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# Colonization, Pathogenicity, Host Susceptibility and Therapeutics for *Staphylococcus aureus*: What is the Clinical Relevance?<sup>1</sup>

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# Abstract

*Staphylococcus aureus* is a human commensal that can also cause a broad spectrum of clinical disease. Factors associated with clinical disease are myriad and dynamic and include pathogen virulence, antimicrobial resistance and host susceptibility. Additionally, infection control measures aimed at the environmental niches of *S. aureus* and therapeutic advances continue to impact upon the incidence and outcomes of staphylococcal infections. This review article focuses on the clinical relevance of advances in our understanding of staphylococcal colonization, virulence, host susceptibility and therapeutics.

Over the past decade key developments have arisen. First, rates of nosocomial methicillin-resistant *S. aureus* (MRSA) infections have significantly declined in many countries. Second, we have made great strides in our understanding of the molecular pathogenesis of *S. aureus* in general and community-associated MRSA in particular. Third, host risk factors for invasive staphylococcal infections, such as advancing age, increasing numbers of invasive medical interventions, and a growing proportion of patients with healthcare contact, remain dynamic. Finally, several new antimicrobial agents active against MRSA have become available for clinical use.

Humans and *S. aureus* co-exist and the dynamic interface between host, pathogen and our attempts to influence these interactions will continue to rapidly change. Although progress has been made in the past decade, we are likely to face further surprises such as the recent waves of community-associated MRSA.

# Keywords

Staphylococcus aureus; methicillin-resistant; MRSA; epidemiology; pathogenesis; treatment

# Background

The epidemiology and clinical manifestations of any infectious disease are influenced by several factors including the pathogen, the host, the environment and therapeutic advances.

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The story of *Staphylococcus aureus*, an incredibly versatile organism that is normally a human commensal, demonstrates how these factors result in a dynamic and rapidly shifting landscape. This article will begin by defining key concepts relevant to *S. aureus*, followed by a review of recent developments in the interactions between an organism that is becoming more resistant to antibiotics, a host population that is undergoing more medical procedures, and the host population's attempts to change the environment and course of disease.

#### **Clinical syndromes**

*S. aureus* is a gram-positive coccus with numerous virulence factors and the ability to acquire antibiotic resistance determinants [1]. Skin and skin structure infections (SSSI) represent approximately 90% of all *S. aureus* infections and thus the major burden of staphylococcal disease [2–6]. However, infections of the bloodstream, respiratory tract, bone and joint, surgical wounds, and increasingly medical devices are particularly feared due to the high morbidity and mortality and prolonged treatment required. More recently, severe manifestations of community-associated disease such as fulminant sepsis [7], the Waterhouse-Friderichsen syndrome [8] and necrotizing pneumonia [9,10] have become prevalent. The rise of antibiotic resistance has further compromised effectiveness of existing antimicrobial agents. Thus, despite improvements in medical care, mortality from *S. aureus* bacteremia continues to be 20–30% in the developed world [11].

#### Antibiotic resistance

Following the introduction of penicillin in the mid-1940s, penicillin-resistant strains of S. aureus were soon reported [12] and this was followed by a pandemic of penicillin-resistant S. aureus [13]. Although initially prevalent only in hospitals, penicillin-resistance is now present in over 90% of community isolates. These strains produce a plasmid-encoded penicillinase that disrupts the  $\beta$ -lactam ring of penicillin. Methicillin, a penicillinase stable β-lactam, was introduced in the late 1950s; however, reports of methicillin-resistance rapidly appeared [14]. The mechanism of resistance to methicillin involves an altered and low affinity penicillin-binding protein (PBP2a) that is encoded by the mecA gene, which confers a broad resistance against all members of the  $\beta$ -lactam antibiotics. The *mecA* gene is carried on the mobile genetic element staphylococcal chromosome cassette (SCC) with the entire complex termed the SCCmec element. There are now 11 defined variants of SCCmec (types I to XI) (see http://www.sccmec.org/) that differ in size and composition of antimicrobial resistance elements. Typically, multi-resistant nosocomial strains of MRSA harbor SCCmecII and SCCmecIII, which are larger and include multiple resistance determinants. On the other hand, the more recent community-associated MRSA strains harbor the smaller SCCmecIV, which carry fewer resistance elements and thus often retain susceptibility to macrolides, quinolones, tetracyclines, trimethoprim-sulfamethoxazole and lincosamides. Moreover, the smaller size of SCCmecIV has been postulated to allow it to be more mobile and supportive evidence of this is the fact that SCCmecIV has been inserted into multiple lineages of S. aureus whereas SCCmecII and SCCmecIII have only been found in three and two lineages respectively [15].

#### Molecular genotypes

Early studies using phage typing established the utility of discriminating different strains or lineages of *S. aureus* [16]. Currently, the most commonly used techniques for molecular genotyping of *S. aureus* are pulsed-field gel electrophoresis (PFGE) and sequence based methods such as multilocus sequence typing (MLST) and *spa* typing. PFGE involves separating *Sma*I-digested DNA fragments of the genome by size in agarose gel and provides a very fine level of resolution. PFGE is limited by difficulties with inter-laboratory standardizations and portability, although it continues to be widely used in the United States,

and the Centers for Disease Control and Prevention (CDC) has developed a national *S. aureus* PFGE database [17]. Sequence based approaches have the advantage of producing unambiguous and reproducible results that can be compared on web-based databases. MLST involves the sequencing of 450–500bp fragments of seven housekeeping genes [18]. By assigning sequences for each fragment to different alleles, the combination of alleles can be designated to a unique sequence type (ST). There are now over 2000 STs on the *S. aureus* MLST database (http://saureus.mlst.net/). Sequencing the *spa* locus, a rapidly evolving hypervariable region of the genome, is simpler, because it only requires the sequence from one gene, and in general produces results concordant with MLST [19]. More recently, whole genome sequencing with next generation sequencing technologies has become an incredibly powerful means of determining the genetic make-up and relationships between *S. aureus* isolates [20].

The widespread uptake of MLST has allowed a much deeper understanding of the global population structure of *S. aureus*. It appears that there are distinct lineages or clonal complexes (CCs) of *S. aureus* and that these tend to evolve through point mutation rather than recombination [21,22]. However, it is also clear that virulence and resistance genes are frequently acquired through horizontal gene transfer onto what is a relatively stable genetic background [23].

#### Genotypic and epidemiologic definitions

Although molecular genotyping techniques are useful in determining the molecular epidemiology of S. aureus, such techniques are rarely available in standard diagnostic laboratories or sufficiently rapidly performed to guide clinicians in real time. Thus definitions based on epidemiological and resistance phenotype patterns are more commonly used. The CDC definition divides infections into nosocomial (onset of infection >48 hours after hospital admission), community-onset healthcare-associated (onset of infection in the community or <48 hours after hospital admission and the presence of  $\geq 1$  of the following risk factors: a history of hospitalization, surgery, dialysis, or residence in a long-term care facility within one year before the culture date; or the presence of a permanent indwelling catheter or percutaneous medical device at the time of culture; or previous isolation of MRSA), and community-associated (onset of infection in the community or <48 hours after hospital admission with none of the above risk factors) [2]. Resistance phenotype definitions group isolates as multiresistant (resistant to  $\geq 3$  non  $\beta$ -lactam classes of antibiotics) or nonmultiresistant (resistant to <3 non  $\beta$ -lactam classes of antibiotics) [24]. There is obvious overlap between the genotypic, epidemiologic and resistance phenotypic definitions, particularly with the blurring of boundaries and encroachment of initially communityassociated MRSA strains, such as USA300 into hospital environments, and also the increase in breadth of resistance of certain strains [25].

# The environment: colonization and interventions to change the environment

*S. aureus* is a common human commensal. Approximately 30% of healthy adults are colonized with the anterior nares being the typical site of carriage. However, extra-nasal sites of *S. aureus* colonization include the skin, perineum, gastrointestinal tract and the throat. Longitudinal studies have revealed that individuals can be non-carriers, intermittent carriers and persistent carriers [26,27]. The risk of developing a healthcare-associated staphylococcal infection is three to six times increased among nasal carriers with a large bacterial load compared to non-carriers or those with a low bacterial load [28,29] with nasal colonizing strains usually being the source of infection [30,31]. More recently, it has become apparent that the patterns of carriage of community-associated MRSA may differ

from that previously recognized for healthcare-associated MRSA. A substantial proportion of those colonized with community-associated MRSA appear to be colonized at body sites outside of the anterior nares. For example, one study found that 23% of patients colonized with community-associated MRSA were colonized at non-nasal sites (predominantly inguinal regions) [32] and in children with SSSI the rectum was found to be the key site of colonization [33]. Young, healthy individuals appear to be at higher risk of exclusive throat carriage. Of healthy blood donors colonized with *S. aureus*, 30% carried *S. aureus* in the throat only [34]. Furthermore, the household environment has also been found as a reservoir [35]. Even in the Intensive Care Unit (ICU) setting, the throat and rectum are important sites of carriage of MRSA [36].

The clinical importance of understanding the sites, patterns and methods of detection of colonization lies in the ability to intervene through infection control programs and to prevent clinical infections in colonized hosts. For example, various interventions involving nasal decolonization have been shown to be effective in reducing rates of S. aureus bacteremia (SAB) in hemodialysis patients [37], MRSA wound infections post cardiothoracic surgery [38] and surgical site infections [39]. Indeed, for the first time since the global epidemic of MRSA in hospitals, the past five years has seen notable successes in reducing rates of MRSA infection. In three Illinois hospitals between 2003–2007, universal screening of all admissions for MRSA followed by topical decolonization and contact isolation of MRSA colonized patients resulted in a 70% reduction in rates of hospital-associated MRSA infections [40]. In the United States, the Veterans Affairs hospitals universally implemented a "MRSA bundle", consisting of universal active surveillance, contact precautions for those colonized or infected with MRSA, a hand hygiene campaign and programs to stimulate institutional culture change, resulting in a 62% reduction in the incidence of MRSA-related healthcare-associated infections from 1.64 infections per 1000 patient-days to 0.62 per 1000 patient days [41]. The incidence of central line-associated bloodstream infections (CLABSI) due to MRSA in ICUs has significantly reduced from 2001–2007, together with an even greater reduction in CLABSI due to methicillin-susceptible S. aureus (MSSA) [42]. Strikingly, over the period 2005–2008 the incidence of invasive healthcare-associated MRSA infections decreased by 9.4% per year [43].

These reductions in incidence of MRSA infections have not been limited to the United States. In England, following widespread efforts and interventions to combat MRSA, there has been a 57% reduction in MRSA bacteremia from 2004–2008 [44]. In France, sustained reductions over the past decade have also been evident [45], and the Netherlands continues to maintain an incredibly low incidence of MRSA infections [46]. Similarly, in Australia, concerted efforts to improve hand hygiene compliance resulted in significant reductions in rates of MRSA infections and bacteremias on a statewide level [47].

These encouraging indications from the above observational studies need to be tempered by the results of two large and well-designed clinical trials that found no statistically significant reduction in MRSA rates with the use of MRSA nasal surveillance and isolation precautions for MRSA carriers [48,49]. Therefore, it remains unresolved as to which interventions are effective, since multiple interventions targeting MRSA are typically implemented during observational studies. In addition, the publication and implementation of guidelines to prevent CLABSIs and ventilator-associated pneumonias may have occurred concurrently. Edgeworth argues that the decolonization of patients with mupirocin and skin antisepsis agents has likely played an under-recognized role in the success of these programs [44]. Despite these caveats, the balance of evidence indicates that the bundled interventions being administered are proving effective in reducing rates of MRSA in hospitals.

However, an even greater challenge will be to find interventions that are successful in the community where community-associated MRSA appears to be adding to the general burden of staphylococcal disease [50] and where high risk populations may be acting as reservoir and vectors for the amplification and wide-dissemination of community-associated MRSA strains [51,6]. Moreover, traditional approaches to surveillance and decolonization may not apply to community-associated MRSA due to its preponderance for colonization of extranasal sites such as the throat and rectum. Attempts to reduce rates of MRSA infections by screening and nasal decolonization of MRSA carriers in the setting of military and sporting communities have so far proved unsuccessful [52,53].

# The pathogen: Organism related determinants of outcome

*S. aureus* has a large arsenal of virulence factors that allow it to evade host immune responses and to cause clinical disease. These comprise cell surface proteins, extracellular enzymes and exotoxins. Although *in vitro* studies and the use of animal models have provided many insights into the role of these various factors, there has been little to correlate the presence and expression of such factors with clinical disease and outcomes. Indeed, few therapeutic interventions are currently available that have applied this knowledge. Due to the epidemic of USA300 community-associated MRSA, much of the recent research described here has focused on factors associated with USA300, and findings from USA300 may not be applicable to other strains of *S. aureus*. It will be important for similar studies to be conducted with other strains of *S. aureus*.

#### The Panton-Valentine leukocidin controversy

One of the initial observations regarding community-associated MRSA was that the majority of strains harbored the genes encoding for Panton-Valentine leukocidin (PVL). PVL is a bicomponent toxin that forms pores in neutrophils. Early epidemiological studies highlighted the association between PVL and necrotizing pneumonia, furunculosis and severe bone and joint infections [54,9,55]. An early paper described an association between PVL and lethal necrotizing pneumonia in young adults but the overall numbers were small with only 16 cases of PVL+ pneumonia of which 8 were recruited retrospectively [9]. Further cases of necrotizing pneumonia associated with PVL, in both MRSA and MSSA infections and often associated with influenza and poor outcomes, have been reported [56,10,57-59]. In Australia, where non-PVL lineages of community-associated MRSA from clonal complex 1 circulate [60], and also where the majority of PVL+ isolates were found to be MSSA in northern Australia [61], PVL has consistently been linked to furunculosis [61–63], but not to poorer outcomes [61,64]. Recent studies have failed to find an association between the presence of PVL and poorer outcomes in complicated skin and skin structure infections (cSSSI) [65,66] (Tong et al., Abstract C2-1287 at 50<sup>th</sup> Interscience Conference for Antimicrobial Agents and Chemotherapy, Boston, September 12-15, 2010), hospital acquired pneumonia [67] or invasive disease [68].

Conflicting results from experimental studies have also led to a significant degree of controversy regarding the pathogenic role of PVL. In one study using a mouse model, it was reported that PVL was directly causative of necrotizing pneumonia [69]. However, it was subsequently determined that an unintended point mutation in the *agr* promoter of the *S. aureus* isolate used resulted in defective virulence gene regulation and explained the observed phenotype [70]. Furthermore, mammalian neutrophils from different species differ in their susceptibility to PVL. Mouse neutrophils are more resistant than human and rabbit neutrophils to PVL [71–74]. Therefore mouse models may not be appropriate for determining the role of PVL in human disease and could explain the negative findings from several studies [75,76]. However, even the use of rabbit models (rabbit neutrophils are susceptible to PVL), has not resolved the controversy. A rabbit model of necrotizing

pneumonia compared the virulence of a USA300 wild-type strain with that of an isogenic PVL-deletion mutant and found that expression of PVL resulted in increased pathogenicity [77]. Two recent papers using a rabbit SSSI model came to conflicting conclusions about the role of PVL [78,79]. To complicate matters further, Yoong and Pier found in a low-inoculum murine skin abscess model that PVL stimulates a protective host immune response that is abrogated by antibodies to PVL [80]. However, epidemiological evidence linking a history of past PVL infection (using the history of a previous furuncle as a surrogate) with protection against death in PVL-associated pneumonia has also been reported [81]. Thus, the results of a Phase 1–2 vaccine trial of a vaccine containing a PVL component are eagerly awaited (ClinicalTrials.gov NCT01011335, study completed in March 2011).

Apart from PVL, phenol-soluble-modulins (PSMs) and alpha-toxins have been investigated as factors important for lysis of neutrophils. Alpha-toxin is another pore-forming toxin that has been found to be an important virulence factor in murine pneumonia and skin infection models [82,83]. PSMs are amphipathic, alpha-helical peptides that have cytolytic activity. PSMs have been shown to contribute significantly to community-associated MRSA virulence [84] and to enhance the activity of PVL in lysis of human neutrophils [73]. Notably, a recent study has used a rabbit skin infection model to compare the relative virulence of a wildtype USA300 isolate with isogenic deletion mutants for PVL, alpha-toxin and PSMα. These investigators found that alpha-toxin and PSMα played more important roles than PVL as virulence determinants for this USA300 strain [79]. A novel bicomponent leukotoxin named LukGH has recently been identified through cell surface proteomics of USA300 strain LAC [85]. LukGH was found to have cytolytic activity towards neutrophils and is potentially a novel virulence factor for USA300.

In summary, there is strong evidence for an epidemiological link between PVL and furunculosis and necrotizing pneumonia. The outcomes from cSSSI and hospital acquired pneumonia appear to not be different for PVL+ and PVL- disease. This may not be the case with community-acquired PVL+ necrotizing pneumonia, which is often associated with influenza and frequently results in poor outcomes. There continues to be conflicting evidence of the role of PVL in animal models of SSSI and pneumonia but it is becoming clearer that PVL is not the sole or dominant factor contributing to virulence of *S. aureus*.

Nonetheless, some clinicians are now moving towards treating severe PVL+ disease with therapies targeted at the PVL toxin. Published recommendations from France suggest that for severe SSTI, severe bone and joint infections, and necrotizing pneumonia, that antitoxin agents that inhibit protein synthesis such as clindamycin, rifampicin or linezolid should be added to standard bactericidal agents and that intravenous immunoglobulin (IVIg) be considered for necrotizing pneumonia and refractory SSSI [86]. There is evidence that inhibitors of protein synthesis do reduce the production of PVL [87,88], however there is a cautionary note that the production of PSMs may be increased with sub-inhibitory concentrations of such agents [89]. For necrotizing pneumonia, the recommendations acknowledge that the use of IVIg is only supported by *in vitro* data and a handful of case reports [90,91].

#### Staphyloxanthin

*Staphylococcus aureus* is named as such due to the golden color of colonies in growth media (aureus = "golden", Latin). Recent work has demonstrated the importance of the carotenoid pigment, staphyloxanthin, to the ability of *S. aureus* to resist killing by neutrophils. Loss of pigmentation results in reduced virulence in murine skin and sepsis models [92]. Additionally, inhibition of staphyloxanthin biosynthesis resulted in improved clearance of *S. aureus* in a murine intra-peritoneal sepsis model, although no mention is made of an impact on murine survival [93]. Interestingly, an isolate from an early-branching and

phylogenetically divergent *S. aureus* lineage, clonal complex 75, has been shown to lack the genes necessary for synthesis of staphyloxanthin and 126 isolates from this lineage were found to all be non-pigmented [94]. Whether the acquisition of genes for staphyloxanthin biosynthesis was a significant event in the evolution of *S. aureus* as a virulent species remains to be determined.

#### Underlying genetic background

A further question of interest is whether S. aureus strains of distinct lineages differ in their virulence and the clinical diseases caused. Earlier studies using MLST data demonstrated that there was no difference in the populations of S. aureus that caused asymptomatic nasal carriage compared to a diverse range of severe disease [95,96]. Similarly, using amplified fragment length polymorphism analysis, it was shown that all lineages found in carriage isolates were also present in invasive isolates. However, some lineages appeared to be more frequently associated with impetigo isolates [97]. Focused studies have implicated CC5 and CC30 as more likely to cause bacteremia with hematogenous complications [98]. CC30 was also more likely to be a cause of persistent bacteremia [99] and over-represented in isolates from infective endocarditis as compared to SSSIs [100]. The reasons for why CC30 might be over-represented in these severe forms of infection are unknown. Recent evidence demonstrating that the same CC30 isolates that are associated with an increased risk for endocarditis in humans are also paradoxically less likely to cause septic death in two in vivo models implies that these contemporary CC30 isolates may have evolved an increased tendency to bind to host tissues at the expense of specific toxin production (Sharma et al., Abstract B-060/60 at 51st Interscience Conference for Antimicrobial Agents and Chemotherapy, Chicago, September 17-20, 2011). A fascinating study from Deleo and colleagues provides support for this potential explanation. In their work, they elegantly demonstrate how contemporary CC30 isolates, when compared to isolates of the historical hypervirulent 80/81 clone from the 1950s and 60s, contained non-synonymous SNPs in genes encoding accessory gene regulator C (agrC) and alpha hemolysin (*hla*) and had reduced virulence in vivo (DeLeo et al., Proc Natl Acad Sci USA 2011 in press).

Other studies have corroborated this surprising finding that even within monophyletic lineages significant differences in virulence may exist. In a genomic study of 10 USA300 isolates, isolates that were very closely related on a genome level demonstrated substantial differences in their virulence as determined in a murine sepsis model [101]. Similarly, single amino acid polymorphisms in the fibronectin binding protein A of *S. aureus* can result in an increased force of binding between *S. aureus* cells and fibronectin (Lower et al., Proc Natl Acad Sci USA 2011 in press). These single amino acid polymorphisms were found to be more common in SAB isolates from patients with infected cardiac devices compared to those from patients with uninfected cardiac devices. Collectively, these studies suggest that for *S. aureus*, as for other pathogens [102], determining variation at the single nucleotide level across the whole genome will be necessary to truly understand and elucidate associations with virulence in clinical disease.

# The human host: Host related determinants of outcome

Perhaps of greater importance than pathogen virulence factors, many characteristics of the human host, whether intrinsic or acquired, have a major influence on clinical outcomes – ranging from establishing *S. aureus* as an asymptomatic colonizer to deleterious and fulminant invasive staphylococcal infection.

For some time now, researchers and clinicians alike have known that *S. aureus* infections may be associated with specific racial, social and physical attributes of the human host. Age, sex and racial origin are three important characteristics that determine risk of *S. aureus* 

colonization and infection. For example, a secondary analysis of the 2001–2002 National Health and Nutrition Examination Survey (NHANES) in which 9,622 participants contributed microbiologic data showed women and patients aged 65 years and older were more likely to have MRSA colonization [103]. In addition to risk of colonization, older age group was also a risk factor for higher mortality if a patient became infected with *S. aureus*. A recent study of patients with *S. aureus* bacteremia found that patients aged 80 or older had substantially higher mortality rates [104]. The authors of the study went on to conclude that specific interventions to optimize clinical care practices in elderly patients with *S. aureus* bacteremia are essential given the compelling data on the disproportionately high rates of morbidity and mortality.

Racial origin is also a major determinant of risk of S. aureus colonization and infection at the population level. In the same aforementioned NHANES study, white persons were found to have statistically higher risks of S. aureus colonization than Hispanic or Black persons [103]. Despite this lower rate of colonization, African Americans were significantly more likely to develop invasive MRSA infections than whites [43,105]. Indigenous populations of different continents also appear to be at high risk for the acquisition and infection with community-associated MRSA as seen in the Native American, Pacific Islander, and Australian Aboriginal populations [51]. The reasons for these higher infection rates in racially distinct populations are unknown, and likely complex. At least some of these differences in infection rates may relate to sociocultural rather than genetic issues. For example, culturally unique behaviors among different populations may be more relevant in the pathogenesis of some forms of S. aureus infection. Other economic factors such as living in crowded conditions, lack of access to healthcare and suboptimal personal hygiene practices, are also likely to be relevant. In support of the role of these lifestyle issues is the observation that crowding, communal living conditions and sharing of personal items are thought to be important contributors to community-associated MRSA outbreaks among incarcerated inmates [106], military recruits [107] and professional athletes [108].

#### Skin and mucosal protection against S. aureus

There is no greater barrier against *S. aureus* than our skin and mucosal defenses. Data clearly show that *S. aureus* infections occur more frequently when the normal protective defenses of human skin are weakened or if they are breached, especially if there is a high burden of *S. aureus* colonization or if personal hygiene practices are lacking. Early evidence of the protective importance of human skin came from patients who had skin conditions and high rates of *S. aureus* colonization. For instance, a point-prevalence study of 54 patients with atopic dermatitis attending the dermatology clinic of the Children's Hospital of Philadelphia showed that greater than 80% were colonized with MRSA [109] – a colonization rate that is substantially higher than what is expected in the general population (~3%).

Within the community setting, protective skin barriers may be breached as a result of simple trauma during shaving of facial or body hair or from more serious injuries sustained during sports and training. Interestingly, the simple act of shaving body hair was identified as a specific risk factor associated with MRSA infections among college football players during an outbreak in Connecticut [110]. "Turf burns" and other more serious skin abrasions sustained during sports are much more obvious portals of entry for *S. aureus* colonizing the skin. An outbreak of community-associated MRSA SSSI among St. Louis Ram's football players demonstrated that infections often occurred at the sites of skin abrasions and turf burns [111]. The authors of the report also suggested that appropriate treatment and coverage of such skin abrasions may have helped to prevent community-associated MRSA infections seen in the outbreak.

Two other activities in the community setting are associated with high risks of *S. aureus* infection, namely tattooing and injection drug use. Both such activities may occur with contaminated sharps that can introduce *S. aureus* directly into soft tissue and the vascular system, respectively. For example, a 2006 report from the CDC revealed that 34 cases of community-associated MRSA infections were reported in patients who received tattoos from unlicensed tattooists who did not routinely perform hand hygiene, skin antisepsis or adequate disinfection of equipment and surfaces [112].

#### **Immune Deficiencies**

Immune competency is also an important part of the body's defense against *S. aureus*. There are inherited and acquired immunodeficiency states that are well associated with *S. aureus* infections. Inherited defects in white blood cell function or immune responses such as present in chronic granulomatous disease (CGD), Job's syndrome, Chediak-Higashi syndrome, and Wiskott-Aldrich syndrome all predispose to recurrent staphylococcal infections [113]. The key role of II-1 and II-17 in the immune response to *S. aureus* infections of the skin has recently been reviewed [114]. On the other hand, defects in specific cellular immunity are generally not closely associated with infections with *S. aureus*. Even though some studies have shown that HIV-infected patients are at higher risk of *S. aureus* infections, much of the link between *S. aureus* infection in HIV-infected patients are at higher risk or shown that high-risk behaviors. Interestingly, regardless of HIV-infection status, men who have sex with men have emerged as an important and independent risk factor for development of community-associated MRSA infections over the last decade [115].

#### **Medical comorbidities and Healthcare Contact**

Many medical comorbidities have been found to be important and manageable risk factors for *S. aureus* colonization and infection, including peritoneal dialysis or hemodialysis, diabetes mellitus, and rheumatoid arthritis [116]. There are many thoughts as to how these comorbidities increase risk of *S. aureus* infection. First, some of these comorbidities are associated with functional defects of the immune system. For instance, poorly-controlled diabetes mellitus is known to be associated with decreased neutrophil function and decreased cellular chemotaxis [117]. Similarly, patients with uremia or who are receiving dialysis have a degree of neutrophil dysfunction as well as diminished antibody production and opsonic capabilities.

Second, many of the aforementioned comorbidities may result in regular healthcare contact, more hospitalizations and frequent invasive procedures – all of which are well-understood major risk factors for *S. aureus* colonization and invasive disease [118,119]. In particular, breaches of the skin and mucosal defense with intravascular lines and hemodialysis needling are key risk factors. Furthermore, frequent healthcare contact often culminates in antibiotic use (whether appropriate or not), which is also is independently associated with acquisition of MRSA [120].

Finally, comorbid conditions can result in reduced functional status and an inability to independently perform personal hygiene and skin care, leading to higher rates of *S. aureus* colonization and infection. A case-control study at 7 hospitals in the southeastern region of the United States showed that poor functional status (defined as requiring assistance with any activity of daily living) was highly associated with infection with MRSA, particularly surgical site infections [121]. Additionally, if patients with multiple comorbidities and poor functional status were admitted to long-term care facilities (LTCF), the risk of MRSA colonization and infection also increases [122]. Specific characteristics in LTCFs have been

found as significant risk factors for MRSA colonization and infection: low ratio of nurses to patient beds, location of facility in a deprived area, and a stay of more than 6 months in the LTCF.

It is important to remember that modern medicine can offer new diagnostic and therapeutic options through the use of indwelling lines and other invasive procedures. Such opportunities come at a great risk of causing debilitating or even, lethal, *S. aureus* infections. Indeed, there has been a notable shift in the epidemiology of infective endocarditis (IE) over the past 20 years with *S. aureus* now the leading cause of IE in many regions of the world and much of this being healthcare-associated [123]. This sobering reminder underscores the need for increased vigilance and the urgency for a collective and aggressive response to reduce the threat of *S. aureus* infection.

## Therapeutics: attempts to change outcomes

There are several overarching principles in the management of *S. aureus* infections. Two such important principles are grounded in basic science and studies on pathogenesis of staphylococci and the conclusions correlate well with data and outcomes from clinical studies.

*S. aureus* can form biofilm which facilitates its survival and multiplication on infected surfaces and prosthetic. In addition, the biofilm can provide protection against both host immune defenses as well as antibiotic agents, potentially prolonging the duration of organism exposure to antibiotics and promoting the possible opportunity for transfer of antibiotic resistance genes between organisms [124]. Thus, one of the most important management principles for invasive *S. aureus* infection is to completely remove infected tissue and/or prosthetic material [125] – to minimize the ability for *S. aureus* to persist and relapse.

Direct drainage of an abscess is another tenet in the management of *S. aureus* infections [126]. Models of staphylococcal abscess clearly demonstrate the ability of *S. aureus* to replicate in the center of an abscess, separated from surrounding immune cells and protected from high levels of antibiotics [127]. In fact, many smaller *S. aureus* abscesses may be adequately treated with drainage alone; the converse is also true, that treatment failure frequently occurs if antibiotics are used while the staphylococcal abscess remains undrained.

Over the last 30 years, oxacillins (nafcillin and flucloxacillin) and vancomycin have been the stalwarts of antibiotic therapy against MSSA and MRSA respectively. Recently, several new anti-staphylococcal antibiotics have become available. Furthermore, the quest for an effective vaccine to prevent staphylococcal infections continues. The following is a brief overview of recent drug and vaccine developments.

#### Linezolid

Linezolid is a synthetic oxazolidinone that is bacteriostatic against *S. aureus*. It acts by binding to the 23S portion of the 50S subunit of bacterial ribosomes and inhibits protein synthesis. The antimicrobial activity of linezolid is predicted by the 24-hour area under the time-concentration curve to minimum inhibitory concentration (MIC). Interestingly, the drug also displays moderate post-antibiotic effect against *S. aureus* [128].

Linezolid has been approved by the Food and Drug Administration (FDA) for the treatment of nosocomial pneumonia and cSSSI, including those due to MRSA [129]. The retrospective analysis of two randomized controlled trials (RCTs) claimed that linezolid was associated with improved survival and clinical cure compared to vancomycin for nosocomial

pneumonia due to MRSA [130]. However, these findings have been seriously questioned due to flaws in study design and a recent meta-analysis of RCTs involving 1641 patients found that linezolid was not superior to glycopeptide antibiotics [131]. This issue is hoped to be resolved when the results of a RCT of linezolid versus vancomycin for MRSA ventilator-associated pneumonia are published (ClinicalTrials.gov NCT00084266).

There are three important pharmacologic features of linezolid that require caution with prescription. Linezolid has been associated with dose-dependent myelosuppression that is usually reversible. Thrombocytopenia is the commonest pattern of haematologic change. Renal impairment, low baseline platelet count and prolonged use of linezolid beyond 14 days are all major risk factors for linezolid-associated thrombocytopenia. Thus, weekly complete blood counts should be monitored in patients receiving linezolid for more than 7–10 days [132]. Second, manifestations of mitochondrial dysfunction, such as neuropathy and lactic acidosis, have also been infrequently reported with linezolid use. Interestingly, some reports indicated that neuropathy tended to persist even after cessation of linezolid [133]. Third, linezolid is a weak inhibitor of monoamine oxidase (MAO) and can precipitate serotonin toxicity when administered with a non-selective MAO-Inhibitors such as a serotonin-reuptake inhibitor (SSRI) or a serotonin-noradrenergic reuptake inhibitor (SNRI) [134].

Although resistance to linezolid appears to be rare [135], possibly because its high cost limits widespread use, documented mechanisms include the horizontal acquisition of a *cfr* rRNA methyltransferase [136] and point mutations at the target site of 23S rRNA [137] and *rlmN* that encodes for a conserved RNA methyltransferase [138].

Finally, linezolid is available in both oral and intravenous formulations and the bioavailability of the oral form approaches 100%, which should be used whenever possible.

#### Tigecycline

Tigecycline is a glycylcycline antibiotic with acts on the 30S ribosomal subunit and prevents amino acid incorporation into bacterial peptide chains. Tigecycline is structurally related to minocycline and tetracycline, however, studies show that it remains active against most *S. aureus* isolates that are tetracycline resistant [139]. In addition to effective coverage against MRSA, tigecycline has broad gram-negative activity except for *Pseudomonas* and *Proteus spp.*. Currently, tigecycline has FDA approval for three common indications: treatment of cSSSI, complicated intra-abdominal infections and community-acquired pneumonia (CAP).

Tigecycline has several important pharmacological properties that require particular caution and consideration in prescription. First, tigecycline distributes extensively into lung and biliary tissue, however drugs levels are often low in serum, bone and joints and in the cerebrospinal fluid (CSF). Second, tigecycline has a high proportion of gastrointestinal adverse effects; nausea and vomiting have consistently been more frequent in the tigecycline compared to the comparator arms of phase III studies [140]. Finally, tigecycline has a Pregnancy Class D rating which is based on animal studies showing delayed bone development and increased incidence of fetal loss. Thus, it should not be used in pregnant women unless potential benefits clearly outweigh the potential risks to the mother and/or the fetus.

#### Ceftaroline

Ceftaroline is a novel cephalosporin with high affinity for the modified penicillin-binding protein (PBP 2A) and thus has *in vitro* activity against MRSA. Ceftaroline is different to recent new antibiotics with anti-MRSA coverage. First, ceftaroline also has activity against a narrow spectrum of gram-negative organisms; however, there is no intrinsic activity against

*Pseudomonas spp.*, or other gram-negative pathogens that produce beta-lactamases, including AmpCs, extended-spectrum beta-lactamases and *Klebsiella pneumoniae* carbapenemases [141]. Second, there are some emerging data to suggest that ceftaroline might be more efficacious for invasive infections. For example, a recent *in vivo* study indicated that ceftaroline is superior to daptomycin in sterilizing cardiac vegetations infected with *S. aureus* [142].

Two pairs of large RCTs have studied ceftaroline for treatment of cSSSI and CAP. CANVAS 1 & 2 trials were parallel phase III studies that found ceftaroline monotherapy to be non-inferior to the combination of vancomycin plus aztreonam in the treatment of cSSSI, including those infections caused by MRSA [143]. Although FOCUS 1 & 2 studies found ceftaroline to be non-inferior to ceftriaxone for treatment of moderately-severe CAP not requiring intensive care therapy, efficacy of ceftaroline on CAP due to MRSA was not specifically examined [144]. In these randomized studies, ceftaroline was well tolerated and there were no excess adverse effects compared to controls.

#### Daptomycin

Daptomycin is a cyclic lipopeptide with marketing approval for the treatment of cSSSIs and for SAB and right-sided infective endocarditis. There have been two international phase III studies of daptomycin at a dose of 4mg/kg intravenously (IV) for 7–14 days compared to vancomycin or penicillinase-resistant penicillins for cSSSIs [145]. Of clinically evaluable outcomes, daptomycin and comparator arms demonstrated successful treatment in 83% and 84% of patients respectively. Further analyses demonstrated that daptomycin treated patients required a shorter duration of IV therapy with an earlier clinical response [145,146].

In a RCT of daptomycin 6mg/kg IV compared to either, vancomycin or a  $\beta$ -lactam, plus an aminoglycoside, for the first four days of therapy, daptomycin demonstrated non-inferiority for treatment of SAB and right-sided endocarditis [147]. For MRSA bacteremia, the success rate was higher for the daptomycin treated patients (44% versus 32% for daptomycin and standard therapy respectively).

These data suggest that daptomycin is now clearly part of the armamentarium for the treatment of cSSSIs and SAB. Importantly, daptomycin should not be used for treatment of staphylococcal pneumonia as pulmonary surfactant inhibits the action of daptomycin [148] and creatine kinase levels should be monitored weekly during therapy. Also notable, and of significant concern, is that in the registrational multinational trial of daptomycin for SAB, treatment emergent resistance occurred in ~5% of daptomycin treated patients [147]. Others have also reported daptomycin resistance that appears related to heterogenously vancomycin intermediate *S. aureus* (hVISA) and VISA strains [149,150].

#### Telavancin

Telavancin is a lipoglycopeptide that disrupts peptidoglycan synthesis. Phase III clinical trials have been conducted for cSSSI and hospital-acquired pneumonia. The ATLAS studies enrolled 1867 patients with cSSSI and compared telavancin to vancomycin [151]. Of clinically evaluable patients 88% and 87% were cured with televancin and vancomycin respectively and for those with MRSA infections eradication rates were 90% and 85% respectively. Recently, the ATTAIN study found that telavancin was non-inferior to vancomycin for treatment of hospital-acquired pneumonia due to gram-positive pathogens with respective cure rates of 59% and 59% respectively [152].

## Vaccines

Given the large burden of disease due to *S. aureus*, concerns about increasing antimicrobial resistance and the growing numbers of patients with risks for invasive staphylococcal infections, there is a great need for an effective vaccine. However, to date, there have only been disappointing results in clinical trials for staphylococcal vaccines. The general topic of vaccines for *S. aureus* has been thoroughly addressed elsewhere [153–155] and here we briefly comment on human clinical trials conducted in the past 10 years.

StaphVAX<sup>®</sup>, produced by Nabi Biopharmaceuticals, is a polysaccharide conjugate vaccine that contains purified capsular antigens CP5 and CP8. A phase III clinical trial of StaphVAX<sup>®</sup> in hemodialysis patients showed no protection against bacteremia between 3 and 54 weeks, but a post-hoc analysis demonstrated efficacy up to 40 weeks post vaccination [156]. In a second unpublished study involving 3600 hemodialysis patients no protection was observed. There are now studies progressing with the use of a capsular polysaccharide conjugate vaccine (PentaStaph<sup>®</sup>, that has been purchased by GlaxoSmithKline from Nabi) that also contains antigens to teichoic acid and the secreted toxins alpha-toxin and PVL [155].

Merck's V710<sup>®</sup> vaccine contains IsdB, one of many microbial surface components recognizing adhesive matrix molecules (MSCRAMMs) of *S. aureus*. Use of V710 in a phase I study elicited promising immune responses [157] however a phase II/III trial in elective cardiothoracic surgery patients to prevent serious *S. aureus* infections has recently been terminated. Although full results have not yet been released, it is understood that the Data Monitoring Committee recommended termination of the study due to the observation that V710 was unlikely to demonstrate a statistically significant clinical benefit as well as safety concerns

(http://www.merck.com/newsroom/news-release-archive/research-and-development/2011\_0608.html). There is an ongoing trial of V710 in hemodialysis patients.

Passive immunization approaches also have not been successful in clinical trials. INH-A21 (Veronate) is a pooled human immunoglobulin preparation with high antibody titres against clumping factor A (ClfA) that was tested in a phase III trial involving low birth weight infants. It failed to reduce late onset sepsis and SAB in this cohort [158]. Other products now through phase II studies include tefibazumab (monoclonal antibodies to ClfA) [159], Altastaph<sup>®</sup> (antibodies to CP5 and CP8) [160,161] pagibaximab (monoclonal antibodies to lipoteichoic acid) [162] and Aurograb<sup>®</sup> (antibodies to ATP-binding cassette). Pagibaximab has proceeded to a phase 2b/3 trial.

All these results demonstrate the great difficulties in developing a vaccine against *S. aureus*. Questions remain as to the best combination of antigens, and also which patient population should be targeted for the use of vaccines or passive immunotherapy. The conundrum is that high-risk patients such as those on hemodialysis may have relatively poor immune responses to a vaccine.

# Conclusions

As long as *S. aureus* remains a human commensal, humans will continue to be at risk for developing staphylococcal infections. With increases in patient age, comorbidities, healthcare contact, and use of invasive medical procedures, the number of patients at risk of invasive staphylococcal infections will almost certainly increase. Furthermore, *S. aureus* has proven remarkably adaptable to challenges posed through the introduction of antibiotics and we have witnessed multiple waves of resistant *S. aureus* to  $\beta$ -lactams and reports of resistance to new antimicrobial agents continue unabated. Studies into the virulence and

pathogenicity of *S. aureus* have deepened our understanding of these mechanisms and it is hoped that this will translate into new therapeutics that specifically target such virulence factors (although it is likely that the organism will once again adapt to such agents). The good news is that efforts to reduce the incidence of healthcare-associated staphylococcal infections have recently achieved some success and highlights the importance of prevention of what are often devastating and costly infections.

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# References

- Chambers HF, Deleo FR. Waves of resistance: *Staphylococcus aureus* in the antibiotic era. Nat Rev Microbiol. 2009; 7 (9):629–641. [PubMed: 19680247]
- Fridkin SK, Hageman JC, Morrison M, Sanza LT, Como-Sabetti K, Jernigan JA, Harriman K, Harrison LH, Lynfield R, Farley MM. Methicillin-resistant *Staphylococcus aureus* disease in three communities. N Engl J Med. 2005; 352 (14):1436–1444. [PubMed: 15814879]
- Purcell K, Fergie J. Epidemic of community-acquired methicillin-resistant *Staphylococcus aureus* infections: a 14-year study at Driscoll Children's Hospital. Arch Pediatr Adolesc Med. 2005; 159 (10):980–985. [PubMed: 16203945]
- Kaplan SL, Hulten KG, Gonzalez BE, Hammerman WA, Lamberth L, Versalovic J, Mason EO Jr. Three-year surveillance of community-acquired *Staphylococcus aureus* infections in children. Clin Infect Dis. 2005; 40 (12):1785–1791. [PubMed: 15909267]
- 5. Liu C, Graber CJ, Karr M, Diep BA, Basuino L, Schwartz BS, Enright MC, O'Hanlon SJ, Thomas JC, Perdreau-Remington F, Gordon S, Gunthorpe H, Jacobs R, Jensen P, Leoung G, Rumack JS, Chambers HF. A population-based study of the incidence and molecular epidemiology of methicillin-resistant *Staphylococcus aureus* disease in San Francisco, 2004–2005. Clin Infect Dis. 2008; 46 (11):1637–1646. [PubMed: 18433335]
- Tong SY, Bishop EJ, Lilliebridge RA, Cheng AC, Spasova-Penkova Z, Holt DC, Giffard PM, McDonald MI, Currie BJ, Boutlis CS. Community-associated strains of methicillin-resistant *Staphylococcus aureus* and methicillin-susceptible *S. aureus* in Indigenous northern Australia: epidemiology and outcomes. J Infect Dis. 2009; 199 (10):1461–1470. [PubMed: 19392622]
- Centers for Disease Control and Prevention . Four pediatric deaths from community-acquired methicillin-resistant Staphylococcus aureus – Minnesota and North Dakota, 1997–1999. MMWR Morb Mortal Wkly Rep. 1999; 48 (32):707–710. [PubMed: 21033181]
- Adem PV, Montgomery CP, Husain AN, Koogler TK, Arangelovich V, Humilier M, Boyle-Vavra S, Daum RS. *Staphylococcus aureus* sepsis and the Waterhouse-Friderichsen syndrome in children. N Engl J Med. 2005; 353 (12):1245–1251. [PubMed: 16177250]
- Gillet Y, Issartel B, Vanhems P, Fournet JC, Lina G, Bes M, Vandenesch F, Piemont Y, Brousse N, Floret D, Etienne J. Association between *Staphylococcus aureus* strains carrying gene for Panton-Valentine leukocidin and highly lethal necrotising pneumonia in young immunocompetent patients. Lancet. 2002; 359 (9308):753–759. [PubMed: 11888586]
- Tong SY, Anstey NM, Lum GD, Lilliebridge RA, Stephens DP, Currie BJ. Fatal communityassociated methicillin-resistant *Staphylococcus aureus* pneumonia after influenza. Med J Aust. 2008; 188 (1):61. [PubMed: 18205572]
- Turnidge JD, Kotsanas D, Munckhof W, Roberts S, Bennett CM, Nimmo GR, Coombs GW, Murray RJ, Howden B, Johnson PD, Dowling K. *Staphylococcus aureus* bacteraemia: a major cause of mortality in Australia and New Zealand. Med J Aust. 2009; 191 (7):368–373. [PubMed: 19807625]

- Barber M, Rozwadowska-Dowzenko M. Infection by penicillin-resistant staphylococci. Lancet. 1948; 2 (6530):641–644. [PubMed: 18890505]
- Rountree PM, Beard MA. Further observations on infection with phage type 80 staphylococci in Australia. Med J Aust. 1958; 45 (24):789–795. [PubMed: 13612474]
- 14. Jevons MP. "Celbenin"-resistant Staphylococci. Br Med J. 1961; 1:124-125.
- Lina G, Durand G, Berchich C, Short B, Meugnier H, Vandenesch F, Etienne J, Enright MC. Staphylococcal chromosome cassette evolution in *Staphylococcus aureus* inferred from ccr gene complex sequence typing analysis. Clin Microbiol Infect. 2006; 12 (12):1175–1184. [PubMed: 17121623]
- Rountree PM, Freeman BM. Infections caused by a particular phage type of *Staphylococcus aureus*. Med J Aust. 1955; 42 (5):157–161. [PubMed: 13253118]
- McDougal LK, Steward CD, Killgore GE, Chaitram JM, McAllister SK, Tenover FC. Pulsed-field gel electrophoresis typing of oxacillin-resistant *Staphylococcus aureus* isolates from the United States: establishing a national database. J Clin Microbiol. 2003; 41 (11):5113–5120. [PubMed: 14605147]
- Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG. Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. J Clin Microbiol. 2000; 38 (3):1008–1015. [PubMed: 10698988]
- Shopsin B, Gomez M, Montgomery SO, Smith DH, Waddington M, Dodge DE, Bost DA, Riehman M, Naidich S, Kreiswirth BN. Evaluation of protein A gene polymorphic region DNA sequencing for typing of *Staphylococcus aureus* strains. J Clin Microbiol. 1999; 37 (11):3556– 3563. [PubMed: 10523551]
- 20. Harris SR, Feil EJ, Holden MT, Quail MA, Nickerson EK, Chantratita N, Gardete S, Tavares A, Day N, Lindsay JA, Edgeworth JD, de Lencastre H, Parkhill J, Peacock SJ, Bentley SD. Evolution of MRSA during hospital transmission and intercontinental spread. Science. 2010; 327 (5964): 469–474. [PubMed: 20093474]
- Feil EJ, Li BC, Aanensen DM, Hanage WP, Spratt BG. eBURST: Inferring Patterns of Evolutionary Descent among Clusters of Related Bacterial Genotypes from Multilocus Sequence Typing Data. J Bacteriol. 2004; 186 (5):1518–1530. [PubMed: 14973027]
- Willems RJ, Hanage WP, Bessen DE, Feil EJ. Population biology of Gram-positive pathogens: high-risk clones for dissemination of antibiotic resistance. FEMS Microbiol Rev. 2011; 35 (5): 872–900. [PubMed: 21658083]
- Lindsay JA, Holden MT. Understanding the rise of the superbug: investigation of the evolution and genomic variation of *Staphylococcus aureus*. Funct Integr Genomics. 2006; 6 (3):186–201. [PubMed: 16453141]
- 24. O'Brien FG, Lim TT, Chong FN, Coombs GW, Enright MC, Robinson DA, Monk A, Said-Salim B, Kreiswirth BN, Grubb WB. Diversity among community isolates of methicillin-resistant *Staphylococcus aureus* in Australia. J Clin Microbiol. 2004; 42 (7):3185–3190. [PubMed: 15243080]
- 25. Diep BA, Chambers HF, Graber CJ, Szumowski JD, Miller LG, Han LL, Chen JH, Lin F, Lin J, Phan TH, Carleton HA, McDougal LK, Tenover FC, Cohen DE, Mayer KH, Sensabaugh GF, Perdreau-Remington F. Emergence of multidrug-resistant, community-associated, methicillinresistant *Staphylococcus aureus* clone USA300 in men who have sex with men. Ann Intern Med. 2008; 148 (4):249–257. [PubMed: 18283202]
- 26. Eriksen NH, Espersen F, Rosdahl VT, Jensen K. Carriage of *Staphylococcus aureus* among 104 healthy persons during a 19-month period. Epidemiol Infect. 1995; 115 (1):51–60. [PubMed: 7641838]
- VandenBergh MF, Yzerman EP, van Belkum A, Boelens HA, Sijmons M, Verbrugh HA. Followup of *Staphylococcus aureus* nasal carriage after 8 years: redefining the persistent carrier state. J Clin Microbiol. 1999; 37 (10):3133–3140. [PubMed: 10488166]
- Luzar MA, Coles GA, Faller B, Slingeneyer A, Dah GD, Briat C, Wone C, Knefati Y, Kessler M, Peluso F. *Staphylococcus aureus* nasal carriage and infection in patients on continuous ambulatory peritoneal dialysis. N Engl J Med. 1990; 322 (8):505–509. [PubMed: 2300122]

- Kluytmans JA, Mouton JW, Ijzerman EP, Vandenbroucke-Grauls CM, Maat AW, Wagenvoort JH, Verbrugh HA. Nasal carriage of *Staphylococcus aureus* as a major risk factor for wound infections after cardiac surgery. J Infect Dis. 1995; 171 (1):216–219. [PubMed: 7798667]
- von Eiff C, Becker K, Machka K, Stammer H, Peters G. Nasal carriage as a source of Staphylococcus aureus bacteremia. Study Group. N Engl J Med. 2001; 344 (1):11–16. [PubMed: 11136954]
- Wertheim HF, Vos MC, Ott A, van Belkum A, Voss A, Kluytmans JA, van Keulen PH, Vandenbroucke-Grauls CM, Meester MH, Verbrugh HA. Risk and outcome of nosocomial *Staphylococcus aureus* bacteraemia in nasal carriers versus non-carriers. Lancet. 2004; 364 (9435):703–705. [PubMed: 15325835]
- 32. Yang ES, Tan J, Eells S, Rieg G, Tagudar G, Miller LG. Body site colonization in patients with community-associated methicillin-resistant *Staphylococcus aureus* and other types of *S. aureus* skin infections. Clin Microbiol Infect. 2010; 16 (5):425–431. [PubMed: 19689469]
- Faden H, Lesse AJ, Trask J, Hill JA, Hess DJ, Dryja D, Lee YH. Importance of colonization site in the current epidemic of staphylococcal skin abscesses. Pediatrics. 2010; 125 (3):e618–624. [PubMed: 20156893]
- Mertz D, Frei R, Periat N, Zimmerli M, Battegay M, Fluckiger U, Widmer AF. Exclusive *Staphylococcus aureus* throat carriage: at-risk populations. Arch Intern Med. 2009; 169 (2):172– 178. [PubMed: 19171814]
- 35. Uhlemann AC, Knox J, Miller M, Hafer C, Vasquez G, Ryan M, Vavagiakis P, Shi Q, Lowy FD. The Environment as an Unrecognized Reservoir for Community-Associated Methicillin Resistant *Staphylococcus aureus* USA300: A Case-Control Study. PLoS ONE. 2011; 6 (7):e22407. [PubMed: 21818321]
- 36. Batra R, Eziefula AC, Wyncoll D, Edgeworth J. Throat and rectal swabs may have an important role in MRSA screening of critically ill patients. Intensive Care Med. 2008; 34 (9):1703–1706. [PubMed: 18500421]
- Tacconelli E, Carmeli Y, Aizer A, Ferreira G, Foreman MG, D'Agata EM. Mupirocin prophylaxis to prevent *Staphylococcus aureus* infection in patients undergoing dialysis: a meta-analysis. Clin Infect Dis. 2003; 37 (12):1629–1638. [PubMed: 14689344]
- Walsh EE, Greene L, Kirshner R. Sustained reduction in methicillin-resistant *Staphylococcus aureus* wound infections after cardiothoracic surgery. Arch Intern Med. 2011; 171 (1):68–73. [PubMed: 20837818]
- Bode LG, Kluytmans JA, Wertheim HF, Bogaers D, Vandenbroucke-Grauls CM, Roosendaal R, Troelstra A, Box AT, Voss A, van der Tweel I, van Belkum A, Verbrugh HA, Vos MC. Preventing surgical-site infections in nasal carriers of *Staphylococcus aureus*. N Engl J Med. 2010; 362 (1):9–17. [PubMed: 20054045]
- Robicsek A, Beaumont JL, Paule SM, Hacek DM, Thomson RB Jr, Kaul KL, King P, Peterson LR. Universal Surveillance for Methicillin-Resistant *Staphylococcus aureus* in 3 Affiliated Hospitals. Ann Intern Med. 2008; 148 (6):409–418. [PubMed: 18347349]
- 41. Jain R, Kralovic SM, Evans ME, Ambrose M, Simbartl LA, Obrosky DS, Render ML, Freyberg RW, Jernigan JA, Muder RR, Miller LJ, Roselle GA. Veterans Affairs initiative to prevent methicillin-resistant *Staphylococcus aureus* infections. N Engl J Med. 2011; 364 (15):1419–1430. [PubMed: 21488764]
- Burton DC, Edwards JR, Horan TC, Jernigan JA, Fridkin SK. Methicillin-resistant *Staphylococcus aureus* central line-associated bloodstream infections in US intensive care units, 1997–2007. JAMA. 2009; 301 (7):727–736. [PubMed: 19224749]
- Kallen AJ, Mu Y, Bulens S, Reingold A, Petit S, Gershman K, Ray SM, Harrison LH, Lynfield R, Dumyati G, Townes JM, Schaffner W, Patel PR, Fridkin SK. Health care-associated invasive MRSA infections, 2005–2008. JAMA. 2010; 304 (6):641–648. [PubMed: 20699455]
- Edgeworth JD. Has decolonization played a central role in the decline in UK methicillin-resistant *Staphylococcus aureus* transmission? A focus on evidence from intensive care. J Antimicrob Chemother. 2011; 66(Suppl 2):ii41–47. [PubMed: 20852273]
- 45. Jarlier V, Trystram D, Brun-Buisson C, Fournier S, Carbonne A, Marty L, Andremont A, Arlet G, Buu-Hoi A, Carlet J, Decre D, Gottot S, Gutmann L, Joly-Guillou ML, Legrand P, Nicolas-

Chanoine MH, Soussy CJ, Wolf M, Lucet JC, Aggoune M, Brucker G, Regnier B. Curbing methicillin-resistant *Staphylococcus aureus* in 38 French hospitals through a 15-year institutional control program. Arch Intern Med. 2010; 170 (6):552–559. [PubMed: 20308642]

- 46. Vos MC, Behrendt MD, Melles DC, Mollema FP, de Groot W, Parlevliet G, Ott A, Horst-Kreft D, van Belkum A, Verbrugh HA. 5 years of experience implementing a methicillin-resistant *Staphylococcus aureus* search and destroy policy at the largest university medical center in the Netherlands. Infect Control Hosp Epidemiol. 2009; 30 (10):977–984. [PubMed: 19712031]
- 47. Grayson ML, Jarvie LJ, Martin R, Johnson PD, Jodoin ME, McMullan C, Gregory RH, Bellis K, Cunnington K, Wilson FL, Quin D, Kelly AM. Significant reductions in methicillin-resistant *Staphylococcus aureus* bacteraemia and clinical isolates associated with a multisite, hand hygiene culture-change program and subsequent successful statewide roll-out. Med J Aust. 2008; 188 (11): 633–640. [PubMed: 18513171]
- Huskins WC, Huckabee CM, O'Grady NP, Murray P, Kopetskie H, Zimmer L, Walker ME, Sinkowitz-Cochran RL, Jernigan JA, Samore M, Wallace D, Goldmann DA. Intervention to reduce transmission of resistant bacteria in intensive care. N Engl J Med. 2011; 364 (15):1407– 1418. [PubMed: 21488763]
- Harbarth S, Fankhauser C, Schrenzel J, Christenson J, Gervaz P, Bandiera-Clerc C, Renzi G, Vernaz N, Sax H, Pittet D. Universal screening for methicillin-resistant *Staphylococcus aureus* at hospital admission and nosocomial infection in surgical patients. JAMA. 2008; 299 (10):1149– 1157. [PubMed: 18334690]
- Mostofsky E, Lipsitch M, Regev-Yochay G. Is methicillin-resistant *Staphylococcus aureus* replacing methicillin-susceptible *S. aureus*? J Antimicrob Chemother. 2011; 66 (10):2199–2214. [PubMed: 21737459]
- Tong SY, McDonald MI, Holt DC, Currie BJ. Global implications of the emergence of community-associated methicillin-resistant *Staphylococcus aureus* in Indigenous populations. Clin Infect Dis. 2008; 46 (12):1871–1878. [PubMed: 18462175]
- 52. Ellis MW, Griffith ME, Dooley DP, McLean JC, Jorgensen JH, Patterson JE, Davis KA, Hawley JS, Regules JA, Rivard RG, Gray PJ, Ceremuga JM, Dejoseph MA, Hospenthal DR. Targeted intranasal mupirocin to prevent colonization and infection by community-associated methicillin-resistant *Staphylococcus aureus* strains in soldiers: a cluster randomized controlled trial. Antimicrob Agents Chemother. 2007; 51 (10):3591–3598. [PubMed: 17682105]
- Garza D, Sungar G, Johnston T, Rolston B, Ferguson JD, Matheson GO. Ineffectiveness of surveillance to control community-acquired methicillin-resistant *Staphylococcus aureus* in a professional football team. Clin J Sport Med. 2009; 19 (6):498–501. [PubMed: 19898079]
- 54. Lina G, Piemont Y, Godail-Gamot F, Bes M, Peter MO, Gauduchon V, Vandenesch F, Etienne J. Involvement of Panton-Valentine leukocidin-producing *Staphylococcus aureus* in primary skin infections and pneumonia. Clin Infect Dis. 1999; 29 (5):1128–1132. [PubMed: 10524952]
- Dohin B, Gillet Y, Kohler R, Lina G, Vandenesch F, Vanhems P, Floret D, Etienne J. Pediatric bone and joint infections caused by Panton-Valentine leukocidin-positive *Staphylococcus aureus*. Pediatr Infect Dis J. 2007; 26 (11):1042–1048. [PubMed: 17984813]
- Centers for Disease Control and Prevention . Severe methicillin-resistant *Staphylococcus aureus* community-acquired pneumonia associated with influenza--Louisiana and Georgia, December 2006-January 2007. MMWR Morb Mortal Wkly Rep. 2007; 56 (14):325–329. [PubMed: 17431376]
- Swaminathan A, Massasso D, Gotis-Graham I, Gosbell I. Fulminant methicillin-sensitive Staphylococcus aureus infection in a healthy adolescent, highlighting 'Panton-Valentine leucocidin syndrome'. Intern Med J. 2006; 36 (11):744–747. [PubMed: 17040363]
- Gillet Y, Vanhems P, Lina G, Bes M, Vandenesch F, Floret D, Etienne J. Factors Predicting Mortality in Necrotizing Community-Acquired Pneumonia Caused by *Staphylococcus aureus* Containing Panton-Valentine Leukocidin. Clin Infect Dis. 2007; 45 (3):315–321. [PubMed: 17599308]
- 59. Francis JS, Doherty MC, Lopatin U, Johnston CP, Sinha G, Ross T, Cai M, Hansel NN, Perl T, Ticehurst JR, Carroll K, Thomas DL, Nuermberger E, Bartlett JG. Severe community-onset pneumonia in healthy adults caused by methicillin-resistant *Staphylococcus aureus* carrying the Panton-Valentine leukocidin genes. Clin Infect Dis. 2005; 40 (1):100–107. [PubMed: 15614698]

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- Nimmo GR, Coombs GW, Pearson JC, O'Brien FG, Christiansen KJ, Turnidge JD, Gosbell IB, Collignon P, McLaws ML. Methicillin-resistant *Staphylococcus aureus* in the Australian community: an evolving epidemic. Med J Aust. 2006; 184 (8):384–388. [PubMed: 16618236]
- Tong SY, Lilliebridge RA, Bishop EJ, Cheng AC, Holt DC, McDonald MI, Giffard PM, Currie BJ, Boutlis CS. Clinical correlates of Panton-Valentine leukocidin (PVL), PVL isoforms, and clonal complex in the *Staphylococcus aureus* population of Northern Australia. J Infect Dis. 2010; 202 (5):760–769. [PubMed: 20662623]
- 62. Munckhof WJ, Nimmo GR, Carney J, Schooneveldt JM, Huygens F, Inman-Bamber J, Tong E, Morton A, Giffard P. Methicillin-susceptible, non-multiresistant methicillin-resistant and multiresistant methicillin-resistant *Staphylococcus aureus* infections: a clinical, epidemiological and microbiological comparative study. Eur J Clin Microbiol Infect Dis. 2008; 27 (5):355–364. [PubMed: 18278529]
- 63. Nimmo GR, Schooneveldt JM, Sutherland JL, Power S, Olesen D, Selvey C, Beard F, Jones M, Paterson DL. Epidemiology of non-multiresistant methicillin-resistant *Staphylococcus aureus* infection in Queensland, Australia: associations with indigenous populations and Panton-Valentine leukocidin. Eur J Clin Microbiol Infect Dis. 2010; 29 (10):1253–1259. [PubMed: 20556466]
- 64. Wehrhahn MC, Robinson JO, Pearson JC, O'Brien FG, Tan HL, Coombs GW, Pascoe EM, Lee R, Salvaris P, Salvaris R, New D, Murray RJ. Clinical and laboratory features of invasive community-onset methicillin-resistant *Staphylococcus aureus* infection: a prospective case-control study. Eur J Clin Microbiol Infect Dis. 2010; 29 (8):1025–1033. [PubMed: 20549534]
- 65. Campbell SJ, Deshmukh HS, Nelson CL, Bae IG, Stryjewski ME, Federspiel JJ, Tonthat GT, Rude TH, Barriere SL, Corey R, Fowler VG Jr. Genotypic characteristics of *Staphylococcus aureus* isolates from a multinational trial of complicated skin and skin structure infections. J Clin Microbiol. 2008; 46 (2):678–684. [PubMed: 18077636]
- 66. Bae IG, Tonthat GT, Stryjewski ME, Rude TH, Reilly LF, Barriere SL, Genter FC, Corey GR, Fowler VG Jr. Presence of genes encoding the Panton-Valentine leukocidin exotoxin is not the primary determinant of outcome in patients with complicated skin and skin structure infections due to methicillin-resistant *Staphylococcus aureus*: results of a multinational trial. J Clin Microbiol. 2009; 47 (12):3952–3957. [PubMed: 19846653]
- 67. Peyrani P, Allen M, Wiemken TL, Haque NZ, Zervos MJ, Ford KD, Scerpella EG, Mangino JE, Kett DH, Ramirez JA. Severity of Disease and Clinical Outcomes in Patients With Hospital-Acquired Pneumonia Due to Methicillin-Resistant *Staphylococcus aureus* Strains Not Influenced by the Presence of the Panton-Valentine Leukocidin Gene. Clin Infect Dis. 2011; 53 (8):766–771. [PubMed: 21880581]
- 68. Nickerson EK, Wuthiekanun V, Wongsuvan G, Limmathurosakul D, Srisamang P, Mahavanakul W, Thaipadungpanit J, Shah KR, Arayawichanont A, Amornchai P, Thanwisai A, Day NP, Peacock SJ. Factors predicting and reducing mortality in patients with invasive *Staphylococcus aureus* disease in a developing country. PLoS ONE. 2009; 4 (8):e6512. [PubMed: 19652705]
- Labandeira-Rey M, Couzon F, Boisset S, Brown EL, Bes M, Benito Y, Barbu EM, Vazquez V, Hook M, Etienne J, Vandenesch F, Bowden MG. *Staphylococcus aureus* Panton-Valentine leukocidin causes necrotizing pneumonia. Science. 2007; 315 (5815):1130–1133. [PubMed: 17234914]
- 70. Villaruz AE, Bubeck Wardenburg J, Khan BA, Whitney AR, Sturdevant DE, Gardner DJ, DeLeo FR, Otto M. A point mutation in the *agr* locus rather than expression of the Panton-Valentine leukocidin caused previously reported phenotypes in *Staphylococcus aureus* pneumonia and gene regulation. J Infect Dis. 2009; 200 (5):724–734. [PubMed: 19604047]
- 71. Gladstone GP, Van Heyningen WE. Staphylococcal leucocidins. Br J Exp Pathol. 1957; 38 (2): 123–137. [PubMed: 13426414]
- Szmigielski S, Prevost G, Monteil H, Colin DA, Jeljaszewicz J. Leukocidal toxins of staphylococci. Zentralbl Bakteriol. 1999; 289 (2):185–201. [PubMed: 10360319]
- Hongo I, Baba T, Oishi K, Morimoto Y, Ito T, Hiramatsu K. Phenol-soluble modulin alpha 3 enhances the human neutrophil lysis mediated by Panton-Valentine leukocidin. J Infect Dis. 2009; 200 (5):715–723. [PubMed: 19653829]

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- Loffler B, Hussain M, Grundmeier M, Bruck M, Holzinger D, Varga G, Roth J, Kahl BC, Proctor RA, Peters G. *Staphylococcus aureus* Panton-Valentine leukocidin is a very potent cytotoxic factor for human neutrophils. PLoS Pathog. 2010; 6 (1):e1000715. [PubMed: 20072612]
- 75. Voyich JM, Otto M, Mathema B, Braughton KR, Whitney AR, Welty D, Long RD, Dorward DW, Gardner DJ, Lina G, Kreiswirth BN, DeLeo FR. Is Panton-Valentine leukocidin the major virulence determinant in community-associated methicillin-resistant *Staphylococcus aureus* disease? J Infect Dis. 2006; 194 (12):1761–1770. [PubMed: 17109350]
- 76. Bubeck Wardenburg J, Palazzolo-Ballance AM, Otto M, Schneewind O, DeLeo FR. Panton-Valentine leukocidin is not a virulence determinant in murine models of community-associated methicillin-resistant *Staphylococcus aureus* disease. J Infect Dis. 2008; 198 (8):1166–1170. [PubMed: 18729780]
- 77. Diep BA, Chan L, Tattevin P, Kajikawa O, Martin TR, Basuino L, Mai TT, Marbach H, Braughton KR, Whitney AR, Gardner DJ, Fan X, Tseng CW, Liu GY, Badiou C, Etienne J, Lina G, Matthay MA, DeLeo FR, Chambers HF. Polymorphonuclear leukocytes mediate *Staphylococcus aureus* Panton-Valentine leukocidin-induced lung inflammation and injury. Proc Natl Acad Sci U S A. 2010; 107 (12):5587–5592. [PubMed: 20231457]
- 78. Lipinska U, Hermans K, Meulemans L, Dumitrescu O, Badiou C, Duchateau L, Haesebrouck F, Etienne J, Lina G. Panton-Valentine Leukocidin Does Play a Role in the Early Stage of *Staphylococcus aureus* Skin Infections: A Rabbit Model. PLoS ONE. 2011; 6 (8):e22864. [PubMed: 21850240]
- Kobayashi SD, Malachowa N, Whitney AR, Braughton KR, Gardner DJ, Long D, Bubeck Wardenburg J, Schneewind O, Otto M, Deleo FR. Comparative Analysis of USA300 Virulence Determinants in a Rabbit Model of Skin and Soft Tissue Infection. J Infect Dis. 2011; 204 (6): 937–941. [PubMed: 21849291]
- Yoong P, Pier GB. Antibody-mediated enhancement of community-acquired methicillin-resistant *Staphylococcus aureus* infection. Proc Natl Acad Sci U S A. 2010; 107 (5):2241–2246. [PubMed: 20133867]
- Rasigade JP, Sicot N, Laurent F, Lina G, Vandenesch F, Etienne J. A history of Panton-Valentine leukocidin (PVL)-associated infection protects against death in PVL-associated pneumonia. Vaccine. 2011; 29 (25):4185–4186. [PubMed: 21527300]
- Bubeck Wardenburg J, Bae T, Otto M, Deleo FR, Schneewind O. Poring over pores: alphahemolysin and Panton-Valentine leukocidin in *Staphylococcus aureus* pneumonia. Nat Med. 2007; 13 (12):1405–1406. [PubMed: 18064027]
- Kennedy AD, Bubeck Wardenburg J, Gardner DJ, Long D, Whitney AR, Braughton KR, Schneewind O, DeLeo FR. Targeting of alpha-hemolysin by active or passive immunization decreases severity of USA300 skin infection in a mouse model. J Infect Dis. 2010; 202 (7):1050– 1058. [PubMed: 20726702]
- 84. Wang R, Braughton KR, Kretschmer D, Bach TH, Queck SY, Li M, Kennedy AD, Dorward DW, Klebanoff SJ, Peschel A, Deleo FR, Otto M. Identification of novel cytolytic peptides as key virulence determinants for community-associated MRSA. Nat Med. 2007; 13 (12):1510–1514. [PubMed: 17994102]
- Ventura CL, Malachowa N, Hammer CH, Nardone GA, Robinson MA, Kobayashi SD, DeLeo FR. Identification of a novel *Staphylococcus aureus* two-component leukotoxin using cell surface proteomics. PLoS ONE. 2010; 5 (7):e11634. [PubMed: 20661294]
- 86. Gillet Y, Dumitrescu O, Tristan A, Dauwalder O, Javouhey E, Floret D, Vandenesch F, Etienne J, Lina G. Pragmatic management of Panton-Valentine leukocidin-associated staphylococcal diseases. Int J Antimicrob Agents. 2011
- Dumitrescu O, Badiou C, Bes M, Reverdy ME, Vandenesch F, Etienne J, Lina G. Effect of antibiotics, alone and in combination, on Panton-Valentine leukocidin production by a *Staphylococcus aureus* reference strain. Clin Microbiol Infect. 2008; 14 (4):384–388. [PubMed: 18261123]
- Dumitrescu O, Boisset S, Badiou C, Bes M, Benito Y, Reverdy ME, Vandenesch F, Etienne J, Lina G. Effect of antibiotics on *Staphylococcus aureus* producing Panton-Valentine leukocidin. Antimicrob Agents Chemother. 2007; 51 (4):1515–1519. [PubMed: 17242137]

- Joo HS, Chan JL, Cheung GY, Otto M. Subinhibitory concentrations of protein synthesisinhibiting antibiotics promote increased expression of the agr virulence regulator and production of phenol-soluble modulin cytolysins in community-associated methicillin-resistant *Staphylococcus aureus*. Antimicrob Agents Chemother. 2010; 54 (11):4942–4944. [PubMed: 20713669]
- 90. Rouzic N, Janvier F, Libert N, Javouhey E, Lina G, Nizou JY, Pasquier P, Stamm D, Brinquin L, Pelletier C, Vandenesch F, Floret D, Etienne J, Gillet Y. Prompt and successful toxin-targeting treatment of three patients with necrotizing pneumonia due to *Staphylococcus aureus* strains carrying the Panton-Valentine leukocidin genes. J Clin Microbiol. 2010; 48 (5):1952–1955. [PubMed: 20129956]
- Obando I, Valderrabanos ES, Millan JA, Neth OW. Necrotising pneumonia due to influenza A (H1N1) and community-acquired methicillin-resistant *Staphylococcus aureus* clone USA300: successful management of the first documented paediatric case. Arch Dis Child. 2010; 95 (4):305– 306. [PubMed: 20335240]
- 92. Liu GY, Essex A, Buchanan JT, Datta V, Hoffman HM, Bastian JF, Fierer J, Nizet V. *Staphylococcus aureus* golden pigment impairs neutrophil killing and promotes virulence through its antioxidant activity. J Exp Med. 2005; 202 (2):209–215. [PubMed: 16009720]
- 93. Liu CI, Liu GY, Song Y, Yin F, Hensler ME, Jeng WY, Nizet V, Wang AH, Oldfield E. A cholesterol biosynthesis inhibitor blocks *Staphylococcus aureus* virulence. Science. 2008; 319 (5868):1391–1394. [PubMed: 18276850]
- 94. Holt DC, Holden MT, Tong SY, Castillo-Ramirez S, Clarke L, Quail MA, Currie BJ, Parkhill J, Bentley SD, Feil EJ, Giffard PM. A Very Early-Branching *Staphylococcus aureus* Lineage Lacking the Carotenoid Pigment Staphyloxanthin. Genome Biol Evol. 2011; 3:881–895. [PubMed: 21813488]
- 95. Day NPJ, Moore CE, Enright MC, Berendt AR, Smith JM, Murphy MF, Peacock SJ, Spratt BG, Feil EJ. A link between virulence and ecological abundance in natural populations of *Staphylococcus aureus*. Science. 2001; 292 (5514):114–116. [PubMed: 11292876]
- 96. Day NP, Moore CE, Enright MC, Berendt AP, Smith JM, Murphy MF, Peacock SJ, Spratt BG, Feil EJ. Retraction. Science. 2002; 295 (5557):971. [PubMed: 11887789]
- 97. Melles DC, Gorkink RF, Boelens HA, Snijders SV, Peeters JK, Moorhouse MJ, van der Spek PJ, van Leeuwen WB, Simons G, Verbrugh HA, van Belkum A. Natural population dynamics and expansion of pathogenic clones of *Staphylococcus aureus*. J Clin Invest. 2004; 114 (12):1732–1740. [PubMed: 15599398]
- 98. Fowler VG Jr, Nelson CL, McIntyre LM, Kreiswirth BN, Monk A, Archer GL, Federspiel J, Naidich S, Remortel B, Rude T, Brown P, Reller LB, Corey GR, Gill SR. Potential associations between hematogenous complications and bacterial genotype in *Staphylococcus aureus* infection. J Infect Dis. 2007; 196 (5):738–747. [PubMed: 17674317]
- 99. Xiong YQ, Fowler VG, Yeaman MR, Perdreau-Remington F, Kreiswirth BN, Bayer AS. Phenotypic and genotypic characteristics of persistent methicillin-resistant *Staphylococcus aureus* bacteremia *in vitro* and in an experimental endocarditis model. J Infect Dis. 2009; 199 (2):201– 208. [PubMed: 19086913]
- 100. Nienaber JJ, Sharma Kuinkel BK, Clarke-Pearson M, Lamlertthon S, Park L, Rude TH, Barriere S, Woods CW, Chu VH, Marin M, Bukovski S, Garcia P, Corey GR, Korman T, Doco-Lecompte T, Murdoch DR, Reller LB, Fowler VG Jr. Methicillin-Susceptible *Staphylococcus aureus* Endocarditis Isolates Are Associated With Clonal Complex 30 Genotype and a Distinct Repertoire of Enterotoxins and Adhesins. J Infect Dis. 2011; 204 (5):704–713. [PubMed: 21844296]
- 101. Kennedy AD, Otto M, Braughton KR, Whitney AR, Chen L, Mathema B, Mediavilla JR, Byrne KA, Parkins LD, Tenover FC, Kreiswirth BN, Musser JM, DeLeo FR. Epidemic community-associated methicillin-resistant Staphylococcus aureus: Recent clonal expansion and diversification. Proc Natl Acad Sci U S A. 2008; 105 (4):1327–1332. [PubMed: 18216255]
- 102. Shea PR, Beres SB, Flores AR, Ewbank AL, Gonzalez-Lugo JH, Martagon-Rosado AJ, Martinez-Gutierrez JC, Rehman HA, Serrano-Gonzalez M, Fittipaldi N, Ayers SD, Webb P, Willey BM, Low DE, Musser JM. Distinct signatures of diversifying selection revealed by genome analysis

of respiratory tract and invasive bacterial populations. Proc Natl Acad Sci U S A. 2011; 108 (12): 5039–5044. [PubMed: 21383167]

- 103. Graham PL 3rd, Lin SX, Larson EL. A U.S. population-based survey of *Staphylococcus aureus* colonization. Ann Intern Med. 2006; 144 (5):318–325. [PubMed: 16520472]
- 104. Big C, Malani PN. *Staphylococcus aureus* bloodstream infections in older adults: clinical outcomes and risk factors for in-hospital mortality. J Am Geriatr Soc. 2010; 58 (2):300–305. [PubMed: 20070420]
- 105. Klevens RM, Morrison MA, Nadle J, Petit S, Gershman K, Ray S, Harrison LH, Lynfield R, Dumyati G, Townes JM, Craig AS, Zell ER, Fosheim GE, McDougal LK, Carey RB, Fridkin SK. for the Active Bacterial Core surveillance MI. Invasive Methicillin-Resistant *Staphylococcus aureus* Infections in the United States. JAMA. 2007; 298 (15):1763–1771. [PubMed: 17940231]
- 106. Centers for Disease Control and Prevention . Methicillin-resistant *Staphylococcus aureus* infections in correctional facilities---Georgia, California, and Texas, 2001–2003. MMWR Morb Mortal Wkly Rep. 2003; 52 (41):992–996. [PubMed: 14561958]
- 107. Aiello AE, Lowy FD, Wright LN, Larson EL. Meticillin-resistant *Staphylococcus aureus* among US prisoners and military personnel: review and recommendations for future studies. Lancet Infect Dis. 2006; 6 (6):335–341. [PubMed: 16728319]
- Kirkland EB, Adams BB. Methicillin-resistant *Staphylococcus aureus* and athletes. J Am Acad Dermatol. 2008; 59 (3):494–502. [PubMed: 18550208]
- 109. Suh L, Coffin S, Leckerman KH, Gelfand JM, Honig PJ, Yan AC. Methicillin-resistant *Staphylococcus aureus* colonization in children with atopic dermatitis. Pediatr Dermatol. 2008; 25 (5):528–534. [PubMed: 18950393]
- 110. Begier EM, Frenette K, Barrett NL, Mshar P, Petit S, Boxrud DJ, Watkins-Colwell K, Wheeler S, Cebelinski EA, Glennen A, Nguyen D, Hadler JL. A high-morbidity outbreak of methicillin-resistant *Staphylococcus aureus* among players on a college football team, facilitated by cosmetic body shaving and turf burns. Clin Infect Dis. 2004; 39 (10):1446–1453. [PubMed: 15546080]
- 111. Kazakova SV, Hageman JC, Matava M, Srinivasan A, Phelan L, Garfinkel B, Boo T, McAllister S, Anderson J, Jensen B, Dodson D, Lonsway D, McDougal LK, Arduino M, Fraser VJ, Killgore G, Tenover FC, Cody S, Jernigan DB. A clone of methicillin-resistant *Staphylococcus aureus* among professional football players. N Engl J Med. 2005; 352 (5):468–475. [PubMed: 15689585]
- 112. Centers for Disease Control and Prevention . Methicillin-resistant *Staphylococcus aureus* skin infections among tattoo recipients--Ohio, Kentucky, and Vermont, 2004–2005. MMWR Morb Mortal Wkly Rep. 2006; 55 (24):677–679. [PubMed: 16791134]
- 113. Moreillon, P.; Que, Y.; Glauser, M. Staphylococcus aureus (including staphylococcal toxic shock). In: Mandell, G.; Bennett, J.; Dolin, R., editors. Principles and Practice of Infectious Diseases. 6. Churchill Livingstone; Philadelphia, PA: 2005. p. 2321
- Miller LS, Cho JS. Immunity against *Staphylococcus aureus* cutaneous infections. Nat Rev Immunol. 2011; 11 (8):505–518. [PubMed: 21720387]
- 115. Mathews WC, Caperna JC, Barber RE, Torriani FJ, Miller LG, May S, McCutchan JA. Incidence of and risk factors for clinically significant methicillin-resistant *Staphylococcus aureus* infection in a cohort of HIV-infected adults. J Acquir Immune Defic Syndr. 2005; 40 (2):155–160. [PubMed: 16186732]
- 116. Jacobsson G, Dashti S, Wahlberg T, Andersson R. The epidemiology of and risk factors for invasive *Staphylococcus aureus* infections in western Sweden. Scand J Infect Dis. 2007; 39 (1): 6–13. [PubMed: 17366006]
- 117. Mowat A, Baum J. Chemotaxis of polymorphonuclear leukocytes from patients with diabetes mellitus. N Engl J Med. 1971; 284 (12):621–627. [PubMed: 5545603]
- 118. Schentag JJ, Hyatt JM, Carr JR, Paladino JA, Birmingham MC, Zimmer GS, Cumbo TJ. Genesis of methicillin-resistant *Staphylococcus aureus* (MRSA), how treatment of MRSA infections has selected for vancomycin-resistant *Enterococcus faecium*, and the importance of antibiotic management and infection control. Clin Infect Dis. 1998; 26 (5):1204–1214. [PubMed: 9597254]

- Harinstein L, Schafer J, D'Amico F. Risk factors associated with the conversion of meticillinresistant *Staphylococcus aureus* colonisation to healthcare-associated infection. J Hosp Infect. 2011; 79 (3):194–197. [PubMed: 21640432]
- 120. Muller AA, Mauny F, Bertin M, Cornette C, Lopez-Lozano JM, Viel JF, Talon DR, Bertrand X. Relationship between spread of methicillin-resistant *Staphylococcus aureus* and antimicrobial use in a French university hospital. Clin Infect Dis. 2003; 36 (8):971–978. [PubMed: 12684908]
- 121. Anderson DJ, Chen LF, Schmader KE, Sexton DJ, Choi Y, Link K, Sloane R, Kaye KS. Poor functional status as a risk factor for surgical site infection due to methicillin-resistant *Staphylococcus aureus*. Infect Control Hosp Epidemiol. 2008; 29 (9):832–839. [PubMed: 18665820]
- 122. Manzur A, Gudiol F. Methicillin-resistant *Staphylococcus aureus* in long-term-care facilities. Clin Microbiol Infect. 2009; 15(Suppl 7):26–30. [PubMed: 19951331]
- 123. Fowler VG Jr, Miro JM, Hoen B, Cabell CH, Abrutyn E, Rubinstein E, Corey GR, Spelman D, Bradley SF, Barsic B, Pappas PA, Anstrom KJ, Wray D, Fortes CQ, Anguera I, Athan E, Jones P, van der Meer JT, Elliott TS, Levine DP, Bayer AS. *Staphylococcus aureus* endocarditis: a consequence of medical progress. JAMA. 2005; 293 (24):3012–3021. [PubMed: 15972563]
- 124. Archer NK, Mazaitis MJ, Costerton JW, Leid JG, Powers ME, Shirtliff ME. *Staphylococcus aureus* biofilms: Properties, regulation, and roles in human disease. Virulence. 2011; 2(5)
- 125. Shaklee J, Zerr DM, Elward A, Newland J, Leckerman K, Asti L, Guth R, Bass J, Selvarangan R, Coffin S, Zaoutis T. Improving surveillance for pediatric *Clostridium difficile* infection: derivation and validation of an accurate case-finding tool. Pediatr Infect Dis J. 2011; 30 (3):e38–40. [PubMed: 21079527]
- 126. Liu C, Bayer A, Cosgrove SE, Daum RS, Fridkin SK, Gorwitz RJ, Kaplan SL, Karchmer AW, Levine DP, Murray BE, MJR, Talan DA, Chambers HF. Clinical practice guidelines by the infectious diseases society of america for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children: executive summary. Clin Infect Dis. 2011; 52 (3):285– 292. [PubMed: 21217178]
- 127. Cheng AG, DeDent AC, Schneewind O, Missiakas D. A play in four acts: *Staphylococcus aureus* abscess formation. Trends Microbiol. 2011; 19 (5):225–232. [PubMed: 21353779]
- 128. Rybak MJ, Cappelletty DM, Moldovan T, Aeschlimann JR, Kaatz GW. Comparative in vitro activities and postantibiotic effects of the oxazolidinone compounds eperezolid (PNU-100592) and linezolid (PNU-100766) versus vancomycin against *Staphylococcus aureus*, coagulasenegative staphylococci, *Enterococcus faecalis*, and *Enterococcus faecium*. Antimicrob Agents Chemother. 1998; 42 (3):721–724. [PubMed: 9517963]
- Weigelt J, Itani K, Stevens D, Lau W, Dryden M, Knirsch C. Linezolid versus vancomycin in treatment of complicated skin and soft tissue infections. Antimicrob Agents Chemother. 2005; 49 (6):2260–2266. [PubMed: 15917519]
- 130. Wunderink RG, Rello J, Cammarata SK, Croos-Dabrera RV, Kollef MH. Linezolid vs vancomycin: analysis of two double-blind studies of patients with methicillin-resistant *Staphylococcus aureus* nosocomial pneumonia. Chest. 2003; 124 (5):1789–1797. [PubMed: 14605050]
- 131. Walkey AJ, O'Donnell MR, Wiener RS. Linezolid vs glycopeptide antibiotics for the treatment of suspected methicillin-resistant *Staphylococcus aureus* nosocomial pneumonia: a meta-analysis of randomized controlled trials. Chest. 2011; 139 (5):1148–1155. [PubMed: 20864609]
- 132. Gerson SL, Kaplan SL, Bruss JB, Le V, Arellano FM, Hafkin B, Kuter DJ. Hematologic effects of linezolid: summary of clinical experience. Antimicrob Agents Chemother. 2002; 46 (8):2723– 2726. [PubMed: 12121967]
- 133. Legout L, Senneville E, Gomel JJ, Yazdanpanah Y, Mouton Y. Linezolid-induced neuropathy. Clin Infect Dis. 2004; 38 (5):767–768. [PubMed: 14986270]
- 134. Lawrence KR, Adra M, Gillman PK. Serotonin toxicity associated with the use of linezolid: a review of postmarketing data. Clin Infect Dis. 2006; 42 (11):1578–1583. [PubMed: 16652315]
- 135. Farrell DJ, Mendes RE, Ross JE, Sader HS, Jones RN. LEADER Program results for 2009: an activity and spectrum analysis of linezolid using 6,414 clinical isolates from 56 medical centers

in the United States. Antimicrob Agents Chemother. 2011; 55 (8):3684–3690. [PubMed: 21670176]

- 136. Morales G, Picazo JJ, Baos E, Candel FJ, Arribi A, Pelaez B, Andrade R, de la Torre MA, Fereres J, Sanchez-Garcia M. Resistance to linezolid is mediated by the *cfr* gene in the first report of an outbreak of linezolid-resistant *Staphylococcus aureus*. Clin Infect Dis. 2010; 50 (6):821–825. [PubMed: 20144045]
- Wilson P, Andrews JA, Charlesworth R, Walesby R, Singer M, Farrell DJ, Robbins M. Linezolid resistance in clinical isolates of *Staphylococcus aureus*. J Antimicrob Chemother. 2003; 51 (1): 186–188. [PubMed: 12493812]
- 138. Gao W, Chua K, Davies JK, Newton HJ, Seemann T, Harrison PF, Holmes NE, Rhee HW, Hong JI, Hartland EL, Stinear TP, Howden BP. Two novel point mutations in clinical *Staphylococcus aureus* reduce linezolid susceptibility and switch on the stringent response to promote persistent infection. PLoS Pathogens. 2010; 6 (6):e1000944. [PubMed: 20548948]
- 139. Greer ND. Tigecycline (Tygacil): the first in the glycylcycline class of antibiotics. Proc (Bayl Univ Med Cent). 2006; 19 (2):155–161. [PubMed: 16609746]
- 140. Florescu I, Beuran M, Dimov R, Razbadauskas A, Bochan M, Fichev G, Dukart G, Babinchak T, Cooper CA, Ellis-Grosse EJ, Dartois N, Gandjini H. Efficacy and safety of tigecycline compared with vancomycin or linezolid for treatment of serious infections with methicillin-resistant *Staphylococcus aureus* or vancomycin-resistant enterococci: a Phase 3, multicentre, doubleblind, randomized study. J Antimicrob Chemother. 2008; 62(Suppl 1):i17–28. [PubMed: 18684703]
- 141. DiMondi VP, Drew RH, Chen LF. Ceftaroline fosamil for treatment of community-acquired pneumonia: findings from FOCUS 1 and 2 and potential role in therapy. Expert Rev Anti Infect Ther. 2011; 9 (8):567–572. [PubMed: 21819323]
- 142. Jacqueline C, Amador G, Batard E, Le Mabecque V, Miegeville AF, Biek D, Caillon J, Potel G. Comparison of ceftaroline fosamil, daptomycin and tigecycline in an experimental rabbit endocarditis model caused by methicillin-susceptible, methicillin-resistant and glycopeptideintermediate *Staphylococcus aureus*. J Antimicrob Chemother. 2011; 66 (4):863–866. [PubMed: 21393213]
- 143. Corey GR, Wilcox M, Talbot GH, Friedland HD, Baculik T, Witherell GW, Critchley I, Das AF, Thye D. Integrated analysis of CANVAS 1 and 2: phase 3, multicenter, randomized, doubleblind studies to evaluate the safety and efficacy of ceftaroline versus vancomycin plus aztreonam in complicated skin and skin-structure infection. Clin Infect Dis. 2010; 51 (6):641–650. [PubMed: 20695801]
- 144. File TM Jr, Low DE, Eckburg PB, Talbot GH, Friedland HD, Lee J, Llorens L, Critchley I, Thye D. Integrated analysis of FOCUS 1 and FOCUS 2: randomized, doubled-blinded, multicenter phase 3 trials of the efficacy and safety of ceftaroline fosamil versus ceftriaxone in patients with community-acquired pneumonia. Clin Infect Dis. 2010; 51 (12):1395–1405. [PubMed: 21067350]
- 145. Arbeit RD, Maki D, Tally FP, Campanaro E, Eisenstein BI. The safety and efficacy of daptomycin for the treatment of complicated skin and skin-structure infections. Clin Infect Dis. 2004; 38 (12):1673–1681. [PubMed: 15227611]
- 146. Krige JE, Lindfield K, Friedrich L, Otradovec C, Martone WJ, Katz DE, Tally F. Effectiveness and duration of daptomycin therapy in resolving clinical symptoms in the treatment of complicated skin and skin structure infections. Curr Med Res Opin. 2007; 23 (9):2147–2156. [PubMed: 17669231]
- 147. Fowler VG Jr, Boucher HW, Corey GR, Abrutyn E, Karchmer AW, Rupp ME, Levine DP, Chambers HF, Tally FP, Vigliani GA, Cabell CH, Link AS, DeMeyer I, Filler SG, Zervos M, Cook P, Parsonnet J, Bernstein JM, Price CS, Forrest GN, Fatkenheuer G, Gareca M, Rehm SJ, Brodt HR, Tice A, Cosgrove SE. Daptomycin versus standard therapy for bacteremia and endocarditis caused by *Staphylococcus aureus*. N Engl J Med. 2006; 355 (7):653–665. [PubMed: 16914701]
- 148. Silverman JA, Mortin LI, Vanpraagh AD, Li T, Alder J. Inhibition of daptomycin by pulmonary surfactant: in vitro modeling and clinical impact. J Infect Dis. 2005; 191 (12):2149–2152. [PubMed: 15898002]

- 149. van Hal SJ, Paterson DL, Gosbell IB. Emergence of daptomycin resistance following vancomycin-unresponsive *Staphylococcus aureus* bacteraemia in a daptomycin-naive patient--a review of the literature. Eur J Clin Microbiol Infect Dis. 2011; 30 (5):603–610. [PubMed: 21191627]
- 150. Kelley PG, Gao W, Ward PB, Howden BP. Daptomycin non-susceptibility in vancomycinintermediate *Staphylococcus aureus* (VISA) and heterogeneous-VISA Z (hVISA): implications for therapy after vancomycin treatment failure. J Antimicrob Chemother. 2011; 66 (5):1057– 1060. [PubMed: 21393156]
- 151. Stryjewski ME, Graham DR, Wilson SE, O'Riordan W, Young D, Lentnek A, Ross DP, Fowler VG, Hopkins A, Friedland HD, Barriere SL, Kitt MM, Corey GR. Telavancin versus vancomycin for the treatment of complicated skin and skin-structure infections caused by gram-positive organisms. Clin Infect Dis. 2008; 46 (11):1683–1693. [PubMed: 18444791]
- 152. Rubinstein E, Lalani T, Corey GR, Kanafani ZA, Nannini EC, Rocha MG, Rahav G, Niederman MS, Kollef MH, Shorr AF, Lee PC, Lentnek AL, Luna CM, Fagon JY, Torres A, Kitt MM, Genter FC, Barriere SL, Friedland HD, Stryjewski ME. Telavancin versus vancomycin for hospital-acquired pneumonia due to gram-positive pathogens. Clin Infect Dis. 2011; 52 (1):31–40. [PubMed: 21148517]
- 153. Verkaik NJ, van Wamel WJ, van Belkum A. Immunotherapeutic approaches against *Staphylococcus aureus*. Immunotherapy. 2011; 3 (9):1063–1073. [PubMed: 21913829]
- 154. Schaffer AC, Lee JC. Staphylococcal vaccines and immunotherapies. Infect Dis Clin North Am. 2009; 23 (1):153–171. [PubMed: 19135920]
- 155. Broughan J, Anderson R, Anderson AS. Strategies for and advances in the development of *Staphylococcus aureus* prophylactic vaccines. Expert Rev Vaccines. 2011; 10 (5):695–708. [PubMed: 21604989]
- 156. Shinefield H, Black S, Fattom A, Horwith G, Rasgon S, Ordonez J, Yeoh H, Law D, Robbins JB, Schneerson R, Muenz L, Fuller S, Johnson J, Fireman B, Alcorn H, Naso R. Use of a *Staphylococcus aureus* conjugate vaccine in patients receiving hemodialysis. N Engl J Med. 2002; 346 (7):491–496. [PubMed: 11844850]
- 157. Harro C, Betts R, Orenstein W, Kwak EJ, Greenberg HE, Onorato MT, Hartzel J, Lipka J, DiNubile MJ, Kartsonis N. Safety and immunogenicity of a novel *Staphylococcus aureus* vaccine: results from the first study of the vaccine dose range in humans. Clin Vaccine Immunol. 2010; 17 (12):1868–1874. [PubMed: 20943877]
- 158. DeJonge M, Burchfield D, Bloom B, Duenas M, Walker W, Polak M, Jung E, Millard D, Schelonka R, Eyal F, Morris A, Kapik B, Roberson D, Kesler K, Patti J, Hetherington S. Clinical trial of safety and efficacy of INH-A21 for the prevention of nosocomial staphylococcal bloodstream infection in premature infants. J Pediatr. 2007; 151(3):260–265. 265, e261. [PubMed: 17719934]
- 159. Weems JJ Jr, Steinberg JP, Filler S, Baddley JW, Corey GR, Sampathkumar P, Winston L, John JF, Kubin CJ, Talwani R, Moore T, Patti JM, Hetherington S, Texter M, Wenzel E, Kelley VA, Fowler VG Jr. Phase II, randomized, double-blind, multicenter study comparing the safety and pharmacokinetics of tefibazumab to placebo for treatment of *Staphylococcus aureus* bacteremia. Antimicrob Agents Chemother. 2006; 50 (8):2751–2755. [PubMed: 16870768]
- 160. Benjamin DK, Schelonka R, White R, Holley HP, Bifano E, Cummings J, Adcock K, Kaufman D, Puppala B, Riedel P, Hall B, White J, Cotton CM. A blinded, randomized, multicenter study of an intravenous *Staphylococcus aureus* immune globulin. J Perinatol. 2006; 26 (5):290–295. [PubMed: 16598296]
- 161. Rupp ME, Holley HP Jr, Lutz J, Dicpinigaitis PV, Woods CW, Levine DP, Veney N, Fowler VG Jr. Phase II, randomized, multicenter, double-blind, placebo-controlled trial of a polyclonal anti-*Staphylococcus aureus* capsular polysaccharide immune globulin in treatment of *Staphylococcus aureus* bacteremia. Antimicrob Agents Chemother. 2007; 51 (12):4249–4254. [PubMed: 17893153]
- 162. Weisman LE, Thackray HM, Steinhorn RH, Walsh WF, Lassiter HA, Dhanireddy R, Brozanski BS, Palmer KG, Trautman MS, Escobedo M, Meissner HC, Sasidharan P, Fretz J, Kokai-Kun JF, Kramer WG, Fischer GW, Mond JJ. A randomized study of a monoclonal antibody

(pagibaximab) to prevent staphylococcal sepsis. Pediatrics. 2011; 128 (2):271–279. [PubMed: 21788224]