

## NIH Public Access

Author Manuscript

Arch Pediatr Adolesc Med. Author manuscript; available in PMC 2012 January 25.

#### Published in final edited form as:

Arch Pediatr Adolesc Med. 2010 December; 164(12): 1173–1175. doi:10.1001/archpediatrics.2010.233.

### Nasopharyngeal carriage of *Streptococcus pneumoniae* in very low birth weight infants after administration of heptavalent pneumococcal conjugate vaccine

Jocelyn Y. Ang, MD<sup>1</sup>, Jorge L. Lua, MD<sup>2</sup>, Basim I. Asmar, MD<sup>1</sup>, Seetha Shankaran, MD<sup>2</sup>, Roy J. Heyne, MD<sup>3</sup>, Robert L. Schelonka, MD<sup>4</sup>, Abhik Das, PhD<sup>5</sup>, Lei Li, PhD<sup>6</sup>, Delois M. Jackson, MSc<sup>7</sup>, Rosemary D. Higgins, MD<sup>8</sup>, Carl T. D'Angio, MD<sup>9</sup>, and on behalf of the National Institute of Child Health and Human Development (NICHD) Neonatal Research Network

<sup>1</sup>Division of Infectious Diseases, Department of Pediatrics/ Children's Hospital of Michigan/ Wayne State University, Detroit, Michigan

<sup>2</sup>Division of Neonatal-Perinatal Medicine, Department of Pediatrics/ Children's Hospital of Michigan/ Wayne State University, Detroit, Michigan

<sup>3</sup>University of Texas Southwestern Medical Center, Department of Pediatrics/ Division of Neonatal-Perinatal Medicine, Dallas, Texas

<sup>4</sup> University of Alabama at Birmingham, Department of Pediatrics/ Division of Neonatology, Birmingham, Alabama

<sup>5</sup>Research Triangle Institute (RTI) International, Statistics and Epidemiology Unit, Rockville, MD

Eunice Kennedy Shriver National Institute of Child Health and Human Development – Stephanie Wilson Archer, MA. *RTI International* (U01 HD36790) – W. Kenneth Poole, PhD; Betty K. Hastings; Elizabeth McClure, MEd; Rebecca L. Perritt, MS; Steve Emrich, MS; Kristin Zaterka-Baxter, RN; Carolyn Petrie Huitema, MS; Jamie E. Newman, MPH; Scott E. Schaefer, MS; Jeanette O'Donnell Auman, BS.

**Correspondence:** Jocelyn Y. Ang, MD Children's Hospital of Michigan Carman and Ann Adams Department of Pediatrics Division of Infectious Diseases Wayne State University School of Medicine 3901 Beaubien Blvd. Detroit, MI 48201 Tel: (313) 745-5863 Fax: (313) 996-8846 jang@med.wayne.edu.

**Role of Authors:** On behalf of the network, Drs. Abhik Das (DCC PI) and Lei Li (DCC Statistician) had full access to all the data in the study and take responsibility for the integrity of the data and accuracy of the data analysis. All authors are involved in the interpretation of data, and in the preparation, review, and approval of the manuscript.

We are indebted to our medical and nursing colleagues and the infants and their parents who agreed to take part in this study. The following investigators participated in this study:

NRN Chairs: Alan Jobe, MD PhD, University of Cincinnati (2001-2006); Michael S. Caplan, MD, Northwestern University (2006-2011).

*University of Alabama at Birmingham* Health System and Children's Hospital of Alabama (GCRC M01 RR32, U10 HD34216) – Waldemar A. Carlo, MD; Myriam Peralta-Carcelen, MD MPH; Monica V. Collins, RN BSN MaEd; Shirley S. Cosby, RN BSN; Vivien A. Phillips, RN BSN.

*University of Rochester* Golisano Children's Hospital at Strong (GCRC M01 RR44, U10 HD40521) – Dale L. Phelps, MD; Gary J. Myers, MD; Cassandra A. Horihan, MS; Rosemary L. Jensen; Diane L. Hust, RN PNP.

University of Texas Southwestern Medical Center at Dallas Parkland Health & Hospital System and Children's Medical Center Dallas (GCRC M01 RR633, U10 HD40689) – Charles R. Rosenfeld, MD; Walid A. Salhab, MD; Pablo J. Sanchez, MD; Janet S. Morgan, RN; Jackie F. Hickman, RN; Alicia Guzman; Nancy A. Miller, RN; Gaynelle Hensley, RN.

Wayne State University Hutzel Women's Hospital and Children's Hospital of Michigan (U10 HD21385) – Athina Pappas, MD; Rebecca Bara, RN BSN.

We thank Cynthia G. Whitney, MD, MPH and Bernard Beall, PhD from the Respiratory Diseases Branch, Centers for Disease Control and Prevention (CDC), Atlanta, Georgia for their assistance. Serotyping of pneumococcal isolates was performed in the Streptococcus Laboratory at CDC.

We thank Theresa Painter MA, MT and Christine Wollenweber MT from Detroit Medical Center University Laboratory/ Wayne State University for their assistance in processing nasopharyngeal specimens for culture, storage and shipment.

We thank William Lyman, PhD, Director of Children's Research Center of Michigan (CRCM) Detroit, Michigan for his support.

<sup>6</sup>Research Triangle Institute (RTI) International, Statistics and Epidemiology Unit, Research Triangle Park, North Carolina

<sup>7</sup>Streptococcus Laboratory, Respiratory Diseases Branch, Centers for Disease Control and Prevention (CDC), Atlanta, Georgia

<sup>8</sup>Eunice Kennedy Shriver National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, MD

<sup>9</sup>Strong Children's Research Center, University of Rochester School of Medicine and Dentistry, Department of Pediatrics/ Division of Neonatal-Perinatal Medicine, Rochester, New York

#### Introduction

The effect of pneumococcal conjugate vaccine-7 (PCV-7) in reducing pneumococcal nasopharyngeal (NP) carriage in very low birth weight (VLBW) infants has not been studied. Our primary objective was to characterize NP carriage of *S. pneumoniae* in a group of VLBW infants (401-1500 grams) before administration of first PCV-7 (PRE) and at 4-6 weeks after a 3-dose PCV-7 primary series (POST). We also investigated the correlation between vaccine induced pneumococcal IgG antibody level and pneumococcal NP carriage POST PCV-7.

#### **Methods**

VLBW infants participating in a PCV-7 immunogenicity study<sup>1</sup> were enrolled from 4 NICHD Neonatal Research Network (NRN) sites (Detroit, MI, Rochester, NY, Dallas, TX and Birmingham, AL). The study was approved by NRN and each site's institutional review board. Written informed consent was obtained from each subject's parent/guardian. Infants received PCV-7 at approximately 2, 4 and 6 months of age.

NP cultures were obtained at the PRE and POST visits. Antimicrobial susceptibility testing of all pneumococcal isolates was performed.

Pneumococcal isolates were serotyped at the Centers for Disease Control and Prevention, Atlanta, GA. Serotypes 4, 6B, 9V, 14,18C, 19F, and 23F were classified as vaccine serotypes (VT). Other serotypes were classified as non-VT (NVT).

Anti-pneumococcal antibodies<sup>2</sup> against seven VT were measured at POST visit and  $0.15\mu$ g/ml was chosen as a possible measure of protective level.<sup>3</sup>

Descriptive statistics were used to characterize the study subjects with regard to birth weight, gestational age at birth (GA) and chronologic age (CA) at swab collection, as well as serotype and antimicrobial susceptibilities of pneumococcal isolates.

#### Results

123 of 135 infants enrolled had at least one NP swab obtained; 71 had PRE and 102 had POST NP swab cultures. 50 infants had both PRE and POST NP swabs. Most (44/71=62%) had PRE NP swab done while in NICU, whereas all but one had POST NP swab done after hospital discharge.

The median GA of 123 infants was 28 weeks (range 23-32). The median CA at PRE and POST NP swab collections were 2 months (range 1-3) and 8 months (range 6-10) respectively.

Arch Pediatr Adolesc Med. Author manuscript; available in PMC 2012 January 25.

*S. pneumoniae* was isolated in 4.2% (3/71) PRE and 12.7% (13/102) POST samples. One infant had positive PRE and POST NP cultures with 2 different serotypes; another colonized at PRE had a negative POST NP culture. Among the 50 infants with both PRE and POST NP swabs, 49 had negative PRE NP cultures of whom 8 became colonized at POST (all NVT) (Table 1).

Serotyping was done on 15 of 16 isolates (Table 1). One PRE isolate was a VT; all 12 POST isolates were NVT. Antibiotic susceptibility (Table1) showed one PRE isolate resistant to erythromycin. Of 12 POST isolates, 2 (19A) were resistant to 4 antibiotics, 1 (35B) resistant to penicillin and 1 (non-typeable) resistant to erythromycin.

Serum anti-capsular IgG antibody levels to 7 VT were available for 100 infants who had POST NP swabs. Anti-pneumococcal antibody  $0.15 \,\mu$ g/ml were achieved in 88-99% and varied by serotype. Since all POST pneumococcal isolates were NVT, assessment of NP carriage status based on antibody levels could not be performed.

#### Comment

The pneumococcal NP carriage rate of 12.7 % in our VLBW infants post-PCV-7 vaccination was lower than previously reported during the pre-PCV-7<sup>4</sup> and early post-PCV-7<sup>5</sup> eras and was exclusively due to NVT. Our findings also support the recent observation that NVT serotype 19A is becoming more prevalent. An expanded PCV such as PCV-13 which includes 6 additional serotypes (1, 3, 5, 6A, 7F and 19A) may change that. Regardless of serum pneumococcal anti-capsular antibody levels, all POST pneumococcal NP isolates were NVT, suggesting protection against VT NP carriage at 4-6 weeks POST period.

#### Acknowledgments

**Funding /Support:** The National Institutes of Health and the Eunice Kennedy Shriver National Institute of Child Health and Human Development (NICHD) and Children's Research Center of Michigan (CRCM).

**Role of Sponsor:** The National Institutes of Health and the Eunice Kennedy Shriver National Institute of Child Health and Human Development (NICHD) provided grant support for the Neonatal Research Network's (NRN) PCV-7 Study. The agencies provided overall oversight for study conduct, but all data analyses and interpretation were independent of the funding agencies. The Children's Research Center of Michigan (CRCM) in Detroit, MI provided funding for the nasopharyngeal swab cultures and compensation for nurse coordinators' time.

Data collected at participating NRN sites were transmitted to Research Triangle Institute (RTI) International, the data coordinating center (DCC) for the NRN, which stored, managed, and analyzed the data for this study.

#### References

- D'Angio CT, Heyne RJ, O'Shea TM, et al. on behalf of the NICHD Neonatal Research Network. Heptavalent pneumococcal conjugate vaccine immunogenicity in very-low-birth-weight, premature infants. Pediatr Infect Dis J. Jul; 29(7):600–606. [PubMed: 20234331]
- Wernette CM, Frasch CE, Madore D, et al. Enzyme-linked immunosorbent assay for quantitation of human antibodies to pneumococcal polysaccharides. Clin Diagn Lab Immunol. Jul; 2003 10(4): 514–519.
- Black S, Shinefield H, Fireman B, et al. Efficacy, safety and immunogenicity of heptavalent pneumococcal conjugate vaccine in children. Northern California Kaiser Permanente Vaccine Study Center Group. Pediatr Infect Dis J. Mar; 2000 19(3):187–195. [PubMed: 10749457]
- Ghaffar F, Friedland IR, McCracken GH Jr. Dynamics of nasopharyngeal colonization by Streptococcus pneumoniae. Pediatr Infect Dis J. Jul; 1999 18(7):638–646. [PubMed: 10440444]
- 5. Pelton SI, Loughlin AM, Marchant CD. Seven valent pneumococcal conjugate vaccine immunization in two Boston communities: changes in serotypes and antimicrobial susceptibility

Arch Pediatr Adolesc Med. Author manuscript; available in PMC 2012 January 25.

among Streptococcus pneumoniae isolates. Pediatr Infect Dis J. Nov; 2004 23(11):1015–1022. [PubMed: 15545856]

 Wikler, MA.; Bush, K.; Cockerill, FR., III, et al. Performance Standards for Antimicrobial Susceptibility Testing : Eighteenth Informational Supplement. Clinical and Laboratory Standards Institute; Wayne, PA: 2008. **NIH-PA** Author Manuscript

# Table 1

~	
· · · ·	
÷Ξ	
. <u> </u>	
<u> </u>	
.≘	
q	
<u>o</u>	
2	
~~~	
S	
сa,	
. 3	
4	
2	
5	
· Ħ	
E	
·Ħ	
Б	
a	
·2	
H	
~~~	
Q	
_ <u>P</u>	
$\mathbf{P}$	
5	
Ĕ	
e)	
$\mathbf{S}$	
Ĵ,	
Ē	
50	
·	
õ	
3	
<u> </u>	
늰	
Ч	
.5	
Ū.	
5	
٠Ă	
5	
õ	
Ē	
0	
õ	
ч	
of	
o f	
ce of	
ace of	
lace of	
place of	
d place of	
nd place of	
and place of	
e and place of	
ne and place of	
me and place of	
time and place of	
: time and place of	
s: time and place of	
tes: time and place of	
ates: time and place of	
plates: time and place of	
solates: time and place of	
isolates: time and place of	
) isolates: time and place of	
P) isolates: time and place of	
VP) isolates: time and place of	
(NP) isolates: time and place of	
(NP) isolates: time and place of	
al (NP) isolates: time and place of	
eal (NP) isolates: time and place of	
geal (NP) isolates: time and place of	
ngeal (NP) isolates: time and place of	
yngeal (NP) isolates: time and place of	
rryngeal (NP) isolates: time and place of	
naryngeal (NP) isolates: time and place of	
haryngeal (NP) isolates: time and place of	
pharyngeal (NP) isolates: time and place of	
sopharyngeal (NP) isolates: time and place of	
asopharyngeal (NP) isolates: time and place of	
nasopharyngeal (NP) isolates: time and place of	
l nasopharyngeal (NP) isolates: time and place of	
al nasopharyngeal (NP) isolates: time and place of	
cal nasopharyngeal (NP) isolates: time and place of	
occal nasopharyngeal (NP) isolates: time and place of	
occal nasopharyngeal (NP) isolates: time and place of	
ococcal nasopharyngeal (NP) isolates: time and place of	
nococcal nasopharyngeal (NP) isolates: time and place of	
mococcal nasopharyngeal (NP) isolates: time and place of	
umococcal nasopharyngeal (NP) isolates: time and place of	
eumococcal nasopharyngeal (NP) isolates: time and place of	
'neumococcal nasopharyngeal (NP) isolates: time and place of	

Clindamycin MIC <sup>e</sup> µg/m	0.12 (S)	0.25 (S)	0.5 (I)	0.06 (S)	0.12 (S)	0.12 (S)	>256 (R)	0.25 (S)	>256 (R)	0.125 (S)	0.25 (S)	0.25 (S)	0.25 (S)	0.5 (I)	0.25 (S)	$N/A^{\mathcal{B}}$
Erythromycin MIC <sup>e</sup> μg/ml	32 (R)	0.25(S)	0.25(S)	0.06 (S)	0.25 (S)	0.25 (S)	>256 (R)	0.25 (S)	>256 (R)	16 (R)	0.25 (S)	0.25 (S)	0.12 (S)	0.25 (S)	0.25 (S)	N/A <sup>g</sup>
Ceftriaxone MIC <sup>e</sup> μg/ml	0.03 (S)	0.03 (S)	0.016 (S)	1 (S)	0.03 (S)	0.03 (S)	2 (I)	0.12 (S)	2 (I)	0.06 (S)	0.12 (S)	0.12 (S)	0.03 (S)	0.016 (S)	0.03 (S)	$N/A^{\mathcal{G}}$
Penicillin MIC <sup>e</sup> μg/ml	0.12 (I)	0.03 (S)	0.03 (S)	2 (R)	0.03 (S)	0.03 (S)	2 (R)	0.12 (I)	2 (R)	0.38 (I)	0.06 (S)	0.12 (I)	0.06 (S)	0.03 (S)	0.06 (S)	$N/A^{\mathcal{B}}$
Serotype <sup>d</sup>	Non-typeable	38	19F	35B	10A	35F	19A	19A	19A	Non-typeable	6A	6A	38	15C	23A	$N/A^{\mathcal{B}}$
Birth weight in grams	1035	1100	1450	1100	750	1185	1230	910	925	1480	1405	1365	1216	1286	680	650
Post- discharge days when NP swab was obtained <sup>c</sup>	-55 (still in NICU)	10	20	166	122	168	189	139	120	189	823	235	159	208	182	182
NP Swab collection place <sup>b</sup>	NICU	PCP	PCP	PCP	PCP	PCP	PCP	PCP	PCP	PCP	PCP	PCP	PCP	PCP	PCP	PCP
NP swab collection time <sup>a</sup>	Pre	$\mathrm{Pre}^{f}$	Pre	$\operatorname{Post}^{f}$	Post	Post	Post	Post	Post	Post	Post	Post	Post	Post	Post	Post

Arch Pediatr Adolesc Med. Author manuscript; available in PMC 2012 January 25.

"Before the first heptavalent pneumococal conjugate vaccine (PCV-7) dose (PRE); 4-6 weeks after the third PCV-7 dose (POST).

 $b_{\rm Neonatal}$  Intensive Care Unit (NICU) or Primary Care Provider's (PCP) office.

 $^{c}$ The13 POST isolates were recovered at a median of 179 days after hospital discharge (range 120-238 days).

d Serotyped at the Streptococcus Laboratory of Centers for Disease Control and Prevention (CDC), Atlanta, GA using latex agglutination and confirmation by quellung reaction with type-specific pneumococcal antisera.

e<sup>e</sup>E-test method (AB Biodisk, Solna, Sweden); Minimum inhibitory concentration (MIC) in μg/ml, interpreted as: susceptible (S), intermediate (I) and resistant (R).<sup>6</sup>

Г

Ang et al.

 $\mathcal{S}_{\text{Isolate not viable for further testing.}}$ 

 $f_{
m The\ same\ infant.}$ 

Page 6

Arch Pediatr Adolesc Med. Author manuscript; available in PMC 2012 January 25.