BamHI, EcoRI, and AvaI; the presence of the 3' end of 28S rRNA in the restriction enzyme digests was confirmed by Southern hybridization (8) to the α -sarcin RNA fragment(9).

The sequence of the 28S rRNA gene was determined by the Maxam and Gilbert procedure (12, 13). The analysis included a determination of the sequence of almost all

Figure 1. Restriction endonuclease map of a rat 28S rRNA gene and a diagram of the overlapping fragments whose sequence of nucleotides was determined. The upper portion of the figure depicts the restriction endonuclease sites in a rat 28S rRNA gene that were used to generate fragments (lower portion of the figure) for the determination of the sequence of nucleotides. The numbers indicate the first nucleotide in the restriction endonuclease recognition sequence counting from the 5' end of the 28S rRNA gene. Each arrow designates a nucleotide sequence that was determined. The "S" strand is synonymous with the RNA, the "C" strand is complementary. Plasmid DNA was isolated by the clear lysate method (10, 11). The procedures used for the digestion of DNA with restriction enzymes, for the isolation of DNA fragments, and for the labeling of the fragments at the 5' end with [γ - 32 P]ATP using T4 polynucleotide kinase were cited earlier (4). Double-stranded DNA fragments were separated by heating for 2 min at 90 °C in 50% DMSO, 50 mM Tris/HCl, pH 8.3, 50 mM borate, 2 mM EDTA. The presence of portions of the 28S rRNA gene in the fragments was established by Southern hybridization (8) to 32 P-labeled 28S rRNA prior to undertaking the sequence determination. The sequence of nucleotides in the DNA fragments was determined by the Maxam and Gilbert method (12, 13) using thin (0.5 mm) gels of 8, 10, or 20% polyacrylamide (13, 14). There are several short regions of ambiguity: 669 to 675; 777 to 785; 1281 to 1285; 2003 to 2009; 2033 to 2040; and 2058 to 2064. In each case the number of nucleotides differed when read in the two directions due to compression in the sequencing ladder in one of the two gels. We report the sequence with the larger number of nucleotides

of both strands and extensive overlaps of portions of the sequences in separate fragments (Fig. 1). The order of the fragments was approximated from the restriction endonuclease map, and by reference to the sequence of the 5' (3) and 3' ends (15) of rat 28S rRNA.

The sequence of the nucleotides in 28S rRNA was inferred from the sequence of the gene (Fig. 2). Rat 28S rRNA has 4718 nucleotides. The molecular weight is 1.53×10^6 (based on the assumption of a molecular weight of 325 for a nucleotide) somewhat less than the 1.7×10^6 (5230 nucleotides) estimated from physicochemical measurements (16, 17). The guanine and cytosine content of rat 28S rRNA is 67%.

Ribosomal nucleic acids are methylated. We have not identified the nucleotides in rat 28S rRNA that are methylated. However, the pattern of methylated bases in oligonucleotide digests of 28S rRNAs from human (HeLa), hamster (BHK/C13), and mouse (L) cells could hardly be distinguished, i.e. the identity is 99% (18). Moreover, the methylated nucleotides in 28S rRNAs from human (HeLa cells) and from *X. laevis* have at least 95% identity (18). Twenty-nine of the 30 methylation sites in yeast 26S rRNA (19) are conserved in the rat nucleic acid. Thus, the methylated bases in rat 28S rRNA are likely to be the same as those in other large eukaryotic rRNAs.

Comparison of the Sequence of Rat 28S rRNA with other Large Eukaryotic rRNAs. The sequence of nucleotides in yeast 26S rRNA (*Saccharomyces carlsbergensis* (19) and *Saccharomyces cerevisiae* (20)), and in *X. laevis* 28S rRNA (21) have been determined and can be compared with the structure of rat 28S rRNA. Yeast 26S rRNA has 3393 nucleotides whereas rat 28S rRNA has 4718 (Table I). When the sequences of the rat and yeast rRNAs were aligned there were 1946 identities (the same nucleotide at the same position). Thus, the percentage identity is 41 for rat and 57 for yeast; the latter is higher because it has a smaller number of nucleotides (Table I). In general, regions of close identity are interrupted by domains that are quite different (Fig. 3A). We have sought to identify the variable regions as well as sites of insertion of the additional nucleotides (Table II). There are three regions of rat 28S rRNA where large insertions have occurred. They are between positions 430 and 1206 (570 nucleotides), 1977 and 2180 (118 nucleotides), and 2745 and 3328 (440 nucleotides) (Table II). These three inserts account for 1128 or 85% of the 1325 additional nucleotides in rat 28S rRNA. The next largest inserts are of 43 nucleotides in the region 1244 to 1409, and of 53 nucleotides at 3698 to 3753 in the rat sequence (Table II). The regions of variability and of insertions in rat 28S rRNA are rich in guanine and cytosine (Table II). Indeed, the insertions account, in large part, for the difference between the content of these two nucleotides in rat (67%) and yeast (48%; ref. 19).

There is considerably more identity in the structure of rat and *X. laevis* 28S rRNAs than with yeast 26S rRNA. Rat 28S rRNA has 4718 nucleotides, *X. laevis* 28S rRNA has 4110. When the sequences are aligned there are 3248 identities, or 69% for rat and 79%

pCGCGACCUCA	GAUCAGACGU	GGCGACCCGC	UGAAUUUAAG	CAUAUUAGUC	50
AGCGGAGGAA	AAGAAACUAA	CCAGGAUUC	CUCAGUAACG	GCGAGUGAAC	100
AGGGAAGAGC	CCAGCGCCGA	AUCCCCGCCG	CGCGCCGGG	CGCGGAAAU	150
GUGGCGUACG	GAAGACCCAC	UCCCCGGCGC	CGCUCGUGGG	GGGCCCAAGU	200
CCUUCUGAUC	GAGGCCACG	CCGUGGACGG	UGUGAGGCCG	GUAGCGGCC	250
CGGCGCGCCG	GGCGGGUCU	UCCCGGAGUC	GGGUUGCUUG	GGAAUGCAGC	300
CCAAAGCGGG	UGGUAAACUC	CAUCUAAGGC	UAAAUACCGG	CACGAGACCG	350
AUAGCCAACA	AGUACCGUAA	GGGAAAGUUG	AAAAGAACUU	UGAAGAGAGA	400
GUUCAAGAGG	GCGUGAAACC	GUUAAGAGGU	AAACGGGUGG	GGUCCGCGCA	450
GUCCGCCCGG	AGGAUUCAAC	CCGGCGGCGC	GCGCCGGCCG	GCCGGUGGUC	500
CCGGCGGAUC	UUUCCCGCUC	CCCGUUCCUC	CCGACCCUC	CACCCGCGCG	550
UCUCUCUCCC	CCCUCCCCGC	GUCCCGCCGU	CGCCGUCCCC	GCUCCUCCCU	600
CCGGGGGGGU	GUCGGCGGGC	GCUCCGGCGG	CGGGCGCGGG	GUGUGGUGGG	650
GGCGCGCGGG	CGGGGCGGGG	GGUGGGGUCG	GCGGGGACC	GCCCCGGUC	700
GGCGACCGGC	CGCCGCGGGG	CGCACUCCA	CCGUGGCGGU	GCGCCGCGAC	750
CGGCUCCGGG	ACGGCUGGGA	AGGCCCGCGG	GGGAAGGUGG	CUCGGGGGGG	800
GCGGCGUCAC	CCGUGGGCGC	CGGACCACCC	CGCCCCGAGU	GUUACAGCCC	850
CCCGGACGCA	GCGCUCGCGG	AAUCCCGGGG	CCGAGGGAGC	CGGAUACCCG	900
UCGCCGCGCU	CUCCCCCCGG	CCUCUCCCU	CCCGCCCCUC	CCCGUGGGGU	950
GGCGGAAAGG	GGGGCGGUCG	CGGGGGCCGG	GCCGCCCCUC	CCACGGCGCG	1000
ACCGCUCUCC	CACCCCCCGU	GCUCUCGUC	GUCCUCUCG	GGGUCCCGGG	1050
GGCCCGGGGG	GCGGGGCGGA	CUGUCCCAG	UGCGCCCCGG	GCGUCGUCGC	1100
GCCGUCGGGC	CCGGGGGGGC	CGUCGUCACG	CGCUCUCCU	CCCUUCUCG	1150
GGGUGGGGGG	GAGCGAAGCC	GAGCGCACGG	GGUCGGCGGC	GAUGUCGGCU	1200
ACCCACCCGA	CCCUCUUGA	AACACGGACC	AAGGAGUCUA	ACGCGUGCGC	1250
GAGUCAGGGG	CUCGUCCGAA	AGCCGCGGUG	GCGCAAUGAA	GGUGAAGGGC	1300
CCCGUUCCCG	GGGGCCCCGA	GGUGGGAUCC	CGAGGCCUCU	CCAGUCCGCC	1350
GAGGGCGCAC	CACCGGCCCG	UCUCGCCCGC	CGCGCCGGGG	AGGUGGAGCA	1400
CGAGCGUACG	CGUUAGGACC	CGAAAGAUGG	UGAACUAUGC	UUGGGCAGGG	1450
CGAAGCAGAG	GAAACUCUGG	UGGAGGUCCG	UAGCGGUCCU	GACGUGCAAA	1500
UCGGUCGUCC	GACUUGGGUA	UAGGGGCGAA	AGACUAAUCG	AACCAUCUAG	1550
UAGCUGGUUC	CCUCCGAAGU	UUCCUCAGG	AUAGCUGGCG	CUCUCGCAAC	1600

GCGUUCGCUC	GACAACCCGC	AGUUUUUACC	GGUAAAGCGA	AUGAUUAGAG	1650
GUCUUGGGGC	CGAAACGAUC	UCAACCUAUU	CUCAAACUUU	AAAUGGGUAA	1700
GAAGCCCGGC	UCGCUGGCGU	GGAGCCGGGC	GUGGAUGC GA	GUGCCUAGUG	1750
GGCCACUUUU	GGUAAGCAGA	ACUGGCGCUG	CGGGAUGAAC	CGAACGCCGG	1800
GUUAAGGCGC	CCGAUGCCGA	CGCUCAUCAG	ACCCAGAAA	AGGUGUUGGU	1850
UGAUAUAGAC	AGCAGGACGG	UGGCCAUGGA	AGUCGGAAUC	CGCUAGGAGU	1900
GUGUAAACAAC	UCACCUGCCG	AAUCAACUAG	CCCUGAAAAU	GGAUGGCGCU	1950
GGAGCGUCGG	GCCCAUACCC	GGCCGUCGCC	GGCAGUCGGA	ACGGGACGGG	2000
AGCGGCCCGC	GGCGCGCGAC	CCCGGGGGCC	GGGCGGCUC	GGCUUCGGCC	2050
GGCCGCCGCC	CGUCCACCCC	CGGGGUCCC	CCC CGCGCU	CGGGCCCCGC	2100
GGAGCCUACG	CCGCACGAG	UAGGAGGGCC	GCUCGGGUEA	GCCUUGAAGC	2150
CUAGGGCGCG	GGCCCGGGUG	GAGCCGCCGC	AGGUGCAGAU	CUUGGUGGUA	2200
GUAGCAAUA	UUCAAACGAG	AACUUUGAAG	GCCGAAGUGG	AGAAGGGUUC	2250
CAUGUGAACA	GCAGUUGAAC	AUGGGUCAGU	CGGUCCUGAG	AGAUGGGCGA	2300
GUGCCGUUCC	GAAGGGACGG	GCGAUGGCCU	CCGUUGCCCU	CAGCCGAUCG	2350
AAAGGGAGUC	GGGUUCAGAU	CCCCGAAUCC	GGAGUGGCGG	AGAUGGGCGC	2400
CGCGAGGCGU	CCAGUGCCGG	UAACGCGACC	GAUCCCGGAG	AAGCCGGCGG	2450
GAGCCCGGG	AGAGUUCUCU	UUUCUUUGUG	AAGGGCAGGG	CGCCUUGGAA	2500
UGGGUUCGCC	CCGAGAGAGG	GGCCCGUGCC	UUGGAAAGCG	UCGCAGUUCC	2550
GGCGGCGUCC	GGUGAGCUCU	CGCUGGCCCU	UGAAAUCCG	GGGGAGAGGG	2600
UGUAAAUCUC	GCGCCGGGCC	GUACCAUAU	CCGCAGCAGG	UCUCCAAGGU	2650
GAACGCCUCU	GGCAUGUUGG	AACA AUGUAG	GUAAGGGAAG	CGCAAGCCG	2700
GAUCCGU AAC	UUCGGGAUAA	GGAUUGGCUC	UAAGGCUGG	GUCGGUCGGG	2750
CUGGGGCGCG	AAGCGGGGC	UGGGCGCGCG	CCGCGGCUGG	ACGAGGCGCC	2800
GCCGCCCC	UCCACGUCC	GGGAGACCC	CCUCCUUUCC	GCCCGGGCCC	2850
GCCUCCCCU	CUCCCGCGG	GGCCCGCGG	UCCCCCGCU	CGUCGCCGUG	2900
GUCCCCUCCU	CCUCCCUUC	UUCCCCGUCC	GCGGGGGGA	CGGGGCGGGU	2950
GCGGGGGGGC	GCGCGCGCGC	GCGGCCAGG	GGCGGCGGU	CCAACCCCGC	3000
GCGGGCCGGA	GCGGGGGAA	CCCGGGGCC	CCCUGUGGG	GGGGGCCCGG	3050
ACACCCGGGG	GGGACCGGCG	GCGGCGGCGA	CUCUGGACGC	GAGCCGGGCC	3100
CUUCCCGUGG	AUCGCCCCAG	CUGCGGCGGG	CGUCGCGGCC	GCUCCCGGGG	3150
AGCCCGGGCG	GUCGCCCGGC	GGGGUUUUC	UCCGGCCUCG	UCCUCCCCU	3200

UCCCCUCCG	CGGGGUCGGG	GGUUCCCGGG	GUUCGGGGUU	CUCCUCCGCG	3250
CGGCGGUUCC	CCC GCCGGGU	GCGCCCCCG	GGCGGGUUU	CCCGGGCCG	3300
CCCGCCUCGG	CCGGCGCCUA	GCAGCCGACU	UAGAACUGGU	GCGGACUAGG	3350
GGAAUCCGAC	UGUUUAAUUA	AAACAAAGCA	UCGCGAAGGC	CCGCGGCGGG	3400
UGUUGACGCG	AUGUGAUUUC	UGCCAGUGC	UCUGAAUGUC	AAAGUGAAGA	3450
AAUUCAUUGA	AGCGGGGUA	AACGGCGGGA	GUAACUAUGA	CUCUCUUAAG	3500
GUAGCCAAAU	GCCUCGUCAU	CUAAUAGUG	ACGCGCAUGA	AUGGAUGAAC	3550
GAGAUUCCCA	CUGUCCCUAC	CUACUAUCCA	GCGAAACCAC	AGCCAAGGGA	3600
ACGGGUUGG	CGAAUCAGC	GGGAAAGAA	GACCCUGUUG	AGCUUGACUC	3650
UAGUCUGGCA	CGGUGAAGAG	ACAUGAGAGG	UGUAGAAUAA	GUGGGAGGCC	3700
CCGGCGCCC	CCCCGUUCC	CGGAGGGGU	CGGGGCGGG	UCCGCCGCC	3750
UCGCGGGCCG	CCGGUAAAU	ACCACUACUC	UCAUCGUUUU	UUCACUGACC	3800
CGGUGAGGCG	GGGGGCGAG	CCCCGAGGG	CUCUCGUUC	UGGCGCCGAA	3850
CGGUCCGCG	CGCGGGCGG	GCGCGACCG	CUCCGGGGAC	AGUGCCAGGU	3900
GGGAGUUUG	ACUGGGGCG	UACACUGUC	AAACGGUAA	GCAGGUGUCC	3950
UAAGGCGAGC	UCAGGGAGGA	CAGAAACCUC	CCGUGGAGCA	GAAGGGCAA	4000
AGCUCGCUUG	AUCUUGAUUU	UCAGUACGAA	UACAGACCGU	GAAAGCGGG	4050
CCUCACGAUC	CUUCUGACCU	UUUGGUUUU	AAGCAGGAG	UGUCAGAAA	4100
GUUACCACAG	GGAAACUGG	CUUGUGGCG	CCAAGCGUUC	AUAGCGACGU	4150
CGCUUUUUGA	UCCUUCGAUG	UCGGCUCUUC	CUAUCAUUGU	GAAGCAGAAU	4200
UCACCAAGCG	UUGGAUUGUU	CACCCACUAA	UAGGGAACGU	GAGCUGGGUU	4250
UAGACCGUCG	UGAGACAGGU	UAGUUUACC	CUACUGAUGA	UGUGUUGUUG	4300
CCAUGGUAAU	CCUGUCAGU	ACGAGAGGAA	CCGCAGGUUC	AGACAUUUGG	4350
UGUAUGUGCU	UGGCUGAGGA	GCCAAUGGG	CGAAGCUACC	AUCUGUGGGA	4400
UUAUGACUGA	ACGCCUCUAA	GUCAGAAUCC	GCCCAAGCGG	AACGAUACGG	4450
CAGCGCCGAA	GGAGCCUCGG	UUGGCCCCGG	AUAGCCGGCU	CCCCGUCCGU	4500
CCCCGUCCGG	CGGGUCCCCG	CCUCGUCGCC	CCCCGGGAA	ACGGGUGCG	4550
GCCGAAAGG	GGGCCGCCU	CUCGCCGUC	ACGCUAAACG	CACGUUCGUG	4600
UGGAACUUGG	CGCUAAACCA	UUCGUAGACG	ACCUGCUUCU	GGGUCGGGGU	4650
UUCGUACGUA	GCAGAGCAGC	UCCUCGCUG	CGAUCUAUUG	AAAGUCAGCC	4700
CUCGACACAA	GGUUUGU				4718

Figure 2. The sequence of nucleotides in rat 28S rRNA inferred from the structure of a gene.

TABLE I Comparison of the Structure of Rat, *X. laevis* and Yeast Large rRNAs

	<u>Nucleotides (no.)</u>	<u>Different</u> <u>Nucleotides (no.)</u>	<u>Similar</u> <u>Nucleotides (no.)</u>	<u>Identity (%)^a</u>
A. Rat	4718	2772		41
Yeast	3393	1447	1946	57
B. Rat	4718	1470		69
<i>X. laevis</i>	4110	862	3248	79

^aBecause of the difference in the number of nucleotides in the large rRNAs in the three species the percent identity is conditioned by whether the comparison is between the larger and the smaller, or the smaller and larger.

for *X. laevis* (Table I). Once again, we have attempted to locate the variable sequences and to identify where the insertions are (Table III). There are five regions that account for 592 or 97% of the additional 608 nucleotides. They are at positions 492 to 725 (132 nucleotides), 891 to 1170 (101 nucleotides), 1984 to 2097 (102 nucleotides), 2807 to 3065 (167 nucleotides), and 3130 to 3292 (90 nucleotides) (Table III). The inserted sequences, again, are especially rich in guanine and cytosine (Table III). We take notice that there are two deletions in the rat sequence, in regions 1333 to 1352 (25 nucleotides) and 2404

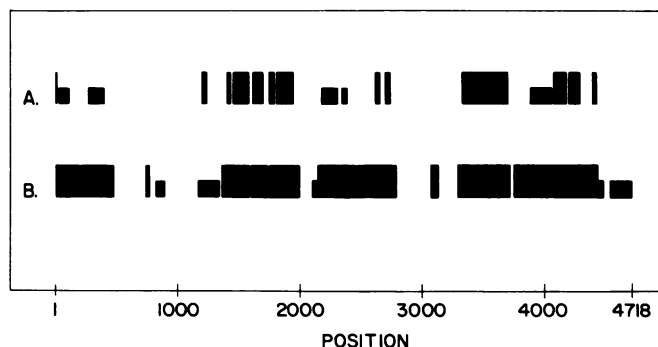


Figure 3. Regions of similarity in the structure of yeast 26S rRNA and *X. laevis* 28S rRNA compared to rat 28S rRNA. The sequences of the large rRNAs from *S. carlsbergensis* (19) and *X. laevis* (21) were compared to that of rat using the computer program of Queen and Korn (37) with the minimum number of matches set at 15 nucleotides. The tall blocks represent regions of yeast (*S. carlsbergensis*) 26S rRNA (A) or *X. laevis* 28S rRNA (B) with 85 to 100% identity with rat 28S rRNA; the short blocks are regions with 75 to 85% identity.

TABLE II. Comparison of Rat 28S rRNA and Yeast 26S rRNA: Variable Regions

Position	Rat		Yeast		G + C (%)	Nucleotides (no.)	Position	Yeast		G + C (%)	Nucleotides Inserted (no.)
	Nucleotides (no.)	G + C (%)	Nucleotides (no.)	Position				Nucleotides (no.)	G + C (%)		
114-275	162	77	152	113-264	47				10		
430-1206	777	81	207	424-630	48				570		
1244-1409	166	75	123	667-789	49				43		
1586-1619	34	71	12	968-979	42				22		
1708-1754	47	77	47	1068-1114	47				0		
1977-2180	204	84	86	1340-1425	60				118		
2316-2343	28	71	19	1560-1578	47				9		
2384-2437	54	74	32	1618-1649	38				22		
2464-2609	146	62	140	1679-1818	51				6		
2745-3328	584	84	144	1954-2097	59				440		
3392-3400	9	100	9	2162-2170	33				0		
3698-3753	56	89	3	2468-2470	33				53		
3779-3887	109	76	95	2494-2588	47				14		
4431-4718	288	66	263	3131-3393	46				25		
		<u>79</u>			<u>51</u>						

TABLE III. Comparison of Rat and *X. laevis* 28S rRNAs: Variable Regions

<u>Position</u>	<u>Rat</u>		<u><i>X. laevis</i></u>		<u>G + C (%)</u>	<u>Nucleotides Inserted (no.)</u>
	<u>Nucleotides (no.)</u>	<u>G + C (%)</u>	<u>Position</u>	<u>Nucleotides (no.)</u>		
492-725	234	84	489-590	102	88	132
796-829	34	85	663-664	0		34
891-1170	280	83	732-910	179	87	101
1302-1318	17	88	1043-1056	14	93	3
1333-1352	20	70	1071-1115	45	93	-25
1594-1617	24	67	1359-1364	6	67	18
1984-2097	114	88	1739-1750	12	67	102
2404-2405	0		2059-2094	36	94	-36
2807-3065	259	86	2502-2593	92	86	167
3130-3292	163	82	2655-2727	73	86	90
3709-3756	48	88	3146-3160	15	67	33
4495-4554	60	83	3904-3956	53	81	7
		<u>85</u>			<u>87</u>	

to 2405 (36 nucleotides). Thus, the increase in size of the large RNAs in the 60S ribosomal subunit is the result of the addition en bloc of skeins of guanine- and cytosine-rich oligonucleotides (Fig. 3B). Nonetheless, the total content of the two nucleotides is very similar in rat (67%) and X. laevis (65%; ref 21) 28S rRNAs.

We note in passing that there are two recognition sequences for the restriction endonuclease BamHI, at positions 1325 and 2700, in the rat 28S rRNA gene (3,7,15). The mouse 28S rRNA gene also has two BamHI sites (22, 23). However, the X. laevis 28S rDNA has only one (21, 24), and yeast none (19, 20). The loss of the second BamHI site in X. laevis 28S rDNA is the result of a guanine to adenine transition at position 2398, changing the recognition sequence GGATCC to AGATCC.

Eukaryotic 80S ribosomes have increased in size during evolution from plants to animals (25, 26). The size of the 40S subunit has remained fairly constant so the increase in size is to be attributed to changes in the 60S subparticle and primarily to an increase in the number of nucleotides in the large RNA. The number of nucleotides in 18S rRNAs from rat (1874; ref. 4), X. laevis (1825; ref. 27), and yeast (1789; ref. 28) are similar, whereas the number in the large rRNAs progressively increase as one goes from yeast (3393; ref. 19, 20), to X. laevis (4110; ref. 21), to rat (4718). It is difficult to provide an explanation for this evolutionary change, or to rationalize the insertion sequences being rich in guanine and cytosine.

The 5.8S rRNA Contact Sites in 28S rRNA. In the large subunit of eukaryotic ribosomes 5.8S rRNA is non-covalently associated with 28S rRNA (29). The association between these RNA molecules is relatively stable, although the complex can be dissociated by agents that disrupt hydrogen bonds. Fragments containing the 5'-terminal 73 nucleotides and the 3'-terminal 83 nucleotides of 5.8S rRNA can associate independently with 28S rRNA (30). However, it is probable that only approximately the 3'-terminal 20 nucleotides and the 5'-terminal 20-30 nucleotides are involved in the interaction with 28S rRNA (31). While the exact location of the junction sites in 28S rRNA have not been established experimentally there is evidence that 5.8S rRNA interacts with a structural domain near the 5' end (23).

One can deduce what the identity of these contact sites might be by examining the proposal for the secondary structure of Escherichia coli 23S rRNA. The 5'-terminal region of E. coli 23S rRNA is structurally homologous to eukaryotic 5.8S rRNA (32, 33), indeed the basic difference between eukaryotes and prokaryotes in this regard is that in the former 5.8S rRNA has been processed out of the transcription unit in a manner that leads to formation of a separate molecule. It follows then, that one can examine the secondary structure of E. coli 23S rRNA (34), locate the region homologous to 5.8S rRNA, and take notice of where this region interacts with the remainder of the structure. The prediction arising from that exercise is that the 3' end of 5.8S rRNA

interacts with the immediate 5' end of 28S rRNA and that the 5' end of 5.8S rRNA is complementary to a region about 300 to 400 nucleotides from the 5' end of 28S rRNA (23). As a result of the determination of the sequence of yeast 26S rRNA, and elucidation of the possible secondary structure (19), the proposal could be tested. Two regions of extensive complementarity between the two nucleic acids were found (19) which are entirely consistent with the prediction. Moreover, an exactly analogous interaction has been suggested for mouse 5.8S and 28S rRNAs predicated on the entire sequence of the former and the 585 nucleotides at the 5' end of the latter (35).

Guided by the earlier work (19, 35) we have searched the sequences of rat 5.8S and 28S rRNAs for regions of complementarity. In conformity with the junction structures proposed for yeast and mouse rRNAs, there are two sites of possible extensive base pairing in rat 5.8S and 28S rRNAs. They encompass, in the first instance, 17 base pairs between the 3' end of 5.8S rRNA (positions 137 to 155) and the 5' end of 28S rRNA (positions 2 to 20); and in the second, 24 base pairs between the 5' end of 5.8S rRNA (positions 4 to 32) and a region of 28S rRNA centered about nucleotide 400. Thus, the results for rat are entirely compatible with the proposals for the contact sites in yeast (19) and mouse rRNAs (35). They do not lend credence to the submission that both ends of 5.8S rRNA interact with the 3' end of Neurospora crassa 25S rRNA (36), although the results do not entirely negate the possibility of some interaction of the 3' end of the large rRNA with either 5.8S rRNA or the 5' end of 28S rRNA.

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