

Codon catalog usage and the genome hypothesis

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ABSTRACT

Frequencies for each of the 61 amino acid codons have been determined in every published mRNA sequence of 50 or more codons. The frequencies are shown for each kind of genome and for each individual gene. A surprising consistency of choices exists among genes of the same or similar genomes. Thus each genome, or kind of genome, appears to possess a "system" for choosing between codons. Frameshift genes, however, have widely different choice strategies from normal genes. Our work indicates that the main factors distinguishing between mRNA sequences relate to choices among degenerate bases. These systematic third base choices can therefore be used to establish a new kind of genetic distance, which reflects differences in coding strategy. The choice patterns we find seem compatible with the idea that the genome and not the individual gene is the unit of selection. Each gene in a genome tends to conform to its species' usage of the codon catalog; this is our genome hypothesis.

INTRODUCTION

The genetic code provides options among codons for all 20 amino acids of protein except methionine and tryptophan, which have single codons (see Figure 1 of reference 1). Choices among synonymous codons do not affect the nature of the protein produced but they may relate to expression of a gene. Indeed, mRNA expressivity (rate of synthesis of a protein and the total amount of it made) may be under the control of degenerate base use in the mRNA. This is still largely an untested hypothesis. It is, however, an important one since choices among third bases have often been considered "neutral", that is, of no influence on fitness of the phenotype. Messenger RNA expressivity may well

turn out to be an adaptive phenomenon.

In order to better understand codon usage, we have studied all published mRNA sequences of more than about 50 codons. We here report three analyses on these sequences. Table 1 first shows number, name, symbol, length in codons and reference for each mRNA. Then Figures 1, 2 and 3 reveal the analyses. In Figure 1 mRNA are combined according to genome type (RNA or DNA virus, animal, etc.). Frequencies per thousand for each of the 61 amino acid codons appear in Figures 1 and 2 (initiator and terminator codons are excluded). Absolute frequencies are had by multiplying by number of codons (Table 1). Figure 2 presents frequencies in each individual gene (unless it is the only example of a genome type, in which case it is in Figure 1). This allows comparison of codon use not only among genes in the same or different genomes, but also between individual genes and the genome types of Figure 1. Finally, messengers with similar patterns of use of the code's degeneracy are seen as neighboring points in Figure 3. The spacings in Figure 3 are projections into two dimensions of distances calculated by correspondence analysis (2, 3) on codon third base frequencies. Other work has shown that the codon frequencies themselves give correspondence analysis distances that depend mostly on systematic choices between degenerate bases (4, 5).

DISCUSSION

It is not evident that distances between mRNA based on codon frequencies would depend mainly on degenerate base use. The other two codon positions could dominate in establishing these distances if proteins were relatively more different among themselves than are their mRNA. Elsewhere (4, 5) we have found that distances between proteins, determined by correspondence analysis on their amino acid frequencies, do not agree with distances between their mRNA, calculated from frequencies for the 61 codons in each messenger. However, frequencies of duet coded amino acids can particularly affect distances between mRNA (5). In order to eliminate all possible influence of protein composition, therefore, distances in Figure 3 were determined solely from quartet codons (1). Each of the eight quartet sets of codons has a complete choice of bases in codon position III. Figure 3 results from correspondence analysis on these quartet third base frequencies in each mRNA.

The structure in Figure 3 can be considered at several levels. First, animals lie to the right and viruses to the left. Next, there is grouping by genome or genome type: All papova virus (SV40 and BKV) genes lie together

(no. 51-56); immunoglobins (IG) fall in a nearby zone (no. 73-78). All mRNA of mammals except immunoglobins (MAM-IG) are grouped to the upper right (no. 79-90). Curiously, however, frameshift genes E of ϕ x174 and G4 (no. 13 and 23) are near this mammal area. Other genes of these phage are mainly to the upper left of the figure. Finally, most IG have different coding strategies from other mammalian genes (compare, for example, MUSBLP and MUSK2, no. 73 and 80).

What do these groupings and distances mean? We can partly answer this question although the exact biological significance is unknown. The horizontal axis of Figure 3 roughly corresponds to GC content of the third bases (5). The rightmost mRNA often use C and G as third bases. Thus, with MUSBLP 90.5% of all quartet codons have C or G in position III, while the value is only 2.8% in the solitary yeast mitochondrial mRNA (no. 64) and 37.0% in MUSK2. This also explains positioning of the above E genes near mammals: their mRNA show more GC in quartet position III (68.9 and 64.7%, respectively) than do other phage mRNA. Likewise, vertical contrast exists between use of A and U. For example, quartet third bases in MUSK2 contain 42.6% A and 20.4% U while those in FDV5 (no. 33) have 4.1% A and 53.1% U. Consequently, in base choices for quartet codons papova virus mRNA resemble IG mRNA but differ greatly from other mammalian genes, for both GC content and use of A (1). Lastly, correspondence analysis groupings as in Figure 3 are very stable, whether the starting data are frequencies for all 61 codons, the 32 quartet codons, or for third bases of all codons or, as here, third bases of quartet codons. Some rationalization of this stable structure is found in the distribution of the isoacceptor tRNA for the codons (5-7). Coordination of codon usage with tRNA gene amplification and expression may eventually aid in understanding the genome hypothesis.

Our general goal in this work is to identify and understand the biological information in nucleic acid sequences. Our hypothesis, however, is so far mostly only descriptive. We observe that mRNA of the same genome are clustered by correspondence analysis on codon frequencies and that their proteins are not similarly grouped by analysis on the amino acid frequencies (4). Indeed, correspondence analysis on the proteins does not suggest classical systematics as does that on mRNA. In this very limited sense, messengers better reflect evolution than proteins do (4, 5). We do not yet know why.

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Table 1 mRNA Portfolio

No.	Species and gene	Symbol	No.codons	Ref.
1	Phage MS2 gene A	MS2A	392	8
2	Phage MS2 coat	MS2C	129	9
3	Phage MS2 replicase	MS2R	544	10
4	Phage Q beta coat	QBVC	79	11
5	Tobacco mosaic virus gene A	TMVA	55	12
6	Tobacco mosaic virus coat	TMVC	158	13
7	Tobacco mosaic virus 30K protein	TMV30K	105	13
8	Turnip yellow mosaic virus coat	TYMC	188	14
9	Phage ϕ X 174 gene A	FIXA	511	15,16
10	Phage ϕ X 174 gene B	FIXB	119	15,16
11	Phage ϕ X 174 gene C	FIXC	85	15,16
12	Phage ϕ X 174 gene D	FIXD	151	15,16
13	Phage ϕ X 174 gene E	FIXE	90	15,16
14	Phage ϕ X 174 gene F	FIXF	422	15,16
15	Phage ϕ X 174 gene G	FIXG	174	15,16
16	Phage ϕ X 174 gene H	FIXH	326	15,16
17	Phage ϕ X 174 gene J	FIXJ	37	15,16
18	Phage ϕ X 174 gene K	FIXK	55	17
19	Phage G4 gene A	VG4A	553	18
20	Phage G4 gene B	VG4B	119	18
21	Phage G4 gene C	VG4C	83	18
22	Phage G4 gene D	VG4D	151	18
23	Phage G4 gene E	VG4E	95	18
24	Phage G4 gene F	VG4F	426	18
25	Phage G4 gene G	VG4G	176	18
26	Phage G4 gene H	VG4H	336	18
27	Phage G4 gene J	VG4J	24	18
28	Phage G4 gene K	VG4K	55	17
29	Phage fd gene 1	FDV1	347	19
30	Phage fd gene 2	FDV2	409	19
31	Phage fd gene 3	FDV3	423	19
32	Phage fd gene 4	FDV4	425	19
33	Phage fd gene 5	FDV5	86	19
34	Phage fd gene 6	FDV6	111	19
35	Phage fd gene 7	FDV7	32	19
36	Phage fd gene 8	FDV8	72	19
37	Phage fd gene 10	FDV10	110	19
38	Phage M13 gene 1	M131	215	20
39	Phage M13 gene 3	M133	423	20
40	Phage M13 gene 4	M134	59	20
41	Phage M13 gene 6	M136	111	20
42	Phage M13 gene 7	M137	32	21
43	Phage M13 gene 9	M139	31	21
44	Phage T7 gene 1	T7VG1	56	22
45	Phage λ gene CI	LAMCI	236	23
46	Phage λ gene CII	LAMCII	96	24
47	Phage λ gene cro	LAMCRO	65	24
48	Phage λ gene O	LAMO	298	25
49	Phage 434 gene CII	434CII	96	26
50	Phage 434 gene cro	434CRO	70	26
51	Simian virus 40 gene T	S4OGT	626	27,28

No.	Species and gene	Symbol	No. codons	Ref.
52	Simian virus 40 gene t	S4OPT	173	27,28
53	Simian virus 40 gene VP1	S4OVP1	361	27,28
54	Simian virus 40 gene VP2	S4OVP2	351	27,28
55	Simian virus 40 gene VP3	S4OVP3	233	27,28
56	Virus BK gene t	BKVPT	171	29
57	Hepatitis B virus surface antigen	HBVSA	225	30,31
58	Escherichia coli lac l	ECOLAC	359	32
59	Escherichia coli ribosomal protein L11	ECOL11	141	33
60	Escherichia coli ribosomal protein L1	ECOL1	233	33
61	Escherichia coli ribosomal protein L10	ECOL10	164	33
62	Escherichia coli ribosomal protein L7/L12	ECO712	120	33
63	Salmonella paratyphi ampr gene	SAPAMP	285	34
64	Saccharomyces subunit 9 mitochondrial ATPase	SACMT9	75	35
65	Saccharomyces iso-1 cytochrome C	SACCC1	108	36
66	Psammechinus miliaris H1 histone	PSMH1	85	37
67	Psammechinus miliaris H2A histone	PSMH2A	123	37
68	Psammechinus miliaris H2B histone	PSMH2B	102	37
69	Psammechinus miliaris H3 histone	PSMH3	135	37
70	Strongylocentrotus purpuratus H2A histone	SPUH2A	123	38
71	Strongylocentrotus purpuratus H3 histone	SPUH3	102	38
72	Chicken ovalbumin	GALOVA	385	39
73	Mouse K2 immunoglobulin	MUSK2	117	40
74	Mouse immunoglobulin light chain MOPC 21	MUSLC	107	41
75	Mouse immunoglobulin λ 1-type light chain	MUSL1	173	42
76	Mouse V λ I1 immunoglobulin	MUSVL2	165	43
77	Mouse MOPC-41 light chain immunoglobulin	MUSVL41	129	44
78	Mouse immunoglobulin γ -1 constant heavy chain	MUSPH21	153	45
79	Mouse hemoglobin beta	MUSBGL	146	46
80	Mouse beta lipotropin	MUSBLP	48	47
81	Rat growth hormone	RATGH	215	48
82	Rat prolactin hormone	RATPLH	132	49
83	Rat preproinsulin	RATPPI	107	50
84	Rabbit hemoglobin alpha	RABAGL	141	51
85	Rabbit hemoglobin beta	RABBGL	146	52
86	Bovine corticotropin beta lipotropin	BOVCBL	264	53
87	Human alpha chorionic gonadotropin	HUMACG	115	54
88	Human hemoglobin beta	HUMBGL	132	55
89	Human pregrowth hormone	HUMPGH	216	56
90	Human chorionic somatomammotropin	HUMCSL	168	57

Nucleic Acids Research

Codon	SS R N A			VIRUS	SS DNA	DS D N A VIRUS			
	ALL MO 1-8	PHAGE 1-4	PLANT 5-8	VIRUS 9-43	ALL 44-57	PHAGE 48-50	PAPPOVA 51-56	HBUSA 57	
Ara CGA	8	6	11	5	8	16	0	0	
CGC	10	15	6	15	14	27	0	4	
CGG	5	6	5	3	1	0	1	4	
CGU	20	25	14	21	9	15	2	9	
AGA	12	4	20	5	15	6	28	4	
AGG	6	3	9	2	15	9	24	0	
Leu CUA	11	11	10	4	10	5	12	31	
CUC	17	20	15	15	9	12	3	31	
CUG	8	11	6	18	17	19	13	31	
CUU	13	14	12	30	23	25	22	18	
UUA	16	17	16	24	20	13	28	18	
UUG	12	6	18	17	16	12	21	18	
Ser UCA	19	15	24	18	11	10	9	27	
UCC	12	13	10	13	7	4	7	22	
UCG	19	21	17	8	4	5	0	13	
UCU	20	22	19	36	16	14	18	18	
AGC	13	14	13	4	10	13	7	4	
AGU	11	4	17	8	14	8	20	22	
Thr ACA	17	7	27	11	16	17	14	27	
ACC	26	23	30	12	20	22	18	22	
ACG	9	9	9	8	4	6	0	13	
ACU	32	32	31	25	18	13	25	13	
Pro CCA	10	8	11	7	13	10	13	40	
CCC	16	10	22	5	8	3	10	31	
CCG	11	14	8	10	8	14	0	9	
CCU	14	18	10	17	17	5	29	22	
Ala GCA	23	29	18	10	22	31	14	4	
GCC	19	18	19	12	11	15	8	4	
GCG	17	20	14	15	11	22	0	4	
GCU	22	24	21	35	40	42	41	13	
Gly GGA	17	12	21	8	18	13	23	27	
GGC	11	16	5	22	11	12	11	9	
GGG	10	16	5	5	11	10	12	18	
GGU	22	34	10	37	14	16	12	9	
Val GUA	16	19	12	13	11	7	15	9	
GUC	27	23	31	14	4	6	2	4	
GUG	16	14	17	8	12	9	15	18	
GUU	30	36	24	37	24	26	23	18	
Lys AAA	28	27	29	44	45	48	47	13	
AAG	28	28	28	19	30	39	24	0	
Asn AAC	34	40	29	18	24	35	13	9	
AAU	28	24	32	27	18	11	26	18	
Gln CAA	17	17	16	26	26	24	29	13	
CAG	21	29	13	21	21	22	20	18	
His CAC	3	5	1	4	4	0	7	4	
CAU	3	3	3	9	9	8	12	0	
Glu GAA	18	11	25	23	36	38	39	0	
GAG	24	26	22	18	32	41	26	9	
Asp GAC	25	15	34	23	24	26	24	4	
GAU	25	19	30	25	28	22	38	9	
Tyr UAC	21	26	16	10	12	10	15	13	
UAU	6	5	8	26	15	11	21	13	
Cys UGC	9	8	10	5	11	3	18	22	
UGU	3	4	2	8	9	3	10	40	
Phe UUC	18	20	16	24	16	22	5	40	
UUU	18	11	25	26	25	16	35	31	
Ile AUA	15	10	19	8	9	5	12	18	
AUC	25	25	24	15	17	29	0	31	
AUU	11	12	11	33	26	26	26	22	
Met AUG	13	12	14	16	29	32	27	22	
Trp UGG	12	16	7	13	23	17	24	58	

Fig. 1(i)

Codon	RS DNA								
	BACT 58-63	MITOCH 64	YEAST 65	ALL ANI 65-70	ANI-MAM 65-72	ALL MAM 73-90	IG 73-78	MAM-IG 79-90	
Arg CGA	3	0	0	4	8	2	7	0	
	CGC	20	0	0	14	26	9	4	12
	CGG	2	0	0	6	5	6	3	8
	CGU	20	0	0	10	21	7	1	9
	AGA	2	13	28	8	14	6	12	3
AGG	0	0	0	11	12	11	9	11	
Leu CUA	3	13	9	9	9	8	11	7	
	CUC	3	0	0	25	27	24	26	24
	CUG	59	0	0	42	21	51	16	68
	CUU	7	0	9	10	16	7	7	7
	UUA	6	147	9	3	1	4	11	1
UUG	6	0	46	8	10	7	10	6	
Ser UCA	3	67	9	10	9	11	24	5	
	UCC	14	0	0	17	11	20	17	22
	UCG	4	0	9	3	2	4	1	5
	UCU	18	0	19	17	17	17	29	11
	AGC	9	0	0	20	19	21	27	18
AGU	6	0	0	15	11	17	33	9	
Thr ACA	4	27	19	13	13	13	26	7	
	ACC	21	0	28	26	28	25	30	23
	ACG	6	0	0	8	10	7	4	9
	ACU	24	0	28	18	13	19	36	11
Pro CCA	6	13	28	12	13	12	20	7	
	CCC	5	0	0	17	12	18	13	21
	CCG	26	0	0	6	2	8	5	9
	CCU	1	13	9	12	11	13	13	12
Ala GCA	40	53	0	16	29	11	20	6	
	GCC	18	13	37	38	49	34	22	40
	GCG	28	0	0	6	6	5	1	8
	GCU	70	67	28	25	31	23	21	24
Gly GGA	4	27	0	14	23	18	20	5	
	GGC	31	0	19	26	21	28	16	34
	GGG	5	0	19	11	13	11	9	11
	GGU	31	107	74	19	19	18	25	15
Val GUA	34	67	0	4	8	3	5	1	
	GUC	9	0	0	19	25	16	23	13
	GUG	14	0	19	31	22	35	17	44
	GUU	42	13	9	9	12	7	7	7
Lys AAA	65	27	56	19	29	15	17	14	
	AAG	11	0	93	54	74	45	24	56
Asn AAC	21	13	46	27	21	29	29	29	
	AAU	7	13	19	9	6	10	13	9
Gln CAA	14	13	19	12	17	10	11	9	
	CAG	23	0	0	31	29	32	30	32
His CAC	6	0	19	19	9	22	8	29	
	CAU	4	0	19	10	9	10	11	10
Glu GAA	57	13	46	22	23	22	21	23	
	GAG	14	0	19	35	33	36	23	43
Asp GAC	30	13	28	23	11	27	22	30	
	GAU	19	0	9	16	11	18	23	16
Tyr UAC	10	0	28	21	23	20	20	20	
	UAU	4	13	19	12	3	15	17	14
Cys UGC	4	0	9	9	1	12	6	16	
	UGU	2	13	19	9	2	11	18	8
Phe UUC	12	80	19	29	18	33	30	35	
	UUU	12	13	19	14	9	16	13	18
Ile AUA	3	0	0	4	3	4	7	3	
	AUC	28	27	19	25	40	20	21	19
AUU	18	93	19	11	10	11	18	8	
Met AUG	29	27	19	16	16	16	14	16	
	UGG	3	0	9	12	1	16	25	12

Fig 1 (ii)

	LOW	LOW	LOW	474	474	S49	S49	S49	S49	REV	ECO	ECO	ECO	ECO	ECO	PSM	PSM	PSM	PSM			
	CT	CT	CT	CT	CT	HT	HT	HT	HT	PT	L19	L19	L19	L19	AMP	AMP	AMP	AMP	AMP			
Arg	31	15	7	31	29	0	0	0	0	0	11	0	0	0	7	0	0	0	0	22		
Asp	4	21	17	17	47	2	0	0	0	0	28	14	21	30	0	25	12	12	33	10	44	
Cys	0	0	0	0	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Glu	4	21	0	20	21	0	0	0	0	0	6	14	26	43	8	25	12	33	10	37	10	
Ala	21	0	0	26	29	19	23	30	41	7	0	0	0	0	0	0	0	0	0	0	22	
Met	4	21	0	14	10	12	14	34	52	18	0	0	0	0	0	0	0	0	0	0	12	
Leu	10	0	0	10	10	17	7	11	0	0	0	0	0	0	0	0	0	0	0	0	15	
Val	4	10	15	10	10	13	20	11	0	0	0	0	0	0	0	0	0	0	0	0	15	
Phe	34	43	0	13	43	20	13	35	11	14	17	41	6	0	0	32	0	24	16	0	37	
Trp	25	0	0	15	0	29	40	19	27	21	35	14	0	0	0	0	0	0	0	0	0	
Ile	4	21	0	14	12	25	20	12	14	0	0	0	0	0	0	0	0	0	0	0	0	
Ser	10	0	0	10	14	14	0	6	11	13	12	11	0	0	0	0	0	0	0	0	0	
Pro	17	0	0	13	0	6	6	6	6	6	17	35	9	6	8	11	12	8	20	7	0	
Thr	0	10	0	10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
His	13	10	0	13	14	14	17	14	31	26	6	19	14	13	18	33	17	36	8	49	22	
Asn	11	10	0	11	10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Gly	17	10	0	17	10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Tyr	0	21	0	17	21	43	19	12	20	26	0	31	28	30	24	0	11	12	8	0	0	
Arg	4	0	0	10	15	7	10	29	21	17	33	31	26	23	0	21	35	8	0	0	0	
Ala	34	10	0	17	10	14	6	10	11	9	6	7	4	0	0	0	0	0	0	0	0	
Pro	0	10	0	10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Cys	4	10	15	10	10	0	0	0	0	0	17	57	26	30	17	11	12	0	0	0	0	
Glu	25	0	0	10	10	19	29	33	26	34	33	10	0	0	0	0	0	0	0	0	0	
Val	35	0	0	10	10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Met	8	53	31	10	31	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Leu	17	53	31	17	52	43	40	17	36	91	39	23	0	0	0	0	0	0	0	0	0	
Val	4	10	15	13	21	29	14	40	33	20	13	18	3	7	0	0	0	0	0	0	0	
Pro	21	10	15	23	0	0	10	6	14	13	18	31	14	47	30	50	14	24	33	20	0	
Cys	31	10	15	30	0	0	13	6	28	14	9	6	11	0	0	0	0	0	0	0	0	
Glu	25	0	0	10	10	29	10	12	14	20	17	0	17	64	39	24	17	28	0	41	29	
Val	4	0	0	10	10	14	13	12	11	20	26	12	0	43	39	37	58	18	24	0	0	
Pro	4	10	0	7	21	0	0	0	0	0	6	25	7	15	12	0	7	15	33	29	0	
Cys	8	10	10	30	10	19	15	28	23	10	0	0	0	0	0	0	0	0	0	0	0	
Glu	47	0	0	10	10	19	15	28	23	10	0	0	0	0	0	0	0	0	0	0	0	
Val	43	53	35	27	45	27	50	45	53	36	14	0	32	28	56	43	62	21	12	8	0	
Met	31	43	53	27	45	43	32	40	17	0	0	0	0	0	0	0	0	0	0	0	0	
Leu	31	45	40	31	14	19	15	19	9	13	6	14	28	53	6	0	18	118	57	50	0	
Val	25	10	0	30	10	0	30	23	30	23	18	19	0	0	0	0	0	0	0	0	0	
Pro	8	21	46	17	21	57	21	17	25	48	52	12	36	7	4	12	8	18	24	34	0	
Cys	30	21	0	27	21	29	22	6	28	17	23	42	43	34	6	0	14	35	8	29	49	
Val	0	0	0	0	0	0	0	0	0	0	9	12	11	0	0	0	0	0	0	0	0	
Pro	4	15	0	0	0	19	17	16	9	12	11	0	0	0	0	0	0	0	0	0	0	
Cys	58	52	31	20	45	41	27	29	56	17	17	35	24	37	33	33	39	54	24	10	10	
Glu	15	45	10	40	31	0	19	13	30	20	21	38	19	35	42	18	42	18	24	16	49	44
Ala	21	10	46	40	21	14	46	64	25	23	30	41	28	7	13	18	0	39	24	10	15	
Trp	13	0	0	14	14	14	20	14	9	13	12	0	0	0	0	0	0	0	0	0	0	
Val	0	45	13	0	0	10	46	11	0	0	0	0	0	0	0	0	0	0	0	0	0	
Cys	8	0	13	0	0	10	46	11	0	0	0	0	0	0	0	0	0	0	0	0	0	
Glu	4	0	0	14	13	17	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Phe	17	21	15	10	21	14	2	12	3	0	0	0	0	0	0	0	0	0	0	0	0	
Trp	34	0	31	12	0	14	53	29	34	29	3	0	28	4	18	17	0	18	17	0	30	
Val	4	10	15	23	42	0	16	16	0	0	0	0	0	0	0	0	0	0	0	0	0	
Met	17	21	31	20	21	57	30	12	33	34	34	0	19	43	30	12	33	25	59	24	59	44
Leu	34	43	15	27	42	29	27	52	22	9	41	25	26	38	25	32	12	0	0	0	0	
Trp	13	31	0	30	31	14	16	35	0	20	26	41	6	0	0	0	0	0	0	0	0	

Fig. 2 (iii)

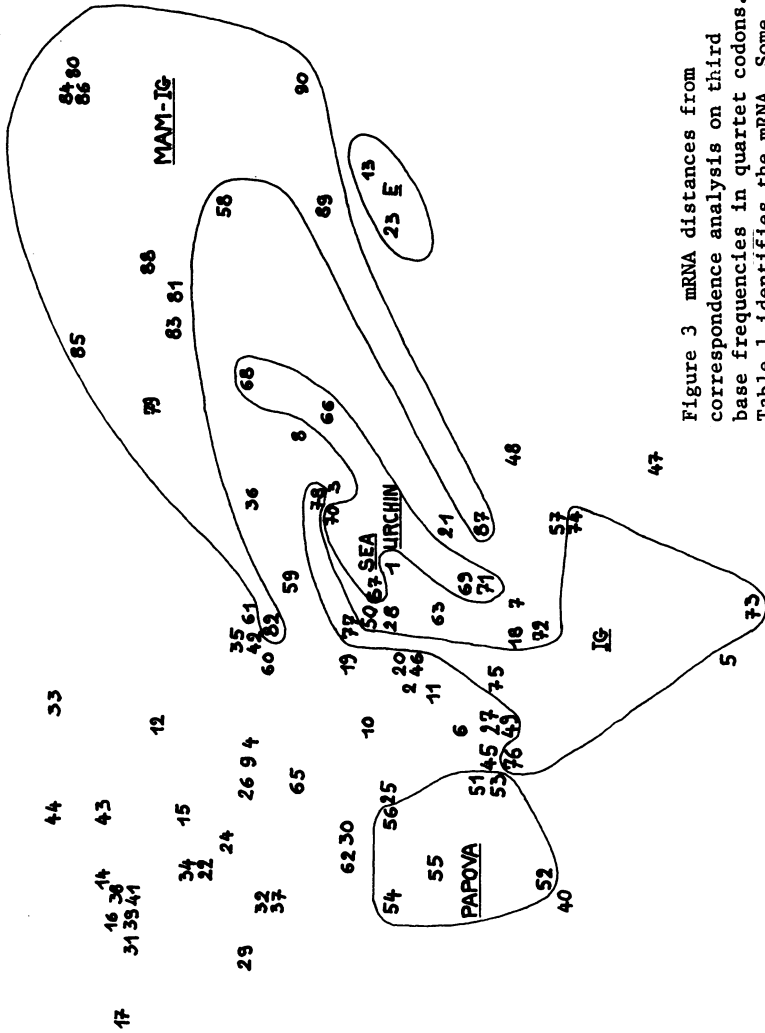


Figure 3 mRNA distances from correspondence analysis on third base frequencies in quartet codons. Table 1 identifies the mRNA. Some interesting genome types have been circled (here "by eye"; elsewhere groupings are made by automatic classification (4, 5)).

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